

# The Trend in Mitigation Strategies of 3-Monochloropropane-1,2-diol and Glycidyl Esters in Edible Vegetable Oils: Today and Tomorrow

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Received: 14 June 2023  
Accepted: 8 August 2024



## SUMMARY

3-Monochloropropane-1,2-diol (3-MCPD) and its esters, which have carcinogenic and genotoxic effects, are contaminants induced by high-temperature that have been detected in refined oils and fatty foods. 3-MCPD esters, chlorinated propanols, were first identified in 1978 in acid-hydrolysed vegetable proteins used as flavour enhancers in many foods. Glycidyl esters (GE) are contaminants that can occur in edible oils during heat treatment and are formed mainly during the deodorisation phase of refining. The International Agency for Research on Cancer has classified 3-MCPD as a 'potential carcinogen for humans' in group 2B. Glycidol has also been classified as group 2A with mutagenic and carcinogenic properties, *i.e.* 'probably carcinogenic to humans'. In addition, glycidol has been classified by the International Agency for Research on Cancer (IARC) as a 'possible human carcinogen' (group 2A). Toxicological studies have shown that these genotoxic and carcinogenic contaminants induced by heat treatment are released in the gastrointestinal tract and cause the formation of tumours. In this review the mechanisms of formation, toxicological effects of 3-MCPD and GE on human health, and methods of their detection are shown. The latest strategies to mitigate and prevent 3-MCPD and GE formation during crude oil production, refining and beyond are also discussed.

**Keywords:** mitigation of 3-MCPD and GE; detection in edible oils; genotoxic and carcinogenic contaminants; reduction of precursor formation

## INTRODUCTION

Fats and oils are macronutrients that can be consumed directly or used in the preparation of food (1–3). Today, vegetable oils constitute 80 % of the fat and oil production worldwide (2,4). Edible oils are obtained from plant seeds or pulp by mechanical, thermal or chemical methods (2). Some foods undergo heat treatment during production, which can lead to the formation of contaminants that can have toxic effects on the consumers (5). Fats and oils are refined to improve their sensory properties and consumer acceptance (2,6). Since higher temperatures are applied during these refining steps, the process temperature is a decisive factor for the sensory, physical and chemical properties of the refined oils (7).

During oil refining, free fatty acids and minor contaminants such as pesticides and polycyclic aromatic hydrocarbons, as well as undesirable flavour and colour, are removed or reduced (6,8). As a side effect, undesirable compounds (*trans* fatty acids, acylglycerol polymers, cyclic fatty acid esters and 3-MCPDE) can be formed in oils at high temperatures used for oil refining (9–12). 3-MCPDE and GE, which have carcinogenic and genotoxic effects, are among the contaminants detected in refined oils (especially palm oil) and fatty foods (13–19). 3-MCPDE consists of acylglycerols released at temperatures above 200 °C in the presence of organic or inorganic chlorinated compounds (2). These compounds then react with chloride ions and are converted into 3-MCPDE (20).

3-MCPDE and GE are formed in vegetable oils especially during the deodorisation step, when their amount highly increases (3,17,21–23). They are relatively higher in palm oil than in refined vegetable oils such as soybean, canola and sunflower (16,24–26).

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Studies have shown that chloropropanols can be formed both in ester form (linked) and in free form (8,23). The free form of 3-MCPDE was first identified by Velíšek *et al.* in acid-hydrolysed vegetable proteins, which are used as flavour enhancers in many foods (14,16,24). It was first discovered in 1980 by Davidek *et al.* (27) in the ester form. In 2001, the European Commission Scientific Committee on Food (SCF) set the maximum content for 3-MCPD in hydrolysed vegetable protein (HVP) and soy sauce at 20 µg/kg (28). In 2006, Zelinková *et al.* (29) found that vegetable fats and oils contain high amounts of 3-MCPDE formed during refining. Later, the tolerable daily intake was set by JECFA (FAO/WHO Joint Experts Committee on Food Additives) and Scientific Committee on Food at 2 µg per kg of body mass (25).

