

# **Dynamic susceptibility contrast‑enhanced perfusion magnetic resonance imaging parameters for predicting**  *MGMT* **promoter methylation and prognostic value in newly diagnosed patients with glioblastoma**

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**Abstract.** O6‑methylguanine DNA methyltransferase *(MGMT)*  promoter methylation is an important clinical biomarker of newly diagnosed glioblastoma. Previous radiological studies using dynamic susceptibility contrast (DSC) magnetic resonance imaging (MRI) perfusion have aimed to predict *MGMT* methylation status non‑invasively in gliomas with radiological characteristics. The possibility of predicting *MGMT* methylation status using DSC‑MRI perfusion with a radiological approach remains controversial. The present study aimed to evaluate the usefulness of MRI perfusion parameters as non‑invasive markers to predict *MGMT* methylation status and prognosis in newly diagnosed glioblastoma patients. Thus, 50 patients with histologically confirmed primary glioblastoma, *IDH*‑wildtype who underwent tumor resection at Osaka University Hospital (Suita, Japan) between January 2017 and January 2023 were included in this study. The mean cerebral blood volume (CBV) ratio (rCBV) and cerebral blood flow (CBF) ratio (rCBF) for tumors with *MGMT* methylation (mean rCBV:2.09 and mean rCBF:3.08) were significantly higher compared with those for tumors without *MGMT* methylation (mean rCBV:1.33 and mean rCBF:1.85; P<0.05). While patients with *MGMT* methylation had longer progression‑free survival (PFS) compared

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with those without *MGMT* methylation (P<0.05), there was no significant difference in OS with or without *MGMT* methylation  $(P=0.06)$ . By contrast, there was no association between MRI perfusion parameters and OS or PFS in patients with glioblastoma. Furthermore, the association between CBV, CBF, *MGMT* promotor methylation status, OS, and PFS were explored. There was no significant prognostic difference between low vascularity tumors (rCBV <1.3 or rCBF <1.8) with or without *MGMT* methylation. On the other hand, high vascularity tumors (rCBF ≥1.8) with *MGMT* promotor methylation were associated to longer OS and PFS compared with those without. However, there was no association between *MGMT* methylation status and OS or PFS in patients with high rCBV (rCBV  $\geq$ 1.3). The present study indicated that CBV and CBF could be used to predict the *MGMT* methylation status in glioblastomas. However, the prognostic value of tumor vascularity and *MGMT* methylation status may be limited.

### **Introduction**

Glioblastoma is the most common type of malignant brain tumor. The prognosis of glioblastoma is extremely poor, even with standard treatments, such as chemoradiotherapy. O6‑methylguanine DNA methyltransferase *(MGMT)*  promoter methylation is associated with favorable outcomes after temozolomide (TMZ) chemotherapy in patients with newly diagnosed glioblastoma (1). Thus, the evaluation of *MGMT* methylation status is important for the treatment of these patients.

Several radiological studies have shown the potential of conventional magnetic resonance imaging (MRI) to predict the *MGMT* methylation status using image texture and deep learning architectures (2‑7). Dynamic susceptibility contrast (DSC) MRI offers insight into tumor tissue vascularity by analyzing perfusion. Previous radiological studies using

*Key words:* dynamic susceptibility contrast-enhanced perfusion magnetic resonance imaging, *MGMT* promoter methylation, glioblastoma

MRI perfusion have aimed to predict *MGMT* methylation status noninvasively in gliomas using radiological characteristics (5,8‑11) and radiomics (12‑15). Some reports revealed that DSC‑MRI could be used as a noninvasive technique to predict genetic mutations preoperatively without surgical specimen, and to determine molecular characteristics such as *IDH* mutation status and methylation status of the *MGMT* promoter in glioblastomas (5,8‑10,16). In contrast, other reports have indicated that cerebral blood volume (CBV) did not differ significantly between tumors with methylated or unmethylated *MGMT* (17,18). The possibility of predicting *MGMT* methylation status from DSC‑MRI perfusion using a radiological approach remains controversial, and there is no expert consensus regarding clinical use.

