

REVIEW

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Large-fiber neuropathy in Parkinson's disease: a narrative review

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Abstract

Background Numerous studies reported a higher prevalence of polyneuropathy (PNP) in patients with Parkinson's disease (PD) compared to the general population. Importantly, PNP symptoms can aggravate both motor and sensory disturbances in PD patients and negatively impact the disease course. Recent analyses indicate distinct PNP patterns in PD.

Main text This review aims to provide an overview of the current insights into etiological factors, diagnostic methods, and management strategies of large fiber neuropathy in PD. Despite the higher prevalence, the causes of PNP in PD are still not fully understood. A genetic predisposition can underlie PNP onset in PD. Main research attention is focused on long-term levodopa exposure which is suggested to increase PNP risk by depletion of methylation cofactors such as vitamin B12 and accumulation of homocysteine that altogether can alter peripheral nerve homeostasis. Beyond a potential "iatrogenic" cause, alpha-synuclein deposition has been detected in sural nerve fibers that could contribute to peripheral neuronal degeneration as part of the systemic manifestation of PD. Whereas mild axonal sensory PNP predominates in PD, a considerable proportion of patients also show motor and upper limb nerve involvement. Intriguingly, a correlation between PNP severity and PD severity has been demonstrated. Therefore, PNP screening involving clinical and instrument-based assessments should be implemented in the clinical routine for early detection and monitoring. Given the etiological uncertainty, therapeutic or preventive options remain limited. Vitamin supplementation and use of catechol-O-methyltransferase-inhibitors can be taken into consideration.

Conclusion PNP is increasingly recognized as a complicating comorbidity of PD patients. Long-term, large-scale prospective studies are required to elucidate the causative factors for the development and progression of PD-associated PNP to optimize treatment approaches. The overall systemic role of "idiopathic" PNP in PD and a putative association with the progression of neurodegeneration should also be investigated further.

Keywords Parkinson's disease, Polyneuropathy, Large fiber neuropathy, Etiology, Diagnostics, Management

Background

Parkinson's disease (PD) is currently one of the fastest-growing neurological conditions worldwide [1, 2]. Deposition of α -synuclein is closely associated with neurodegeneration, which is not limited to striatal dopaminergic and other central nervous regions but extends to peripheral organs and the peripheral nervous system [3]. Based on its multisystemic nature, PD manifests with a broad spectrum of motor and non-motor symptoms. Polyneuropathy (PNP) represents a peripheral nerve dysfunction involving sensory, motor, and autonomic

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domains [4]. According to the affected nerve structure and type of fibers, PNP can be categorized into axonal versus demyelinating subtype and small (unmyelinated A δ and C fibers) versus large fiber neuropathy (myelinated A α and A β fibers). For this review, we focus on large fiber neuropathy when referring to PNP.

The first association between PD and PNP was observed in 1991 by a pharmacological study of oral levodopa where PNP was reported as an adverse event [5]. Subsequent studies revealed a higher coincidence of PNP in PD patients, mostly as axonal, predominantly sensory PNP. PNP prevalence varies greatly between studies, ranging from 4.8 to 62% [6–8], whereas PNP rate in the general population of similar age is estimated to be approximately 8% [4, 6]. These divergences in PNP rates could be due to differences in the study population (age, disease duration, therapeutic regimen) and applied diagnostic PNP criteria [9]. PNP symptoms include disturbed sensations such as numbness, paresthesia and pain, sensory ataxia, and muscle weakness and can partially overlap with PD symptoms. For example, pain is an unspecific, yet frequent symptom of PD patients among which neuropathic characteristics can indicate a central, but also a peripheral origin, with a negative impact on the quality of life [10]. Moreover, balance and gait difficulties in PD patients can be aggravated by PNP comorbidity leading to an increased risk for falls [11, 12]. Therefore, this review aims to comprehensively summarize the current views on etiological factors, diagnostics, and treatment options of large fiber neuropathy in PD. For this purpose, a selective literature search was performed using PubMed based on the main search terms “Parkinson’s” in combination with “polyneuropathy”, “peripheral neuropathy” or “large fiber neuropathy”.

Neuropathology

While large-fiber neuropathy has been more attributed to extrinsic factors, small-fiber neuropathy is considered an intrinsic feature of PD as evidenced by cutaneous denervation and α -synuclein deposits. Doppler et al. reported depositions of phosphorylated α -synuclein in somatosensory and autonomic nerve fibers in skin biopsies of PD patients accompanied by a significant reduction of intraepidermal small nerve fibers [13]. Intraepidermal small nerve fiber density was found to be reduced in untreated as well as levodopa-treated PD patients compared to controls [14]. In contrast, Podgorny et al. did not observe any difference in epidermal nerve densities between early untreated PD patients and controls [15]. Instead, small-fiber neuropathy in PD was demonstrated by reduced corneal nerve fiber densities suggesting corneal confocal microscopy as a more sensitive tool for early detection of preclinical small fiber loss. In

a longitudinal study, a higher oligomeric α -synuclein load was detected in skin biopsies of patients with PD and multiple system atrophy compared to controls and tauopathies, yielding a high diagnostic performance for synucleinopathies [16]. Interestingly, PD patients displayed a small nerve fiber pathology with progressive denervation after two years of follow-up, highlighting the potential of small-fiber neuropathy as a progression marker in PD.

Due to their invasiveness, nerve biopsies for evaluation of large-fiber nerve alterations were reserved for individual severe PNP cases under enteral or high-dose oral levodopa therapy. Sural nerve biopsies revealed a severe axonal degeneration with a marked reduction in myelinated nerve fiber density, accompanied by endoneurial edema and myelin debris in endoneurial macrophages [17]. Inflammatory changes with perivascular lymphocytic cuffing have also been described [18]. In the superficial peroneal nerve, intra-axial ubiquitin aggregates were found to be more numerous in PD patients with PNP compared to those without PD as possible correlates of the underlying neurodegenerative process [19]. For the first time, Zhang et al. verified the deposition of phosphorylated α -synuclein in sural nerves *in vivo* in all 16 PD patients and none of the 15 controls [20]. Expression of α -synuclein was mainly located in Schwann cells, but scarcely in axons. The same study group demonstrated peripheral nerve inflammation by increased expression of glial fibrillary acidic protein and inflammatory cytokines (IL-1 β , IL-6, TNF- α) in activated Schwann cells of PD patients irrespective of the presence of sensory disturbances [21]. Different expression patterns of phosphorylated α -synuclein and tau protein in sural nerve biopsy even allowed differentiation of PD from the atypical Parkinsonian syndromes multiple system atrophy and progressive supranuclear paralysis [22]. In summary, these findings support the hypothesis of a peripheral origin of α -synuclein pathology and the multisystemic nature of PD, yet the pathogenicity of α -synuclein deposition needs to be clarified. Therefore, clinicopathological correlations by means of nerve biopsy are mandatory to validate peripheral α -synuclein pathology as a diagnostic biomarker and further elucidate the underlying pathomechanisms of PNP in PD.

Etiology

Despite the higher PNP rate in PD, the exact linkage between these two disease entities is still unclear. Age represents a risk factor for developing both PD and PNP. Ceravolo et al. calculated an increase of PNP risk in PD by approximately 8% for each year of age [6]. Age-dependent nutritional deficiencies such as decreased vitamin B12 levels have been discussed to partly influence the onset

of PNP in PD [23]. Yet, the PNP rate in PD is higher than in age-matched controls indicating a higher susceptibility of the PD population. The leading causes of PNP are diabetes, followed by alcohol abuse, toxic agents, vitamin deficiencies, immune-mediated causes, and hereditary factors, whereas 20–30% of PNP cases remain idiopathic [24]. Toth et al. found that 30% of PD patients with PNP had a defined cause for PNP including diabetes, monoclonal gammopathy, and chronic inflammatory demyelinating peripheral neuropathy [25]. The PD population shows a range of comorbidities including cardiovascular diseases and diabetes [26, 27]. Interestingly, diabetes has been associated with increased PD risk [28]. Evidence supports the idea of shared pathomechanisms allowing the repurposing of antidiabetic agents like glucagon-like peptide-1 receptor agonists for neuroprotective effects in PD. Focusing on truly idiopathic PNP will allow a better understanding of the relationship between PD and PNP. However, PNP-causing comorbidities should be taken into consideration to provide a more holistic estimation of PNP prevalence in PD.