The latest European Commission Regulation set the limit value for GE in vegetable oils at 1 mg/kg (26). Over time, these contaminants have been detected in most processed foods such as meat, milk, grain products, malt products, coffee, fried cheese, smoked foods and biscuits (2,8,13,15,17,28,29). The study carried out by Wöhrlin *et al.* (30) showed that 3-MCPDEs can even be found in infant food formula. It is confirmed that high amounts of 3-MCPDE in foods, especially in refined vegetable oils and fats, can cause health problems due to their toxicity in the free form (3). According to the report of The German Federal Institute for Risk Assessment (BfR), 3-MCPD is completely degraded from its esters in the gastrointestinal tract (3). Some studies have also shown that these compounds change from ester to free forms (17,31,32). In addition, studies by Abraham *et al.* (33) and Andres *et al.* (34) reported that the relative bioavailability of 3-MCPDE was 86%. According to the Joint FAO/WHO Expert Committee on Food Additives (JECFA), the main target organ of 3-MCPD toxicity is the kidneys, causing nephropathy, tubular hyperplasia and adenomas (35,36). The International Agency for Research on Cancer (IARC) classifies 3-MCPDE as carcinogenic compounds (31,32,37,38). Studies carried out on experimental

mice have shown that 3-MCPDE in free form have nephrotoxic effects and cause the formation of tumours. The International Agency for Research on Cancer classified 3-MCPD as a 'potential carcinogen for humans' in group 2B (39). On the other hand, glycidol with mutagenic and carcinogenic properties was classified in group 2A and labelled as 'most likely carcinogenic to humans' (17,40–42).

In the last decade, scientific interest in MCPDE and GE has increased. A literature search was carried out in the Web of Science database using the keywords 'MCPD' and 'glycidyl esters' for the period between 2000 and 2024. The results were sorted according to the relevance of the search criteria. Those related to disciplines other than food technology were excluded. If the proceedings are ignored, access to 529 distinct published journal articles is possible. The published articles were categorised into five main topics, as shown in Fig. 1. The topic that received the most attention was the toxicity, bioavailability and adverse health effects of 3-MCPD and GE. In addition, several articles focused on the methods of analysis and validation. Since 2011, the number of publications has increased significantly, and it stands out as the year with the highest number of published review papers. The blue bar shows the number of studies published on the mitigation of 3-MCPD and GE in oils and fats. Among the newly developed methods, the combination of ultrasound to accelerate the esterification part and n-hepta-fluorobutyrylimidazole (HFBI) derivatisation during dispersive liquid-liquid microextraction (DLLME) has a positive effect on both cost and environmental friendliness (43).

Foods and refined vegetable oils containing 3-MCPD and its esters, which have carcinogenic and mutagenic effects, can be consumed directly. International committees and studies have demonstrated the adverse effects of these contaminants on health. For this reason, it is crucial to prevent the formation of 3-MCPD and its esters in refined vegetable oils and foods or to reduce their amounts. Unlike other

