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Low detection of H5N1 virus in commercial chickens with a low-level of vaccination coverage against H5N1 virus infection in Bangladesh

Sukanta Chowdhury^{1*} , Mohammad Enayet Hossain¹, Rashedul Hasan², Mojnu Miah³, Sajal Kanti Biswas¹, Md Mahmudul Hasan⁴, Probir Kumar Ghosh¹, Jenifar Quaiyum Ami¹, Akash Saha¹, Sumon Ghosh⁵, Mahmudur Rahman⁶, Fahmida Chowdhury¹ and Mohammed Ziaur Rahman¹

Abstract

Background Bangladesh has reported > 560 H5N1 outbreaks in poultry and eight human cases since 2007. Commercial chicken farms were mostly affected. Commercial chicken farms across the country use imported vaccines against H5N1 virus; however, these vaccines did not use local circulatory isolates of H5N1 virus. Vaccination may have limited effectiveness in chicken because of the mismatch in terms of subtypes and clades. To test this, we conducted a mixed-method study to assess the impact of ongoing vaccination against H5N1 virus on H5N1 viral shedding through freshly dropped feces of chickens raised in commercial farms that exclusively vaccinated or did not vaccinate their chickens.

Methods Initially, we collected vaccination coverage data from all active farms in a subdistrict of each of eight division. In each district, 25 vaccinated and 25 non-vaccinated chicken farms were selected randomly for sample collection. All samples were tested to detect avian influenza viruses using rRT-PCR.

Results A total of 5092 poultry farms were surveyed; among them 1284 (25%) chicken farms administered vaccine against H5N1 virus. In total 21 of 400 tested farms (5%) had chicken feces samples that tested positive for AIVs; of these three were positive for H5N1 subtype of clade 2.3.2.1. Out of three H5N1 positive farms, 1 (33%) was vaccinated and 2 (67%) were unvaccinated. The chicken farms that administered vaccine against H5N1 was found protective for the detection of H5N1 viral RNA (aOR 0.39, 95% CI: 0.32–0.48). The H5N1 isolates of clade 2.3.2.1 sequenced in this study formed a cluster with the vaccine strain A/duck/Guangdong/S1322/2010 (H5N1) [Re-6].

Conclusions The overall low vaccination coverage with low detection of H5N1 virus in commercial chickens makes it difficult to assess the effectiveness of the vaccine in reducing H5N1 viral shedding.

Keywords HPAI, H5N1, Commercial chicken, Vaccine, Bangladesh

*Correspondence:

Sukanta Chowdhury
sukanta@icddr.org

Full list of author information is available at the end of the article



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Introduction

Highly pathogenic avian influenza (HPAI) viruses can cause severe infections in poultry and humans [1–3]. Enhanced biosecurity measures, stamping out, and movement restrictions have been implemented conventionally to control H5N1 epidemics in poultry [4]. The HPAI H5N1 virus is endemic in several Asian countries, including China, Egypt, Indonesia, India, Hong Kong, Vietnam and Bangladesh, where vaccination programs against HPAI viruses in poultry have been implemented [5]. However, there is ongoing debate among scientists about vaccine effectiveness as HPAI outbreaks have been reported among poultry in countries that vaccinate [5–11].

In 2012, the Government of Bangladesh approved vaccines against A(H5N1) in commercial chickens, and now vaccines are available throughout the country [12]. Three different types of vaccines have been imported from Europe; RE-6 and H5N2 inactivated vaccine, which are used in commercial chickens, and the rHVT-H5 vectored vaccine, which is used in day-old-chicks [13]. The proportion of chicken farms that are being vaccinated in Bangladesh has not been reported. The lack of information on vaccination coverage indicates inadequate monitoring. Viruses from Bangladesh were not used in these imported vaccines. RE-6 inactivated vaccine contains clade 2.3.2.1b of influenza A(H5N1) viruses, A(H5N2) inactivated vaccine contains A/Potsdam/1986(H5N2) virus; and the rHVT-H5 vectored vaccine contains clade 2.2 of A(H5N1) viruses [13]. Though different types of H5N1 virus clades are circulating in poultry in Bangladesh since 2007, the predominant clade of circulating H5N1 virus was 2.3.2.1 since 2011 [14, 15]. The virus isolates used in these vaccines may not be matched with local influenza A(H5N1) strains perfectly, and that may produce suboptimal protection in commercial chickens. This lower level of immunity may exert evolutionary pressure on the virus, and such newly emerged strains may pose a risk for the emergence of novel strains to the human population [13]. Vaccination has also raised concerns about the possibility of difficult to detect asymptomatic spread of HPAI viruses in chickens with some immunity.

To ensure high levels of herd immunity, 60–80% of H5N1 vaccine coverage is needed in a chicken population [7, 16]. In Bangladesh, there is a need to estimate the vaccination coverage in commercial chicken farms and investigate the impact of the current H5N1 vaccination programs on H5N1 viral shedding to the environment and its possible impacts on the diversity of circulating avian influenza viruses. This study estimated coverage of avian influenza vaccine in poultry farms in Bangladesh and assessed the impact of ongoing H5N1 vaccination

on avian influenza virus shedding including H5N1 virus to the environment and examined molecular changes of avian influenza viruses isolated from vaccinated and unvaccinated chickens.

