#### **REVIEW ARTICLE**



# Allogenic haematopoietic stem cell transplantation in VEXAS: A review of 33 patients

Syed B. Ali<sup>1,2,3,4</sup> · Carmelo Gurnari<sup>5,6</sup>

Received: 27 July 2024 / Revised: 21 September 2024 / Accepted: 23 September 2024 / Published online: 30 September 2024 © Crown 2024

# Abstract

Vacuolation, E1 enzyme, X-linked, autoinflammatory, somatic (VEXAS) syndrome is a multisystem disease due to a genetic mutation in the ubiquitin-activating enzyme (*UBA1*). Allogeneic haematopoietic stem cell transplantation (allo-HSCT) offers both therapeutic and cure but also carries significant risks. A review of VEXAS and HSCT cases was undertaken. Thirty-three patients were identified; majority males (n=32, 97.0%), median time from symptoms to HSCT: 3 years (IQR 2.0–4.8) and median age of 59 years (IQR 52.5–65.5). *UBA1* mutation Met41Thr was most common (11/32, 34.4%). The median variant allele frequency was 56.5% (IQR 43.0–73.5) with no correlation with increasing age. Prior to HSCT, 4.5 (IQR 2.8–6) treatments were trialled. Peripheral blood HSCT (30/31, 96.8%) and HLA-matched, unrelated donor (18/32, 56.3%) were most common. Conditioning regimens varied, with reduced intensity treatment with fludarabine as a co-agent most frequently administered (12/31, 38.7%). Both acute and/or chronic GVHD (18/32, 56.3%) and infections were common (12/32, 37.5%). Overall, 27 individuals (81.8%) were alive, and those undergoing HSCT prospectively had median follow up of 9 months (IQR 3.8–14.4). Of the six deceased, infection was implicated in four. In 11 cases with post-HSCT molecular data, a complete eradication of *UBA1* mutation was reported. In summary, while consensus treatment strategy regarding VEXAS is lacking, this review highlights HSCT may remain not only a therapeutic option but also enable cure. However, considerations regarding comorbidities, concurrent haematological disorders as well as overall risks of GVHD and infections need to be made.

#### Key points

• Very few reported prospective cases of VEXAS and allogeneic haematopoietic stem cell transplantation (allo-HSCT) have been reported.

• While risks of graft versus host disease and infection remain barriers, this treatment modality remains an option for selected patients.

• Allo-HSCT is the only treatment strategy which can remove the UBA1 mutation.

Keywords VEXAS Syndrome  $\cdot$  Somatic mutation  $\cdot$  Transplantation  $\cdot$  Allogenic stem cell transplantation  $\cdot$  Myelodysplatic syndrome

# Introduction

Vacuolation, E1 enzyme, X-linked, autoinflammatory, somatic (VEXAS) syndrome is a multisystem disease arising

Syed B. Ali syed.ali@sa.gov.au

- <sup>1</sup> Department of Clinical Immunology and Allergy, Flinders Medical Centre, Bedford Park, South Australia, Australia
- <sup>2</sup> School of Medicine and Public Health, Flinders University, Bedford Park, South Australia, Australia
- <sup>3</sup> Department of Clinical Immunology and Allergy, Royal Adelaide Hospital, Adelaide, South Australia, Australia

due to a genetic mutation in the ubiquitin-activating enzyme (UBAI) on the X-chromosome [1]. Several pathways including protein homeostasis and cell signalling are affected with dysregulation of ubiquitination processes and activation of inflammation [2, 3].

- <sup>4</sup> School of Medicine and Biomedical Sciences, University of Adelaide, Adelaide, South Australia, Australia
- <sup>5</sup> Department of Translational Hematology and Oncology Research, Taussig Cancer Institute, Cleveland Clinic, Cleveland, OH, USA
- <sup>6</sup> Department of Biomedicine and Prevention, University of Rome Tor Vergata, Rome, Italy

Clinical Rheumatology (2024) 43:3565-3575

Somatic mutations in the *UBA1* gene have a reported prevalence of 0.007% [4]. While canonical phenotypes of male gender and age above 60 years are well described, there are increasing reports of the disease affecting females or a younger age of presentation [5]. As concurrent haematological disease such as myelodysplastic syndrome (MDS) may occur with VEXAS, the presence of a somatic *UBA1* mutation is now being detected in patients with longstanding diagnoses of MDS pre-existing the discovery of VEXAS [6].

Treatment paradigms for VEXAS focus on targeting proinflammatory pathways, but given the relatively recent discovery of the disease and its heterogeneous manifestations, clear guidelines are lacking. Glucocorticoids form the foundation of treatment and additional treatments such as conventional disease-modifying antirheumatic drugs (csDMARDs) are frequently used, although most have demonstrated poor efficacy [7]. Biologic therapies targeting the IL1 and IL6 cytokines have also been used but with variable efficacy [7]. Recently, oral Janus kinase inhibitors have gained popularity due to simultaneous targeting of multiple cytokine targets; however, a systematic review reported complete remission in only 33% and partial remission in 27.3% from this treatment [7]. Concurrent haematological diseases, such as MDS is often addressed with azacitidine, a hypomethylating targeted treatment which is has a direct cytotoxic effect [8].

More importantly, given low response rates with treatment strategies which do not eliminate the *UBA1* mutation, patients may have transfusion dependency for symptomatic anaemia. Due to the somatic nature of the mutation, variant allele frequency may also increase over time, possibly contributing to progressive disease. Furthermore, as the 5-year survival rate is as low as 63% [9], treatment strategies need to address improving both mortality and morbidity. This is particularly the case as patient population is older, with more comorbidities, making immunosuppressant therapies more problematic in terms of adverse effects as well as interactions.

