# scientific reports

### OPEN



## Mathematical modeling predicts that endemics by generalist insects are eradicated if nearly all plants produce constitutive defense

Suman Chakraborty<sup>1,2</sup>, Shalu Dwivedi<sup>1</sup> & Stefan Schuster<sup>1⊠</sup>

Plants with constitutive defense chemicals exist widely in nature. The phenomenon is backed by abundant data from plant chemical ecology. Sufficient data are also available to conclude that plant defenses act as deterrent and repellent to attacking herbivores, particularly deleterious generalist insects. In the wild, generalist species are usually not endemic, meaning they are not restricted to certain plant species in a region. Therefore, our objective is to inspect theoretically whether evolution of chemical defenses in all plant species eradicate an endemic by any generalist species. The objective is addressed by developing deterministic ordinary differential equations under the following conditions: Plants without constitutive defenses are susceptible to oviposition by generalist insects, while they become defended against generalists by storing chemical defenses. From the models, we explicitly obtain that a generalist-free stable state is only possible if the vast majority of all plant individuals have chemical defenses. The model also allows one to predict the highest possible percentage of undefended plant individuals, which may be considered as free-riders.

**Keywords** Susceptible plants, Constitutive defense, Mathematical modeling, Generalist insects, Endemic, Non-endemic, Stability analysis, Plant-herbivore interactions

Plants store chemical defenses constitutively during their normal course of development<sup>1–3</sup>. Ample evidences of such defenses are available in nature<sup>2,3</sup>, as reviewed in the subsection 'Examples of plant defenses' below. Crucial roles of these constitutive defense compounds are to deter and repel deleterious generalist insects<sup>4–8</sup>. Several experimental studies tested the effects of plant defense. There is a slight difference in the meaning of plant deterrent and repellent chemicals. A plant deterrent is a chemical that prevents feeding and oviposition by insects, while a plant repellent is a chemical that causes insects to move away from its source<sup>5,9,10</sup>. If plants do not evolve constitutive defenses, then they are susceptible to generalist insects for oviposition. The phenomenon is expressed by Fig. 1.

According to some estimations, more than 100,000 different secondary compounds are found in plants<sup>11–13</sup>. In chemical ecology, a plausible theory could be that all individuals of every plant species have constitutive chemical defenses, even plants edible by humans<sup>3,5,11,14</sup>. Edible plants do involve some defense chemicals, which can, however, be tolerated due to their low concentrations and are often appreciated as flavors, like in black and white mustard or the numerous varieties of cabbage.

Although the theory that all plants have chemical defenses is confirmed by plentiful experimental evidence<sup>3</sup>, it has not been mathematically assessed yet. So, in this study, a proof of this theory is sought by building deterministic models, based on autonomous ordinary differential equations (ODEs). Specifically, the research question is whether any generalist insect species can become endemic if nearly all plant individuals (of all species) store chemical defenses. To be noted, generalist insects have a high diet breadth, but they do not feed permanently on specific plant species of a particular region<sup>15–18</sup>, which makes them non-endemic in the wild. This ability of being non-endemic or using multiple resources could have ecological and evolutionary advantage to generalists. Previous studies explained that generalists increase their fitness through access to various host plant species<sup>18–21</sup>. Although models have been proposed to understand the role of volatile and non-volatile organic compounds in plant-herbivore interactions<sup>22–25</sup>, it is not exactly clear how the non-endemic behavior of generalists is correlated to the multifaceted (ecological) roles of plant toxins.

<sup>1</sup>Department of Bioinformatics, Matthias Schleiden Institute, Friedrich Schiller University Jena, Ernst-Abbe-Pl. 2, Jena 07743, Thuringia, Germany. <sup>2</sup>International Max Planck Research School 'Chemical Communication in Ecological Systems', Jena 07745, Thuringia, Germany. <sup>Se</sup>email: stefan.schu@uni-jena.de



Fig. 1. Plant defense vs generalist insects. Pictures drawn in "AutoDraw" (Google Creative Lab).



Fig. 2. Basic scheme of the susceptible-exploited-insects model.

The main contributions of this study are two models and conclusions derived from them:

- 1. In the first model, a hypothetical situation is considered, where plants do not have constitutive defense compounds. This is done to investigate whether plants can avoid a generalist endemic without evolving chemical defenses. As depicted in Fig. 1, generalists oviposit on plants if defense compounds are absent. The undefended plants (of any species) are distinguished into two sections: Susceptible plants and exploited plants. Susceptible plants are likely to be exploited through oviposition by generalist insects, whereas exploited plants already have insect eggs (singly or in clutches) laid on them. The growth of the insect population is directly determined by the mean number of laid eggs. The flux across the compartments is expressed by the flowchart shown in Fig. 2. This model is influenced by the classic SIR model in epidemiology<sup>26–29</sup>.
- 2. The second model is based on the first one and includes, in addition, the phenomenon that new plants can germinate with chemical defenses and, thus, become defended against generalists. Defended plants are not susceptible to generalists, as expressed in Fig. 1. To formulate this model, the technique used for SIR models with vaccination (SIRV) or immunization<sup>27-30</sup> is applied, where the total plant population includes the susceptible, exploited and defended plants. In case of plants, immunization could be the process of evolving chemical defenses to become defended against generalist herbivores<sup>13,31</sup>. From this model, one can investigate what percentage of plants have to germinate with chemical defenses to avoid a generalist endemic. That would theoretically explain the widespread occurrence of constitutive defense in plants<sup>32-36</sup>.

The paper structure is as follows: Models are built in two subsections of the "Models" section. Next, simulated figures are provided in the Illustration of results section. Thereafter, the results of the models and future perspectives are expressed in the Discussion. Finally, the Conclusion is devoted to the main message of this study, model limitations and possible application domains. The required calculations for the stability analysis of the models are given separately in the Supplementary Information.

#### Examples of plant defenses

Some examples of plants with defenses are provided by proanthocyanidins (PAs) in bilberry, peanuts, plums, cranberries, curry, and cinnamon plants<sup>37</sup>; flavonoids in fruits, vegetables, tea, cocoa and wine<sup>38</sup>; glucosinolates in the Brassicaceae plant family (including cabbage, rape and black mustard)<sup>39,40</sup>; caffeine in coffee, tea, cacao and kola plants<sup>41,42</sup>; nicotine in tobacco<sup>43</sup>; cannabinoids in cannabis<sup>44</sup>; terpenes and formylated phloroglucinol

compounds (FPCs) in rose gum (*Eucalyptus grandis*)<sup>45</sup>; morphine in opium poppy<sup>46</sup>; cardenolides in milkweed<sup>47</sup>; atropine in deadly nightsade<sup>3</sup>; mimosine in Persian silk (Mimosa) and river tamarind (Leucaena) trees<sup>48,49</sup>; coniine in hemlock<sup>50</sup>; hydrogen cyanide (*HCN*) in cyanogenic plants, such as almonds, cassava, bamboo etc.<sup>51</sup>; colchicine in naked ladies (*Colchicum autumnale*) plant<sup>52</sup> and so on.