Methods

Study sites

We conducted this study in eight divisions (Dhaka, Chattogram, Rajshahi, Khulna, Barishal, Sylhet, Rangpur, and Mymensingh) of Bangladesh from June 2021 to February 2022. In each division, we identified a sub-district with the largest number of poultry farms were surveyed for baseline data collection (Fig. 1). Samples were collected from selected poultry farms. Poultry farm statistics were collected from the Department of Livestock Services to identify eight sub-districts across eight divisions for the survey.

Study design

Initially, we conducted a baseline survey to collect information on H5N1 vaccination coverage of commercial poultry farms. With the local government livestock officials' help, the study field team visited every commercial poultry farm in the eight selected sub-districts with a large number of commercial poultry farms to collect data. We defined poultry farms as those farms that raise chicken, duck, geese, turkey, pigeon or quail in an intensive farming system (raising a large number of poultry in controlled environments). Commercial chicken farms included commercial layer, commercial broiler, parent stock/breeder, and Sonali chicken. Sonali is the crossbred between Rhode Island Red cocks and Fayoumi hens [17]. Farm size was categorized as scale small (<1000 poultry), medium (1001–2000 poultry) and large (>2000 poultry) [17]. The field staff interviewed selected farm owners to collect data using a structured questionnaire. Questionnaire included two sections to collect data on farm demographics and avian influenza vaccination status [including A(H5N1) and A(H9N2)].

Based on the baseline survey data, commercial chicken farms were categorized into two groups; vaccinated and unvaccinated. On the day of the farm visit, those farms that reported vaccinating their current chicken flocks against A(H5N1) or A(H5N2) viruses were categorized as having vaccinated flocks. Commercial chicken farms having only a history of vaccine intake at hatchery or breeder farms against H5N1 or H5N2 was not considered for this study. Chicken flocks with obvious evidence of vaccination history against the H5N1 or H5N2 were prioritized for this study. Considering the above-mentioned criteria, we prepared a final list of vaccinated chicken farms for each of eight sub-districts. Another list was prepared for unvaccinated farms for each sub-district. Those chicken