Allogeneic haematopoietic stem cell transplantation (allo-HSCT) is a possible therapeutic option with the attractive notion of replacing progenitor cells affected by the *UBA1* somatic mutation, thereby enabling disease remission [10]. However, as allo-HSCT carries both high rates of morbidity and mortality, upfront treatment might not always be desirable. There is a risk with delaying allo-HSCT; however, in that with accumulating inflammatory burden and immunosuppression, patients may become too frail to be considered for allo-HSCT.

Given the everchanging landscape of VEXAS and notable difficulties in achieving controlled remission with treatments thus far, a narrative review of allo-HSCT in this disease entity was performed. The purpose of this review is to provide clinicians with current data, raising awareness about the risks and benefits of allo-HSCT. This is of particular importance as clear guidelines on transplantation in the setting of VEXAS are lacking. A review of the literature was completed with search terms "VEXAS" and "transplantation" using PubMed and Embase online databases in April 2024. All publications were reviewed for the search period January 2020 to April 2024. Only those with peer-reviewed publications and full text availability were included. Duplicate publications, including cases already reported, as well as abstracts from meetings were excluded.

Detailed data collection including demographics, *UBA1* somatic mutation, variant allele frequency, clinical presentation, delay of diagnosis from initial symptoms, treatment modalities leading up to transplantation, as well as transplantation regimen and outcomes were recorded. Corresponding authors were directly contacted to obtain missing data and incorporated only if received, otherwise left blank.

Case series and reports were further sub-grouped for purposes of analysis as follows: transplantation following diagnosis of VEXAS, i.e., prospectively (group 1), or patients who had previously undergone transplantation for other haematological diseases such as MDS in whom the diagnosis of VEXAS was made retrospectively (group 2).

# Results

A total of 12 publications were identified globally: France (n = 3), USA (n = 2), Canada (n = 1), Czech Republic (n = 1), Italy (n = 1), Netherlands (n = 1), Spain (n = 1), Sweden (n = 1), and UK (n = 1). These publications comprised a total of 33 individuals: 23 patients in group 1, and 10 patients in group 2.

Clinical details are summarised in Table 1. The median age at transplantation was 59.0 years (IQR 52.5–65.5). There were no significant differences between the ages of patients in the two groups (58.6 vs. 59.5 years, p = 0.79). Most individuals were males (n = 32, 97.0%).

The median time between symptoms and transplantation was 3.0 years (IQR 2.0–4.8) with no significant difference between the two groups (both at 3.0 years, p = 0.87).

Somatic *UBA1* mutation data was available for 32 patients. Of these, Met41Thr was most common (n = 11, 34.4%), followed by Met41Val (n = 9, 28.2%) and Met41Leu (n = 6, 18.8%) respectively. Eighteen cases had variant allele frequency reported; median of 56.5% (IQR 43.0–73.5), and no significant relationship with increasing age was observed (p = 0.88).

For group 1, VEXAS was the sole diagnosis in nine (39.1%), while the remainder had additional haematological disorders: MDS (n = 13, 56.5%) and multiple myeloma (n = 1, 4.3%). Data regarding treatment for transplantation

was available for 22/23 individuals, and in these patients, several lines of treatment were implemented (median 4.5, IQR 2.8–6). Of these corticosteroids and csDMARDs were common, both used in 14 cases (63.6%). For this prospective transplantation, 14/23 had data available. Of these, VEXAS progression with inflammatory symptoms with or without transfusion dependency (n=9, 64.3%) were most common (Table 1).

Detailed clinical information regarding transplantation of all individuals is also summarised in Table 1. For those with data available, stem cell source was chiefly peripheral blood (30/31, 96.8%), and HLA-matched, unrelated donor (18/32, 56.3%) were most common donor types. Conditioning regimens varied, with reduced intensity treatment with fludarabine as a co-agent most frequently administered (n = 12/31, 38.7%). Similarly, there was also variation in graft versus host disease (GVHD) prophylaxis, of which ciclosporin was co-administered with other immunosuppressives in just over half of the cohort (n = 19/32, 59.4%).

Of the 32 patients with data available on post-transplant complications, more than half had acute and/or chronic GVHD (n = 18/32, 56.3%) (Table 1). Complications of infection were reported in 12/32 individuals (37.5%).

Overall, 27 individuals (81.8%) were alive at the time of publication. In group 1, 20/23 (86.9%) were alive with median follow up of 9.0 months (IQR 3.8–14.4). Of the three deceased patients (13.0%), information was available for two with infection reported as the cause. Two of the three had the Met41Val mutation, with the other case with Met41Thr mutation. In group 2, three of ten (30%) were deceased, in which infection were attributed to two, while the other was from solid organ neoplasia (Table 1).

Notably, in 11 cases with post-HSCT molecular data, a complete eradication of *UBA1* mutation was reported following the transplant procedure, thereby establishing its potentiality for complete cure. Furthermore, in some cases, VEXAS phenotype was reported to be controlled even with persistent mixed chimerism [11].

### Discussion

This narrative review provides up-to-date clinical information about the small number of published cases regarding transplantation in VEXAS. The reported cases were either from centres in North American or Europe, which may reflect firstly the limited experience of transplantation in autoinflammatory diseases, and secondly testing availability for the *UBA1* mutation in some regions.

A clear consensus treatment strategy is still lacking for VEXAS, perhaps due to the heterogeneous nature of the disease, with manifestations which can require the involvement of multiple different medical specialties. Glucocorticoids are the major treatment modality; however, often, high doses over a prolonged period are needed, thereby exposing patients to adverse effects, including dependency on unacceptably high doses (i.e.,  $\geq$  30-mg prednisolone 30 mg) in up to 11% of cases [7]. In this review, the average time between symptoms and transplantation was 3.6 years, indicating that patients may often experience waxing and waning symptoms that might contribute to the morbidity associated with this disease. In addition, several other factors to consider in this context include delays in diagnosis, particularly with availability of UBA1 genetic testing and recognition of vacuoles within bone marrow raising possibility of diagnosis, as well as access to transplantation services, lack of local experience, and familiarity and persistence with traditional paradigms of immunosuppression even if ineffective. Remarkably, most VEXAS diagnoses in the recent era result from UBA1 testing of patients already followed up for rheumatologic and haematologic disorders even years prior to the actual suspicion, given the very recent discovery of the disease. Data on lag of VEXAS diagnosis in the current era where UBA1 testing is available and clinical awareness is more accurate are needed to establish whether the pleomorphic nature of the disease rather than its novelty constitute reason for the observed delay in establishing a proper diagnosis.