Some examples regarding the effects of plant defenses on insects are given in the following. Proanthocyanidins act as feeding deterrents to the gypsy moth (*Lymantria dispar*), brown-tail moth (*Euproctis chrysorrhoea*) and winter moth (*Operophtera brumata*)<sup>53–55</sup>; flavonoids deter African armyworm (*Spodoptera exempta*), fall armyworm (*Spodoptera frugiperda*) and African cotton leafworm *Spodoptera littoralis*<sup>56,57</sup>; isoflavonoids are deterrents to cotton bollworm (*Helicoverpa armigera*), African cotton leafworm and fall armyworm (*S. frugiperda*)<sup>58</sup>; glucosinolates are deterrent and repellent to cabbage moth (*Mamestra brassicae*), southern armyworm (*Spodoptera eridania*), cabbage looper (*Trichoplusia ni*) and green peach aphid (*Myzus persicae*)<sup>40</sup>; caffeine deters beet armyworms and cotton aphids<sup>59,60</sup>; nicotine, cannabinoids and morphine can be deterrents to many insect herbivores<sup>43,61,62</sup>, and terpenes in lavender are repellent to the clothes moth<sup>63</sup>. Lavender oil is commercially used by humans to protect clothes from this insect<sup>63</sup>. Furthermore, cardenolides deter *Nephila* spiders<sup>64</sup>; atropine is a feeding deterrent to gypsy moth larvae, *Lymantria dispar*<sup>65</sup>; mimosine can suppress insect growth<sup>66,67</sup>; coniine and hydrogen cyanide (*HCN*) are poisonous to several groups of insects<sup>68,69</sup>; colchicine sterilizes *Bactrocera tau* (walker) fly and kills *Melanoplus differentialis* and *Gryllus assimilis*<sup>70,71</sup> etc.

#### Models

The deterministic dynamical system is based on three variables, S, E and I. S is the number of susceptible plants, E is the number of exploited plants and I is the insect population at time t. The sum of susceptible and exploited plants is equal to the total plant population in the first model:

$$S + E = N \tag{1}$$

where *N* denotes the total number of plants in a region. Moreover, initially, all plants are susceptible to generalist insects, so that the initial value of *S* reads:

$$S_0 = N \tag{2}$$

Therefore,  $E_0 = 0$  from Eq. (1). Insects can emerge (e.g. by invasion) in a group or singly initially, i.e.  $I_0 > 0$ .

#### Model excluding plant defense

The first model is developed from the fluxes between the *S*, *E* and *I* compartments, including demography and re-susceptibility in plants, shown by the scheme in Fig. 3. So, it is named as *SEI* model. In that Figure,  $\eta$  is the oviposition number (constant), i.e. the number of plants exploited by an insect per day,  $\beta$  is the deposition number (constant), i.e. the mean number of eggs deposited by an insect per exploited plant per day and  $\gamma$  is the natural death rate (constant) for an insect, which is the reciprocal of the average lifespan  $(1/\gamma)$  of an insect.

In order to keep S + E = N constant, we assume that the germination (reproductive growth) and death rate constants (per capita) are the same in plants, denoted by  $\mu$ . Although somewhat artificial, this assumption is also made in many SIR type models to keep the total population constant<sup>29,30</sup>. Exploited plants can be re-susceptible to insects attack, because a plant can be exploited (by insects) multiple times in its lifetime. Let  $\alpha$  be the per capita rate at which exploited plants become re-susceptible. The rate equations are:



**Fig. 3**. Scheme of the *SEI* model (3) with demographic effect and re-susceptibility in plants. Variables and parameters are explained in the text.

$$\frac{dS}{dt} = \mu N - \eta I \frac{S}{N} - \mu S + \alpha E$$
(3a)

$$\frac{dE}{dt} = \eta I \frac{S}{N} - \mu E - \alpha E \tag{3b}$$

$$\frac{dI}{dt} = \beta \eta I \frac{S}{N} - \gamma I \tag{3c}$$

where the germination rate constant of plants is proportional to the total plant population (*N*) and all new germinated plants are susceptible as well. Since S + E = N (Eq. (1)), the sum of the above Eqs. (3a) and (3b) gives:

$$\frac{dN}{dt} = 0 \tag{4}$$

Equation (4) confirms that the total plant population is a constant if the rate constants of plant germination and death are equal. The normalized term S/N in Eqs. (3a), (3b) and (3c) represents the prevalence of susceptibility. From the commencement of the attack, the insect population increases in Eq. (3c) if:

$$\frac{dI}{dt} > 0$$

which is, under the condition  $I \neq 0$ , equivalent to:

$$S > \frac{N\gamma}{\eta\beta} \Rightarrow R_0 > \frac{N}{S}, \quad \text{where } R_0 = \frac{\eta\beta}{\gamma}$$
 (5)

Since  $S \leq N$  from Eqs. (1) and (2), Eq. (5) leads to:

$$R_0 > 1$$
 (6)

where  $R_0$  is the reproduction number of an insect, i.e. the number of viable eggs laid by an insect in its lifetime. Since an insect can lay between 100 and 2000 eggs (mean values) in its lifetime<sup>72–75</sup>, we obtain  $R_0 \gg 1$ . So, the insect population (*I*) grows initially from the commencement of attack.

The model (3) has a non-endemic equilibrium (oviposition and generalist free):

$$H^*_{Free} = (S^*, E^*, I^*) = (N, 0, 0)$$
(7)

 $H_{Free}^*$  is asymptotically stable for  $R_0 < 1$ , shown in the Supplementary Information S1 and illustrated in Figs. 5 and 6A. However, since all insects lay an exceptionally high number of eggs<sup>74</sup>,  $R_0 < 1$  is practically impossible when host plants (N) are abundant in a region. In contrast,  $H_{Free}^*$  becomes an unstable equilibrium (a saddle point) for  $R_0 > 1$ , as illustrated in Fig. 6B and the Supplementary Information S1. The instability of  $H_{Free}^*$  for  $R_0 > 1$  explains that plants cannot eradicate the generalists. Moreover, the model (3) has an endemic equilibrium (including oviposition and generalist insects) for  $R_0 > 1$ :

$$H_{Endemic}^{*} = \left(\frac{N}{R_{0}}, \frac{N(R_{0}-1)}{R_{0}}, \frac{N(\mu+\alpha)(R_{0}-1)}{\eta}\right)$$
(8)

 $H^*_{Endemic}$  is an asymptotically stable equilibrium, which is either a stable node or a stable focus, the proof is given in the Supplementary Information S2. It is important to note that all three variables are positive in the



**Fig. 5.** Time-courses (A, B) of model (3) converge to  $H^*_{Free}$  when  $R_0 < 1$ . Parameters:  $S_0 = 100, \eta = 0.05, \beta = 0.5, \gamma = 1/20, \mu = 0.05$  and  $\alpha = 0.01$ .