The potential impact of DSC-MRI perfusion in the prediction of *MGMT* methylation status in glioblastoma remains disputed. In previous reports, elevated CBV has been associated with decreased survival of glioblastoma patients (19‑22). In contrast, *MGMT* methylation status is highly correlated with survival in glioblastomas with moderate vascularity, but not in those with high vascularity (17,23). Furthermore, patients with glioblastomas showing stable or increasing CBV following chemoradiotherapy experienced significantly improved PFS, particularly in those cases presenting *MGMT* methylation (24).

This study aimed to evaluate the possibility of using DSC‑MRI perfusion as a non‑invasive method to predict *MGMT* methylation status and prognosis in newly diagnosed glioblastoma patients.

## **Materials and methods**

*Study design and patient selection.* This retrospective study was approved by the Clinical Research Review Committee of Osaka University (Approval No. 22302). The inclusion criteria were as follows: patients who i) have definite pathological results; ii) have MRI images available, including conventional MRI and DSC-MRI, before surgery; iii) did not undergo radiotherapy or chemotherapy before MRI examination; iv) have an available *MGMT* promoter methylation status. Patients with recurrent tumors, tumors with unsatisfactory images, and young patients aged less than 18 years old were excluded from the study. Fifty patients with histologically confirmed primary glioblastoma, *IDH*‑wildtype (according to the 2021 World Health Organization International Histological Classification of Tumors) who underwent tumor resection at our institution between January 2017 and January 2023 were included in the study (34 men and 16 women; median age, 70.5 years; Table I). All patients were diagnosed according to the 2021 guidelines, regardless of the resection date. DSC‑MRI and conventional MRI pulse sequences were acquired preoperatively for all patients. All patients underwent surgical resection with concomitant TMZ treatment and radiotherapy, followed by adjuvant TMZ treatment. Tumor samples were collected after resection.

*Magnetic resonance imaging.* All images, including axial T1-, T2-, and T2\*-weighted images, fluid-attenuated inversion recovery, and contrast-enhanced T1-weighted sequences (T1Gd) were obtained using a 3‑T MR unit (DISCOVERY MR Table I. Characteristics of patients with glioblastoma.



750; GE Healthcare, Milwaukee, WI, USA) with a 32‑channel head coil. Perfusion MRI was performed using a T2\*‑weighted, single-shot, gradient-recalled, echo-planar imaging (GRE EPI) sequence. The perfusion MRI sequence parameters were as follows: repetition time/echo time, 2000/13.3 ms; matrix,  $128x128$ ; flip angle, 60; section thickness, 5 mm; and acquisition time, 90 sec. The contrast, a standard dose of 0.1 mmol/kg body weight of meglumine gadoterate (Guerbet Japan, Tokyo, Japan), was injected at a rate of 2‑3 ml/s, followed by saline flush using a power injector.

*Imaging analysis.* Imaging analysis was performed using Synapse Vincent (Fuji Medical Systems, Tokyo, Japan) in perfusion mode. A single region of interest (ROI) with a diameter of 5 mm (Fig. 1) was set manually on the solid part in each of the enhancing tumor regions from every patient, avoiding areas of cyst formation, hemorrhage, and large vessels, as per previous reports (16,25,26), and the values of CBV, cerebral blood flow (CBF), and mean transit time (MTT) were calculated. The ROI was manually set in the contralateral normal area for each enhanced tumor. The contralateral area was normal and contained no tumor infiltration.

Disease‑to‑normal ratios were calculated by dividing the values of CBV, CBF, and MTT for the tumors by the values of







the contralateral normal area, and described as rCBV, rCBE, and rMTT, respectively.

Genomic DNA extraction. Tumor samples were immediately frozen and stored at ‑80˚C or immersed in RNAlater Stabilization Solution (Thermo Fisher Scientific, Waltham, MA). Genomic DNA was extracted using a DNeasy Blood & Tissue Kit (Qiagen, Valencia, CA, USA) or NucleoSpin Tissue (Macherey-Nagel, Düren, Germany), as described previously (27).