There is a pressing need for novel approaches to VEXAS treatment, distinct from traditional paradigms of treating inflammatory diseases or managing haematological malignancies. Recognising that VEXAS patients are of an older demographic, frequently with multiple comorbidities, the timing of definitive treatments such as allo-HSCT is of utmost importance. Management strategies need to include regular and objective assessments of disease activity and treatment success or failure, to ensure that delays to appropriate escalation are minimised. With growing experience with allo-HSCT, there may be less of a need to try prolonged glucocorticoids and/or DMARDs prior to considering transplant. Furthermore, it appears that the specific UBA1 mutation may have prognostic value, with a large cohort of 116 patients showing that Met41Val mutations had poor outcomes with a 5-year survival of 76.7%, in comparison to Met41Thr and Met41Leu at 83% and 100% respectively [12]. Consequently, there is the potential for therapeutic paradigms which are customised to different levels of genetic risk.

Although allo-HSCT remains the only curative treatment option and should be considered early, its non-negligible morbidity and mortality risk must be carefully considered. In this context, the decision is recommended to be undertaken by multidisciplinary and experienced team, which comprehensively assess patients, especially in this disease where age and comorbidities are common [13]. Interestingly,

Reference	Age, sex	Clinical presen-	Time between	Mutation, variant allele fre-	Concurrent	Lines, and types	Transplantation	_		GvHD monthylowic	Post-transplantation	tation
		phenotype	symptoms and transplant (years)	ducity (%)		or u cauncin pre-transplant	Indication	Type, donor	Conditioning	pupuytaxis	Complica- tions	Follow-up (months) alive /dead
VEXAS and pro	spective tran	VEXAS and prospective transplantation (group 1)	1)									
Stiburkova et al. July 2023, Czech Repub- lic[31]	61 M	Fever, rash, arthritis, lym- phadenopathy	7	c.1420G > C;pGly477Ala, N/A	SQM	5; CS, MTX, Infx Tofa, Tcz	MDS and VEXAS progression: transfusion support, inflamma- tory	PB, MUD	S-flu, treo	ATG, Cyc, MMF	liN	8, alive
Gurmari et al. Decem- ber 2023, Italy[14]	W 69	Fever, rash, arthritis, lung, VTE	3.8	c.122 T > C; pMet41Thr, 50%	MDS	13; csDMARD, bDMARD, tsDMARD, Azac	Ν/Α	PB, MUD	R-flu, bu	ATG, Cyc, MTX	Nil	18.6, alive
	53 M	Arthritis, VTE	0.75	c.118-2A > C, N/A	SQM	6; csDMARD, bDMARD, Azac	N/A	PB, MUD	S-flu, bu	ATG, Cyc, MTX	Grade I aGVHD	1.1, alive
	52 M	Fever, arthritis, chondritis, rash, VTE, lung	4	c.122 T > C; pMet41Thr, 43%	Nil	5; csDMARD, bDMARD	NA	PB, MMUD	S-flu, mel, TT	ATG, MMF, TCRab/ CD19 depletion	Nil	18.3, alive
	52 M	Fever, rash, arthritis	1.9	c.121A > C; pMet41Leu, N/A	Nil	2, csDMARD	NA	PB, MUD	R-flu, bu	ATG, CNI, MMF	Grade IV aGVHD, Limited cGVHD	10.9, alive
	67 M	Rash	б	c.121A > G; pMet41Val, N/A	Nil	N/A	N/A	PB, MUD	R-flu, mel	Alem, Cyc	Limited cGVHD	12.2, dead
	65 M	Fever, rash, chondritis, lung, VTE	4.67	c.122 T > C; pMet41Thr, 73%	MDS	2; csDMARD	N/A	PB, MUD	R-flu, treo	ATG, Cyc. MTX	Grade I aGVHD	14.4, alive
	59 M	N/A	4.3	c.167C>T; pSer56Phe, 90%	MDS	2; Azac	N/A	PB, MRD	R-flu, bu, cy, amsa, arac	ATG, CNI, MMF	Grade II aGVHD	29.1, alive
	59 M	Arthritis, chon- dritis	5.1	c.118-1G>C; N/A	SOM	5; csDMARD, bDMARD, tsDMARD, Azac	N/A	PB, MMRD	R-flu, bu, TT	Cy, CNI, MMF	Grade II aGVHD	4.3, alive
	59 M	Fever, rash, arthritis, VTE	4.75	c.122 T > C; pMet41Thr, 75%	MDS	1; tsDMARD	N/A	PB, MUD	S-flu, treo	ATG, Cyc, MTX	Nil	0.9, alive
Guerineau et al. April 2024, France[32]	29 M	Fever, rash, myositis, epididymitis	Т	c.121A > G; pMet41Val, 63%	MDS	3; CS, Azac Venetoclax	VEXAS pro- gression: inflamma- torv	PB, MRD	Bu, cy	ATG, Cyc, MTX	Grade III aGVHD, cGVHD	24, alive