**Fig.** 7. Time-course of model (3) when  $R_0 > 1$ . (**A**, **B**) Case where  $H^*_{Endemic}$  is a stable node. Parameters:  $\eta = 4, \beta = 0.5, \gamma = 1/20, \mu = 0.05$  and  $\alpha = 0.01$ . (**C**, **D**) Case where  $H^*_{Endemic}$  is a stable focus. Parameters:  $\eta = 2, \beta = 0.5, \gamma = 1/10, \mu = 0.005$  and  $\alpha = 0.001$ .

endemic state. This implies, in particular, that even in the long run, a certain percentage of plants is susceptible. The reason is that permanently, some plants are germinating (with birth rate constant  $\mu$ ) and some are returning to the susceptible state (with rate constant  $\alpha$ ). Persistence of the insect population creates a problem for plants, because then the insects continue damaging plants in the stable endemic state, shown by the time-courses in Fig. 7. Therefore, the conditions of model 3 are not sufficient to stop an endemic by generalists. However, generalists are usually non-endemic in the wild<sup>19,76-79</sup>, including generalist insect species<sup>15–18</sup>. So, an advancement is made in model (3) to verify whether evolution of constitutive defenses in plants can end the generalist endemic.

#### Model including plant defense

A new parameter  $\sigma$  is introduced, representing the per capita rate at which new plants germinate with chemical defenses. These defended plants, denoted by the compartment *D*, are not hosts to generalists. Obviously,  $\sigma \leq \mu$  (with  $\mu$  being the per capita reproduction or germination rate constant). The fluxes between the compartments are as shown in Fig. 3 with an additional flux to the compartment of defended plants (*D*) and a modification in the flux entering the susceptible compartment, expressed by the scheme in Fig. 4.

The rate equations are:

$$\frac{dS}{dt} = (\mu - \sigma)N - \frac{\eta IS}{N} - \mu S + \alpha E$$
(9a)

$$\frac{dE}{dt} = \frac{\eta IS}{N} - (\mu + \alpha) E \tag{9b}$$

$$\frac{dD}{dt} = \sigma N - \mu D \tag{9c}$$

$$\frac{dI}{dt} = \frac{\beta \eta I S}{N} - \gamma I \tag{9d}$$

where the germinating plants enter the susceptible compartment at a per capita rate  $(\mu - \sigma)$ , shown in Fig. 4. The model (9) has an additional ODE representing the growth rate in defended plants (*D*), Eq. (9c). This main structural difference between models (3) and (9) allows us to assess the benefit of chemical defense evolved in plants. This new model (9) is called *SEDI* model, where the total plant population (*N*) is equal to the sum of susceptible, exploited and defended plants (S + E + D = N). Thus, the sum of Eqs. (9a), (9b) and (9c) gives:

$$\frac{N}{tt} = 0 \tag{10}$$

Eq. (10) proves that the total plant population is a constant in model (9) if the germination and death rate constants (per capita) are assumed to be equal.

The four-dimensional model (9) can be reduced to three dimensions, because D is the only variable occurring in the ODE (9c). The solution of that equation is not needed (although it can easily be found analytically) since the implicit functions of S, E and I in the right-hand sides of the ODEs (9a), (9b) and (9d) are sufficient to analyze the behavior of model (9) in view of equilibria in  $\mathbb{R}^3$  and their stability. The model (9) has two possible equilibrium points. The non-endemic (oviposition and insect free) equilibrium is:

$$Q_{Free}^* = ((1-p)N, 0, 0) \tag{11}$$

where  $p = \frac{\sigma}{\mu}$  is the fraction of germinated plants (per capita) that evolved constitutive defense. To be noted  $p \in [0, 1]$ , because  $\sigma \leq \mu$ . Interestingly, some susceptible plants ((1 - p)N) are not oviposited by generalist insects for p < 1.  $Q_{Free}^*$  is asymptotically stable for  $R_0 < 1$ , proved in the Supplementary Information S3 and illustrated by Figs. 9 and 10B. For  $R_0 > 1$ ,  $Q_{Free}^*$  is asymptotically stable if and only if:

$$p > 1 - \frac{1}{R_0}$$
 (12)



**Fig. 4**. Scheme of the *SEDI* model (9) with demographic effect, re-susceptibility and germinated plants defended against generalists. Variables and parameters are explained in the text.



Fig. 9. Time-courses (A, B) of model (9) converge to  $Q^*_{Free}$  when  $R_0 < 1$ . Parameters are the same as in Fig. 8 except for  $\eta = 0.05$ .



**Fig. 10**. Phase portraits of  $Q^*_{Free}$ , obtained from model (9). (A) Stable for  $R_0 > 1$  and  $p > 1 - \frac{1}{R_0}$ . Parameters:  $\eta = 4, \beta = 0.5, \gamma = 1/20, p = \sigma/\mu = 0.99$  and  $\alpha = 0.01$ . (B) Stable for  $R_0 < 1$ . Parameters are the same as in Fig. 9.



**Fig. 8.** Time-courses of model (9) converge to  $Q^*_{Free}$  when  $p > 1 - \frac{1}{R_0}$ . Parameters:  $\eta = 4, \beta = 0.5, \gamma = 1/20, \mu = 0.05, \sigma = 0.049$  and  $\alpha = 0.01$ .

In the extreme case p = 0, this is equivalent to the condition  $R_0 < 1$  mentioned above. Calculations of deriving  $Q_{Free}^*$  and the proof of its asymptotic stability for  $R_0 > 1$  are given in the Supplementary Information S3. The time-course and phase-portrait of stable  $Q_{Free}^*$  at  $R_0 > 1$  are illustrated by Figs. 8 and 10A, respectively.

The endemic equilibrium of the model (9) is:

$$Q_{Endemic}^{*} = \left(\frac{N}{R_{0}}, \left(1 - p - \frac{1}{R_{0}}\right)N, \left(1 - p - \frac{1}{R_{0}}\right)\frac{(\mu + \alpha)NR_{0}}{\eta}\right)$$
(13)

Generalists are non-endemic in the wild if the endemic equilibrium (13) does not exist at all, i.e. the insect and oviposition free equilibrium ( $Q_{Free}^*$ ) is the only possible stable equilibrium. This is achieved if and only if the inequality (12) holds true, because exploited plants and generalists would be negative in the equilibrium (13) for  $p > 1 - 1/R_0$ . In contrast, the endemic equilibrium of generalists exists and is stable (either a stable node or a stable focus) if  $p < 1 - 1/R_0$ , shown by the time-courses in Fig. 11. The derivation and stability analysis of  $Q_{Endemic}^*$  are given in the Supplementary information S4. Although  $Q_{Free}^*$  is stable for  $R_0 < 1$  shown by Figs. 9 and 10B, the case is only feasible if the host plants of generalist insects are quite low in a region.