*MGMT promoter methylation analysis.* The methylation status of the *MGMT* promoter (accession number: NM\_002412.5) was assessed using quantitative methylation-specific PCR (qMSP). Purified DNA was subjected to bisulfite modification by an EZ DNA Methylation‑Gold Kit (Zymo Research, Irvine, CA), according to the manufacturer's instructions. The qMSP was performed on a QuantStudio12K Flex Real‑Time PCR System (Thermo Fisher Scientific) with POWER SYBR® Green PCR Master Mix (Thermo Fisher Scientific). The bisulfite-modified DNA was amplified using specific primers for each methylated or unmethylated molecule as listed in Table II. Real-time PCR conditions were as follows: 95°C for 10 min followed by 45 cycles of 95˚C for 10 sec, and 60˚C for 60 sec. The quantification of methylated and unmethylated sequences was performed by employing the standard curve method as previously described. In the dissociation curve analysis, heterogeneity of the amplified methylated and unmethylated molecules was assessed from melting temperature. The mean  $\pm$  standard deviation of methylation value was calculated from triplicate PCRs. We used a 1% cut-off value for the determination of *MGMT* methylation based on an outcome‑based study of newly diagnosed GBMs as mentioned in our previous publications (27,28). Sequences of primers used for quantitative methylation‑specific PCR are provided in Table II.

*Statistical analysis.* Statistical analyses were performed using Prism version 9 (GraphPad Software, San Diego, CA, USA). Results were considered statistically significant at a P*‑*value of <0.05. The unpaired t‑test was used for comparisons between two groups. Receiver operating characteristic (ROC) curve analysis was performed to compare the performance of each imaging parameter based on each ROI in distinguishing tumors with *MGMT* methylation from those without *MGMT* methylation. The Kaplan‑Meier method was used to derive OS and PFS curves.

We also attempted to construct a model based on three perfusion parameters to determine *MGMT* methylation status in glioblastomas by performing multiple logistic regression.

## **Results**

*Perfusion MRI parameters and MGMT methylation status.*  The mean rCBV for tumors with *MGMT* methylation (2.09; range, 0.72–5.14) was significantly higher than that for tumors without *MGMT* methylation (1.33; range, 0.51-2.78; P<0.005). The mean rCBF for tumors with *MGMT* methylation (3.08; range, 0.49-11.03) was significantly higher than that for tumors without *MGMT* methylation (1.85; range, 0.65‑4.77; P<0.05). In contrast, the rMTT for tumors with and without *MGMT* methylation did not differ (Fig. 2, Table III).

Receiver operating characteristic (ROC) analysis showed that the rCBV [area under the curve (AUC)=0.7484] and rCBF (AUC=0.6883) were more effective in distinguishing between tumors with and without *MGMT* methylation than the rMTT (AUC=0.5406; Fig. 3).

We attempted to construct a model based on three perfusion parameters to determine *MGMT* methylation status in glioblastomas by performing multiple logistic regression. The following predictive formula, created using parameters derived from the multiple logistic regression, was obtained to estimate the probability of *MGMT* methylation (probability range: 0 to 1) for each ROI:

log\_odds=0.01832 + 4.743 \* rCBV + 1.034 \* rCBF + 4.214 \* rMTT odds=exp(log\_odds)

Probability=odds/(ones(size(odds)) + odds)

*Prognosis according to MGMT methylation status.* The PFS and OS were 15.0 months and 24.9 months, respectively, in the patients with *MGMT* methylation, and 8.5 months and 15.2 months, respectively, in the patients without *MGMT* methylation (Fig. 4 and Table IV). Patients with *MGMT* meth‑ ylation had longer PFS than those without *MGMT* methylation (P<0.05), but there was no significant difference in OS between patients with and without *MGMT* methylation (P=0.06).

*Prognosis according to perfusion MRI parameters.* In contrast, there was no association between perfusion MRI parameters and OS or PFS in patients with glioblastoma (Fig. 5 and Table IV).