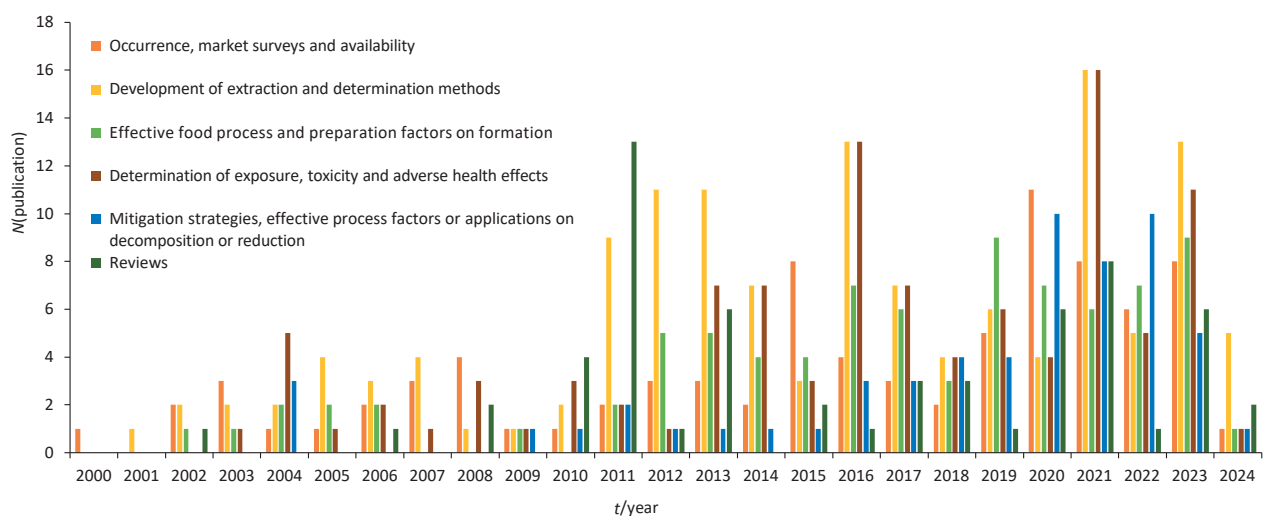


Fig. 1. Distribution of published scientific articles indexed in different categories in the Web of Science database between 2000 and 2024

studies on this topic, this review provides detailed data on the formation of 3-MCPDE and their toxic effects, analytical extraction and determination methods, reduction strategies, preventive/reduction procedures that can be applied in the extraction and refining of oils and fats, and current regulations. Even though we found 529 articles in the literature search, we only selected and summarised recently published studies on specific topics. While we have primarily summarised recently published studies, we have also cited older, important studies that provide relevant background information on the topic.

## TOXICITY OF 3-MCPD AND GE, AND RELATED REGULATIONS

3-Monochloropropane-1,2-diol (3-MCPD) and (2,3-epoxy-1-propanol) glycidol and their esters (3-MCPDE and GE), contaminants in food processing that are harmful to human health, are formed during the refining of vegetable fats and oils, especially during deodorisation at high temperatures. Previous studies have shown that these components can be found in baby food, margarine, coffee, soy sauce, bakery products, soup, meat, milk, malt products, fried cheese and smoked foods. 3-MCPD and GE have been declared as ingredients that jeopardise food safety worldwide and cause numerous health problems. Studies have shown that 3-MCPD, when consumed regularly in substantial amounts, impairs male fertility, kidney function and body mass of rats (Fig. 2) (3,44–46). 1,3-DCP, the ester of 3-MCPD, has been found to be a genotoxic, hepatotoxic and carcinogenic substance that is classified as a human carcinogen (46). The International Agency for Research on Cancer (IARC) has defined 3-MCPD as a group 2B 'substance with possible carcinogenic effects on humans' and GE as a group 2A 'substance with possible carcinogenic effects on humans' (47). Studies on the potential toxic effects of 3-MCPDE and GE have not yet been completed and many unknowns on this topic remain. Various toxicological studies have reported that long-term consumption of high

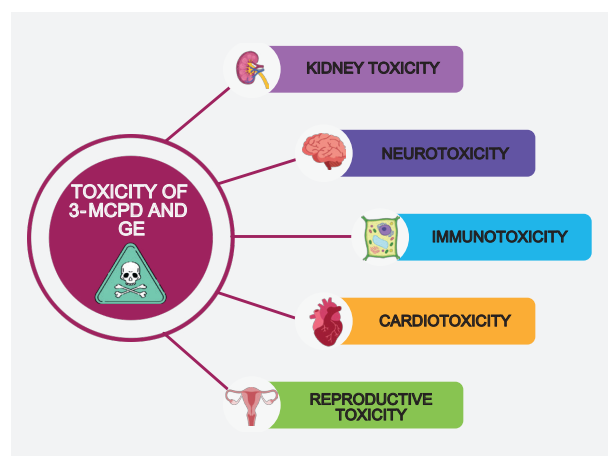


Fig. 2. Toxicity of 3-monochloropropane-1,2-diol (3-MCPD) and glycidyl ester (GE)

