## The potential diagram for oxygen at pH 7

## Paul M. WOOD

Department of Biochemistry, University of Bristol, School of Medical Sciences, Bristol BS8 1TD, U.K.

Successive one-electron reductions of molecular oxygen yield the superoxide radical  $(O_2^{-})$  H<sub>2</sub>O<sub>2</sub>, the hydroxyl radical (OH<sup>+</sup>) and water. Redox potentials at pH 7 for one-, two- and four-electron couples involving these states are presented as a potential diagram. The significance of each of these potentials is explained. The complete potential diagram enables complex systems to be rationalized, such as production of OH<sup>+</sup> by H<sub>2</sub>O<sub>2</sub> plus Fe<sup>3+</sup>.

In inorganic chemistry, diagrams showing potentials between different oxidation states of an element are widely used as summaries and teaching aids. These potentials are variously known as redox potentials, oxidation-reduction potentials and standard electrode potentials.

Fig. 1 presents a potential diagram for oxygen at the biochemical standard state of pH 7. Successive oneelectron reductions of molecular oxygen give the superoxide radical anion  $(O_2^{-*})$ ,  $H_2O_2$ , the hydroxyl radical (OH<sup>\*</sup>) and water. The standard sign conventions lead to redox potentials being cited for reductive reactions:

Oxidized +  $n e^- \rightarrow$  reduced (Clark, 1960)

However, the potential diagram is equally useful for discussion of oxidations.

The voltages shown apply to standard states; as explained by myself (Wood, 1987), these approximate to  $10^5$  Pa (0.987 atm) of O<sub>2</sub>, 1 mol·l<sup>-1</sup> concentrations of O<sub>2</sub><sup>--</sup>, H<sub>2</sub>O<sub>2</sub> and OH<sup>+</sup>, and pure liquid water. The values given are derived from diagrams for pH 0 and pH 14 in a recent compilation (Hoare, 1985). Data for OH<sup>+</sup> are based on the mean of two recent determinations:  $E_0$ (OH<sup>+</sup>/H<sub>2</sub>O) = +2.72 V (Schwarz & Dodson, 1984) and +2.74 V (Kläning *et al.*, 1985). As an example of the conversion to pH 7, consider  $E_0$  (O<sub>2</sub>/H<sub>2</sub>O<sub>2</sub>) = +0.695 V. The equation is written as a reduction:

$$O_2 + 2 H^+ + 2e^- \rightarrow H_2O_2$$

For this reaction,  $\Delta G'_0 = \Delta G_0 - \mathbf{R}T \ln [\mathrm{H}^+]^2$ . Since

$$\Delta G = -nEF$$

where F is Faraday's constant (96485 J·V<sup>-1</sup>·mol<sup>-1</sup>),  $E'_0 = E_0 + (\mathbf{R}T/2F) \ln 10^{-14}$ . For 25 °C,  $E'_0 = E_0 - 0.414$  V. In converting to pH 7, care must be taken to choose appropriate protonation states, or include pK values (Hoare, 1985). Thus  $O_2^{-*}$  becomes protonated at acid pH  $(O_2^{-*} + H^+ \rightarrow HO_2^{-}, pK = 4.8)$ , whereas OH and  $H_2O_2$  both lose a proton at high pH (HO<sup>•</sup>  $\rightarrow O^{-*} + H^+$ , pK = 11.8;  $H_2O_2 \rightarrow HO_2^{-+} + H^+$ , pK = 11.7). The potentials shown in the diagram all fall with increasing pH, apart from  $E_0$  ( $O_2/O_2^{-*}$ ), which is pH-independent.

A useful property of potential diagrams is that alternative connections between different states have the

same value for  $\sum nE$ , where *n* is the number of electrons transferred in a reductive step with redox potential *E*. For example,  $-0.33+0.89 = 2 \times (+0.28)$ . As has been mentioned, for each step,  $\Delta G = -nEF$ . Alternative pathways must have the same overall  $\Delta G$  change, implying the same value for  $\sum nE$ .