🖄 Springer

Table 1 (continued)	inued)											
Reference	Age, sex	Clinical presen-	Time between	Mutation, variant allele fre-	Concurrent	Lines, and types	Transplantation			GvHD monthylavie	Post-transplantation	tion
		phenotype	symptoms and transplant (years)	ducircy ( 10)		or u caunent pre-transplant	Indication	Type, donor	Conditioning	рюриунахиз	Complica- tions	Follow-up (months) alive /dead
Diarra et al. Febru- ary 2022, France[33]	50 M	Rash, chondri- tis, lung, DVT	7	c.122 T > C; pMet41Thr, 67%	NDS	6; CS, dapsone, colchi- cine, ana canakinumab, siltuximab	VEXAS pro- gression: inflamma- tory	PB, MUD	S-flu, bu,	ATG, Cyc, MTX	Infections: bacterial catheter	3, alive
Loschi et al. Novem- ber 2021, France[34]	70 M	Rash, arthritis	a	c.121A > C; pMct41Leu, N/A	SCIM	8; CS, HCQ, thalidomide, MTX, infx, ana, usteki- numab, IVIG, bara, cy, ruxo	Likely MDS progression as Increased transfusion support and inflam- matory symptoms controlled	PB, MMUD	R-flu, bu	Cy, Cyc, MMF	Grades I and II aGVHD (gastroin- testinal and cutaneous respec- tively)	4, alive
van Leeuwen- Kerkhoff et al. August 2022, Nether- lands[35]	50 M	Fever, rash, chondritis, arthritis	ς	e.122 T>C; pMet41Thr, 43%	Nil	5; CS, aza, RTX, Canaki- numab	VEXAS progres- sion: steroid dependency	PB, MMUD	Flu, mel, thio	ATG, MMF, Canaki - numab	Grade 1 aGVHD <i>Infections:</i> CMV	9, alive
Al-Hakim et al. September 2022, UK[29]	51 M	Fever, Rash, Lung	0	c.121A > G; pMet41Val, 50%	SQM	4;CS, ana, bara colchicine	VEXAS pro- gression: inflamma- tory symp- toms and transfusion dependency	PB, MRD	Flu, bu, thio	Cyc, Tac, MMF	<i>Infection:</i> Salmonella, pseu- domonas	0.4, death due to infection
	67 M	Fever, rash, PE	ς	c.121A > G; pMet41Val, 24%	Nil	3; CS, MTX, HCQ	VEXAS pro- gression: inflam- matory symptoms	PB, MUD	Flu, mel	Alem, Cyc	aGVHD, HLH Infection: EBV	5, alive
	61 M	Fever, rash, lung	6	c.122 T>C; pMet41Thr, 50%	II	6; CS, MTX, aza, MMF, ana, tcz, bara	VEXAS pro- gression: inflamma- tory symp- toms and transfusion dependency	PB, MRD	Flu, treo	Cyc, MMF	Myelitis, optic neu- ropathy <i>Infection:</i> Clostridium difficile	<ol> <li>death due to sepsis, multiorgan failure and cardiac arrest</li> </ol>

Reference	Age, sex	Clinical presen-			Concurrent	Lines, and types	Transplantation	-		GvHD	Post-transplantation	ation
		tation/disease phenotype	symptoms and transplant (years)	quency (%)	haem disease	ot treatment pre-transplant	Indication	Type, donor	Conditioning	prophylaxis	Complica- tions	Follow-up (months) alive /dead
Mangaonkar et al. Novem- ber 2022, USA[15]	63 M	Relapsing polychondritis	N/A	c.122 T>C; pMet41Thr, N/A	SCIM	11, CS, MTX, MMF, decit- abine, HCQ, testosterone, cyc, RTX, tcz, adalimumab, abatacept, lenalidomide	MDS and VEXAS progression: transfusion dependency and inflam- matory symptoms	PB, MUD	Flu, mel	Cy, Tac, MMF	Mucositis Infection: bacteraemia (organism N/A)	16.2, alive
	60 M	Urticarial vasculitis	A/A	c.121A > G; pMet41 Val, N/A	Nil	3; CS, dapsone, Oma, MMF	VEXAS pro- gression: inflam- matory symptoms	PB, MRD	Flu, mel	Cy, Tac, MMF	Nil	12.8, alive
	59 M	Arthritis	Ν/Α	c.118-1G>C (splice variant); N/A	Nil	7; CS, MTX, leftunomide, adalimumab, etanercept, infx, tcz	VEXAS pro- gression: inflam- matory symptoms	PB, MRD	Flu, mel	Tac, MTX	Drug induced rash	3.8, alive
	74 M	N/A	N/A	c.122 T > C, pMet41Tht, N/A	ĨŻ	3; CS, ruxo, tez	VEXAS pro- gression: inflam- matory symptoms	PB, MUD	Flu, mel	Cy, tac, MMF	Grade 1 aGVHD Mucositis, <i>Infection:</i> Clostridium difficile, Escherichia coli	2.9, alive
	49 M	Relapsing polychondritis	A/A	c.122 T > C; pMet41Tht, N/A	MDS	l; CS	MDS and VEXAS progression: inflam- matory symptoms	PB, MUD	Flu, mel	Cy, tac, MMF	Mild dermal hypersen- siti vity reaction	9.6, ali ve
Obiorah et al. August 2021, USA[36] Transplantation a	69 M and retrospec	Obiorah et al. 69 M N/A N/A N/A August 2021, USA[36] Transplantation and retrospective diagnosis of VEXAS (group 2)	N/A EXAS (group 2)	c.121A > C; pMet41Leu, 86%	MM	3; CS, lena- lidomide, bortezomib	MM-incom- plete response	N/A	N/A	N/A	MM relapse	6, alive
Al-Hakim et al. September 2022, UK[29]	62 M	Fever, rash, relapsing chondritis		c.121A > C; pMet41Leu, 40%	SOM	1; CS	Likely both VEXAS and MDS: Transfusion	PB, MUD	Flu, bu	ATG, cyc	Grade I cGVHD	40, alive