Table III. Correlation between MRI perfusion parameters and *MGMT* promoter methylation status in patients with glioblastoma.

CBV, cerebral blood volume; CBF, cerebral blood flow; MTT, mean transit time.



Figure 1. Tumor in right basal ganglia: (A) Contrast-enhanced T1-weighted image; (B) cerebral blood volume; (C) cerebral blood flow. The blue circle stands for the region of interest delineated.



Figure 2. Boxplots of (A) rCBV), (B) rCBF) and (C) rMTT in tumors with or without *MGMT* methylation. Box plots show statistical differences in rCBV and rCBF in tumors with or without *MGMT* methylation. By contrast, rMTT for tumors with and without *MGMT* methylation did not differ. rCBV, cerebral blood volume ratio; rCBF, cerebral blood flow ratio; rMTT, MTT ratio; MGMT, O6‑methylguanine DNA methyltransferase.

*Prognosis according to MGMT methylation status and perfu‑ sion MRI parameters.* The study investigated the significance in PFS and OS differences between the following two groups: low vascularity tumors with *MGMT* methylation and low vascularity tumors without *MGMT* methylation (Figs. 6, 7 and Table IV). Juan‑Albarracín *et al* reported that significant differences were observed in the Kaplan‑Meier estimated survival functions for

populations divided based on the median rCBV and rCBF (29). They indicated that the median rCBV and rCBF were found to be the relevant prognostic markers in patients with glioblastoma. Previous studies assessed the combined role of tumor vascularity, estimated from perfusion MRI, and *MGMT* methylation status on OS in patients with glioblastoma (17,23). The classification of tumor vascularity was based on the median rCBV





Figure 3. Receiver operating characteristic curve showed more reliable predictions that distinguished between tumors with and without *MGMT* methylation in glioblastomas in terms of (A) rCBV, (B) rCBF and (C) rMTT. rCBV, cerebral blood volume ratio; rCBF, cerebral blood flow ratio; rMTT, MTT ratio; MGMT, O6‑methylguanine DNA methyltransferase; AUC, area under the curve.



Figure 4. Kaplan‑Meier (A) OS and (B) PFS curves of the patients with glioblastoma grouped according to *MGMT* methylation status. The patients with *MGMT* methylation (n=22) had longer PFS compared with those without *MGMT* methylation (n=28) (P<0.05), but there was no significant difference between patients with and without *MGMT* methylation in OS. OS, Overall survival; PFS, progression-free survival; MGMT, O6-methylguanine DNA methyltransferase.

and rCBF values reported by Juan‑Albarracín' *et al* (17,23). We validated thresholds calculated from the current study cohort based on previous reports (17,23,29) and defined the vascular groups using the median rCBV and rCBF. There was no significant association between *MGMT* methylation status and prognosis in patients with low vascularity tumors (rCBV <1.3 or rCBF <1.8). We also evaluated differences in PFS and OS in high vascularity tumors (rCBV  $\geq$ 1.3 or rCBF  $\geq$ 1.8) with methylated and unmethylated *MGMT* promoters. There was no association between *MGMT* methylation status and OS or PFS in patients with high rCBV (rCBV  $\geq$ 1.3). On the other hand, high vascularity tumors (rCBF ≥1.8) with *MGMT* methylation were associated to longer OS and PFS compared to those without *MGMT* methylation (P<0.05).