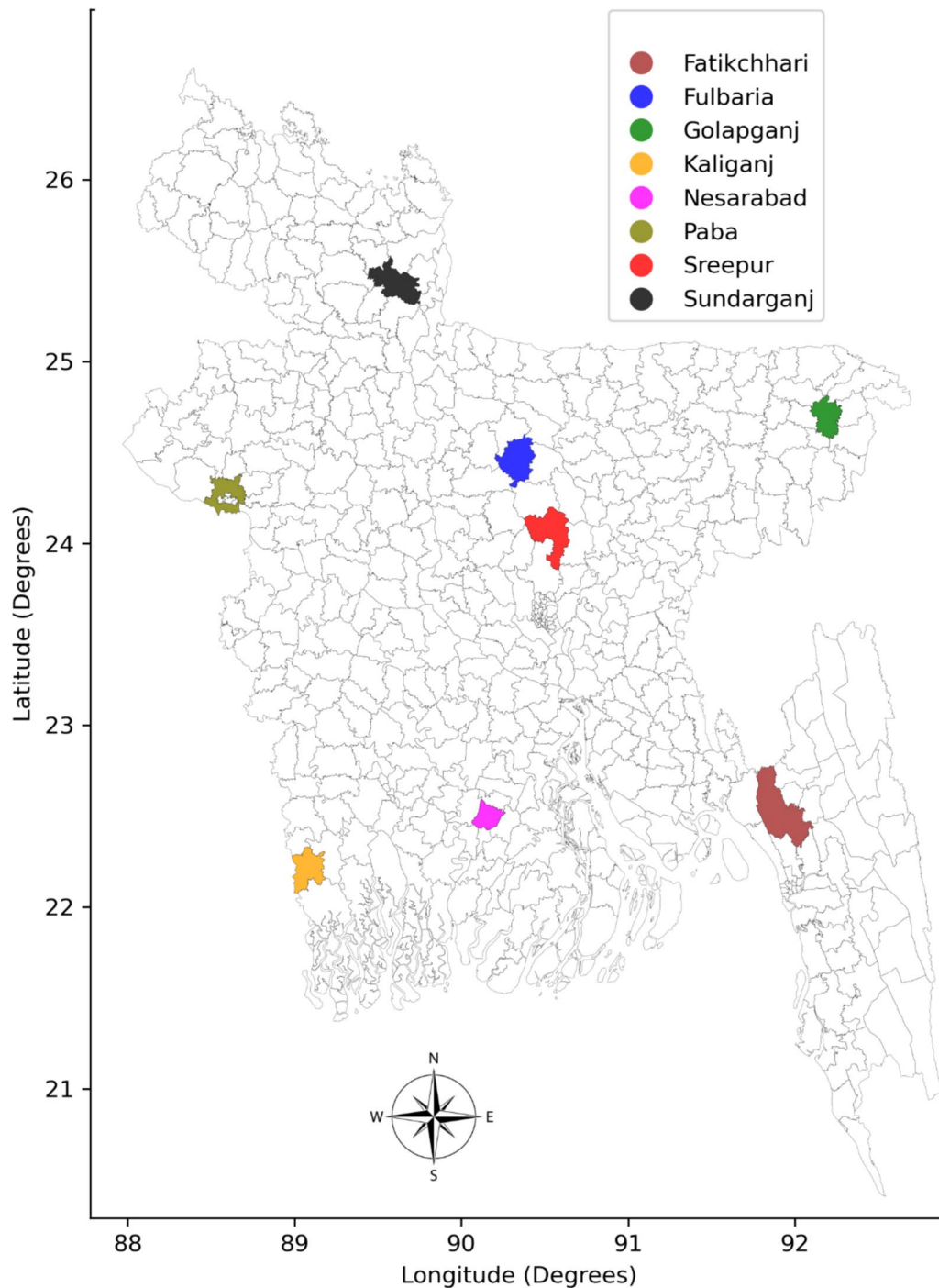


Fig. 1 Study sites for baseline data collection of commercial poultry farms

farms that have never administered the vaccines against A(H5N1) or A(H5N2) or A(H9N2) in their current chicken flocks or did not administer vaccine at the hatchery was considered unvaccinated chicken farms.

A total of 400 (200 vaccinated and 200 unvaccinated) farms were identified from eight sub-districts under

eight divisions through a two-stage selection process: matching and simple randomization. In each sub-district, total 50 farms (25 vaccinated and 25 unvaccinated) were enrolled for sample and further data collection. In the first step, geographical location (same sub-district) and production systems (layer or broiler)

were used to match vaccinated and unvaccinated farms. After farm matching, simple random sampling was used to identify pairs of vaccinated and unvaccinated farm. Every selected farm was visited twice fortnightly for fecal sample collection between December 2021 and February 2022. In each visit, 10 swab sticks were used to collect freshly laid feces from 10 chickens of the selected flock, and the 10 swabs were then combined to prepare a fecal pool sample. A total of 800 fecal pool were collected from 400 selected farms. In vaccinated flock, swabs were taken at least 14 days after vaccination.

Detection and sequencing of avian influenza viruses

All collected samples were tested to detect avian influenza viruses subtypes H5, H7 and H9. The icddr, b (International Centre for Diarrheal Diseases Research, Bangladesh) One health laboratory extracted total RNA from 100 μ l of each pool swab specimen from the 800 collected pool samples in VTM, using RNase kit (Qiagen, Cat 74106) according to the manufacturer's instruction. The extracted RNA was screened for detection of matrix gene of influenza A viruses using the FDA-approved TaqMan qRT-PCR (real-time reverse transcription polymerase chain reaction) assay [18]. Primers and probes specific for matrix (M) gene were included to detect any of the 16 types of avian influenza A viruses. To identify influenza A H5, H7 and H9 subtypes, hemagglutinin (HA) gene specific-primers and probes were used [18]. Full genome sequencing was performed for selected influenza A-positive samples using Illumina MiSeq technology, as previously described [19]. All sequences were uploaded to the Global Initiative on Sharing All Influenza Data.

Phylogenetic analyses

The HA reference nucleotide sequences of H5 and H9 subtypes were obtained from the online repository Global Initiative of Sharing All Influenza Data (GISAID) and Bacterial and Viral Bioinformatics Resource Center (BV-BRC) [20]. Full-length reference sequences of the vaccine strains related to H5 and H9 subtypes of HA genes were included for phylogenetic analysis. These sequences were aligned using the ClustalW program within BioEdit software [21]. The alignment was then utilized to construct a phylogenetic tree using the Maximum Likelihood approach using MEGA v.7.0, employing the General Time Reversible (GTR) substitution model with gamma distribution rates [22]. The replicability of the maximum-likelihood tree at each node was assessed by bootstrap analysis of 1000 replications.

Statistical analysis

We performed descriptive analysis to summarize the farm demographics, vaccination status and biosecurity practices of commercial chickens. Flock morbidity and mortality were estimated for vaccinated and unvaccinated farms separately. The proportion of avian influenza viruses (AIVs) and AIV (H5) for vaccinated and unvaccinated flocks were estimated separately. Chi-square test was performed to estimate the significance difference of different avian influenza viruses subtypes occurrence between vaccinated and unvaccinated flock. We estimated odds ratios (OR) and 95% confidence intervals (CI) to identify associations between vaccination status and AIV (H5) detection. The variables with a p -value of <0.05 in univariate analysis were selected for multivariate analysis. Backward stepwise selection of variables with a significance level of 0.05 was used to construct final models. Variables were retained or removed from the model after considering the p -value of <0.05 . We estimated the adjusted odds ratios (aOR) using multivariate logistic regression, combining all significant variables.