Genetics

The occurrence of PNP has been reported in hereditary forms of parkinsonism [29]. A nerve conduction study revealed that 8 of 9 patients with juvenile parkinsonism with PARK2-mutation showed reduced sural nerve amplitudes [30]. PARK2 gene encodes an E3 ubiquitin ligase termed Parkin which plays an important role in proteasomal degradation. Interestingly, PARK2 is expressed in sural nerves [31]. It is hypothesized that proteasomal impairment caused by loss of Parkin function could contribute to neurotoxicity, although the effects on peripheral nerves remain unknown. Another candidate gene to increase susceptibility for PNP could be the MTHFR gene encoding the methylenetetrahydrofolate reductase (MTHFR), an essential enzyme for homocysteine metabolism [32]. Mutation of the MTHFR gene could lead to an elevation of homocysteine levels which is associated with an increased risk for PNP due to its peripheral toxic effects [33]. For PD patients, this mutation is particularly critical since it can aggravate levodopa-induced hyperhomocysteinemia. Moreover, an association of MTHFR polymorphisms with increased risk for developing PD is under debate.

Levodopa metabolism and vitamin deficits

In their pioneering study, Toth et al. demonstrated a significant association between cumulative lifetime levodopa dosage and fasting methylmalonic acid (MMA) levels with PNP severity [25]. Since then, further studies supported the hypothesis that levodopa may play a

causative role in the development of PNP in PD, possibly related to its metabolic products [6, 32, 34, 35].

Levodopa's metabolic cycle starts with methylation by catechol-O-methyltransferase (COMT) using S-adenosylmethionine (SAM) as a methyl donor [32]. Demethylation of SAM produces S-adenosylhomocysteine (SAH) which is hydrolyzed to homocysteine. Homocysteine enters either the remethylation pathway requiring vitamin B12 and folate cofactors or the trans-sulfuration pathway for the production of cysteine involving vitamin B6 as a cofactor. The latter pathway leads to the formation of L-methylmalonyl-coenzyme A which in case of vitamin B12 deficiency results in the accumulation of MMA, an indirect marker of vitamin B12 deficiency. Consequently, chronic levodopa intake can lead to the accumulation of homocysteine and MMA as well as depletion of vitamin B6, B12, and folate, each of which can alter peripheral nerve homeostasis. Homocysteine can exert neurotoxicity by overstimulating glutamate receptors, promoting oxidative stress and DNA hypomethylation, and is linked to various neurological and cardiovascular conditions [36]. Vitamin B6 plays a key role in the synthesis of neurotransmitters, whereas vitamin B12 is particularly involved in nerve regeneration and remyelination [37].

In a large multicenter study with 330 PD patients and 137 controls, PNP risk was stratified according to levodopa exposure [6]. Overall, 19.40% of patients with ≥ 3 years of levodopa exposure, 6.80% with < 3 years of levodopa exposure, 4.82% in the dopa-naïve group, and 8.76% in the control group were diagnosed with PNP-identifying levodopa as a main risk factor. The risk of PNP was 2.38-fold higher in the long-term treated group compared to the control group. This finding is supported by a study of a Romanian cohort reporting a significantly higher PNP prevalence in PD subjects on levodopa treatment compared to untreated PD subjects [38]. In this study, lower vitamin B12 levels correlated with higher levodopa daily dose and treatment, levodopa dose correlated inversely with the nerve amplitudes. In contrast, Shahrizaila et al. observed no difference in the PNP rate between levodopa-naïve and levodopa-treated PD patients [39]. Another study confirmed a higher PNP prevalence in the treated group compared to the untreated group, though concluded after regression analysis, that the levodopa effect was only contributory and surpassed by age and folate levels [23]. Higher homocysteine levels have been reported in PD patients with PNP correlating with levodopa intake [40], but also in drug-naïve de novo PD patients suggesting that homocysteine is independently elevated in PD [41]. A study of an Indian cohort found no difference in homocysteine, vitamin B12, and folate levels

between PD patients and controls, however, the study population exhibited a low overall PNP rate of 9.68% [42].

The impact of levodopa therapy on PNP risk has been shown to be more pronounced with levodopa/carbidopa intestinal gel (LCIG) infusion. A systematic review reported a higher occurrence of PNP under LCIG (42.1%) in comparison to oral levodopa treatment (30.2%) [32]. PNP characteristics differed between the different administration modes as LCIG-treated subjects displayed a sensorimotor PNP with sub-/acute onset, rarely with demyelinating features, compared to the more commonly observed mild chronic axonal predominantly sensory PNP in levodopa-treated patients. In a prospective study of 23 PD patients, two patients developed a subacute PNP, two patients a chronic PNP, and seven patients a subclinical PNP two years after starting LCIG therapy [43]. Mancini et al. studied PNP characteristics under different therapeutic regimens including LCIG, oral levodopa, and other dopaminergic medications [44]. PNP prevalence was shown to be increased under levodopa therapy, independent of the route of administration. The majority of cases exhibited a subacute sensory PNP, whereas 29% of LCIG patients manifested an acute demyelinating motor form. Vitamin B12 and folate levels were significantly lower, homocysteine levels were significantly higher in levodopa-treated compared with non-levodopa-treated patients. Loens et al. found that only vitamin B6 and homocysteine levels correlated with levodopa dose and concluded that PNP risk increased with higher levodopa dose accumulated by longer disease duration and greater bioavailability due to intestinal administration [45]. Furthermore, PNP has been associated with weight loss, which is a frequent side effect of LCIG therapy [46]. It has been suggested that the viscous enteral gel might hamper intestinal membrane function leading to malabsorption that could result in vitamin B and folate deficiencies further exacerbating PNP severity [17, 47]. Recently, continuous subcutaneous foslevodopa/foscarbidopa infusion has been introduced as an additional treatment option for advanced PD. So far, over the short term of 12 weeks, the incidence of weight loss and PNP was found to be equally low between oral and subcutaneous levodopa application forms (1% weight loss, 3% PNP in both groups) [48]. Only long-term data will answer the question of whether subcutaneous application differs from intestinal application of levodopa in terms of PNP risk. Taken together, these findings point towards a strong relationship between chronic levodopa intake and PNP risk, although the exact causal pathomechanisms and associations with one-carbon metabolites need to be further elaborated. Publications addressing PNP prevalence and associated findings in PD are listed in Table 1.

Diagnosics

According to the American Academy of Neurology (AAN) recommendations, the most accurate diagnosis of distal PNP is obtained from the combination of clinical symptoms, signs, and electrodiagnostic findings [51]. Signs are considered to better predict PNP than symptoms, whereas electrodiagnostic studies would provide a higher level of specificity to PNP diagnosis. Nerve conduction studies in the PD population have used different diagnostic approaches to define PNP. For example, some studies applied the recommended PNP criteria by AAN [6, 34, 42], and other studies solely relied on the electrodiagnostic findings for PNP diagnosis causing a great variability of PNP rates [7, 12, 41]. For better comparability among studies, a standardized implementation of the AAN PNP criteria would be preferable. Yet, electrodiagnostic studies can capture early subclinical nerve alterations particularly useful for longitudinal evaluation of PNP progression that is required to better understand PNP involvement in PD.