## **Discussion**

Our study indicates that CBV and CBF can be used to predict the *MGMT* methylation status in glioblastomas. According to our results, the rCBV and rCBF in tumors with *MGMT*

methylation were higher than those in tumors without *MGMT* methylation. The possibility of predicting the *MGMT* methylation status from DSC‑MRI using a radiological approach remains controversial. In previous reports, the CBV derived from DSC‑MRI of glioblastomas with a methylated *MGMT* promoter were reported to be lower than those corresponding to glioblastomas with unmethylated *MGMT* (8,10,16). In contrast, other reports have indicated that CBV does not differ significantly between tumors with methylated and unmethylated *MGMT* (17,18). Using stereotactic image-based histological validation, Song *et al* reported that CBF showed no statistically significant differences between gliomas with and without *MGMT* promoter methylation (30). Perfusion parameters are influenced by the location of the tumor in relation to major blood vessels, heterogeneous vascularization of the tumor, tumor necrosis, and intratumoral cystic changes. The DSC‑MR perfusion technique is known to be affected by the partial volume effect caused by adjacent tissues. Contouring ROI, excluding necrosis and proximate vascular structures, reduces the partial volume effect caused by adjacent

Variable	No. of cases	<b>MST</b>	P-value (log-rank)	<b>PFS</b>	P-value (log-rank)
MGMT promoter methylation status					
Methylated	22	24.9	0.06	15.0	0.03
Unmethylated	28	15.2		8.5	
CBV ratio					
< 1.3	23	19.5	0.53	8.3	0.72
$\geq 1.3$	27	14.7		11.3	
CBF ratio					
$<1.8$	24	22.8	0.10	8.6	0.86
$\geq1.8$	26	13.6		10.9	
CBV ratio $< 1.3$					
MGMT methylated	5	45.7	0.15	15.7	0.23
MGMT unmethylated	18	17.3		8.0	
CBV ratio $\geq 1.3$					
MGMT methylated	17	24.9	0.07	15.0	0.06
MGMT unmethylated	10	13.1		8.6	
CBF ratio $< 1.8$					
MGMT methylated	5	NA	0.22	32.6	0.09
MGMT unmethylated	19	19.5		8.6	
CBF ratio $\geq 1.8$					
MGMT methylated	17	24.9	0.01	15.0	0.04
MGMT unmethylated	9	10.1		8.1	

Table IV. Univariate analyses of median survival time and PFS of patients with glioblastoma.

NA, not applicable.



Figure 5. Kaplan‑Meier survival curves of the patients with glioblastoma grouped according to MR perfusion imaging parameters: (A) rCBV OS and (B) PFS (CBV ≥1.3, n=27; rCBV <1.3, n=27); (C) rCBF OS and (D) PFS (rCBF ≥1.8, n=26; rCBF <1.8, n=24). There was no association between each of the MR perfusion imaging parameters and not only OS but also PFS in patients with glioblastoma. OS, Overall survival; PFS, progression-free survival; MGMT, O6‑methylguanine DNA methyltransferase; rCBV, cerebral blood volume ratio; rCBF, cerebral blood flow ratio.





Figure 6. Kaplan-Meier survival curves of the glioblastoma patients grouped according to *MGMT* methylation status and tumor vascularity based on rCBV: (A)  $rCBV < 1.3$  with *MGMT* methylation (n=5); (B)  $rCBV < 1.3$  without *MGMT* methylation (n=18); (C)  $rCBV \ge 1.3$  with *MGMT* methylation (n=17); (D)  $rCBV \ge 1.3$ without *MGMT* methylation (n=10). There was no association between *MGMT* methylation status and prognosis in lower and higher rCBV. OS, Overall survival; PFS, progression-free survival; MGMT, O6-methylguanine DNA methyltransferase; rCBV, cerebral blood volume ratio; rCBF, cerebral blood flow ratio.



Figure 7. Kaplan–Meier survival curves of the glioblastoma patients grouped according to *MGMT* methylation status and tumor vascularity based on rCBF: (A)  $rCBF < 1.8$  with *MGMT* methylation (n=5); (B)  $rCBF < 1.8$  without *MGMT* methylation (n=19); (C)  $rCBF \ge 1.8$  with *MGMT* methylation (n=17); (D)  $rCBF \ge 1.8$ without *MGMT* methylation (n=9). There was no association between *MGMT* methylation status, OS and PFS in lower rCBF. However, patients with *MGMT* methylation had longer OS and PFS compared with those without *MGMT* methylation in higher vascularized tumors (rCBF ≥1.8; P<0.05). OS, Overall survival; PFS, progression-free survival; MGMT, O6-methylguanine DNA methyltransferase; rCBV, cerebral blood volume ratio; rCBF, cerebral blood flow ratio.