 $\textcircled{ } \underline{ \widehat{ } }$  Springer

Reference	Age, sex	Clinical presen-		Mutation, variant allele fre-	Concurrent	Lines, and types	Transplantation	_		GvHD	Post-transplantation	tation
		tation/disease phenotype	symptoms and transplant (years)	quency (%)	haem disease	of treatment pre-transplant	Indication	Type, donor	Conditioning	prophylaxis	Complica- tions	Follow-up (months) alive /dead
Diarra et al. Febru- ary 2022, France[33]	46 M	Fever, rash, vasculitis, orchitis	m	c.121A > G; pMet41 Val, 73%	SQM	6; CS, ana, dapsone, canakinumab, aza, HCQ	VEXAS pro- gression: inflamma- tory—vas- culitis	PB, MUD	S-flu, bu	ATG, cyc, MTX	cGVHD	32, alive
	59 M	Rash	m	c.121A > G; pMet41 Val, 88%	MDS, Myelofi- brosis	8; CS, CP, IVIG, RTX, danazol, ana, dapsone, canakinumab	VEXAS pro- gression: inflam- matory – vasculitis	BM, MRD	R-flu, bu	Cyc, MTX	cGVHD	67, alive
	65 M	Rash, vasculitis, chondritis	0	c.121A > G; pMet41Leu, 43%	SOM	7; CS, MTX, ana, canaki- numab, tcz, IVIG, aza	VEXAS pro- gression	PB, MUD	R-flu, bu	Cy, MMF, cyc	Grade I aGVHD <i>Infections:</i> BK and CMV	38, alive
	58 M	Chondritis, lung	0	c.121A > G; pMet41Val, N/A	SQIM	5;CS, tcz, adaliimumab, aza, ruxo	VEXAS pro- gression	PB, MRD	R-flu, bu, thio	Cy, MMF, cyc	Grades I and II aGVHD (cutane- ous and gastrointes- tinal respec- tively) <i>Infections:</i> bacterial catheter	5, alive
	55 M	Chondritis	Ś	c.121A > G; pMet41Val, N/A	MDS, Myelofi- brosis	4, CS, MMF, colchicine, aza	VEXAS pro- gression	PB, MUD	Bu, CPA, ATG	Cyc, MTX	Grade III a GVHD <i>Infections:</i> bacterial catheter, fusarium	4, death due to infection
Cherniawsky et al. Decem- ber 2022, Canada[37]	68F	Arthritis, relapsing polychondritis	Ś	N/A	SQM	5; CS, MTX, aza, etaner- cept, cyc	MDS pro- gression: transfusion dependency	PB, MRD	R-bu, flu	Cy, MMF, tac	Infection: varicella	13, alive
	61 M	Fever, rash, lung, relaps- ing polychon- dritis, DVT	Ξ	c.122 T > C; pMet41Thr, N/A	IIN	8; CS, Aza, MTX, tcz, Infx, ana, RTX, bara	Likely VEXAS progression: inflam- matory evundoms	PB, MRD	R-bu, flu	Cy, MMF, tac	Infection: fungal (invasive)	1, death due to infection

Reference	Age, sex	Clinical presen-	Time between	Clinical presen- Time between Mutation, variant allele fre-	Concurrent	Lines, and types Transplantation	Transplantation			GvHD	Post-transplantation	ation
		tation/disease phenotype	symptoms and transplant (years)	quency (%)	haem disease	of treatment pre-transplant	Indication	Type, donor	Type, donor Conditioning	prophylaxis	Complica- tions	Follow-up (months) alive /dead
Mascaro et al. August 2023, Spain[38]	55 M	Fever, rash	m	c.118-1G>C; 72%	NA	I; CS	MDS and VEXAS progression	PB, MUD	Flu, bu	Cyc, MMF	'Dulid to Mod' GVHD	Mild to Mod' 84, death due GVHD infection after bilateral lower limb amputation secondary to acute arterial ischaemia
Gunnarsson et al. July 2023, Swe- den[39]	66 M	Portal vein thrombosis	2	c.121A > C; pMet41Leu, N/A	MDS, MM I; Azac	1; Azac	MM—incom- N/A, MUD plete response	N/A, MUD	N/A	N/A	N/A	12, alive

🖄 Springer

donor, MUD matched unrelated donor, N/A not available, NSAID non-steroid anti-inflammatory drug, PB peripheral blood, R reduced intensity, RTX rituximab, ruxo ruxolitinib, S standard row, bu busulfan, cGVHD chronic graft versus host disease, CMV cytomegalovirus, CNI calcineurin inhibitor, CS corticosteroids, bDMARDS biologic disease-modifying antirheumatic drugs, Infx infliximab, mel melphalan, MDS myelodysplastic syndrome, MM multiple myeloma, MMF mycophenolate, MTX methotrexate, MMUD mismatched unrelated donor, MRD matched related csDMARDS conventional synthetic disease-modifying antirheumatic drugs, tsDMARDS targeted synthetic disease-modifying antirheumatic drugs, cy cyclophosphamide, cyc ciclosporine, DVT deep venous thromobosis, EBV Epstein Barr virus, flu fludarabine, HCQ hydroxychloroquine, HLH haemophagocytic lymphohistiocytosis, IVIG intravenous immunoglobulin, oma omalizumab, mar Daricitinio, om done azacitaunie, *puru* azatnioprine, azad aGVHD Acute graft versus host disease, alem alemtuzumab, amsa amsacrine, Ana anakinra, AIG anutinymocyte gioouiin, aza intensity, Tac tacrolimus, Tcz tocilizumab, tofa tofacitinib, Thio thiotepa, Treo treosulphan, Tx transplant due to the known refractory nature of VEXAS, others have suggested evaluating for a suitable haematopoietic stem cell donor at the time of VEXAS for patients who might be seemed appropriate for transplantation [13]. Collectively, given the limited number of cases concentrated in USA and European centres, lack of prospective transplantation in VEXAS alone, and long-term outcomes, clarity on when, who, and how to transplant is urgently needed to inform a holistic, multidisciplinary approach [14].