Clinical scores

Since PD-specific rating scales only cover a few aspects of neuropathic manifestations and are rather unspecific, common clinical scores used for the evaluation of diabetic neuropathy can be applied for PNP screening in PD patients focusing on large fiber neuropathy [52]. In this review, relevant PNP scoring systems that have been utilized for the investigation of PD-related PNP are introduced. The Neuropathy Symptom Score (NSS) and the Neuropathy Disability Score (NDS) were originally proposed by Dyck et al. to assess the quality and severity of neuropathic symptoms and neurological deficits [53]. These scoring systems have been modified over time to facilitate clinical application [54]. The NSS is a patient questionnaire categorized in symptomatology, localization, exacerbating, and improving circumstances. Symptom severity is classified into mild (3–4 points), moderate (5–6 points) and severe neuropathy (7–10 points). Kühn et al. reported that 86% of PD patients experienced at least one neuropathic symptom in the NSS [7]. NSS also correlated with motor (UPDRS part II and III) and non-motor (NMS-Quest, PDQ-39) PD scores. The NDS is based on the clinical examination of neuropathic signs on both sides including ankle jerk reflex, vibration, pinprick, and temperature sensation. The maximum total score is ten points. The clinical scores NSS and NDS were found to correlate with electrophysiological data in diabetic neuropathy [55]. The Toronto Clinical Scoring System (TCSS) is a neuropathy assessment instrument combining the scoring of all three domains of neuropathic symptoms, reflex status, and sensory testing, with a maximum

Table 1 Overview of PNP status in PD

Authors	Medication	Samples	Examined nerves	PNP prevalence	Associated findings
Toth et al. [25]	Oral LD	500 PD	Sural, peroneal, tibial	9.8% of PD patients	Hcy/MMA↑ Cumulative LD dose and MMA level were associated with PNP severity
Toth et al. [34]	Oral LD	58 PD, 58 controls	Peroneal, tibial, median, ulnar	55% of PD patients vs. 9% of controls	Age↑, UPDRS scores↑, Hcy/MMA ↑ PNP severity correlated with LD exposure and MMA level
Ceravalo et al. [6]	Oral LD	330 PD, 137 controls	Sural, peroneal	19.40% of PD patients with long LD exposure, 6.80% with short LD exposure, 4.82% with no LD exposure vs. 8.76% of controls	LD dose↑, Hcy↑, vitamin B12↓
Shahrzaila et al. [39]	Oral LD vs LD-naïve	51 PD	Sural, tibial, median, ulnar	23% of LD-treated vs. 2.4% of LD-naïve PD patients (↔)	MMA/Hcy ↔
De Araújo et al. [49]	Oral LD	38 PD, 16 PS	Peroneal, tibial, sural	34.2% of PD patients vs. 23.5% of patients with parkinsonism other than PD (↔)	EMG abnormalities correlated with age and Hoehn & Yahr stage
Vanta et al. [38]	Oral LD vs LD-naïve	73 PD	sural, peroneal, tibial	67.3% of LD-treated vs. 4.8% of LD-naïve PD patients	Vitamin B12 level correlated with LD daily dose and duration of treatment, LD dose correlated with sural nerve amplitude
Lee and Baik [41]	LD-naïve	105 de novo PD	Median, ulnar, sural, peroneal	22.8% of PD patients	Hcy↑
Kühn et al. [7]	Oral LD	50 PD	Tibial, median, sural	62% of PD patients	Motor nerve amplitudes correlate with Hoehn & Yahr stage and UPDRS Ill score
Mathukumalli et al. [42]	Oral LD	93 PD, 70 controls	median, ulnar, peroneal, tibial, sural	7.53% of PD patients vs. 4.29% in controls (↔)	Vitamin B12/Hcy ↔ between cases and controls
Ramachandran et al. [35]	Oral LD	154 PD	median, ulnar, peroneal, tibial, sural	18.2% of PD patients	Disease duration and severity, cumulative LD dose and hcy level were associated with PNP
Corrà et al. [12]	Oral LD	99 PD	Sural, tibial, peroneal, ulnar, radial, median	12.5% of PD patients	Presence of PNP was associated with gait and balance parameters
Jugel et al. [46]	Oral PD medication vs. LCG	30 PD	median, tibial, sural, peroneal	66.7% of PD patients with oral PD medication vs. 100% LCG-treated patients	Degree of neuropathic change correlated with weight loss and LD dose in LCG group
Mancini et al. [44]	Oral LD vs. LCG vs. non-LD	150 PD	ulnar, peroneal, tibial, sural	28% of LCG-treated patients vs. 20% with oral LD vs. 6% with non-LD treatment	Vitamin 12↓, folate↓, hcy↑ in LD-treated patients. LD daily dose correlated with Hcy levels
Loens et al. [45]	Oral LD vs. LCG	21 PD	median, peroneal, tibial, sural	100% of LCG-treated patients vs. 72.7% with oral LD	Vitamin B12, folate, hcy ↔ LD dose correlated with vitamin B6 deficiency

Table 1 (continued)

Authors	Medication	Samples	Examined nerves	PNP prevalence	Associated findings
Kwon et al. [50]	Oral LD	41 PD	tibial, sural, peroneal, median, ulnar, radial	65.85% of PD patients	PNP deteriorated in 21.95% of cases. Median sensory nerve was most affected. Sural nerve amplitude correlated with lower quality of life and worse cognition

LD, levodopa; LCIG, levodopa/carbidopa intestinal gel; PS, parkinsonism; Hcy, homocysteine; MMA, methylmalonic acid; UPDRS, Unified Parkinson's Disease Rating Scale; ↑ elevated; ↓ reduced; ↔ not significantly different

score of 19 points [56]. Toth et al. used TCSS to assess PNP severity and found a correlation between PNP severity and PD severity as reflected by the Unified Parkinson's Disease Rating Scale (UPDRS) scores [34].

Electrophysiological diagnostics

Nerve conduction studies (NCS) are a reliable and objective method to evaluate large nerve fiber dysfunction and are therefore regarded as the gold standard for diagnosing and monitoring PNP [57]. Axonal nerve damage, most common for PD-related PNP, is detected by reduced compound muscle action potential (cMAP) in motor nerves or reduced sensory nerve action potential (sNAP). Demyelinating neuropathies have been reported in rare cases of PD patients with acute PNP onset and present with reduced conduction velocities and prolonged distal and F-wave latencies. The simplified NCS protocol of the AAN guidelines recommends the unilateral screening of the sural sensory or peroneal motor NCS [51]. In case of an abnormal finding, NCS should be extended to the ulnar (motor, sensory) and median (sensory) nerve. If a response of a studied nerve is absent, the contralateral nerve can be measured. If no peroneal motor response can be obtained, the ipsilateral tibial nerve can be studied. A nerve conduction abnormality (≥ 99 th or ≤ 1 st percentile) in at least two separate nerves, one of which must be the sural nerve, constitutes the minimal criterion for the electrophysiological diagnosis of a PNP. However, studies deviated from these recommendations since this protocol is time-consuming and requires special expertise leading to discrepant results. Firstly, the range of the examined nerves varied between studies. Some studies only focused on the nerves of the lower limbs neglecting nerve involvement of the upper limbs [6, 40, 49]. In our recently published longitudinal study, we observed a PNP deterioration in 21.95% of PD cases over two years [50]. The median sensory nerve was most frequently affected with the strongest nerve amplitude reduction of 45.0%. Secondly, deviant cut-off reference values were used to determine abnormal NCS. For example, Kühn et al. referred to sural sNAP < 3.6 mV for elderly patients as abnormal [7], whereas Araújo et al. tolerated absent sural responses in patients older than 60 years resulting in lower PNP rate [49]. Furthermore, Kühn et al. increased sensitivity for early PNP detection by choosing the lower value of bilateral conduction. Thirdly, the grading of PNP severity differed between studies. Kühn et al. categorized into mild sensory (reduced sural sNAP only), moderate sensorimotor (additionally reduced tibial cMAP), and severe sensorimotor PNP (additionally reduced median cMAP) resulting in 14 mild, 11 moderate, and 6 severe PD cases. Another study differentiated between 15 mild sensory (reduced sNAP), 8 severe sensory (absent sNAP),