It is interesting to obtain that evolution of constitutive defense in plants depends on the reproduction number  $(R_0)$  of insects. Condition (12) can be interpreted as follows. In order to stop an endemic by generalist insects, not all plant individuals need to store chemical defenses. For example, if a generalist insect species has  $R_0 = 500$ , then  $1 - 1/R_0 = 0.998$ . Therefore, that particular generalist species cannot be endemic if more than 99.8% of all plants germinate with deterrent chemicals. Since  $R_0 \gg 1$  for all insect species<sup>74</sup>, condition (12) also proves that if nearly all plants are germinated with chemical defenses, generalist insects become non-endemic in the wild. This phenomenon is shown by Figs. 8 and 10A. In other words, a few plants can remain undefended because if insects appeared (as a fluctuation of the free equilibrium), then these few plants would be insufficient to enable sufficient reproduction of insects.

#### Illustration of results

#### Discussion

The main results of this study are as follows:

- If plants do not evolve chemical defenses, an endemic by generalist insects happens for  $R_0 > 1$ , Fig. 7. That result means generalists become native herbivores of plants in a certain region. However, as stated before, such phenomenon is not evident in the wild<sup>15-18</sup>.
- If a fraction of all plants are germinated with chemical defenses, but condition (12) is not fulfilled, the generalist endemic persists, Fig. 11. However, this result is not consistent with the observations mentioned in the previous point<sup>15–18</sup>.
- Plants display a generalist-free (non-endemic) stable equilibrium for  $R_0 > 1$  if and only if nearly all plants are germinated with chemical defenses (i.e. constitutive plant defense), expressed by condition (12) and shown



**Fig. 11.** Time-course of model (9) for  $p < 1 - \frac{1}{R_0}$ . (**A**, **B**) Case where  $Q^*_{Endemic}$  is a stable node. Parameters:  $\eta = 4, \beta = 0.5, \gamma = 1/20, \mu = 0.05, \sigma = 0.04$  and  $\alpha = 0.01$ . (**C**, **D**) Case where  $Q^*_{Endemic}$  is a stable focus.  $\eta = 2, \beta = 0.5, \gamma = 1/10, \mu = 0.005, \sigma = 0.001$  and  $\alpha = 0.001$ .

in Figs. 8 and 10A. That practically (though not literally) justifies the theory that all plants have chemical defenses<sup>3,5,11,14</sup> due to the nonexistence of generalist endemic species in the wild<sup>15–18</sup>.

If the reproduction number ( $R_0$ ) of generalist insects in a region is below one, they cannot be endemic, Figs. 9 and 10B. This case is feasible when sufficient host plants are unavailable.Condition (12) is necessary and sufficient for the generalist insect species to be non-endemic. That means  $Q_{Free}^*$  is the only stable steady state if  $p > 1 - 1/R_0$  and  $p > 1 - 1/R_0$  occurs if  $Q_{Free}^*$  is the only stable steady state, where  $R_0 > 1$ . Since we obtained that nearly all plants should germinate with chemical defenses, some exceptions could be possible in plant chemical ecology. These exceptions could be considered as free-riders. The concept of free-riders is often used in game theory<sup>80,81</sup>. However, the costs of producing defense chemicals are neglected for the sake of simplicity. Therefore, the free-riders do not have any physiological advantages in comparison to defended plants. If costs were included, as is done in many game-theoretical models<sup>80,81</sup>, a dilemma occurs. The free-riders then have a higher growth rate and could outcompete defended plants, so that the population becomes susceptible to herbivores. Various ways of resolving the dilemma have been proposed in biological contexts other than plant-herbivore interactions, such as spatial structure<sup>82</sup>.

Interestingly, a few plants can be found that may not invest sufficiently in constitutive defense<sup>35,83,84</sup>. For example, blue lupin (*Lupinus angustiflius*) may not produce any alkaloids even after suffering from herbivory<sup>85</sup> and the invasive plant *Lespedeza cuneata* is inadequately defended constitutively against herbivory<sup>86</sup>. However, such examples are rare in plant chemical ecology. The basic model (3) without plant defense could be written as a model without vital dynamics, that is, without germination and death rates, and still having the conservation relation S + E = N. However, for the model (9) with plant defense, considering the plant germination rate is important to describe constitutive defense, which is present in plants during the normal course of development from germination on<sup>1,2</sup>.

The method is developed under the framework of stability theory. Stability theory is widely used in epidemiology<sup>28-30,87</sup>, eco-epidemiology<sup>88-91</sup>, population dynamics<sup>27,92</sup> and other fields of mathematical biology or ecology<sup>28,29</sup> to determine the asymptotic properties of solutions or equilibrium points in a long interval of time. In this field of study, the biological or ecological phenomena are expressed by a system of differential equations<sup>27,92</sup>. Stable equilibria of that system refer to the fixed points, where the entire system converge<sup>27,29,92</sup>. For example, vaccinating a certain percentage of a population eradicates an epidemic, because vaccination above the threshold makes the endemic-free equilibrium of the SIRV model (SIR model with vaccination) asymptotically stable<sup>28,29</sup>. Similarly, from model (9), it can be claimed that nearly all plants have evolved chemical defenses, because only then the generalist endemic free equilibrium is asymptotically stable.

Generalist insects are polyphagous by nature<sup>93,94</sup>, thus, select a wide range of host plants<sup>16</sup>. Nevertheless, plant defense negatively affects them due to their lack of specialism<sup>95-98</sup>. For example, glucosinolates and their hydrolyzed toxic isothiocyanate products of the Brassicaceae plant family reduce the development rate and cause high mortality of lepidopteran generalists<sup>98,99</sup>, such as *Spodoptera exigua, Spodoptera littoralis, Mamestra brassicae, Trichoplusia ni*, and *Helicoverpa armigera*<sup>96</sup>. Specialist insects, on the other hand, have evolved efficient counter-defense or resistance against host plant toxins<sup>95,97,99</sup>. Moreover, specialist insects use plant defense chemicals as a cue to identify host plants for oviposition<sup>33,36,100-102</sup>. Thus, plant defense could increase the pressure of specialist insect herbivores<sup>33,36</sup>. That is a direct contrast between the selection behavior of specialist vs generalist insects. It is not exactly clear how plants are benefiting against specialists by evolving defenses. However, plant defenses can attract the natural enemies of insect herbivores<sup>36,103</sup>, which leads to the tritrophic interaction<sup>104</sup>. That can be a possible explanation.