Results

Demographic characteristics of surveyed poultry farms

A total of 5092 farm owners were interviewed from eight sub-districts under eight divisions to collect baseline information. Majority of the farms were enrolled from Sreepur (18%, $n=898$). Majority of the enrolled farms reared layer chicken (50%, $n=2565$), followed by broiler chicken (35%, $n=1802$) and Sonali chicken (10%, 486). Most of the farms (52%, $n=2664$) were small in size having less than 1000 poultry. According to the farmers self-report, more than 75% ($n=3876$) of the farms were not registered by Government and 46% of the farmers had experience for chicken farming from 1 to 5 years. The average chicken population per farm was comparatively higher in layer farms ($n=1862$) than broiler (1099) and Sonali (1248) farms (Table 1).

Vaccination coverage against H5N1 virus

Overall, more than 95% of the poultry farms initially surveyed reported using some type of vaccines. Vaccine coverage varied by vaccine type (e.g. avian influenza, Newcastle disease, fowl pox and Marek's) and production type (e.g. broiler, layer and Sonali). Overall, 1284 (25%) farms administered vaccine against AIV/H5 virus in their current chicken flocks. Among the vaccinated farms, 1276 (99%) were layer chicken farms. Of the 2565 surveyed layer chicken farms, 1276 (50%) farms used vaccine against H5N1 virus in their current batch. The vaccination against H5N1 virus in broiler and Sonali chicken

Table 1 Demographic characteristics of poultry farms, Bangladesh, June 2021 to February 2022 (N = 5092)

Characteristics	n (%)	95% CI
Sub-district wise poultry farms enrollment		
Sreepur	898 (18)	17–19
Fulbaria	894 (18)	17–19
Fatikchhari	647 (13)	12–14
Sundarganj	616 (12)	11–13
Paba	576 (11)	10–12
Kaliganj	549 (11)	10–12
Nesarabad	521 (10)	9–11
Golapganj	391 (8)	7–8
Types of poultry farms		
Broiler chicken	1802 (35)	34–37
Layer chicken	2565 (50)	49–52
Sonali chicken	486 (10)	9–10
Duck and Geese	189 (4)	3–4
Native chicken	22 (1)	0–1
Others	28 (1)	1–2
Farm size		
Small (≤ 1000 poultry)	2664 (52)	51–54
Medium (1001–2000 poultry)	1633 (32)	31–33
Large (> 2000 poultry)	795 (16)	15–17
Number of registered farms by Government		
Yes	1216 (24)	23–25
No	3876 (76)	75–77
Farm longevity		
≤ 3 years	1381 (27)	26–28
4–6 years	1386 (27)	26–28
7–9 years	756 (15)	14–16
10–12 years	844 (17)	16–18
≥ 13 years	725 (14)	13–15
Farming experience by farmers		
1–5 years	2352 (46)	45–48
6–10 years	1616 (32)	30–33
11–20 years	1026 (20)	19–21
> 20 years	98 (2)	1–2
Average number broiler chicken per farms, mean, (SD, range)	1099 (± 1587 , 100–50000)	
Average number layer chicken per farms, mean, (SD, range)	1862 (± 2456 , 79–60000)	
Average number Sonali chicken per farms, mean, (SD, range)	1248 (± 1215 , 100–10000)	
Average number duck per farms, mean, (SD, range)	251 (± 257 , 24–2600)	
Average numbers of farm workers per farm, mean, (SD, range)	2.3 (± 1.02 , 1–24)	

farms was extremely low ($n=8$). A substantial number of layer chicken farms (24%, $n=607$) reported of using vaccine against H9N2 virus (Table 2).

Characteristics of sampled chicken farms

A total of 400 chicken farms (200 vaccinated and 200 unvaccinated) were selected from eight divisions. Most of vaccinated (49%, $n=197$) and unvaccinated (49%,

$n=195$) farms were layer chicken farms, and the remainder were Sonali farms. Three different categories of farms such as small ($n=151$), medium ($n=136$) and large ($n=113$) scale were included in vaccinated and unvaccinated arms. Most of the chickens in both vaccinated (41%, $n=164$) and unvaccinated (37%, $n=149$) arms were aged less than 12 months. Average number of layer chickens per farm in vaccinated and unvaccinated arms

Table 2 History vaccination in poultry farms, Bangladesh, June 2021 to February 2022 (N = 5092)

Vaccination characteristics	Number of farms performed vaccination		p value
	Yes n(%)	No n(%)	
Previously used vaccines to poultry			
Administered any vaccine	4892 (96)	200 (4)	< 0.001
Administered vaccine against avian influenza	1311 (26)	3781 (74)	
Administered vaccine against Newcastle disease	4770 (94)	322 (6)	
Administered vaccine against infectious bursal disease	4501 (88)	591 (12)	
Administered vaccine against fowl pox	2310 (45)	2782 (55)	
Administered vaccine against fowl cholera	2310 (45)	2782 (55)	
Administered vaccine against Marek's	2243 (44)	2849 (56)	
Administered vaccine against duck plague	154 (3)	4938 (97)	
Administered vaccine against H5N1 virus to their current commercial chicken flock			
Broiler	2 (1)	1802 (100)	< 0.001
Layer	1276 (50)	1289 (50)	
Sonali	6 (1)	480 (99)	
Administered vaccine against H9N2 virus to their current commercial chicken flock			
Broiler	-	1802 (100)	< 0.001
Layer	607 (24)	1958 (76)	
Sonali	2 (1)	484 (100)	

was 2454 and 1550, respectively. The average number of Sonali chicken per farm in vaccinated and unvaccinated arms was 1683 and 1316, respectively (Table 3).