and 5 sensorimotor PNP (reduced sNAP and cMAP) [35]. These findings altogether illustrate the challenges of the inconsistencies of the PNP definition and thus diminished study comparability. An extended electrophysiological status is yet of importance to evaluate PNP severity since previous findings indicate a significant involvement also of motor and upper limb nerve impairment. Several studies have reported an association between the extent of PNP severity and PD severity (UPDRS, Hoehn and Yahr stage) [7, 34, 35]. Kühn et al. found a reduced tibial nerve amplitude in PD patients with higher Hoehn and Yahr stages and compromised UPDRS III score [7]. In the longitudinal analysis, nerve amplitudes correlated with higher motor and also non-motor scores at baseline (PD-related quality of life, cognition scores) [50]. Although no statistically significant correlation could be established between PNP progression and PD progression over the disease course, a significant deterioration of both conditions could be observed. Therefore, large fiber neuropathy can affect PD severity and parallel PD progression as a manifestation of the underlying peripheral neurodegenerative process. Merola et al. suggested PNP as an independent peripheral marker of a severe PD phenotype associated with worse cognitive, axial motor, autonomic, and nonmotor features [58]. Further longitudinal studies of nerve conduction abnormalities are urgently needed to investigate the natural course of PNP in PD. Screening and monitoring of PD patients for PNP comorbidity should be implemented into clinical routine and be particularly considered when PD patients experience clinical worsening. Needle electromyography (EMG) may serve as a supplementary instrument to determine the chronicity of an axonal neuropathic process [57]. The presence of spontaneous muscle fiber activity at rest indicates denervation, whereas reinnervation is assessed by the motor unit action potential on voluntary muscle contraction. Since this technique is laborious and painful for the patient, its indication should be considered carefully only if relevant additional information can be obtained, for example in case of acute PNP onset during LCIG treatment and monitoring such PNP course.

Peripheral nerve imaging

High-resolution nerve ultrasound (HRUS) is a non-invasive modality to visualize morphologic alterations of peripheral nerves [59]. Kühn et al. applied HRUS for the first time to investigate PD-related PNP [7]. In their study, a higher prevalence of enlarged cross-sectional areas was detected in PD patients with PNP compared to those without PD, mostly at typical entrapment sites without clinical or electrophysiological correlate suggesting an increased nerve vulnerability in PD [7]. Magnetic resonance (MR) neurography is an imaging method to

capture the morphological changes and precise location of nerve injury using MR imaging. This method has been already established as a supportive diagnostic tool for chronic inflammatory demyelinating polyradiculoneuropathy [60] but has not been studied for PD-related PNP so far. Allowing a meticulous depiction of nerve alterations, this non-invasive method might be also useful as a tool for evaluating PNP in PD and should be studied in the future.

Nerve biopsy

A nerve biopsy, typically of the sural nerve, is an invasive procedure that leaves the patients with a sensory deficit. It should be considered as a final step in the diagnostic work-up of neuropathies of unknown origin and if an inflammatory or other potentially treatable etiology is suspected [61]. As previously discussed, for research purposes, nerve biopsies are indispensable since they can provide important *in vivo* clinicopathological correlations and clarify the relevance of peripheral α -synuclein deposition that truly helps unravel the pathogenic link between PNP and PD.

Management

Symptomatic approach

Regarding clinical management, symptomatic and causative therapeutic approaches can be distinguished. PNP can cause neuropathic pain that significantly impacts quality of life and therefore requires symptomatic relief. The Neuropathic Pain Special Interest Group proposed gabapentinoids gabapentin and pregabalin, tricyclic antidepressants (TCAs), and serotonin-norepinephrine reuptake inhibitors (SNRI) venlafaxine and duloxetine as first-line drugs for neuropathic pain irrespective of the cause [62]. Gabapentin was found to have positive effects on PD motor symptoms advocating its use for neuropathic pain with additional motor benefits [63, 64]. Furthermore, duloxetine was tested for treatment of central pain in an open-label trial and 13 of 20 PD patients reported varying degrees of pain relief [65]. Despite their effectiveness, TCAs should be used with caution due to the anticholinergic side effects that can cause sedation, deteriorate cognition, or induce psychosis, particularly in the elderly more advanced PD patients. Topical treatment with lidocaine or capsaicin can be considered a second-line option. In severe cases, strong opioids are recommended as third-line drugs. In a double-blind placebo-controlled phase 2 study, treatment with prolonged-release oxycodone-naloxone showed a positive effect on PD-related severe pain [66]. Gait disturbance and postural instability in PD can be aggravated by comorbidity with PNP. Physical exercise with a special focus on balance and gait training has shown great therapeutic

potential to improve proprioception, gait performance, postural control, and prevention of falls in patients with peripheral neuropathy [67], but also in patients with PD [68]. The beneficial effects of exercise intervention for neuropathy were demonstrated by an experimental study of rodents following peripheral nerve injuries, showing increased axon regeneration, muscle reinnervation and increased expression of neurotrophic factors [69]. Therefore, exercise should be implemented as an integral therapeutic part targeting gait and balance disturbances as overlapping symptoms of PD and PNP. The optimal fall prevention protocol for individuals with PD and PNP needs to be determined in further studies.

Causative approach

Since levodopa-induced depletion of methyl group-donating vitamins and cofactors and accumulation of homocysteine and MMA are implied in the pathogenesis of PD-related PNP, treatment approaches are based on vitamin supplementation and use of COMT-inhibitors. In one study, PD patients with idiopathic PNP and patients with idiopathic PNP only identified with abnormalities in cobalamin, homocysteine, or MMA levels and were treated with monthly intramuscular injection of 1000 μ g cobalamin for 1–2 years [25]. In most cases, the abnormal values were normalized upon treatment. Clinical assessment and electrophysiological parameters were stable at follow-up in the PD with PNP subgroup compared to the mild PNP deterioration in the other subgroup. The authors interpreted that one explanation for this finding could be an iatrogenic cause of levodopa-induced vitamin B12 dysmetabolism, alternatively, PNP could represent a peripheral nervous system manifestation of PD. In an open-label study, 30 PD patients were treated on an oral combination supplementation of vitamin B12, folic acid, vitamin B6, and vitamin B2 for 10 days a month since LCIG started and throughout the follow-up [70]. The authors reported a low incidence of PNP (19%) and only mild electrophysiological progression in patients with pre-existing PNP. Ultimately, despite normal vitamin levels and appropriate supplement therapy, the development of PNP cannot be prevented, however, the course of PNP progression could be attenuated.

There are recommendations for PD patients undergoing LCIG therapy [71, 72]. Before starting LCIG, 6 months after, and once a year thereafter, laboratory assessments should be performed including vitamins B12, B6, and folic acid. Since 50% of cobalamin-deficient patients present with normal serum cobalamin, determination of homocysteine and MMA levels is suggested (the latter is more specific as an early marker of functional vitamin B12 deficiency). NCS status should be evaluated at baseline and once yearly. For high-risk

patients (pre-existing neuropathy, advanced PD) prophylactic vitamin B12 supplementation can be considered. If laboratory abnormalities or PNP diagnosis are present, supplementation therapy should be initiated. A common regimen for vitamin B12 would start with 1000 µg intramuscularly for 5–7 days followed by a monthly 1000 µg intramuscular injection. Folate can be administered orally at the dosage of 5 mg daily. Vitamin B6 supplementation is not recommended because it interferes with the actions of the decarboxylase inhibitor and has neurotoxic properties in excess levels [32].