The indirect role of plant defense compounds and their subsequent products is attracting predators and parasitoids of deleterious insects<sup>33,36,103–105</sup>. For example, in *Pieris rapae* infested *Arabidopsis* plants, nitriles (a less toxic hydrolyzed products of glucosinolates) can recruit the parasitoid wasp *Cotesia rubecula*<sup>106</sup>; isothiocyanates in the Brassica plants can recruit *Trichogramma chilonis* wasps during infestation by *Plutella xylostella*<sup>107</sup>; entomopathogenic nematodes are recruited by insect-damaged maize roots<sup>108</sup>. Tritrophic interaction by herbivore-induced plant volatiles (HIPVs) has been modelled before<sup>109,110</sup>, where the emergence of natural enemies reduce the deleterious insect population. Although in this study, the indirect role of plant defenses is not considered, it could be an interesting refinement of the model (9).

A study conducted on microorganisms showed that the generalist genera are older than specialist genera in an evolutionary timescale<sup>111</sup>. Therefore, it is possible that the fundamental cause for plant species evolving chemical defenses was to eradicate the deleterious generalists<sup>4–8</sup>. However, for a holistic understanding, it is important to investigate the plants' benefit of evolving chemical defenses against different group of deleterious insects (generalists and specialists) together<sup>33,36</sup>. Especially, it is known that specialists cause damage by being locally endemic to specific niches<sup>111,112</sup>. This leaves us with multiple questions: Whether tritrophic interaction is capable of ending a specialist-driven endemic, whether the chemical defense of plants is responsible for the endemics by specialists etc. It is worthwhile tackling these questions by mathematical modeling in the future.

#### Conclusion

From the main results of this study, it can be concluded that generalist insects are non-endemic in the wild, because nearly all plants are germinated with secondary metabolites. However, that does not mean generalists become extinct. They can survive with a low reproduction number ( $R_0 < 1$ ) on host plants. Although clear results are obtained, the main limitation of these models is the assumption of a constant total population of plants, i.e. S + E = N in model (3) and S + E + D = N in model (9), where N is constant. It would be intriguing to verify the results by advancing these models to describe overall growing plant populations<sup>109,110</sup>, in analogy to vital dynamics in SIR models<sup>29,30</sup>.

The study fits within the context of plant protection, insect behavior and pest control issues, which could be interesting not only for wildlife practitioners, but also for crop protection. Since generalists are only temporary visitors, crop protection measures should be more relevant against endemic specialist insect pests<sup>36</sup>. Several pest control methods, such as application of insecticides<sup>113</sup>, intercropping or mixed cropping<sup>114</sup>, sterile insect technique (SIT)<sup>115</sup> etc. are practiced to save crop plants from deleterious insects. Including these control measures will be promising extensions to the model (9), which could expedite the eradication process of insect pests. Moreover, since plants are susceptible to both specialists and generalists, it would be significant to expand the model for both groups of insect (pest) species.

#### Data availability

All data generated or analyzed during this study are included in this published article [and its supplementary information file].

Received: 11 June 2024; Accepted: 30 September 2024 Published online: 28 October 2024

#### References

- 1. Dicke, M. Induced responses to herbivory by r. karban and i.t. baldwin. Trends Ecol. Evol. 13(2), 83 (1998).
- Textor, S. & Gershenzon, J. Herbivore induction of the glucosinolate-myrosinase defense system: Major trends, biochemical bases and ecological significance. *Phytochem. Rev.* 8(1), 149–170 (2009).
- 3. Vickery, M. Plant poisons: their occurrence, biochemistry and physiological properties. Sci. Prog. 93, 181-221 (2010).
- 4. Fraenkel, G. S. Raison d'etre of secondary plant substances. Science 129(3361), 1466-1470 (1959).
- Bernays, E. & Chapman, R. The evolution of deterrent responses in plant-feeding insects. In Perspectives in Chemoreception and Behavior. Proceedings in Life Sciences (eds. Chapman, R. F. et al.) (Springer, 1987).
- 6. Wittstock, U. & Gershenzon, J. Constitutive plant toxins and their role in defense against herbivores and pathogens. *Curr. Opin. Plant Biol.* **5**, 300–307 (2002).
- 7. Hartmann, T. From waste products to ecochemicals: Fifty years research of plant secondary metabolism. *Phytochemistry* **68**(22), 2831–2846 (2007).
- 8. War, A. R. et al. Mechanisms of plant defense against insect herbivores. Plant Signal. Behav. 7(10), 1306–1320 (2012).
- Dethier, V. G., Browne, B. L. & Smith, C. N. The designation of chemicals in terms of the responses they elicit from insects. J. Econ. Entomol. 53(1), 134–136 (1960).
- 10. Klun, J. A., Khrimian, A. & Debboun, M. Repellent and deterrent effects of SS220, picaridin, and deet suppress human blood feeding by *Aedes aegypti, Anopheles stephensi*, and *Phlebotomus papatasi*. J. Med. Entomol. **43**(1), 34–39 (2006).
- 11. Swain, T. Secondary compounds as protective agents. Annu. Rev. Plant Physiol. 28(1), 479-501 (1977).
- Singer, A. C., Crowley, D. E. & Thompson, I. P. Secondary plant metabolites in phytoremediation and biotransformation. *Trends Biotechnol.* 21(3), 123–130 (2003).
- Divekar, P. A. et al. Plant secondary metabolites as defense tools against herbivores for sustainable crop protection. Int. J. Mol. Sci. 23(5), 2690 (2022).
- Gershenzon, J. & Ullah, C. Plants protect themselves from herbivores by optimizing the distribution of chemical defenses. Proc. Natl. Acad. Sci. 119(4), 2120277119 (2022).
- 15. Walter, G. H. & Benfield, M. D. Temporal host plant use in three polyphagous heliothinae, with special reference to Helicoverpa punctigera (wallengren) (noctuidae: Lepidoptera). *Austral Ecol.* **19**, 458–465 (1994).
- 16. Ali, J. G. & Agrawal, A. A. Specialist versus generalist insect herbivores and plant defense. Trends Plant Sci. 17(5), 293-302 (2012).
- 17. Wang, Y. et al. Higher plasticity in feeding preference of a generalist than a specialist: Experiments with two closely related helicoverpa species. *Sci. Rep.* **7**, 17876 (2017).
- Rafter, M. A. & Walter, G. H. Generalising about generalists? a perspective on the role of pattern and process in investigating herbivorous insects that use multiple host species. *Arthropod-Plant Interact* 14, 1–20 (2020).
- 19. Futuyma, D. J. & Moreno, G. The evolution of ecological specialization. Annu. Rev. Ecol. Syst. 19, 207-233 (1988).
- Mason, P. A., Wilkes, S. R., Lill, J. T. & Singer, M. S. Abundance trumps quality: Bi-trophic performance and parasitism risk fail to explain host use in the fall webworm. Oikos 120, 1509–1518 (2011).
- Murphy, S. M. & Loewy, K. J. Trade-offs in host choice of an herbivorous insect based on parasitism and larval performance. Oecologia 179, 741–751 (2015).
- Buonomo, B., Giannino, F., Saussure, S. & Venturino, E. Effects of limited volatiles release by plants in tritrophic interactions. Math. Biosci. Eng. 16(5), 3331-3344 (2019).
- Mondal, R., Kesh, D & Mukherjee, D. Role of induced volatile emission modelling tritrophic interaction. *Differ. Equ. Dyn. Syst.* 30, 749–765 (2022).
- 24. Yuan, H. et al. Development of a widely targeted volatilomics method for profiling volatilomes in plants. *Mol. Plant* 15(1), 189–202 (2022).
- 25. Mezzomo, P. et al. Variation in induced responses in volatile and non-volatile metabolites among sixwillow species: Do willow species share responses to herbivory?. *Phytochem.* 226, 114222 (2024).
- Kermack, W. O. & McKendrick, A. G. A contribution to the mathematical theory of epidemics. Proc. R. Soc. Lond. Ser. A 115(772), 700–721 (1927).
- 27. Murray, J. D. Mathematical Biology. I An Introduction 3rd edn. (Springer, 2002).
- 28. Brauer, F. & Castillo-Chavez, C. Mathematical Models in Population Biology and Epidemiology 2nd edn. (Springer, 2001).
- 29. Martcheva, M. An Introduction to Mathematical Epidemiology 1st edn. (Springer, 2015).
- Makinde, O. D. Adomian decomposition approach to a SIR epidemic model with constant vaccinationstrategy. Appl. Math. Comput. 184(2), 842–848 (2007).
- 31. Kothari, I. L. & Patel, M. Plant immunization. Indian J. Exp. Biol. 42(3), 244-252 (2004).
- 32. Jong, T. J. Why fast-growing plants do not bother about defence. Oikos 74(3), 545-548 (1995).
- 33. Van der Meijden, E. Plant defence, an evolutionary dilemma: Contrasting effects of (specialist andgeneralist) herbivores and natural enemies. *Entomol. Exp. Appl.* **80**(1), 307–310 (1996).
- 34. Åström, M. & Lundberg, P. Plant defence and stochastic risk of herbivory. Evol. Ecol. 8(3), 288-298 (1994).
- Ito, K. & Sakai, S. Optimal defense strategy against herbivory in plants: Conditions selecting for induced defense, constitutive defense, and no-defense. J. Theor. Biol. 260(3), 453–459 (2009).
- Chakraborty, S., Gershenzon, J. & Schuster, S. Selection pressure by specialist and generalist insectherbivores leads to optimal constitutive plant defense, a mathematical model. *Ecol. Evol.* 13, 10763 (2023).
- Yu, D., Huang, T., Tian, B. & Zhan, J. Advances in biosynthesis and biological functions of proanthocyanidins in horticultural plants. *Foods* 9(12), 1774 (2020).