Detection, identification and sequencing of AIVs in vaccinated and unvaccinated farms

Of the 800 fecal pools, avian influenza A viral RNA was detected in 24 (3%) pooled samples. Overall, 21 different farms (5%) had at least one fecal pool sample test positive for avian influenza. Samples from three farms were confirmed for H5 subtype specific viral RNA, 17 farms were confirmed for H9 subtype specific viral RNA and sample from one farm remained unsubtypable (not positive for H5, H7 or H9). Of the three H5 positive farms, one had history of vaccination against H5N1 virus. Among the 17 H9 positive farms, seven farms had history of vaccination against H5N1 virus (Table 4). A total of 28 avian influenza positive samples were sequenced; two of them were identified as H5N1 subtype and 26 samples were H9N2 subtype (Fig. 2, Supp. Figures 1 and 2).

Phylogenetic analysis

Phylogenetic analysis of the H5 sequences indicated that the viruses belonged to clade 2.3.2.1 and formed a close cluster together with the vaccine strain A/duck/Guangdong/S1322/2010 (H5N1) [Re-6] but not closely related with other vaccine strains (Fig. 1). Phylogenetic analysis of the H9N2 strains identified in this study, when compared with the vaccine strain (Ceva, a G1-like Middle

East H9N2 isolate), suggests that the isolates formed distinct clusters, indicating their genetic diversity (Supp. Figure 2).

Association between avian influenza virus detection and vaccination status

In univariate analyses, chicken farms that had a history of vaccination against H5N1 virus (OR 0.49, 95% CI: 0.44–0.55) were less likely to be positive for avian influenza A/H5 viral RNA compared with farms that did not have these characteristics. In multivariate regression analysis, chicken farms that administered vaccine against H5N1 to chicken (aOR 0.39, 95% CI: 0.32–0.48) was found protective for the detection of H5N1 viral RNA. In the multivariate regression model, clustering due to geographical locations of farms was adjusted along with other demographics and biosecurity variables (Table 5).

Discussion

This study explored the H5N1 vaccination coverage in eight sub-districts under eight divisions in Bangladesh. The vaccination coverage against H5N1 virus in their current chicken flock was below the optimal standard (60–80% vaccination coverage). The low level of vaccination coverage reported by this study raised a concern about the effectiveness of ongoing vaccination programme for significant reduction of H5N1 virus transmission in poultry population, as previous studies indicate that achieving 60–80% vaccination coverage is necessary to maintain

Table 3 Demographic characteristics of sampled poultry farms, Bangladesh, June 2021 to February 2022 (N=400)

Characteristics	Vaccinated farms n (%)	Unvaccinated farms n (%)
Division wise poultry farms enrollment		
Dhaka	25 (13)	25 (13)
Chattogram	25 (13)	25 (13)
Sylhet	25 (13)	25 (13)
Rajshahi	25 (13)	25 (13)
Khulna	25 (13)	25 (13)
Barishal	25 (13)	25 (13)
Rangpur	25 (13)	25 (13)
Mymensingh	25 (13)	25 (13)
Types of poultry farms		
Layer chicken	197 (49)	195 (49)
Sonali chicken	3 (1)	5 (1)
Farm size		
Small (≤ 1000 poultry)	64 (16)	87 (22)
Medium (1001–2000 poultry)	64 (16)	72 (18)
Large (> 2000 poultry)	72 (18)	41 (10)
Farm longevity		
≤ 12 months	164 (41)	149 (37)
13–24 months	35 (9)	47 (12)
≥ 25 months	1 (1)	4 (1)
Farming experience by farmers		
1–5 years	64 (16)	70 (18)
6–10 years	66 (17)	73 (18)
11–20 years	60 (15)	54 (14)
> 20 years	10 (3)	3 (1)
Average number layer chicken per farms, mean, (SD, range)	2454 (± 3983 , 200–50000)	1550 (± 1412 , 71–10000)
Average number Sonali chicken per farms, mean, (SD, range)	1683 (± 1013 , 180–3000)	1316 (± 1176 , 550–2500)

high levels of flock immunity [7, 16, 23]. Among H5N1 endemic countries, Hong Kong reported the highest vaccination coverage at 86%, followed by Egypt at 70%, Vietnam at 52%, Mongolia at 51%, and China at 47% between 2002 and 2010 [24]. To our knowledge, this is the first study to assess the H5N1 vaccination coverage. Stakeholders in commercial chicken production should focus on enhancing H5N1 vaccination coverage. Additionally, it is crucial for policymakers and health officials in both human and animal health to ensure that H5N1 vaccination does not introduce any further risks to human health.