Methylation of levodopa by the enzyme COMT represents a critical initial step for vitamin B12 depletion and homocysteine accumulation. Therefore, COMT inhibition could have a protective effect on restoring these metabolic imbalances and thus preventing the development of PNP in PD patients. In a multicenter study of 197 PD patients, a significantly lower PNP prevalence was observed in PD patients co-administered with the COMT-inhibitor entacapone for at least 18 months (5.7%) compared to those without COMT-inhibitor under levodopa exposure for at least 3 years (19.4%) [73]. Higher serum vitamin B12 levels and

lower homocysteine levels were measured in patients on COMT inhibitors. In contrast, a study of a heterogenous PD cohort undergoing different therapeutic regimens did not find a difference in PNP prevalence, vitamin B12, and homocysteine levels in patients on COMT-inhibitor [44]. However, the latter study did not specify the treatment duration of levodopa or COMT-inhibitor, which could be relevant to the possible changes in the PNP status. Higher levodopa bioavailability is achieved by the COMT-inhibitor opicapone compared to entacapone, whereas both compounds could prevent an increase in homocysteine [74]. Levodopa-entacapone-carbidopa intestinal gel (LECIG) is a novel infusion therapy option for advanced PD containing the COMT-inhibitor entacapone in addition to levodopa and carbidopa. Notably, a single-center study of 30 PD patients reported no clinically diagnosed cases of PNP and no increase in homocysteine level during the first 6 months of LECIG treatment [75]. However, the authors conceded that many patients used over-the-counter vitamin supplements, and no routine NCS was performed. Figure 1 summarizes diagnostic and therapeutic options for PNP in PD.

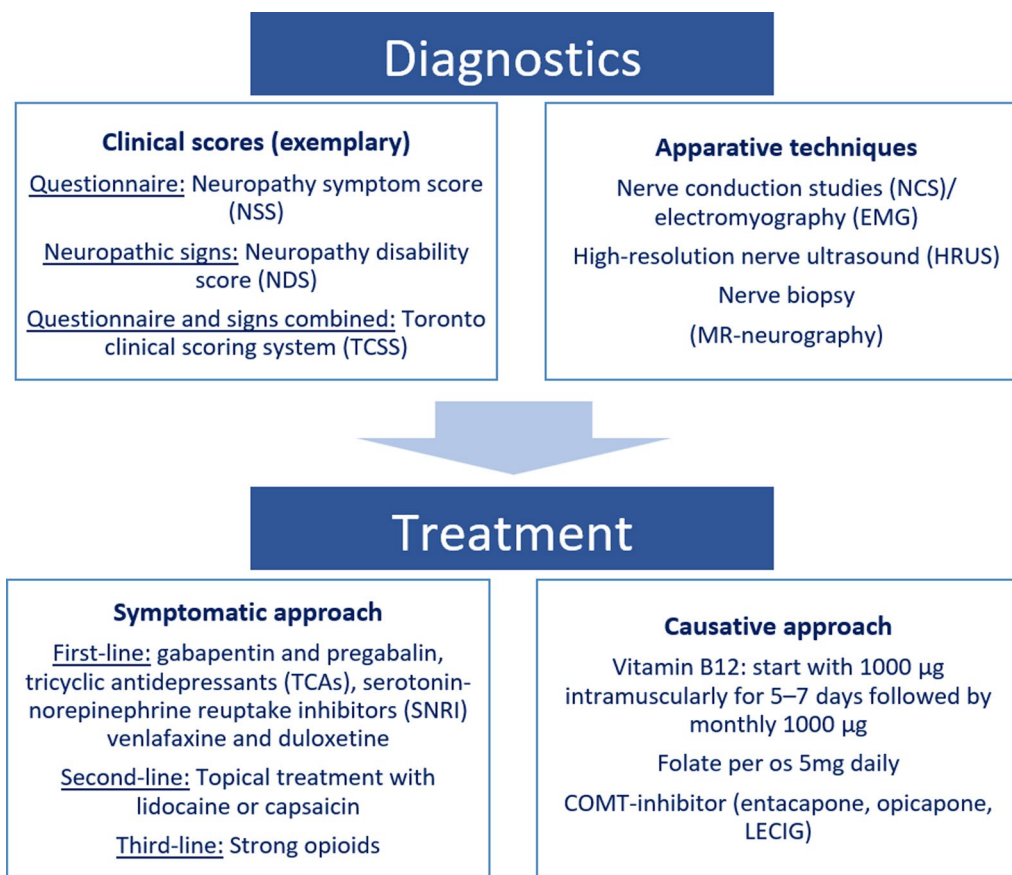


Fig. 1 Diagnostic and therapeutic options for PNP in PD

Conclusions

The higher PNP prevalence in PD indicates a pathophysiological relationship between these distinct conditions probably as a result of a complex interplay between genetic predisposition, extrinsic risk factors influencing peripheral nerve homeostasis, and an intrinsic peripheral neurodegenerative process. Evidence suggests that the cumulative exposure to levodopa is critical for PNP risk. More studies on dopa-naïve PD patients are required to unmask a truly “idiopathic” form of PNP. To further elucidate pathomechanisms, neuropathologic correlations are mandatory despite invasiveness. The pathogenic role of α -synuclein deposits in peripheral nerves needs to be clarified. PNP should be recognized as an important aspect of PD and therefore adequately detected and monitored by comprehensive NCS and clinical scores. Large-scale prospective studies will help understand PNP progression in PD and validate biomarker potential, stratify risk factors for developing PD, and evaluate the protective role of vitamin supplementation and use of COMT inhibitors. The long-term results of the novel PD treatment options such as subcutaneous levodopa infusion therapy and LECIG remain to be seen.

Abbreviations

AAN	American Academy of Neurology
COMT	Catechol-O-methyltransferase
CMAP	Compound muscle action potential
EMG	Electromyography
HRUS	High-resolution ultrasound
LCIG	Levodopa/carbidopa intestinal gel
LECIG	Levodopa-entacapone-carbidopa intestinal gel
MMA	Methylmalonic acid
MR	Magnetic resonance
MTHFR	Methylenetetrahydrofolate reductase
NCS	Nerve conduction studies
NDS	Neuropathy Disability Score
NSS	Neuropathy Symptom Score
PD	Parkinson's disease
PNP	Polyneuropathy
SAH	S-adenosylhomocysteine
SAM	S-adenosylmethionine
SNAP	Sensory nerve action potential
SNRI	Serotonin-norepinephrine reuptake inhibitor
TCA	Tricyclic antidepressant
TCSS	Toronto Clinical Scoring System
UPDRS	Unified Parkinson's Disease Rating Scale

Acknowledgements

Not applicable.

Author contributions

EK was involved in the conception and design of the work and analysis and interpretation of data and has drafted the work. JS contributed to the conception of the work. RS contributed to the conception of the work and substantively revised it. RG and KP were involved in the critical revision of the manuscript. LT was involved in study concept and design, analysis and interpretation, and critical revision of the manuscript. Each author has approved the submitted version and agreed both to be personally accountable for the author's own contributions and to ensure that questions related to the accuracy or integrity of any part of the work, even ones in which the author was not personally involved, are appropriately investigated, resolved, and the resolution documented in the literature.

Funding

Open Access funding enabled and organized by Projekt DEAL. This research received no specific grant from any funding agency in the public, commercial, or not-for-profit sectors.

Availability of data and materials

All data generated or analysed during this study are included in this published article.