doses of these compounds leads to nephritic and reproductive organ failure and hyperplasia in particular (3,25,33,34,36,48–50). In a study on mice, neurotoxic effects such as paralysis were observed after short-term exposure to a high dose of 50 mg/kg per day (49). Almost all 3-MCPDE have been reported to convert to the free form under the influence of lipase in the gastrointestinal tract (GIT), which causes toxicity (34,51–55). Similarly, GE are predominantly hydrolysed to the genotoxic carcinogen glycidol, which can mediate detrimental effects in the gastrointestinal tract (21,55). In addition, 3-MCPD has been found to cause severe proteinuria and glycosuria as well as acute glomerulonephritis, renal and testicular tumourigenesis in mice (56–59). Studies on mice have shown that free 3-MCPD is not absorbed by intestinal cells but passes through the cell surface and causes renal tubular carcinoma and Leydig cell tumour formation in mature male rats, effectively in the kidney and reproductive system (34,48,52). One of the potential toxicity mechanisms of 3-MCPD is its oxidative metabolism to chlorolactaldehyde and chlorolactic acid, which causes oxidative stress and disrupts the glycolytic pathway and energy production (60). GE, which are listed in group 2A, have been classified as genotoxic compounds in *in vivo* and *in vitro* studies (21,42,49). In a study in rats, glycidol administered at a dose of 200 mg/kg per day for 28 days did not cause neurotoxicity but it was reported to cause renal toxicity at repeated doses of 150–400 mg/kg (61–63). Even the lowest dose of GE exposure affected the reproductive organs in rats, and EFSA reported that daily intake of 25 mg/kg caused a 36 % decrease in sperm count (49). Another toxicological study in rats revealed that exposure to 3-MCPD and GE caused lung and heart toxicity. It has also been reported that exposure to glycidol can lead to ovarian dysfunction, colon disease and several carcinomas (55). Another study also reported that lung lesions, including alveolar oedema and hyperaemia or haemorrhagic areas, were observed in Swiss mice exposed to 3-MCPD (36).

The report published by EFSA (49) in 2016 includes the chemical structures, formation mechanisms, amounts in different foods, direct and indirect analysis methods and toxicological properties of 3-MCPD and GE. EFSA has set the maximum content of 3-MCPD in soy sauce and hydrolysed vegetable proteins at 20 µg/kg (25). Additionally, the JECFA set the daily tolerable intake of 3-MCPD amount per body mass at 4 mg/kg (35). Due to the carcinogenic and genotoxic effects of glycidol, EFSA has recommended the application of a 'limit/margin of exposure (MoE)' approach for glycidyl esters and reported that foods with MoE greater than 25 000 should be considered risky. In this context, the regulation was amended in 2018 to specify that the GE value in vegetable oils and baby/child food should not exceed 1000 µg/kg (25,49).

Moreover, the European Union published a new regulation effective on 1 January 2021 (64). According to this regulation, the maximum amount of 3-MCPD and GE in oils should be 1.25 and 1 mg/kg, respectively. It was emphasised that the

mass fractions of 3-MCPD and GE in baby products should not exceed 0.75 and 0.5 mg/kg, respectively. It has been reported that maximum amounts of 3-MCPD and GE in infant formula, namely in follow-on milk in liquid form should not exceed 0.125 and 0.05 mg/kg, and in follow-on milk in powder form should not exceed 0.015 and 0.006 mg/kg, respectively (64).