- 38. Panche, A. N., Diwan, A. D., Chandra, S. R. Flavonoids: An overview. J. Nutr. Sci. 29(5), 47 (2016).
- 39. Halkier, B. A. & Gershenzon, J. Biology and biochemistry of glucosinolates. Annu. Rev. Plant Biol. 57, 303-333 (2006).
  - Hopkins, R. J., van Dam, N. M. & van Loon, J. J. A. Role of glucosinolates in insect plant relationships and multitrophic interactions. Annu. Rev. Entomol. 54, 57–83 (2009).
  - 41. Ashihara, H. & Crozier, A. Biosynthesis and metabolism of caffeine and related purine alkaloids in plants. *Adv. Bot. Res.* 30, 117–205 (1999).
  - 42. Kim, Y. S. & Sano, H. Pathogen resistance of transgenic tobacco plants producing caffeine. Phytochemistry 69(4), 882-888 (2008).
  - 43. Steppuhn, A., Gase, K., Krock. B., Halitschke, R., Baldwin. I. T. Nicotine's defensive function in nature. *PLoS Biol.* 2(8), 217 (2004). 44. Lambert, D. M. & Fowler, C. J. The endocannabinoid system: Drug targets, lead compounds, and potential therapeutic
  - applications. J. Med. Chem. 48(16), 5059–5087 (2005). 45. Henery, M. L., Wallis, I. R., Stone, C. & Foley, W. J. Methyl jasmonate does not induce changes inEucalyptus grandis leaves that
  - 45. There y, M. L., Wans, T. K., Stone, C. & Poley, W. J. Methyl astholiae does not induce changes including planting leaves that alter the effect of constitutive defences on larvae of a specialistherbivore. *Oecologia* 156(4), 847–859 (2008).
  - Zulak, K. G., Khan, M. F., Alcantara, J., Schriemer, D. C. & Facchini, P. J. Plant defense responses inopium poppy cell cultures revealed by liquid chromatography-tandem mass spectrometry proteomics. *Mol. Cell. Proteom.* 8(1), 86–98 (2009).
  - Agrawal, A. A., Petschenka, G., Bingham, R.A., Weber, M.G., & Rasmann, S. Toxic cardenolides: Chemical ecology and coevolution of specialized plant-herbivore interactions. *New Phytol.* 194(1), 28–45 (2012).
  - 48. Adams, R. & Jones, V. V. The structure of leucenol. J. Am. Chem. Soc. 69(7), 1803-1805 (1947).
  - 49. Honda, M. D. H. & Borthakur, D. Mimosine facilitates metallic cation uptake by plants through formation of mimosine-cation complexes. *Plant Mol. Biol.* **102**, 431–445 (2020).
  - Barceloux, D. G. Poison hemlock (Conium maculatum L.). Medical Toxicology of Natural Substances: Foods, Fungi, Medicinal Herbs, Plants, and Venomous Animals 796–799 (Wiley, 2008).
  - 51. Moller, B. L. Functional diversifications of cyanogenic glucosides. Curr. Opin. Plant Biol. 13, 338-347 (2010).
  - Rousseau, G., Clément, J. Fezard, J. B. & Laribi, S. Intoxication au colchique par confusion avec l'ail desours (Colchicum poisoning by confusion with wild garlic (Allium ursinum)). *Rev. Med. Interne* 43(9), 559–561 (2022).
  - 53. Feeny, P. P. Effect of oak leaf tannins on larval growth of the winter moth *Operophtera brumata*. J. Insect Physiol. 14, 805–817 (1968).
  - 54. Bernays, E. A. Plant tannins and insect herbivores: An appraisal. Ecol. Entomol. 6, 353-360 (1981).
  - Pradit, N., Mescher, M. C., Wang, Y., Vorsa, N. & Rodriguez-Saona, C. Phytoplasma infection ofcranberries benefits non-vector phytophagous insects. Front. Ecol. Evol. 7, 181 (2019).
  - Simmonds, M. S. J., Blaney, W. M. & Fellows, L. E. Behavioural and electrophysiological study of antifeedant mechanisms associated with polyhydroxy alkaloids. J. Chem. Ecol. 16, 3167–3196 (1990).
  - Wahba, T. F., El-Bassyouni, G. T., El-Shamy, A. M. & Wahba, M. N., Nanoinert diatomaceous andemamectin benzoate: Enhancing wheat protection against fall armyworms for sustainablemanagement. S. Afr. J. Bot. 169, 413–425 (2024).
  - Simmonds, M. S. J. & Stevenson, P. C. Effects of isoflavonoids from cicer on larvae of Heliocoverpaarmigera. J. Chem. Ecol. 27, 965–977 (2001).
  - Nathanson, J. A. Caffeine and related methylxanthines: Possible naturally occurring pesticides. Science 226(4671), 184–187 (1984).
  - Kim, Y.-S., Lim, S., Kang, K.-K. & Jung, Y.-J, Resistance against beet armyworms and cotton aphids incaffeine-producing transgenic chrysanthemum. *Plant Biotechnol.* 28(4), 393–395 (2011).