Declarations

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

Received: 12 August 2024 Accepted: 9 October 2024

Published online: 28 October 2024

References

- Dorsey, E. R., Sherer, T., Okun, M. S., & Bloem, B. R. (2018). The emerging evidence of the parkinson pandemic. *Journal of Parkinson's Disease*, 8(Suppl 1), S3-8.
- Steinmetz, J. D., Seeher, K. M., Schiess, N., Nichols, E., Cao, B., Servili, C., Cavallera, V., Cousin, E., Hagins, H., Moberg, M. E., & Mehlman, M. L. (2024). Global, regional, and national burden of disorders affecting the nervous system, 1990–2021: A systematic analysis for the global burden of disease study 2021. *The Lancet Neurology*, 23(4), 344–381.
- Beach, T. G., Adler, C. H., Sue, L. I., Vedders, L., Lue, L., White, C. L., Akiyama, H., Caviness, J. N., Shill, H. A., Sabbagh, M. N., & Walker, D. G. (2010). Multi-organ distribution of phosphorylated α -synuclein histopathology in subjects with Lewy body disorders. *Acta neuropathologica*, 119(6), 689–702.
- Misra, U. K., Kalita, J., & Nair, P. P. (2008). Diagnostic approach to peripheral neuropathy. *Annals of Indian Academy of Neurology*, 11(2), 89–97.
- Bulling, M. T., Wing, L. M., & Burns, R. J. (1991). Controlled release levodopa/carbidopa (Sinemet CR4) in Parkinson's disease—an open evaluation of efficacy and safety. *Australian and New Zealand Journal of Medicine*, 21(4), 397–400.
- Ceravolo, R., Cossu, G., Di Bandettini Poggio, M., Santoro, L., Barone, P., Zibetti, M., Frosini, D., Nicoletti, V., Manganelli, F., Iodice, R., Picillo, M., Merola, A., Lopiano, L., Paribello, A., Manca, D., Melis, M., Marchese, R., Borelli, P., Mereu, A., ... Bonuccelli, U. (2013). Neuropathy and levodopa in Parkinson's disease: Evidence from a multicenter study. *Movement Disorders: Official Journal of the Movement Disorder Society*, 28(10), 1391–1397.
- Kühn, E., Averdunk, P., Huckemann, S., Müller, K., Biesalski, A.-S., Zum Hof Berge, F., Motte, J., Fisse, A. L., Schneider-Gold, C., Gold, R., Pitarokoili, K., & Tönges, L. (2020). Correlates of polyneuropathy in Parkinson's disease. *Annals of Clinical and Translational Neurology*, 7(10), 1898–1907.
- Zis, P., Grünewald, R. A., Chaudhuri, R. K., & Hadjivassiliou, M. (2017). Peripheral neuropathy in idiopathic Parkinson's disease: A systematic review. *Journal of the Neurological Sciences*, 378, 204–209.
- Comi, C., Magistrelli, L., Oggioni, G. D., Carecchio, M., Fleetwood, T., Cantello, R., Mancini, F., & Antonini, A. (2014). Peripheral nervous system involvement in Parkinson's disease: Evidence and controversies. *Parkinsonism & Related Disorders*, 20(12), 1329–1334.
- Mylius, V., Ciampi, D., de Andrade, R. G., Cury, M. T., Ehr, U., Eggert, K. M., Beer, S., Kesselring, J., Stamelou, M., Oertel, W. H., Möller, J. C., & Lefaucheur, J.-P. (2015). Pain in Parkinson's disease: Current concepts and a new diagnostic algorithm. *Movement Disorders Clinical Practice*, 2(4), 357–364.