The effectiveness of vaccines against avian influenza strains relies on strong cross-reactivity with the cocirculating avian influenza virus strains [25]. Interestingly, all H5 vaccines used in Bangladesh were imported by local pharmaceutical companies and none of these vaccines contained viruses isolated locally [12, 13]. The mismatch between field viruses and vaccine strains may produce suboptimal level of immunity that trigger

genetic changes and the evolution of new avian influenza viruses that may pose a risk for the emergence of novel strains to the human population. Phylogenetic analysis of the H5 sequences revealed that the viruses belonged to clade 2.3.2.1 and formed a close cluster together with the vaccine strain A/duck/Guangdong/S1322/2010 (H5N1) [Re-6]. The decline of clade 2.2 and EA-nonGsGD after administration of rHVT-H5 vectored vaccine and Potsdam/1986 H5N2 inactivated vaccine, respectively indicates a significant reduction in H5N1 lineage diversity in vaccinated poultry [26]. However, two H5N1 isolates of clade 2.3.2.1 in this study survived post-vaccination of RE-6 inactivated vaccine containing clade 2.3.2.1 suggesting the possibility of selection and persistence of the viruses of this clade. Evolutionary changes in A(H5N1) viruses were also observed in countries using vaccines against A(H5N1) when compared to countries that never used A(H5N1) vaccine in poultry [8]. In Vietnam, low protection levels were detected in ducks and high viral shedding occurred more than 7 days after an

Table 4 Farm level avian influenza virus detection in chickens from vaccinated and unvaccinated farms, Bangladesh, January 2022 to February 2022 (N=400)

Divisions	H5N1 Vaccination status	Number of farms tested	No. (%) of farms positive for influenza A	No. (%) of farm positive for influenza A/H5	No. (%) of farms positive for influenza A/H9
Dhaka	Vaccinated	25	2	-	2
	Unvaccinated	25	1	-	1
Chattogram	Vaccinated	25	-	-	-
	Unvaccinated	25	-	-	-
Sylhet	Vaccinated	25	-	-	-
	Unvaccinated	25	2	-	2
Rajshahi	Vaccinated	25	3	1	3
	Unvaccinated	25	6	2	5
Khulna	Vaccinated	25	1	-	1
	Unvaccinated	25	-	-	-
Barishal	Vaccinated	25	-	-	-
	Unvaccinated	25	1	-	1
Rangpur	Vaccinated	25	2	-	1
	Unvaccinated	25	1	-	1
Mymensingh	Vaccinated	25	1	-	1
	Unvaccinated	25	1	-	1
Total		400	21 (5%)	3 (1%)	19 (5%)

A(H5N1) challenge with mortality [9]. In Egypt, surveillance detected continuous circulation of avian influenza viruses including the A(H5N1) viruses in vaccinated commercial and backyard poultry [10]. Another study in Egypt revealed imported vaccine-mediated antigenic drift evolution of influenza A (H5N1) viruses in vaccinated commercial chickens [11].

Over the study period, we detected only three farms tested positive for HPAI virus (H5N1). This result indicates that farm level H5N1 circulation was very low in both vaccinated and unvaccinated farms during the sampling period. The ongoing live bird market based avian influenza surveillance detected comparatively higher number of poultry (3%) positive for H5 subtype during 2021–2022 (personal communication). This difference suggests that live bird markets may be more likely to sustain circulation of AIVs than farms. The low detection of H5N1 virus found in this study limit our efforts to evaluate the effectiveness of avian influenza vaccine.

Our research results suggested that layer chicken farmers demonstrated a greater inclination to administer the H5N1 vaccine in comparison to those who raise broilers and Sonali chickens. Like our study findings, difference in vaccination coverage was reported in three different countries during 2004 to 2009. In Egypt, broiler chicken (78%) was mostly vaccinated than layer chicken (32%) [24]. In Pakistan, the highest H5 vaccination coverage was recorded in breeder chicken (26%) compared to layer

chicken (3%) and no vaccination to broiler chicken [24]. On the contrary, the highest vaccination coverage was reported by Vietnam in meat duck (90%) than broiler chicken (52%) [24]. The high vaccination coverage was observed in layer chicken farms in this study could be due to the significant investment in layer chicken farms, longer lifespans of layer chicken compared to broilers, and heightened awareness among layer farmers compared to those raising broiler and Sonali chickens.

Bangladesh has reported only a few H5N1 outbreaks in poultry since 2013, compared to the period between 2007 and 2012 [12, 27]. This low number of H5N1 outbreaks could be due the implementation of countrywide vaccination in commercial chicken farms. In contrast, the ongoing live bird market based avian influenza surveillance has consistently detected AIVs in poultry from 2007 to the present day. These inconsistent outcomes have raised questions about the efficacy of existing H5N1 vaccination efforts, the reliability of outbreak reporting systems, and the role of live bird markets in sustaining the transmission of avian influenza viruses.