Previously, reports have indicated patient selection as follows: age 75 years or less, genetically confirmed to have *UBA1* mutation consistent with VEXAS and meeting at least one of the following: (a) severe, glucocorticoid refractory, recurrent inflammatory symptoms, (b) persistent ( $\geq$  3 months) cytopenias, including the need for packed red blood cell transfusion and/or platelet transfusions, and (c) coexistent myeloid malignancy or clonal abnormalities predictive of myeloid transformation [15]. A prospective phase 2 clinical trial (NCT05027945) has begun, which will hopefully shed further light on the indications for allo-HSCT in VEXAS [16].

Given the disease's demographics, a comprehensive assessment is required including organ function, infection risk, personal motivation, and social support [13]. The average age of patients included in this review was below 60 years, with the oldest being 74 years old. Hence, the current literature on allo-HSCT may not be directly applicable to older patients. It is also worth noting that in the literature on haematological malignancy, comorbidities, and performance status independently predict patient outcomes [17, 18].

In addition to patient selection, reduced intensity conditioning regimens and advances in GVHD prophylaxis may be helpful in informing the approach to older patients [19]. Currently, most centres have based allo-HSCT induction therapy for VEXAS on protocols for non-malignant conditions [20, 21]. Regarding conditioning regimens in haematological malignancies, a reduced dosing regimen comprising fludarabine and an alkylating agent has shown comparable efficacy to traditional myeloablative conditioning, with notable advantages including better morbidity and toxicity profiles [22, 23]. On the other hand, as VEXAS might be classified as a non-malignant disease, with the allo-HSCT regimens derived from such disease entities, it has been reported that when compared to malignant disease processes, regimens for non-malignant diseases has a threefold higher risk of graft failure compared to malignant diseases, and lower risk with higher conditioning intensity regimens [24]. In group 1, reduced conditioning regimen was performed in just under 40%.

In our meta-analytic data, near 50% of patients received allo-HSCT from an HLA-matched unrelated donor and GVHD, both acute and chronic were reported in many of the cases (n = 20, 57.1%). Treosulfan, an alkylating agent, has been shown to improve overall survival rates in older patients with MDS and had a mild toxicity profile [25]. In this review, only four individuals were treated with treosulfan, of which two had either grade I or II acute GVHD. Further considerations to reduce graft failure have been proposed targeting the T cells where prominent activation may be observed, although controlling T cell overactivation prior to transplant would be desirable, it may not be realistic or feasible in clinical practice [26].

Ultimately, larger studies, evaluating both VEXAS with and without concurrent haematological malignancy are required along with detailed patient selection criteria. In the setting of an alternate rheumatological or autoimmune disease and transplantation for non-VEXAS cases, overall survival at 70%, and transplant-related mortality at 20.5% have been reported [27]. For patients with VEXAS alone and transplantation, this comprised of less than a third of patients in group 1, while remainder had concurrent haematological malignancy. Additionally, role of allo-HSCT patients with VEXAS and MDS requires further exploration, as this mode of treatment in lower-risk MDS are not favourable at 3 years, with overall survival at 58% and transplant-related mortality at 30%, respectively [28].

In the collected cases, the median time following transplant was relatively short, at only 10.9 months, with the longest follow-up in the prospective cases at 29.1 months. Therefore, while theoretically, transplantation may eliminate the *UBA1* mutation, further prospectively collected longterm data is needed. In terms of mortality, for group 1, there were three deaths (13.0%) with data available for one for which infection was implicated. It has also been reported that poor outcomes might also be observed due to chronicity of symptoms, in addition to advancing age and comorbidities, but also complications from years of immunosuppressive treatments [29].

A final remark pertains to the scarcity of data regarding post-HSCT monitoring in VEXAS. At least 6 patients revealed disappearance of *UBA1* mutations as early as 3–6 months after transplant, paralleling the reversion of the disease clinical phenotype [14]. Methods to track longitudinal clonal dynamics in VEXAS are not standardised and both Sanger (with the sensitivity caveat) or digital droplet PCR have been used [30]. As to chimerism, if a full chimerism is required even in cases without haematological malignancies is another unsolved matter as mixed chimerism has been reported to be linked with resolution of the disease, albeit with short follow-up thereby precluding the evaluation of whether the HSCT conditioning regimen may still be the reason of VEXAS phenotype reversion.

In summary, VEXAS syndrome is a complex, multifaceted disease entity often occurring with concurrent haematological malignancy. While allo-HSCT offers a promising treatment option, long-term data on acute and chronic complications, as well as guidelines around patient selection and personalised conditioning regimens are needed. A focused multidisciplinary approach is warranted to encompass the heterogeneity of VEXAS manifestations. Although the last 4 years have been a time of great proliferation of our understanding of VEXAS, there remains much to discover. Large, international collaborations with prospective data collection and rigorous clinical trials are required to delineate how best to diagnose and treat VEXAS despite its protean presentations.

Author contribution SA developed concept, data collection, and manuscript preparation. CG planned and organised revision.

**Funding** Open Access funding enabled and organized by CAUL and its Member Institutions.

# Declarations

Competing interests The authors declare no competing interests.

**Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.

# References

- Beck DB, Ferrada MA, Sikora KA, Ombrello AK, Collins JC, Pei W et al (2020) Somatic mutations in UBA1 and severe adult-onset autoinflammatory disease. N Engl J Med 383(27):2628–2638. https://doi.org/10.1056/NEJMoa2026834
- Kao RL, Jacobsen AA, Billington CJ, Yohe SL, Beckman AK, Vercellotti GM et al (2022) A case of VEXAS syndrome associated with EBV-associated hemophagocytic lymphohistiocytosis. Blood Cells Mol Dis 93:102636. https://doi.org/10.1016/j.bcmd. 2021.102636
- Çetin G, Klafack S, Studencka-Turski M, Krüger E, Ebstein F (2021) The ubiquitin-proteasome system in immune cells. Biomolecules 11(1):60. https://doi.org/10.3390/biom11010060
- Beck DB, Bodian DL, Shah V, Mirshahi UL, Kim J, Ding Y et al (2023) Estimated prevalence and clinical manifestations of UBA1 variants associated with VEXAS syndrome in a clinical population. JAMA 329(4):318–324. https://doi.org/10.1001/jama.2022. 24836
- 5. Kosmider O, Possémé C, Templé M, Corneau A, Carbone F, Duroyon E et al (2024) VEXAS syndrome is characterized by

inflammasome activation and monocyte dysregulation. Nat Commun 15(1):910. https://doi.org/10.1038/s41467-024-44811-4

- Sirenko M, Bernard E, Creignou M, Domenico D, Farina A, Arango Ossa JE et al (2024) Molecular and clinical presentation of UBA1-mutated myelodysplastic syndromes. Blood e2023023723. https://doi.org/10.1182/blood.2023023723
- Boyadzhieva Z, Ruffer N, Kötter I, Krusche M (2023) How to treat VEXAS syndrome: a systematic review on effectiveness and safety of current treatment strategies. Rheumatology (Oxford) 62(11):3518–3525. https://doi.org/10.1093/rheumatology/kead240
- Keating GM (2012) Azacitidine: a review of its use in the management of myelodysplastic syndromes/acute myeloid leukaemia. Drugs 72(8):1111–1136. https://doi.org/10.2165/11209430-00000 0000-00000
- 9. Bourbon E, Heiblig M, Gerfaud Valentin M, Barba T, Durel C-A, Lega JC et al (2021) Therapeutic options in VEXAS syndrome: insights from a retrospective series. Blood 137(26):3682–3684. https://doi.org/10.1182/blood.2020010177
- Gurnari C, McLornan DP (2022) Update on VEXAS and role of allogeneic bone marrow transplant: considerations on behalf of the Chronic Malignancies Working Party of the EBMT. Bone Marrow Transplant 57(11):1642–1648. https://doi.org/10.1038/ s41409-022-01774-8
- Mangaonkar A, Langer KJ, Lasho T, Finke C, Litzow MR, Hogan WJ et al (2024) Pilot prospective study of reduced intensity conditioning allogeneic hematopoietic stem cell transplantation in patients with Vexas syndrome. Transplant Cell Ther 30(2):S292. https://doi.org/10.1016/j.jtct.2023.12.397
- Georgin-Lavialle S, Terrier B, Guedon AF, Heiblig M, Comont T, Lazaro E et al (2022) Further characterization of clinical and laboratory features in VEXAS syndrome: large-scale analysis of a multicentre case series of 116 French patients. Br J Dermatol 186(3):564–574. https://doi.org/10.1111/bjd.20805
- Bruno A, Gurnari C, Alexander T, Snowden JA, Greco R (2023) Autoimmune manifestations in VEXAS: opportunities for integration and pitfalls to interpretation. J Allergy Clin Immunol 151(5):1204–1214. https://doi.org/10.1016/j.jaci.2023.02.017
- Gurnari C, Koster L, Baaij L, Heiblig M, Yakoub-Agha I, Collin M et al (2024) Allogeneic hematopoietic cell transplantation for VEXAS syndrome: results of a multicenter study of the EBMT. Blood Adv 8(6):1444–1448. https://doi.org/10.1182/bloodadvan ces.2023012478
- Mangaonkar AA, Langer KJ, Lasho TL, Finke C, Litzow MR, Hogan WJ et al (2023) Reduced intensity conditioning allogeneic hematopoietic stem cell transplantation in VEXAS syndrome: data from a prospective series of patients. Am J Hematol 98(2):e28–e31. https://doi.org/10.1002/ajh.26786
- NCT05027945: A phase II study of allogeneic hematopoietic stem cell transplant for subjects with VEXAS (vacuoles, E1 enzyme, X-linked, autoinflammatory, somatic) syndrome: ClinicalTrials. gov. https://clinicaltrials.gov/ct2/show/NCT05027945. Accessed 2/7/24 2024
- Jang RW, Caraiscos VB, Swami N, Banerjee S, Mak E, Kaya E et al (2014) Simple prognostic model for patients with advanced cancer based on performance status. J Oncol Pract 10(5):e335-341. https://doi.org/10.1200/jop.2014.001457
- Piccirillo JF, Tierney RM, Costas I, Grove L, Spitznagel EL (2004) Prognostic importance of comorbidity in a hospital-based cancer registry. JAMA 291(20):2441–2447. https://doi.org/10. 1001/jama.291.20.2441
- Penack O, Peczynski C, Mohty M, Yakoub-Agha I, Styczynski J, Montoto S et al (2020) How much has allogeneic stem cell transplant-related mortality improved since the 1980s? A retrospective