11. Beaulieu, M. L., Müller, M. L. T. M., & Bohnen, N. I. (2018). Peripheral neuropathy is associated with more frequent falls in Parkinson's disease. *Parkinsonism & Related Disorders*, *54*, 46–50.
12. Corrà, M. F., Vila-Chã, N., Sardoeira, A., Hansen, C., Sousa, A. P., Reis, I., Sambayeta, F., Damásio, J., Calejo, M., Schicketmueller, A., Laranjinha, I., Salgado, P., Taipa, R., Magalhães, R., Correia, M., Maetzler, W., & Maia, L. F. (2023). Peripheral neuropathy in Parkinson's disease: Prevalence and functional impact on gait and balance. *Brain : A Journal of Neurology*, *146*(1), 225–236.
13. Doppler, K., Ebert, S., Üçeyler, N., Trenkwalder, C., Ebntheuer, J., Volkmann, J., & Sommer, C. (2014). Cutaneous neuropathy in Parkinson's disease: A window into brain pathology. *Acta neuropathologica*, *128*(1), 99–109.
14. Nolano, M., Provitera, V., Manganelli, F., Iodice, R., Stancanelli, A., Caporaso, G., Saltalamacchia, A., Califano, F., Lanzillo, B., Picillo, M., Barone, P., & Santoro, L. (2017). Loss of cutaneous large and small fibers in naive and l-dopa-treated PD patients. *Neurology*, *89*(8), 776–784.
15. Podgorny, P. J., Suchowersky, O., Romanchuk, K. G., & Feasby, T. E. (2016). Evidence for small fiber neuropathy in early Parkinson's disease. *Parkinsonism & Related Disorders*, *28*, 94–99.
16. Vacchi, E., Senese, C., Chiaro, G., Disanto, G., Pinton, S., Morandi, S., Bertaina, I., Bianco, G., Staedler, C., Galati, S., Gobbi, C., Kaelin-Lang, A., & Melli, G. (2021). Alpha-synuclein oligomers and small nerve fiber pathology in skin are potential biomarkers of Parkinson's disease. *NPJ Parkinson's Disease*, *7*(1), 119.
17. Urban, P. P., Wellach, I., Faiss, S., Layer, P., Rosenkranz, T., Knop, K., & Weis, J. (2010). Subacute axonal neuropathy in Parkinson's disease with cobalamin and vitamin B6 deficiency under duodopa therapy. *Movement disorders: Official journal of the Movement Disorder Society*, *25*(11), 1748–1752.
18. Kimber, T., Blumbergs, P., & Thompson, P. (2013). Severe ataxic polyneuropathy associated with chronic levodopa use in Parkinson's disease. *Parkinsonism & Related Disorders*, *19*(9), 847–849.
19. Vital, A., Meissner, W. G., Canron, M.-H., Martin-Negrier, M.-L., Bezard, E., Tison, F., & Vital, C. (2014). Intra-axonal protein aggregation in the peripheral nervous system. *Journal of the Peripheral Nervous System : JPNS*, *19*(1), 44–49.
20. Zhang, H., Zhu, L., Sun, L., Zhi, Y., Ding, J., Yuan, Y.-S., Shen, F.-F., Li, X., Ji, P., Wang, Z., Niu, Q., & Zhang, K.-Z. (2019). Phosphorylated α -synuclein deposits in sural nerve deriving from Schwann cells: A biomarker for Parkinson's disease. *Parkinsonism & Related Disorders*, *60*, 57–63.
21. Zhang, H., Wu, J., Shen, F.-F., Yuan, Y.-S., Li, X., Ji, P., Zhu, L., Sun, L., Ding, J., Niu, Q., & Zhang, K.-Z. (2019). Activated Schwann cells and increased inflammatory cytokines IL-1 β , IL-6, and TNF- α in patients' sural nerve are lack of tight relationship with specific sensory disturbances in Parkinson's disease. *CNS Neuroscience & Therapeutics*, *26*(5), 518–526.
22. Rong, Z., Shen, F., Wang, Y., Sun, L., Wu, J., Zhang, H., Yuan, Y., Jiang, W., Li, X., Ji, P., & Zhang, K. (2021). Phosphorylated α -synuclein and phosphorylated tau-protein in sural nerves may contribute to differentiate Parkinson's disease from multiple system atrophy and progressive supranuclear paralysis. *Neuroscience Letters*, *756*, 135964.
23. Rajabally, Y. A., & Martey, J. (2013). Levodopa, vitamins, ageing and the neuropathy of Parkinson's disease. *Journal of Neurology*, *260*(11), 2844–2848.
24. Hanewinkel, R., van Oijen, M., Ikram, M. A., & van Doorn, P. A. (2015). The epidemiology and risk factors of chronic polyneuropathy. *European Journal of Epidemiology*, *31*, 5–20.
25. Toth, C., Brown, M. S., Furtado, S., Suchowersky, O., & Zochodne, D. (2008). Neuropathy as a potential complication of levodopa use in Parkinson's disease. *Movement Disorders: Official Journal of the Movement Disorder Society*, *23*(13), 1850–1859.
26. Greten, S., Müller-Funogea, J. I., Wegner, F., Höglinger, G. U., Simon, N., Junius-Walker, U., Gerbel, S., Krause, O., & Klietz, M. (2021). Drug safety profiles in geriatric patients with Parkinson's disease using the FORTA (Fit FOR The Aged) classification: results from a mono-centric retrospective analysis. *Journal of Neural Transmission*, *128*(1), 49–60.
27. Klietz, M., Greten, S., Wegner, F., & Höglinger, G. U. (2019). Safety and tolerability of pharmacotherapies for Parkinson's disease in geriatric patients. *Drugs & Aging*, *36*(6), 511–530.
28. Labandeira, C. M., Fraga-Bau, A., Arias Ron, D., Alvarez-Rodriguez, E., Vicente-Alba, P., Lago-Garma, J., & Rodriguez-Perez, A. I. (2022). Parkinson's disease and diabetes mellitus: Common mechanisms and treatment repurposing. *Neural Regeneration Research*, *17*(8), 1652–1658.
29. Vital, A., Lepreux, S., & Vital, C. (2014). Peripheral neuropathy and parkinsonism: A large clinical and pathogenic spectrum. *Journal of the Peripheral Nervous System: JPNS*, *19*(4), 333–342.
30. Ohsawa, Y., Kurokawa, K., Sonoo, M., Yamada, H., Hemmi, S., Iwatsuki, K., Hagiwara, H., Murakami, T., Shirabe, T., Shimizu, T., & Sunada, Y. (2005). Reduced amplitude of the sural nerve sensory action potential in PARK2 patients. *Neurology*, *65*(3), 459–462.
31. Abbruzzese, G., Pigullo, S., Schenone, A., Bellone, E., Marchese, R., Di Maria, E., Benedetti, L., Ciotti, P., Nobbio, L., Bonifati, V., Ajmar, F., & Mandich, P. (2004). Does parkin play a role in the peripheral nervous system? A family report. *Movement Disorders: Official Journal of the Movement Disorder Society*, *19*(8), 978–981.
32. Romagnolo, A., Merola, A., Artusi, C. A., Rizzone, M. G., Zibetti, M., & Lopiano, L. (2019). Levodopa-induced neuropathy: A systematic review. *Movement Disorders Clinical Practice*, *6*(2), 96–103.
33. Dulski, J., Uitti, R. J., Ross, O. A., & Wszolek, Z. K. (2022). Genetic architecture of Parkinson's disease subtypes – Review of the literature. *Frontiers in Aging Neuroscience*, *14*, 1023574.
34. Toth, C., Breithaupt, K., Ge, S., Duan, Y., Terris, J. M., Thiessen, A., Wiebe, S., Zochodne, D. W., & Suchowersky, O. (2010). Levodopa, methylmalonic acid, and neuropathy in idiopathic Parkinson disease. *Annals of Neurology*, *68*(1), 28–36.
35. Ramachandran, A., Jose, J., Gafoor, V. A., Das, S., & Balaram, N. (2022). Prevalence and risk factors of peripheral neuropathy in Parkinson's disease. *Annals of Indian Academy of Neurology*, *25*(6), 1109–1115.
36. Obeid, R., & Herrmann, W. (2006). Mechanisms of homocysteine neurotoxicity in neurodegenerative diseases with special reference to dementia. *FEBS Letters*, *580*(13), 2994–3005.
37. Baltrusch, S. (2021). The role of neurotropic B vitamins in nerve regeneration. *BioMed Research International*, *2021*, 9968228.
38. Vanta, O. M., Tohanean, N., Pintea, S., & Perju-Dumbrava, L. (2019). Large-fiber neuropathy in Parkinson's disease: Clinical, biological, and electroneurographic assessment of a Romanian cohort. *Journal of Clinical Medicine*, *8*(10), 1533.
39. Shahrazaila, N., Mahamad, U. A., Yap, A.-C., Choo, Y.-M., Marras, C., & Lim, S.-Y. (2013). Is chronic levodopa therapy associated with distal symmetric polyneuropathy in Parkinson's disease? *Parkinsonism & Related Disorders*, *19*(3), 391–393.
40. Müller, T., Renger, K., & Kuhn, W. (2004). Levodopa-associated increase of homocysteine levels and sural axonal neurodegeneration. *Archives of Neurology*, *61*(5), 657–660.
41. Lee, J. J., & Baik, J. S. (2020). Peripheral Neuropathy in de novo Patients with Parkinson's disease. *Yonsei Medical Journal*, *61*(12), 1050–1053.
42. Mathukumalli, N. L., Kandadai, M. R., Shaik, J. A., Kanikannan, M. A., & Borgohain, R. (2020). Serum B12, homocysteine levels, and their effect on peripheral neuropathy in Parkinson's disease: Indian cohort. *Annals of Indian Academy of Neurology*, *23*(1), 48–53.
43. Merola, A., Romagnolo, A., Zibetti, M., Bernardini, A., Cocito, D., & Lopiano, L. (2016). Peripheral neuropathy associated with levodopa-carbidopa intestinal infusion: A long-term prospective assessment. *European Journal of Neurology*, *23*(3), 501–509.
44. Mancini, F., Comi, C., Oggioni, G. D., Pacchetti, C., Calandrella, D., Coletti Moja, M., Riboldazzi, G., Tunesi, S., Dal Fante, M., Manfredi, L., Lacerenza, M., Cantello, R., & Antonini, A. (2014). Prevalence and features of peripheral neuropathy in Parkinson's disease patients under different therapeutic regimens. *Parkinsonism & Related Disorders*, *20*(1), 27–31.
45. Loens, S., Chorbazdzheva, E., Kleimann, A., Dressler, D., & Schrader, C. (2017). Effects of levodopa/carbidopa intestinal gel versus oral levodopa/carbidopa on B vitamin levels and neuropathy. *Brain and Behavior*, *7*(5), 698.
46. Jugel, C., Ehlen, F., Taskin, B., Marzinzik, F., Müller, T., & Klostermann, F. (2013). Neuropathy in Parkinson's disease patients with intestinal levodopa infusion versus oral drugs. *PLoS ONE*, *8*(6), e66639.
47. Klostermann, F., Jugel, C., Müller, T., & Marzinzik, F. (2012). Malnutritional neuropathy under intestinal levodopa infusion. *Journal of Neural Transmission*, *119*(3), 369–372.
48. Soileau, M. J., Aldred, J., Budur, K., Fisseha, N., Fung, V. S., Jeong, A., Kimber, T. E., Klos, K., Litvan, I., O'Neill, D., Robieson, W. Z., Spindler, M. A., Standaert, D. G., Talapala, S., Vaou, E. O., Zheng, H., Facheris, M. F., & Hauser, R. A.