This study had several limitations. Only commercial layer and Sonali chicken farms were sampled to detect viral shedding. We did not find any broiler farms that had history of H5N1 vaccination during sampling. Therefore, this study findings only represent the effectiveness of H5N1 vaccination in layer and Sonali chicken. Detection of very low number of farms for H5N1 virus limit

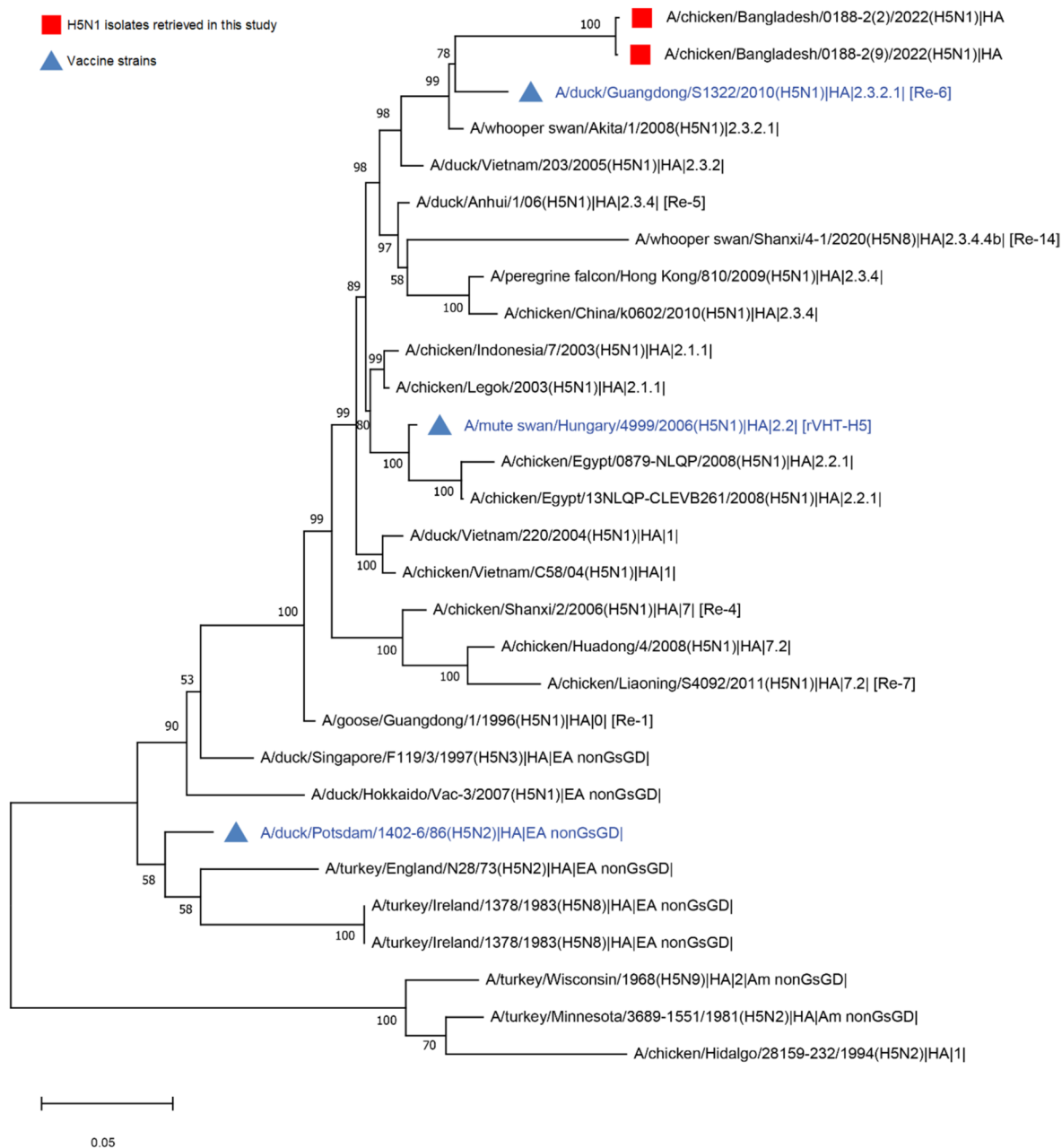


Fig. 2 The phylogenetic tree based on the maximum likelihood approach depicts the relationship of H5N1 isolates sequenced in this study with reference sequences of vaccine strains representing different clades. The analysis involved 29 sequences, with a final dataset comprising a total of 1672 aligned positions. Red squares represent the isolates retrieved in this study, whereas blue triangles indicate the vaccine strains used or licensed for use in Bangladesh. The tree is drawn to scale, indicating a substitution rate of 0.050 per nucleotide position

our efforts to evaluate the effectiveness of avian influenza vaccine.