tissues. As mentioned in previous reports (16,25,26), ROIs were drawn to avoid calcification, blood products, dense bone, or large vessels to ensure the accuracy of the measurements. The size of the ROI for the solid part in our study was smaller than that in previous studies (16,25,26). As glioblastomas are heterogeneous tumors, the ROIs in our study were accurately set on the solid part, which contained only enhancing tumor core lesions in each tumor region (Fig. 1) to exclude the effect of tumor heterogeneity. Therefore, rCBV and rCBF may be affected by the definition of the size of ROI. There is a possibility that the prediction of *MGMT* methylation status could be heavily affected by the method used for ROI design. However, it is still unclear whether rCBV and rCBF were affected by the small ROI or the *MGMT* methylation. Although it is desirable to perform a regression analysis to clarify whether ROI or *MGMT* methylation factors were corrected, this makes it very difficult to perform the mentioned analysis, since multiple ROIs or ROIs of different sizes were not set throughout the course of our study.

Meanwhile, Hegi *et al* have suggested that the methylation status of the *MGMT* promoter may have prognostic value and, additionally, may be a clinically relevant predictor of the benefit of TMZ chemotherapy (1). HIF‑1 was discovered as a molecular target associated with intratumoral hypoxia (31). As previously demonstrated, HIF-1 $\alpha$  silencing dramatically increases sensitivity to TMZ *in vivo* (32). Tang *et al* showed that the inhibition of HIF‑1α through knock‑down sensitizes glioma cells to TMZ, with a decrease in MGMT expression (33). Persano er al. showed that HIF-1 $\alpha$  suppression promotes the downregulation of MGMT, and this is sufficient to override glioblastoma resistance to TMZ (34). In the present study, glioblastomas with *MGMT* promoter methylation showed higher rCBV and rCBF than those without. Glioblastomas with maintained perfusion and oxygenation levels may have suppressed HIF-1 $\alpha$ expression and downregulated MGMT expression, and may be susceptible to TMZ treatment.