  - 61. Park, S. H. et al. Contrasting roles of cannabidiol as an insecticide and rescuing agent for ethanolinduceddeath in the tobacco hornworm Manduca sexta. *Sci. Rep.* **9**, 10481 (2019).
  - 62. Howe, G. A. & Jander, G. Plant immunity to insect herbivores. Annu. Rev. Plant Biol. 59, 41-66 (2008).
  - Plarre, R., et al. Effects of oil of cloves and citronellol, two commercially available repellents, against the webbing clothes moth Tineola bisselliella. Hum (lepidoptera: Tineidae). Anzeiger Schsadlingskunde Pflanzenschutz Umweltschutz 70, 45–50 (1997).
  - Petschenka, G., Bramer, C., Pankoke, H. & Dobler, S. Evidence for a deterrent effect of cardenolides onNephila spiders. Basic Appl. Ecol. 12(3), 260–267 (2011).
  - Shields, V. D. C. et al. The effect of varying alkaloid concentrations on the feeding behavior of gypsymoth larvae, *Lymantria dispar* (L.) (lepidoptera: Lymantriidae). Arthropod Plant Interact. 2(2), 101–107 (2008).
  - Ishaaya, I., Hirashima, A., Yablonski, S., Tawata, S. & Morifusa, E. Mimosine, a nonprotein amino acid, inhibits growth and enzyme systems in *Tribolium castaneum*. Pestic. Biochem. Physiol. 39, 35–42 (1991).
  - Nguyen, B. C. Q., Chompoo, J. & Tawata, S. Insecticidal and nematicidal activities of novel mimosine derivatives. *Molecules* 20, 16741–16756 (2015).
  - 68. Robin, M. Yellow pitcher plant or trumpets (*Sarracenia flava*). United States Department of Agriculture. United States Department of Agriculture Forest Service (Retrieved 3 May 2015).
  - 69. Polunin, N. High-concentration hydrogen cyanide fumigation of fungi and bacteria. Nature 150, 682-684 (1942).
  - 70. Dooley, T. P. The influence of colchicine on the germ cells of insects, *Melanoplus differentialis* and *Gryllus assimilis*, with special reference to the cytoplasmic inclusions. *Proc. Iowa Acad. Sci.* **46**(1), 445–446 (1939).
  - 71. Li, X. et al. Effects of four chemosterilants on Bactrocera tau. Ecotoxicol. Environ. Saf. 243, 114028 (2022).
  - 72. Clausen, C. P. Entomophagous Insects (McGraw-Hill, 1940).
  - 73. Wilson, E. O. The Insect Societies (Harvard University Press, 1971).
  - 74. Hinton, H. E. Biology of Insect Eggs (Pergamon Press, 1981).
  - 75. Holldöbler, B. & Wilson, E. O. The Ants (Harvard University Press, 1990).
  - 76. Devictor, V. et al. Defining and measuring ecological specialization. J. Appl. Ecol. 47, 15-25 (2010).
  - 77. Poisot, T., Bever, J. D., Nemri, A., Thrall, P. H. & Hochberg, M. E. A conceptual framework for the evolution of ecological specialisation. *Ecol. Lett.* 14, 841–851 (2011).
  - Büchi, L. & Vuilleumier, S. Coexistence of specialist and generalist species is shaped by dispersal and environmental factors. Am. Nat. 183(5), 612–624 (2014).
  - 79. Kneitel, J. M. Occupancy and environmental responses of habitat specialists and generalists depend on dispersal traits. *Ecosphere* **9**(3), 02143 (2018).
  - 80. Denison, R. F. et al. Cooperation in the rhizosphere and the "free rider" problem. Ecology 84, 838-845 (2003).
  - Delton, A. W., Cosmides, L., Guemo, M., Robertson T. E. & Tooby, J. The psychosemantics of free riding: Dissecting the architecture of a moral concept. J. Pers. Soc. Psychol. 102(6), 1252–1270 (2012).
  - Schuster, S. et al. Cooperation and cheating in microbial exoenzyme production—theoretical analysis for biotechnological applications. *Biotechnol. J.* 5, 751–758 (2010).
  - 83. Strauss, S. Y. & Agrawal, A. A. The ecology and evolution of plant tolerance to herbivory. Trends Ecol. Evol. 14, 179-185 (1999).
  - Maron, J. L. & Crone, E. Herbivory: Effects on plant abundance, distribution and population growth. Proc. R. Soc. B: Biol. Sci. 273(1601), 2575–2584 (2006).
  - del Pilar Vilariño, M., Mareggiani, G., Grass, M. Y., Leicach, S. R. & Ravetta, D. A. Post damage alkaloid concentration in sweet and bitter lupin varieties and its effect on subsequent herbivory. J. Appl. Entomol. 129, 233–238 (2005).
  - Beaton, L. L., Van Zandt, P. A., Esselman, E. J. & Knight, T. M. Comparison of the herbivore defense and competitive ability of ancestral and modern genotypes of an invasive plant, *Lespedeza cuneata*. Oikos 120, 1413–1419 (2011).