The following paragraphs will discuss the steps constituting the diagram in turn. Put simply, a reductive step on the diagram implies oxidation of a donor with a similar or lower redox potential, whereas an oxidative step on the diagram implies reduction of an acceptor with a similar or higher redox potential.

The complete four-electron reduction,  $O_2 + 4H^+ + 4e^- \rightarrow 2H_2O$ ,  $E'_0(O_2/H_2O) = +0.815$  V, constitutes the terminal reaction of the aerobic respiratory chain. The converse reaction provides the electron source for plant photosynthesis.

The reduction of molecular oxygen to  $H_2O_2$  is less energetically favourable than the complete reduction to water, but requires only two electrons in place of four. For this reason  $H_2O_2$  is the product of oxygen reduction by many oxidases.

The high redox potential for  $H_2O_2$  reduction to water  $(E'_0 = +1.349 \text{ V})$  implies a latent oxidizing power, put to use by peroxidases. For many peroxidases,  $H_2O_2$  reacts with the resting ferric enzyme to yield a 'Compound I', in which the iron has undergone a one-electron oxidation to a ferryl (Fe<sup>IV</sup>=O)<sup>2+</sup> state, whereas the second electron is abstracted from another group close by (Dunford & Stillman, 1976; Penner-Hahn *et al.*, 1983):

$$H_2O_2 + P \cdot Fe^{3+} \rightarrow H_2O + P^{+} \cdot (Fe^{IV} = O)^{2+}$$

The two one-electron redox couples,  $(P^{+*}/P)$  and  $(Fe^{IV}/Fe^{III})$ , may both have potentials approaching +1.0 V (Hayashi & Yamazaki, 1979). This is sufficient for oneelectron oxidation of many aromatic compounds to yield a radical cation, for instance during lignin biosynthesis or lignin breakdown by wood-rotting fungi (Prince & Stiefel, 1987).

The preceding two paragraphs have considered the two-electron couples  $O_2/H_2O_2$  and  $H_2O_2/H_2O$ . The large differences in redox potential between these processes provides a driving force for  $H_2O_2$  dismutation, catalysed by catalase.

Abbreviations and symbols used:  $O_2^{-1}$ , superoxide radical; OH<sup>+</sup>, hydroxyl radical; HO<sub>2</sub><sup>+</sup>, hydroperoxo radical; O<sup>-+</sup>, oxygen anion radical; HO<sub>2</sub><sup>-</sup>, peroxide anion; OH<sup>-</sup>, hydroxyl anion;  $E_0$ , standard electrode potential;  $\Delta G$ , molar Gibbs energy change;  $E'_0$ , electrode potential standard at pH 7.0;  $\Delta G'_0$ , standard Gibbs energy at pH 7.0; F, Faraday constant; E, redox potential; n, number of electrons transferred;  $E_{m,7}$ , midpoint potential in a redox titration at pH 7.0.



Fig. 1. Potential diagram for oxygen at pH 7

Turning to one-electron couples, the reduction of molecular oxygen to  $O_2^{-*}$  has a lower redox potential than for  $H_2O_2$  formation. However, only a single electron need be transferred. For this reason,  $O_2^{-*}$  formation is a characteristic of autoxidation. By contrast,  $O_2^{-*}$  production by neutrophils is a deliberate act of biological warfare (Rossi, 1986). The diagram shows two redox potentials for the molecular oxygen/superoxide couple  $(O_2/O_2^{-*})$ . I (Wood, 1987) have discussed the relationship between these values and why both are useful. The redox potential relative to  $10^5$  Pa of  $O_2$  is *the* standard potential,  $E_0$   $(O_2/O_2^{-*}) = -0.33$  V. However, the redox potential relative to 1 mol of  $O_2 \cdot 1^{-1}$ ,  $E_0$   $[O_{2(sq.)}/O_2^{-*}] = -0.16$  V, is relevant for reversible one-electron exchange:

$$O_2 + D \rightleftharpoons O_2^{--} + D^+$$

Superoxide dismutases contain metal ions with oneelectron redox potentials between those for  $O_2^{-}$  oxidation and reduction, enabling dismutation to take place. For example, the copper atoms of CuZnsuperoxide dismutase have an  $E_{m,7}$  of +0.42 V (Fee & DiCorleto, 1973).