Conclusions

The H5N1 vaccination coverage was below standard and limited to only layer chickens. The detection of the H5N1 virus was lower in both vaccinated and

unvaccinated chicken flocks, which constrained the study’s ability to evaluate the vaccine’s effectiveness in reducing HPAI H5N1 viral shedding. Phylogenetic analysis suggests Re-6 vaccine can protect current circulatory H5N1 virus better than other available vaccines. Regular monitoring is needed to assess the vaccination coverage, impact of ongoing avian influenza

Table 5 Farm level avian influenza subtype H5 detection, H5N1/H9N2 vaccination status and other biosecurity measures January 2022 to February 2022 (N=400)

Variables	No. (%) farms	No. (%) farms positive for avian influenza A/H5	OR (95% CI)	P value	Adjusted OR (95% CI)	P value
H5N1 Vaccination status						
No	200	2	Ref.			
Yes	200	1	0.49 (0.45–0.55)	< 0.001	0.39 (0.32–0.48)	< 0.001
H9N2 Vaccination status						
No	349	3	Ref.			
Yes	51	0	Undefined			
Types of chicken						
Exotic layer chicken	393	3	Ref.			
Sonali	7	-	Undefined	-		
Farm size						
Small (\leq 1000 chicken)	151	-	Ref.			
Medium (1001–2000 chicken)	136	3	Undefined			
Large ($>$ 2000 chicken)	113	-	Undefined			
Presence of fence around farm						
No	241	3	Ref.			
Yes	159	0	Undefined			
Cleaning farm premises						
Daily	72		Ref.			
Every alternative day	193	2	Undefined			
Weekly	49		Undefined			
Fortnightly	17	1	Undefined			
Monthly	69		Undefined			
Disinfection inside farm premises						
Daily	53		Ref.			
Every alternative day	120		Undefined			
Weekly	187	2	Undefined			
Fortnightly	11		Undefined			
Monthly	22	1	Undefined			
No disinfection	7		Undefined			
Drainage system						
Nonfunctional or absent	246	3	Ref.			
Functional	154	0	Undefined			
Slaughtering chicken within farm yard						
No	249	1	Ref.			
Yes	151	2	3.33 (2.42–4.56)	< 0.001	3.36 (2.36–4.77)	< 0.001
Using separate shoes during entering inside farm						
No	56	1	Ref.			
Yes	344	2	0.32 (0.15–0.67)	0.003	0.34 (0.14–0.85)	< 0.022
Raising multiple poultry species adjacent to the farm						
No	396	3	Ref.			
Yes	4	0	Undefined			
Access within farm yard by outside transports						
No	89	0	Ref.			
Yes	311	3	Undefined			

vaccination on viral shedding, genetic reassortment and emergence of new influenza strains of public health importance.

Supplementary Information

The online version contains supplementary material available at <https://doi.org/10.1186/s42522-024-00119-3>.

Supplementary Material 1: Supplementary Fig. 1. The phylogenetic tree based on the maximum likelihood approach shows the relationship of H9N2 isolates sequenced in this study, as indicated by red squares. The sequences of previously reported H9N2 viruses from Bangladesh are indicated by blue squares. The analysis involved a total of 64 sequences, with a final dataset comprising 1682 aligned positions. The tree is drawn to scale, indicating a substitution rate of 0.050 per nucleotide position. Supplementary Fig. 2. The phylogenetic tree based on the maximum likelihood approach illustrates the relationship between the H9N2 isolates of this study and the commercially available H9N2 vaccines, which are developed from specified vaccine strains. The H9N2 vaccines currently marketed in Bangladesh are denoted by blue triangles. This analysis incorporated 41 sequences in total, resulting in a final dataset of 1469 aligned positions. The tree is drawn to scale, representing a substitution rate of 2.00 per nucleotide position.

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Authors' contributions

S.C., M.E.H., S.K.B., S.G., M.R., F.C. and M.Z.R. participated in conceptualization, methodology, laboratory work, and preparation of the manuscript. S.C., M.E.H., S.K.B., S.G., P.K.G., M.R., F.C. and M.Z.R. participated in methodology, supervision and manuscript editions. S.C., M.E.H., R.H., M.M., M.M.H., J.Q.A., A.S. and M.Z.R. contributed to the supervision of laboratory work, data analysis and manuscript editions. All authors have read and approved the final manuscript for publication.

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Data availability

Data are available and shared from the corresponding author upon reasonable request.

Declarations

Ethics approval and consent to participate

This protocol was approved by the Institutional Review Board of icddr, b (PR-20140). All the respondents above 18 years of age were asked to provide informed, written consent to participate in this study.

Consent for publication

Not applicable.

Competing interests

The authors have no conflict of interest to declare.

Author details

¹International Centre for Diarrheal Diseases Research, Bangladesh (icddr,b), Dhaka, Bangladesh. ²Department of Chemistry and Biochemistry, New Mexico State University, Las Cruces, NM, USA. ³Department of Biochemistry and Molecular Biology, University of Arkansas for Medical Sciences, Little Rock, AR 72205, USA. ⁴The Bredesen Center for Interdisciplinary Research

and Graduate Education, The University of Tennessee, Knoxville, USA. ⁵Department of Public Health, The University of Tennessee, Knoxville, USA. ⁶EMPHNET, Dhaka, Bangladesh.

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