## THE EFFECTS OF THE PRECURSORS ON THE FORMATION OF 3-MCPDE AND GE

It has been reported that for the formation of 3-MCPD and esters, hydrolysed triacylglycerols (TAG) should be primarily degraded into glycerol, mono- and diacylglycerols (MAG/DAG) and then converted to MCPD in the presence of chloride ions of glycidyl esters (GE) together with the cyclic acyloxonium molecule, which is formed from these substances at high temperature (57,65). The studies and the International Life Sciences Institute (ILSI) identified MAG, DAG and chloride ions as precursors for the formation of these compounds and determined the process temperature and duration as the effective parameters for the formation of these compounds (21,66–78). In edible vegetable oils, MAG and DAG are formed during maturation, harvesting or transportation of oily fruit seeds from TAGs by lipase enzyme activity, and they are also formed during high-temperature deodorisation phase during oil refining (72,73). Therefore, DAG and MAG contents in different oils or the same oils obtained from different locations may also differ (41). Fruit oils are more sensitive to hydrolysis reactions than seed oils due to their excess water. Hence, DAG content in the fruit oils is higher than in the seed oils. Palm oil obtained from palm fruits, for example, can have a DAG content between 4 and 12 % (74). High rates of MAG and DAG, which are formed due to the hydrolysis reaction, lead to the formation of 3-MCPD and GE (75). In their study, Zelinková *et al.* (29) suggested that there could be a correlation between the DAG content and the MCPD amounts in refined edible oils. The reason for this is that the highest MCPD contents were found in fruit oils such as olive and palm oil. It is known that fruit extract oils have a higher DAG content than seed oils due to the faster formation of lipolytic reactions during harvesting (77,78). The studies have shown that DAGs, precursors of 3-MCPD and GE, have a stronger effect than MAGs under deodorising conditions. Compared to the same amount of mono- and diglycerides, the formation of 3-MCPD was found to be about twice as high as that of diglycerides (5,18). Another study found that the GE content in oils with high DAG content was 12 to 43 times higher than that in edible oils containing mainly TAGs (79). The formation of GE is directly related to high temperatures and the contact time at these temperatures, and they can be formed from diacylglycerols without requiring the presence of chlorinated compounds. In addition, the highest GE content was found in refined palm oil compared to other vegetable oils (20,77). Masukawa *et al.* (79) found that the GE content was 10 times higher in refined commercial oils with high DAG content (87 %) than in oils with low DAG content (3.9–6.8 %). In addition,

Destailats *et al.* (80) found that both MAG and DAG can cause the formation of GE during thermal treatment. It has been reported that GE is mainly present in refined palm oil and palm-based fractions and the GE content increases in direct proportion to the duration and temperature of the deodorisation process (6,20,29).

3-MCPD is formed by the reaction between chloride and lipid sources. This chloride source can either be the food itself or the material that comes into contact with it, which can be chlorinated water or additional salt during production (18, 34,81–85). The mineral chlorine is abundant in nature. It is known that organic and inorganic chloride sources are the potential precursor compounds in the formation of MCPD. Vegetable oils contain chloride compounds in addition to TAG, DAG, MAG and free fatty acids. Destailats *et al.* (80) determined that many inorganic chlorine sources were found in higher amounts (mg/kg) in crude palm oil than in other vegetable oils. Chlorides from formed during crude oil pressing and extraction, and free hydrogen chlorides from the bleaching earth pass into the oil (86). The type, quality, composition and production conditions of the crude oil also affect the chlorine quantity, as do MAGs and DAGs (87–89). The interaction of acylglycerols with chlorine has been reported to cause the formation of 3-MCPDE during the refining of edible oils, especially in the deodorisation step (29,74,80,82,89–91). In addition to the effects of chlorine and MAG/DAG, the processing temperature and duration are other factors that influence the mechanism of the formation of 3-MCPD esters in food (57, 65,68). Studies have shown that the content of contaminants increase rapidly at temperatures above 200 °C, but a balance between their formation and breakdown is reached with increasing processing time (77,78,90–93). GE are unstable molecules, but 3-MCPDs are stable. After the 3-MCPD compounds are formed, their stability depends on the pH and ambient temperature. The formation of 3-MCPD increases with higher pH and temperature. The concentration of 3-MCPDE in vegetable oils depends on their composition and deodorisation. In the study conducted by Franke *et al.* (89) on canola oil, the 3-MCPD content in oil containing only 2 % DAG and 0.1 % free fatty acids increased from 0.4 to 1.0 mg/kg during deodorisation. In palm oil with 5 % DAG and 0.24 % free fatty acids, this rate increased from 1.0 to 4.4 mg/kg (89). On the other hand, Shimizu *et al.* (72) subjected tri-, di- and monooleins to a temperature of 240 °C in the presence and absence of chloride ions to determine the effect of the precursors that trigger the formation of 3-MCPD and GE. They found that 3-MCPDE formed from mono- and dioleins in the presence of chloride ions, while GE formed in the absence of chloride ions. The study showed that di- and monoolein are effective precursors of 3-MCPD and GE in addition to the chloride ion. It was also observed that triolein remained stable during the temperature test and caused the formation of a very small amount of esters (72). In another study, it was found that ester formation was higher at temperatures above 229–240 °C (80).