The  $O_2^{-\cdot}/H_2O_2$  couple has a high redox potential. For this reason, the one-electron oxidation of  $H_2O_2$  to  $O_2^{-\cdot}$ can only be brought about by strong oxidizing agents. As an example, consider the effect of high concentrations of  $H_2O_2$  on horseradish peroxidase. The enzyme is converted into a state known as 'Compound III' (Dunford & Stillman, 1976; Nakajima & Yamazaki, 1987). This is an oxy complex, but lacks the stability of oxymyoglobin. Its formation takes place from Compound II, which contains Fe<sup>IV</sup>:

$$H_2O_2 + (Fe^{IV} = O)^{2+} \rightarrow H_2O + (Fe^{3+} O_2^{-+})^{2+}$$
 (1)

$$(Fe^{3+} \cdot O_2^{-})^{2+} \rightleftharpoons Fe^{3+} + O_2^{-}$$
 (2)

These two reactions represent a one-electron oxidation of  $H_2O_2$ , made possible by the high potential of the (Compound II/ferriperoxidase) redox couple, namely +0.96 V (Hayashi & Yamazaki, 1979).

In 1894, Fenton described the strong oxidizing properties of a mixture of  $H_2O_2$  and  $Fe^{2+}$  (Fenton's reagent). The active species was shown by Haber & Weiss (1934) to be OH<sup>•</sup>, formed by one-electron transfer from Fe<sup>II</sup>:

1

$$H_2O_2 + Fe^{2+} \rightarrow OH^{-} + HO^{-} + Fe^{3+}$$
 (3)

A wide range of states of ferrous iron are active (Cohen, 1985). At pH 7, many iron complexes have a redox potential near 0 V [e.g. EDTA, +0.114; oxalate, +0.002; protoporphyrin-IX (free haem), about -0.12 V; Clark (1960)], making electron transfer to H<sub>2</sub>O<sub>2</sub> a 'downhill'

reaction. The traces of unprotected iron present in many biological systems can be sufficient, provided there is a means of regenerating the ferrous state (Cohen, 1985). This can be accomplished by one-electron reduction of  $Fe^{3+}$  by  $O_2^{--}$  (McCord & Day, 1978):

$$O_2^{-\cdot} + Fe^{3+} \underset{k_{-4}}{\stackrel{\star}{\leftrightarrow}} O_2 + Fe^{2+}$$
(4)

Provided the Fe<sup>III</sup>/Fe<sup>II</sup> redox potential is above -0.16 V [ $E_0$  (O<sub>2(aq.)</sub>/O<sub>2</sub><sup>-•</sup>)],  $k_4$  is greater than  $k_{-4}$  (Wood, 1987). The combination of eqn. (4) plus eqn. (3) is the Haber-Weiss reaction (Haber & Weiss, 1934; Cohen, 1985). OH<sup>•</sup> is liable to be formed whenever O<sub>2</sub><sup>-•</sup> is produced in the absence of adequate levels of catalase and superoxide dismutase (Wolff *et al.*, 1986; Halliwell & Gutteridge, 1986).

A final example shows how the whole potential diagram can be used to rationalize a complex system. If the Fe<sup>2+</sup> of Fenton's reagent is replaced by Fe<sup>3+</sup>, similar oxidations can still take place, although less rapidly (Gutteridge & Wilkins, 1983; Gutteridge & Bannister, 1986). In the literature the activating reaction is often said to be a one-electron oxidation of H<sub>2</sub>O<sub>2</sub>, coupled to reduction of ferric iron (Walling, 1982; Halliwell & Gutteridge, 1984):