- (2022). Safety and efficacy of continuous subcutaneous foslevodopa-foscarbidopa in patients with advanced Parkinson's disease: A randomised, double-blind, active-controlled, phase 3 trial. *The Lancet. Neurology*, 21(12), 1099–1109.
49. de Araújo, D. F., de Melo Neto, A. P., Oliveira, Í. S. C., Brito, B. S., de Araújo, I. T., Barros, I. S., Lima, J. W. O., Horta, W. G., & Gondim, F. A. A. (2016). Small (autonomic) and large fiber neuropathy in Parkinson disease and parkinsonism. *BMC Neurology*, 16, 3. <https://doi.org/10.1186/s12883-016-0667-3>
 50. Kwon, E. H., Bieber, A., Schülken, P., Müller, K., Kühn, E., Averdunk, P., Kools, S., Hilker, L., Kirchgässler, A., Ebner, L., Ortmann, L., Basner, L., Steininger, J., Kleinz, T., Motte, J., Fisse, A. L., Schneider-Gold, C., Gold, R., Scherbaum, R., ... Pitarokouli, K. (2024). Longitudinal evaluation of polyneuropathy in Parkinson's disease. *Journal of Neurology*, 271, 6136.
 51. England, J. D., Gronseth, G. S., Franklin, G., Miller, R. G., Asbury, A. K., Carter, G. T., Cohen, J. A., Fisher, M. A., Howard, J. F., Kinsella, L. J., Latov, N., Lewis, R. A., Low, P. A., & Sumner, A. J. (2005). Distal symmetric polyneuropathy: A definition for clinical research: Report of the American academy of neurology, the American association of electrodiagnostic medicine, and the American academy of physical medicine and rehabilitation. *Neurology*, 64(2), 199–207.
 52. Zhirong Yang, R., Chen, Y. Z., Huang, Y., Hong, T., Sun, F., Ji, L., & Zhan, S. (2018). Scoring systems to screen for diabetic peripheral neuropathy. *Cochrane Database of Systematic Reviews*. <https://doi.org/10.1002/14651858.CD010974.pub2>
 53. Dyck, P. J., Sherman, W. R., Hallcher, L. M., Service, F. J., O'BrienGrina, P. C. L. A., Palumbo, P. J., & Swanson, C. J. (1980). Human diabetic endoneurial sorbitol, fructose, and myo-inositol related to sural nerve morphometry. *Annals of Neurology*, 8(6), 590–596.
 54. Young, M. J., Boulton, A. J., MacLeod, A. F., Williams, D. R., & Sonksen, P. H. (1993). A multicentre study of the prevalence of diabetic peripheral neuropathy in the United Kingdom hospital clinic population. *Diabetologia*, 36(2), 150–154.
 55. Feki, I., & Lefaucheur, J. P. (2001). Correlation between nerve conduction studies and clinical scores in diabetic neuropathy. *Muscle & nerve*, 24(4), 555–558.
 56. Bril, V., & Perkins, B. A. (2002). Validation of the Toronto Clinical Scoring System for diabetic polyneuropathy. *Diabetes Care*, 25(11), 2048–2052.
 57. Chung, T., Prasad, K., & Lloyd, T. E. (2013). Peripheral neuropathy – clinical and electrophysiological considerations. *Neuroimaging Clinics of North America*, 24(1), 49–65.
 58. Merola, A., Rosso, M., Romagnolo, A., Comi, C., Fasano, A., Zibetti, M., Lopez-Castellanos, J. R., Cocito, D., Lopiano, L., & Espay, A. J. (2017). Peripheral neuropathy as marker of severe Parkinson's disease phenotype. *Movement Disorders: Official Journal of the Movement Disorder Society*, 32(8), 1256–1258.
 59. Telleman, J. A., Grimm, A., Goedee, S., Visser, L. H., & Zaidman, C. M. (2018). Nerve ultrasound in polyneuropathies. *Muscle & Nerve*, 57(5), 716–728.
 60. van den Bergh, P. Y. K., van Doorn, P. A., Hadden, R. D. M., Avau, B., Vankrunkelsven, P., Allen, J. A., Attarian, S., Blomkwist-Markens, P. H., Cornblath, D. R., Eftimov, F., Goedee, H. S., Harbo, T., Kuwabara, S., Lewis, R. A., Lunn, M. P., Nobile-Orazio, E., Querol, L., Rajabally, Y. A., Sommer, C., & Topaloglu, H. A. (2021). European academy of neurology/peripheral nerve society guideline on diagnosis and treatment of chronic inflammatory demyelinating polyradiculoneuropathy: Report of a joint task force-second revision. *European Journal of Neurology*, 28(11), 3556–3583.
 61. Sommer, C. L., Brandner, S., Dyck, P. J., Harati, Y., LaCroix, C., Lammens, M., Magy, L., Mellgren, S. I., Morbin, M., Navarro, C., Powell, H. C., Schenone, A. E., Tan, E., Urtizberea, A., & Weis, J. (2010). Peripheral nerve society guideline on processing and evaluation of nerve biopsies. *Journal of the Peripheral Nervous System: JPNS*, 15(3), 164–175.
 62. Finnerup, N. B., Attal, N., Haroutounian, S., McNicol, E., Baron, R., Dworkin, R. H., Gilron, I., Haanpää, M., Hansson, P., Jensen, T. S., Kamerman, P. R., Lund, K., Moore, A., Raja, S. N., Rice, A. S. C., Rowbotham, M., Sena, E., Siddall, P., Smith, B. H., & Wallace, M. (2015). Pharmacotherapy for neuropathic pain in adults: A systematic review and meta-analysis. *The Lancet. Neurology*, 14(2), 162–173.
 63. Olson, W. L., Gruenthal, M., Mueller, M. E., & Olson, W. H. (1997). Gabapentin for parkinsonism: A double-blind, placebo-controlled, crossover trial. *The American Journal of Medicine*, 102(1), 60–66.
 64. van Blercom, N., Lasa, A., Verger, K., Masramón, X., Sastre, V. M., & Linzasoro, G. (2004). Effects of gabapentin on the motor response to levodopa: A double-blind, placebo-controlled, crossover study in patients with complicated Parkinson disease. *Clinical Neuropharmacology*, 27(3), 124–128.
 65. Djaldetti, R., Yust-Katz, S., Kolianov, V., Melamed, E., & Dabby, R. (2007). The effect of duloxetine on primary pain symptoms in Parkinson disease. *Clinical Neuropharmacology*, 30(4), 201–205.
 66. Trenkwalder, C., Chaudhuri, K. R., Martinez-Martin, P., Rascol, O., Ehret, R., Vališ, M., Sători, M., Krygowska-Wajs, A., Marti, M. J., Reimer, K., Oksche, A., Lomax, M., DeCesare, J., & Hopp, M. (2015). Prolonged-release oxycodone-naloxone for treatment of severe pain in patients with Parkinson's disease (PANDA): A double-blind, randomised, placebo-controlled trial. *The Lancet. Neurology*, 14(12), 1161–1170.
 67. Streckmann, F., Balke, M., Cavaletti, G., Toscanelli, A., Bloch, W., Décard, B. F., Lehmann, H. C., & Faude, O. (2022). Exercise and neuropathy: Systematic review with meta-analysis. *Sports Medicine*, 52(5), 1043–1065.
 68. Shen, X., Wong-Yu, I. S. K., & Mak, M. K. Y. (2016). Effects of exercise on falls, balance, and gait ability in Parkinson's disease: A meta-analysis. *Neurorehabilitation and neural repair*, 30(6), 512–527.
 69. Maugeri, G., D'Agata, V., Trovato, B., Roggio, F., Castorina, A., Vecchio, M., Di Rosa, M., & Musumeci, G. (2021). The role of exercise on peripheral nerve regeneration: from animal model to clinical application. *Heliyon*, 7(11), 1.
 70. Rispoli, V., Simioni, V., Capone, J. G., Goffrè Andreasi, N., Preda, F., Sette, E., Tugnoli, V., & Sensi, M. (2017). Peripheral neuropathy in 30 duodopa patients with vitamins B supplementation. *Acta neurologica Scandinavica*, 136(6), 660–667.
 71. Uncini, A., Eleopra, R., & Onofrij, M. (2015). Polyneuropathy associated with duodenal infusion of levodopa in Parkinson's disease: Features, pathogenesis and management. *Journal of neurology, neurosurgery, and psychiatry*, 86(5), 490–495.
 72. Taher, J., Naranian, T., Poon, Y.-Y., Merola, A., Mestre, T., Suchowersky, O., Kulasingam, V., & Fasano, A. (2022). Vitamins and infusion of levodopa-carbidopa intestinal gel. *Le Journal Canadien Des Sciences Neurologiques*, 49(1), 19–28.
 73. Cossu, G., Ceravolo, R., Zibetti, M., Arca, R., Ricchi, V., Paribello, A., Murgia, D., Merola, A., Romagnolo, A., Nicoletti, V., Palermo, G., Mereu, A., Lopiano, L., Melis, M., Abbruzzese, G., & Bonuccelli, U. (2016). Levodopa and neuropathy risk in patients with Parkinson disease: Effect of COMT inhibition. *Parkinsonism & related disorders*, 27, 81–84.
 74. Müller, T., Schlegel, E., Zingler, S., & Thiede, H. M. (2022). Effects of one-day application of levodopa/carbidopa/Entacapone versus levodopa/carbidopa/Opicapone in Parkinson's disease patients. *Cells*, 11(9), 1511.
 75. Viljajarju, V., Mertsalmi, T., Pauls, K. A. M., Koivu, M., Eerola-Rautio, J., Udd, M., & Pekkonen, E. (2024). Levodopa-Entacapone-carbidopa intestinal gel treatment in advanced Parkinson's disease: A single-center study of 30 patients. *Movement Disorders Clinical Practice*, 11(2), 159–165.

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