## THE STRATEGIES FOR MITIGATION OF 3-MCPD AND GE

### *Changes due to the oil extraction parameters*

The type of oil (the presence of glycerol, mono-, di- and triacylglycerols), oil extraction method (temperature and time) and the amount of chlorine and water in the oil are the main factors influencing the formation of 3-MCPD and GE (71,93). The extraction of the oil from fruits with a higher water content than from the seeds and the temperature during the extraction process increase the formation of contaminants (78,79). There are three different ways to reduce the amount of esters in the oils to be refined. The first way is to avoid and/or minimise precursors in the raw material, the second is to adjust and/or optimise the parameters of the refining process and the third is to remove the esters formed in the oil after refining (74). The most important thing is to minimise the amount of precursors in the raw materials. In this way, the ester formation can be kept to a minimum. The reduction of precursors found in the raw material extends to the field, and factors such as climate, soil and growing conditions are effective. Subsequent harvesting practices and storage conditions are also important parameters for the formation of these precursors (31,74,94,95). These different conditions affect the amount of chlorine-containing compounds, which is an important precursor (96). Saline soils, iron(III) chloride and fertilisers (KCl and  $\text{NH}_4\text{Cl}$ ) used in water treatment are among the inorganic sources of chloride (97). In addition, some organochlorine pesticides used to control weeds and pests during cultivation lead to an increase in chlorine amounts in the raw material (74). For this reason, it is recommended to wash the palm fruits with slightly alkaline water to reduce the chlorine content in the product. Studies have shown that there is no difference in the formation of contaminants between the chlorine sources, but that the amount of 3-MCPD increases as the amount of chlorine increases (80,84,90). Storage under unsuitable conditions after harvest also leads to an increase in these contaminants. In oily fruits and seeds, the triglycerides are broken down by lipase into free fatty acids and diglycerides. If lipase activity is high in ripe fruit, this leads to a rapid degradation of triacylglycerols causing the formation of large amounts of diacylglycerol, which also increases the amount of formed esters (21,71,74,89). During the processing of palm fruit bunches into crude palm oil, chlorine catalyses the formation of partial acylglycerols and free fatty acids through lipolytic activity (98). For this reason, palm fruit bunches should be collected quickly after harvest to ensure inactivation of the enzymes, and sterilised without steam. Low temperatures should be used for sterilisation (31,96). Extraction within 48 h of harvest reduced the amount of mono- and diacylglycerols in the oil and resulted in its good quality. In addition, damaged fruits are not processed together with other products to prevent the formation of esters (31).

According to all this knowledge, in order to reduce the formation of 3-MCPDE, either the chlorine-containing

compounds in the fruits should be reduced or these compounds should be removed before refining (74). Chloride-containing compounds can be removed by washing the crude palm oil with polar solvents such as water or water/alcohol mixture. Another efficient way is to wash the oilcake during oil extraction process and then remove chlorine compounds (95,99). Some researchers found that washing crude palm oil with water or water/alcohol mixture reduces 3-MCPD and GE contents (74,95).

### *Reduction of precursors before processing*

As mentioned before, compounds acting as a precursor in the formation of 3-MCPD and GE are MAG/DAG and chloride ions. Elimination of these compounds from the crude vegetable oils before refining will also affect the quantity of 3-MCPD and GE in the end product. The precursors of 3-MCPD and GE in the oil are directly affected by climate, soil, fertilisation, harvesting and storage conditions (95). Chlorinated compounds in the raw material are the most important precursors that play a role in the formation of 3-MCPDE (96). Many researchers have associated them with the use of fertilisers such as ammonium chloride and potassium chloride, and with the presence of chlorine in the irrigation water. The chlorine amount in the raw material depends on the variation, type of soil, growth conditions and differences in the plant genotypes (100). Chlorine-containing chemicals in raw materials can be reduced by using fewer or no pesticides and fertilisers (101). For example, conventional palm production, where fertilisers and pesticides are used that can degrade soil and leave residues, can be replaced with organic palm production without the use of fertilizers, favouring locally adapted systems over synthetic materials (102). Numerous chlorine compounds, including magnesium chloride, calcium chloride, iron(III) chloride and iron(II) chloride, are produced during irrigation and fertilisation (103). Matthäus *et al.* (95) found that washing crude palm oil with water or ethanol reduced the amount of 3-MCPDE by 20–30 %. Also, palm fruit pulp washing before oil extraction reduced 3-MCPDE content by 95 % compared to deodorisation. Yung *et al.* (104) focused on optimising the removal of chloride from crude palm oil by washing with water to minimise the production of 3-MCPDE. By adding 5 % water per oil mass during the washing process, the water-soluble chlorides in crude palm oil are eliminated by up to 76 %, resulting in a 71 % decrease in 3-MCPDE amounts and bringing them below the legal limits. A linear relationship was established between the chloride and the equivalent 3-MCPDE, with a high correlation value ( $R^2$ ) of 0.99. Using the correlations, it is possible to extract 1.0 mg/kg of 3-MCPDE from refined, bleached and deodorised palm oil obtained from crude palm oil with 1.2 mg/kg chloride, using 7 % water during washing. The study also showed a slight decrease in GE of 7 to 11 % after washing with water (104). Ramli *et al.* (105) conducted a larger scale study to reduce 3-MCPDE in palm oil. The study included washing 900 tonnes of crude palm oil in 5–10 % hot water at a temperature of 90–95 °C. Although washing did not have a statistically significant