$$H_2O_2 + Fe^{3+} \rightarrow O_2^{--} + 2H^+ + Fe^{2+}$$

However, as has been stated, the Fe<sup>III</sup>/Fe<sup>II</sup> redox potential at pH 7 is generally near 0 V, compared with +0.89 V for the O<sub>2</sub><sup>-•</sup>/H<sub>2</sub>O<sub>2</sub> couple. As I explained previously (Wood, 1987), a one-electron transfer can have  $\Delta G'_0$  positive, but not *too* positive. For  $\Delta E'_0 = -0.9$  V,  $\Delta G'_0$  will be about +86000 J·mol<sup>-1</sup>. Such a process can scarcely be significant. A more plausible activating reaction would be a two-electron reduction of  $H_2O_2$ :  $E'_0$  $(H_2O_2/H_2O) = +1.349$  V. The iron will either be oxidized to Fe<sup>iv</sup>, with the second electron taken from a complexing ligand (as in a peroxidase), or Fe<sup>v</sup> may be formed transiently (Dunford & Stillman, 1976; Rush & Bielski, 1986). Once a high oxidation state of iron has been produced, there is no difficulty in oxidation of  $H_2O_2$  to superoxide [reactions (1) and (2) above]. Reactions (4) and (3) complete a pathway for hydroxylradical formation, in which all steps are energetically downhill.

## REFERENCES

Clark, W. M. (1960) Oxidation-Reduction Potentials of Organic Systems, Williams and Wilkins, Baltimore

The units are volts (V). The value in parentheses is for 1 mol of  $O_2 \cdot l^{-1}$  as standard state, instead of 10<sup>5</sup> Pa of  $O_2$ . For sources, see the text.

- Cohen, G. (1985) in CRC Handbook of Methods for Oxygen Radical Research (Greenwald, R. A., ed.), pp. 55–64, CRC Press, Boca Raton, Fl
- Dunford, H. B. & Stillman, J. S. (1976) Co-ord. Chem. Rev. 19, 187-251
- Fee, J. A. & DiCorleto, P. E. (1973) Biochemistry 12, 4893-4899
- Fenton, H. J. H. (1894) J. Chem. Soc. 65, 899-910
- Gutteridge, J. M. C. & Bannister, J. V. (1986) Biochem. J. 234, 225-228
- Gutteridge, J. M. C. & Wilkins, S. (1983) Biochim. Biophys. Acta 759, 38-41
- Haber, F. & Weiss, J. (1934) Proc. R. Soc. London Ser. A 147, 332–351
- Halliwell, B. & Gutteridge, J. M. C. (1984) Biochem. J. 219, 1-14
- Halliwell, B. & Gutteridge, J. M. C. (1986) Trends Biochem. Sci. 11, 372–375
- Hayashi, Y. & Yamazaki, I. (1979) J. Biol. Chem. 254, 9101-9106
- Hoare, J. P. (1985) in Standard Potentials in Aqueous Solution (Bard, A. J., Parsons, R. & Jordan, J., eds.), pp. 49–66, IUPAC/Marcel Dekker, New York

Received 29 January 1988; accepted 25 March 1988

- Kläning, U. K., Sehested, K. & Holcman, J. (1985) J. Phys. Chem. 89, 760-763
- McCord, J. M. & Day, E. D. (1978) FEBS Lett. 86, 139-142
- Nakajima, R. & Yamazaki, I. (1987) J. Biol. Chem. 262, 2576–2581
- Penner-Hahn, J. E., McMurry, T. J., Renner, M., Latos-Grazynsky, L., Smith Eble, K., Davis, I. M., Balch, A. L., Groves, J. T., Dawson, J. H. & Hodgson, K. O. (1983) J. Biol. Chem. 258, 12761–12764
- Prince, R. C. & Stiefel, E. I. (1987) Trends Biochem. Sci. 12, 334–335
- Rossi, F. (1986) Biochim. Biophys. Acta 853, 65-89
- Rush, J. D. & Bielski, B. H. J. (1986) J. Am. Chem. Soc. 108, 523-525
- Schwarz, H. A. & Dodson, R. W. (1984) J. Phys. Chem. 88, 3643-3647
- Walling, C. (1982) in Oxidases and Related Redox Systems: Proceedings of the Third International Symposium (King, T. E., Mason, H. S. & Morrison, M., eds.), pp. 85–97, Pergamon Press, New York
- Wolff, S. P., Garner, A. & Dean, R. T. (1986) Trends Biochem. Sci. 11, 27-31
- Wood, P. M. (1987) Trends Biochem. Sci. 12, 250-251