- Laberko A, Sultanova E, Gutovskaya E, Shipitsina I, Shelikhova L, Kurnikova E et al (2019) Mismatched related vs matched unrelated donors in TCRαβ/CD19-depleted HSCT for primary immunodeficiencies. Blood 134(20):1755–1763. https://doi.org/10.1182/blood.2019001757
- Fox TA, Chakraverty R, Burns S, Carpenter B, Thomson K, Lowe D et al (2018) Successful outcome following allogeneic hematopoietic stem cell transplantation in adults with primary immunodeficiency. Blood 131(8):917–931. https://doi.org/10.1182/ blood-2017-09-807487
- 22. Fasslrinner F, Schetelig J, Burchert A, Kramer M, Trenschel R, Hegenbart U et al (2018) Long-term efficacy of reduced-intensity versus myeloablative conditioning before allogeneic haemopoietic cell transplantation in patients with acute myeloid leukaemia in first complete remission: retrospective follow-up of an openlabel, randomised phase 3 trial. Lancet Haematol 5(4):e161–e169. https://doi.org/10.1016/s2352-3026(18)30022-x
- 23. Aoudjhane M, Labopin M, Gorin NC, Shimoni A, Ruutu T, Kolb HJ et al (2005) Comparative outcome of reduced intensity and myeloablative conditioning regimen in HLA identical sibling allogeneic haematopoietic stem cell transplantation for patients older than 50 years of age with acute myeloblastic leukaemia: a retrospective survey from the Acute Leukemia Working Party (ALWP) of the European group for Blood and Marrow Transplantation (EBMT). Leukemia 19(12):2304–2312. https://doi.org/10. 1038/sj.leu.2403967
- Olsson R, Remberger M, Schaffer M, Berggren DM, Svahn BM, Mattsson J et al (2013) Graft failure in the modern era of allogeneic hematopoietic SCT. Bone Marrow Transplant 48(4):537–543. https://doi.org/10.1038/bmt.2012.239
- 25. Beelen DW, Stelljes M, Reményi P, Wagner-Drouet EM, Dreger P, Bethge W et al (2022) Treosulfan compared with reduced-intensity busulfan improves allogeneic hematopoietic cell transplantation outcomes of older acute myeloid leukemia and myelodysplastic syndrome patients: Final analysis of a prospective randomized trial. Am J Hematol 97(8):1023–1034. https://doi.org/10.1002/ajh.26620
- Heiblig M, Patel BA, Groarke EM, Bourbon E, Sujobert P (2021) Toward a pathophysiology inspired treatment of VEXAS syndrome. Semin Hematol 58(4):239–246. https://doi.org/10.1053/j. seminhematol.2021.09.001
- 27. Greco R, Labopin M, Badoglio M, Veys P, Furtado Silva JM, Abinun M et al (2019) Allogeneic HSCT for autoimmune diseases: a retrospective study from the EBMT ADWP, IEWP, and PDWP working parties. Front Immunol 10:1570. https://doi.org/10.3389/ fimmu.2019.01570
- Robin M, Porcher R, Zinke-Cerwenka W, van Biezen A, Volin L, Mufti G et al (2017) Allogeneic haematopoietic stem cell transplant in patients with lower risk myelodysplastic syndrome: a retrospective analysis on behalf of the Chronic Malignancy Working Party of the EBMT. Bone Marrow Transplant 52(2):209–215. https://doi.org/10.1038/bmt.2016.266

- Al-Hakim A, Poulter JA, Mahmoud D, Rose AMS, Elcombe S, Lachmann H et al (2022) Allogeneic haematopoietic stem cell transplantation for VEXAS syndrome: UK experience. Br J Haematol 199(5):777–781. https://doi.org/10.1111/bjh.18488
- Gurnari C, Pascale MR, Vitale A, Diral E, Tomelleri A, Galossi E et al (2024) Diagnostic capabilities, clinical features, and longitudinal UBA1 clonal dynamics of a nationwide VEXAS cohort. Am J Hematol 99(2):254–262. https://doi.org/ 10.1002/ajh.27169
- Stiburkova B, Pavelcova K, Belickova M, Magaziner SJ, Collins JC, Werner A et al (2023) Novel somatic UBA1 variant in a patient with VEXAS syndrome. Arthritis Rheumatol 75(7):1285– 1290. https://doi.org/10.1002/art.42471
- Guerineau H, Kohn M, Al Hamoud A, Sellier J, Osman J, Cabannes-Hamy A (2024) Could it be VEXAS? Ann Hematol 103(6):2169–2171. https://doi.org/10.1007/s00277-024-05750-8
- Diarra A, Duployez N, Fournier E, Preudhomme C, Coiteux V, Magro L et al (2022) Successful allogeneic hematopoietic stem cell transplantation in patients with VEXAS syndrome: a 2-center experience. Blood Adv 6(3):998–1003. https://doi.org/10.1182/ bloodadvances.2021004749
- Loschi M, Roux C, Sudaka I, Ferrero-Vacher C, Marceau-Renaut A, Duployez N et al (2022) Allogeneic stem cell transplantation as a curative therapeutic approach for VEXAS syndrome: a case report. Bone Marrow Transplant 57(2):315–318. https://doi.org/ 10.1038/s41409-021-01544-y
- 35. van Leeuwen-Kerkhoff N, de Witte MA, Heijstek MW, Leavis HL (2022) Case report: up-front allogeneic stem cell transplantation in a patient with the VEXAS syndrome. Br J Haematol 199(3):e12–e15. https://doi.org/10.1111/bjh.18424
- 36. Obiorah IE, Patel BA, Groarke EM, Wang W, Trick M, Ombrello AK et al (2021) Benign and malignant hematologic manifestations in patients with VEXAS syndrome due to somatic mutations in UBA1. Blood Adv 5(16):3203–3215. https://doi.org/10.1182/ bloodadvances.2021004976
- Cherniawsky H, Friedmann J, Nicolson H, Dehghan N, Stubbins RJ, Foltz LM et al (2023) VEXAS syndrome: a review of bone marrow aspirate and biopsies reporting myeloid and erythroid precursor vacuolation. Eur J Haematol 110(6):633–638. https:// doi.org/10.1111/ejh.13944
- Mascaro JM, Rodriguez-Pinto I, Poza G, Mensa-Vilaro A, Fernandez-Martin J, Caminal-Montero L et al (2023) Spanish cohort of VEXAS syndrome: clinical manifestations, outcome of treatments and novel evidences about UBA1 mosaicism. Ann Rheum Dis 82(12):1594–1605. https://doi.org/10.1136/ard-2023-224460
- Gunnarsson K, Pomiano NV, Tesi B, Tobiasson M, Creignou M, Ungerstedt J (2023) VEXAS – nytt autoinflammatoriskt syndrom med bred symtombild. Lakartidningen 119:22024

**Publisher's Note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.