- 87. Adak, D., Majumder, A. & Bairagi, N. Mathematical perspective of covid-19 pandemic: Disease extinction criteria in deterministic and stochastic models. Chaos Soliton. Fract. 42, 110381 (2021).
- Chattopadhyay, J. & Bairagi, N. Pelicans at risk in salton sea-an eco-epidemiological model. Ecol. Model. 136(2-3), 103-112 (2001)
- 89. Chattopadhyay, J. & Pal, S. Viral infection on phytoplankton-zooplankton system-a mathematical model. Ecol. Model. 151(1), 15-28 (2002)
- 90. Chattopadhyay, J., Srinivasu, P. D. N. & Bairagi, N. Pelicans at risk in salton sea-an eco-epidemiological model-ii. Ecol. Model. 167(1-2), 199-211 (2003).
- 91. Pal, S., Chatterjee, S. & Chattopadhyay, J. Role of toxin and nutrient for the occurrence and termination of plankton bloom-results drawn from field observations and a mathematical model. Biosystems 90(1), 87-100 (2007).
- 92. Kot, M. Elements of Mathematical Ecology (Cambridge University Press, 2001).
- 93. Schoonhoven, L. M., van Loon, J. J. A. & Dicke, M. Insect-Plant Biology (Oxford University Press, 2005).
- 94. Mello, M. O. & Silva-Filho, M. C. Plant-insect interactions: An evolutionary arms race between two distinct defense mechanisms. Braz. J. Plant. Physiol. 14, 71-81 (2002).
- 95. Jeschke, V. et al. How glucosinolates affect generalist lepidopteran larvae: Growth, development and glucosinolate metabolism. Front. Plant Sci. 8, 1995 (2017).
- 96. Jeschke, V. et al. So much for glucosinolates: A generalist does survive and develop on brassicas, but at what cost?. Plants 10(5), 962 (2021).
- 97. Chakraborty, S., Gershenzon, J. & Schuster, S. Comparing two strategies of counter-defense against plant toxins: A modeling study on plant-herbivore interactions. Front. Ecol. Evol. 11, 1197757 (2023).
- 98. Chakraborty, S. & Schuster, S. How plant toxins cause early larval mortality in herbivorous insects: An explanation by modeling the net energy curve. *Toxins* 16(2), 72 (2024). 99. Sun, R. et al. Tritrophic metabolism of plant chemical defenses and its effects on herbivore and predator performance. *ELife* 8,
- 51029 (2019).
- 100. Mewis, I. Z., Ulrich, C. H. & Schnitzler, W. H. The role of glucosinolates and their hydrolysis products in oviposition and hostplant finding by cabbage webworm, Hellula undalis. Entomol. Exp. Appl. 105, 129-139 (2002).
- 101. Renwick, J. A, Haribal, M., Gouinguené, S. & Städler, E. Isothiocyanates stimulating oviposition by the diamondback moth, Plutella xylostella. J. Chem. Ecol. 32(4), 755-766 (2006).
- 102. Kos, M. et al. Relative importance of plant-mediated bottom-up and top-down forces on herbivore abundance on Brassica oleracea. Funct. Ecol. 25(5), 1113-1124 (2011).
- 103. Dicke, M. & Baldwin, I. T. The evolutionary context for herbivore-induced plant volatiles: Beyond the 'cry for help'. Trends Plant Sci. 15(3), 167-175 (2010).
- 104. Turlings, T. C. J. & Erb, M. Tritrophic interactions mediated by herbivore-induced plant volatiles: Mechanisms, ecological relevance, and application potential. Annu. Rev. Entomol. 63(1), 433-452 (2018).
- 105. Thaler, J. S. Jasmonate-inducible plant defences cause increased parasitism of herbivores. Nature 399, 686-688 (1999).
- 106. Van Poecke, R. M. P., Posthumus, M. A. & Dicke, M. Herbivore-induced volatile production by arabidopsis thaliana leads to attraction of the parasitoid cotesia rubecula: Chemical, behavioral, and gene-expression analysis. J. Chem. Ecol. 27(10), 1911-1928 (2001).
- 107. Reddy, G. V., Holopainen, J. K. & Guerrero, A. Olfactory responses of plutella xylostella natural enemies to host pheromone, larval frass, and green leaf cabbage volatiles. J. Chem. Ecol. 28(1), 131-143 (2002).
- 108. Rasmann, S. et al. Recruitment of entomopathogenic nematodes by insect-damaged maize roots. Nature 434, 732-737 (2005).
- 109. Liu, Y. et al. Modelling tritrophic interactions mediated by induced defence volatiles. Ecol. Model. 220(23), 3241-3247 (2009).
- 110. Fergola, P. & Wang, W. On the influences of defensive volatiles of plants in tritrophic interactions. J. Biol. Syst. 19(2), 345-363 (2011).
- 111. von Meijenfeldt, F. A. B., Hogeweg, P. & Dutilh, B. E. A social niche breadth score reveals niche range strategies of generalists and specialists. Nat. Ecol. Evol. 7, 768-781 (2023).
- 112. Behroozian, M., Ejtehadi, H., Memariani, F., Pierce, S. & Mesdaghi, M. Are endemic species necessarily ecological specialists? functional variability and niche differentiation of two threatened dianthus species in the montane steppes of northeastern iran. Sci. Rep. 10, 11774 (2020).
- 113. Zhang, Z. et al. Design, synthesis and biological activity of diamide compounds based on 3-substituent of the pyrazole ring†. Pest Manag. Sci. 78(5), 2022–2033 (2022).
- 114. Letourneau, D. K. et al. Does plant diversity benefit agroecosystems? A synthetic review. Ecol. Appl. 21(1), 9-21 (2011).
- 115. Bourtzis, K. & Vreysen, M. J. B. Sterile insect technique (sit) and its applications. Insects 12(7), 638 (2021).

#### Acknowledgements

Stimulating discussions with Abdel H. Halloway and Jonathan Gershenzon are gratefully acknowledged. Financial support by the German Research Foundation (DFG) in the SFB 1127 ChemBioSys project no. 239748522 to S.S. and the Jena School of Microbial Communication (JSMC) to S.D. is gratefully acknowledged. S. C. thanks the IMPRS "Chemical Communication in Ecological Systems" for training and support.

#### Author contributions

S.C. conceptualized the study and developed the models. S.C. and S.D. wrote the original draft, analyzed the models, did the simulations and provided the calculations. S.S. supervised the study and refined the model and manuscript draft. All authors verified the results and reviewed the manuscript.

#### Funding

Open Access funding enabled and organized by Projekt DEAL.

#### Additional information

Supplementary Information The online version contains supplementary material available at https://doi. org/10.1038/s41598-024-74771-0.

Correspondence and requests for materials should be addressed to S.S.

Reprints and permissions information is available at www.nature.com/reprints.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

**Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.

© The Author(s) 2024