In contrast, whether MRI perfusion parameters correlate with the prognosis of glioblastoma remains controversial. Previous studies have shown that CBV (19‑22,35) and CBF (36) have prognostic value. However, no significant association between overall survival time and CBV has been reported in previous studies (37,38). The prognostic correlation between CBV and *MGMT* methylation status may be influenced by conditions such as tumor vascularity and treatment‑induced changes over time. Previous studies have shown a highly significant impact of *MGMT* status on the prognosis of patients with moderately vascularized tumors, but not in patients with highly vascularized tumors (17,23). Goldman *et al* reported that treatment‑induced changes in CBV affect the prognosis of glioblastoma (24). They reported that glioblastomas that showed stable or increasing CBV following chemoradiotherapy were associated to a significantly improved PFS compared to those with decreased CBV following chemoradiotherapy, particularly in those exhibiting *MGMT* methylation (24). Batchelor *et al* found that patients with glioblastoma treated with chemoradiotherapy plus cediranib demonstrated an increase in perfusion and significantly improved survival compared to patients treated with chemoradiotherapy alone. This effect may be due to anti‑angiogenic therapy, normalization of blood flow, and enhancement of drug delivery (39). It has been assumed that the methylation of the *MGMT* promoter induced by the maintained CBV and improved oxygenation enhanced the therapeutic benefits of alkylating agents. In our study, highly vascularized tumors based on rCBF with *MGMT* methylation were associated to longer OS and PFS than those without *MGMT* methylation. High CBF tumors may be less hypoxic, leading to *MGMT* promoter methylation, and improved prognosis with TMZ treatment. The failure to observe a significant difference in OS with and without *MGMT* methylation can be attributed to the small sample size, which reduced the power (40,41). This can be seen from the P-value of 0.06, which is very close to the significance level. Conversely, even with such a low detection power, a significant difference in OS can be confirmed between patients with and without *MGMT* methylation who have more highly vascular tumors (rCBF ≥1.8), which may suggest the idea that *MGMT* methylation status has a stronger effect on OS in cases with higher rCBF. Radiological diagnosis using rCBV and rCBF has the potential to predict *MGMT* methylation status preoperatively, without reliance on surgical specimens. In our study, there was no association between perfusion MRI parameters and OS or PFS in patients with glioblastoma. Furthermore, there was no significant association between *MGMT* methylation status and prognosis in patients with lower vascularity tumors based on both the rCBV and the rCBF and those with more highly vascularized tumors based on rCBV. The measurement of cerebral blood perfusion in DSC‑MRI is based on the assumption that gadolinium‑based contrast agents do not cross the blood‑brain barrier. CBV is calculated by the tissue signal change caused by the gadolinium‑based contrast agent and the arterial input function. Based on the assumption that the gadolinium‑based contrast agents do not cross the blood‑brain barrier, the CBV changes caused by the gadolinium‑based contrast agents are thought to be due to the gadolinium‑based contrast agent stored in the capillaries. However, this assumption does not hold in glioblastoma tumor tissues where the blood‑brain barrier has been disrupted. The value of the CBV calculated by the model described above is ambiguous (42). Conversely, CBF is calculated by dividing CBV by MTT (CBF=CBV/MTT), where MTT is the time taken for the tracer to pass through the region of interest. Thus, CBF compensates somewhat for the blood-brain barrier breakdown. It is possible that calculated values with such technical 'corrections' more sensitively reflect tumor characteristics. It is not certain that the combination of MRI perfusion parameters with *MGMT* methylation status can be used to predict the prognosis of glioblastomas. We are skeptical that the combination of perfusion MRI parameters with *MGMT* methylation status can be used to predict the prognosis of glioblastomas.

A few limitations and caveats in the current study should be noted and addressed. As previously mentioned, this study was limited by its small sample size, leading to potential bias in our results. First, as the ROIs in our study were accurately set on the solid part in each enhancing tumor region to exclude the effect of tumor heterogeneity while avoiding areas of cyst formation, hemorrhage and large vessels, the size of the ROIs was smaller than that in previous studies (16,25,26). Therefore, there is a possibility that the prediction for *MGMT* methylation



status may have been heavily affected by the method used for ROI design. Second, while the methylation status of the *MGMT* promoter may have prognostic value, there was no significant difference in the OS of patients with and without *MGMT* methylation in our study. Future large-scale studies are required to validate the proposed prognostic value of CBF and *MGMT* methylation status.

In conclusion, we aimed to evaluate whether DSC-MRI could be employed as a non‑invasive method to predict *MGMT* methylation status and prognosis in newly diagnosed glioblastoma patients. Our study indicates that rCBV and rCBF can be used to predict the *MGMT* methylation status preoperatively, offering the possibility to change clinical management in patients affected by glioblastoma. However, we are not certain that the combination of MRI perfusion parameters with *MGMT* methylation status can be used to predict prognosis in these patients.

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## **Availability of data and materials**

The data generated in the present study are not publicly available due to them containing information that could compromise research participant privacy/consent but may be requested from the corresponding author.

## **Authors' contributions**

YO conceived and designed the study. DC, YO, RU, HiK, RH, NoK, NaK, YK and HaK acquired the data. DC, YO and SY analyzed and interpreted the data and drafted the manuscript. AA and NT contributed to the methodology for radiological analysis. DC and YO confirmed the authenticity of all the raw data. All authors provided critical revision of the manuscript for important intellectual content. All authors read and approved the final manuscript.

## **Ethics approval and consent to participate**

The study was conducted according to the guidelines of the Declaration of Helsinki and approved by the Ethics Committee of Osaka University Hospital (approval no. 22302). Written informed consent was obtained from all patients.

## **Patient consent for publication**

Patients provided written informed consent for publication of their data.

## **Competing interest**

The authors declare that they have no competing interests.

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