effect ( $p > 0.05$ ) on other oil quality indicators, such as the concentration of free fatty acids (FFA), diacylglycerol (DAG) and degradation of bleaching index (DOBI), a simple hot water washing removed more than 85 % of the total chlorine. The study found that the most effective method to produce refined oil of excellent quality with significantly low content of 3-MCPDE was water washing of crude palm oil both before and after the refining process (105). In another study, washing the crude palm oil with  $V(\text{ethanol}):V(\text{water})=50\%$  before deodorisation reduced the 3-MCPDE content by 30 % (97). Considering all the studies conducted in this framework, it is found that washing a crude vegetable oil with polar solvents helps to remove water-soluble chlorine and reduce the formation of 3-MCPDE (94). In addition, previous research was conducted to reduce 3-MCPDE. The organochlorine components were removed using monoacylglycerols as agents in various oils without the use of a solvent. The study was carried out by mixing and homogenising a crude vegetable oil with monoacylglycerols and separating these components from the oil by crystallisation. Finally, the amount of 3-MCPDE was reduced, as a result of the reaction of the organochlorine compounds with monoacylglycerols, by 60–90 %, depending on the types of oils and the thermal treatment (106).

Unlike 3-MCPDE, the precursors of GE are MAG and DAG, but not chloride ions (95). The studies show that the content of MAG and DAG in crude vegetable oils has a significant effect on the formation of GE. Thus, reducing the formation of GE is considered the most effective method to reduce the amounts of MAG and DAG in vegetable oils. The amount of MAG and DAG in vegetable oils increases due to the hydrolysis of TAGs by lipase enzyme activity (21). Mishandling during harvesting or storage of the oily fruits or seeds causes the product damage and increased lipase activity, which results in a higher MAG and DAG content. Thus, fruits and seeds should be sterilised after harvest to inactivate lipase. However, it is recommended to keep the sterilisation temperature below 120 °C to avoid triggering the formation of MCPDE (28,41,90,99). For oil extraction after harvesting, processing fruits under unfavourable conditions also increases the amount of MAG and DAG in the oil (74,107). The location of the growth also influences the content of 3-MCPD and GE in oily seeds or fruits. The amount of 3-MCPD in oil is usually determined by the concentration of the precursor in raw material before processing (103).

### Strategies applicable during the refining

Vegetable oil can be refined in two ways, namely physical and chemical refining. In both cases, the main purpose of refining is to remove or reduce unwanted/undesirable ingredients. Degumming, bleaching and deodorisation are the main steps in both refining methods. Neutralisation, which is achieved by saponification of free fatty acids and separation of the aqueous soap-stock phase, is the main difference between chemical and physical refining. The refining steps were further investigated to reduce the 3-MCPDE and GE content

in vegetable oil and were generally associated with optimisation approaches. Within this framework, some studies aimed to reduce the 3-MCPDE and GE content by reducing the formation rate through optimisation of the parameters of the refining steps, while others aimed to apply new steps within the refining process (Table 1; 5,20,29,37,38,72,89,108–120).

Washing the oil prior to degumming was reported as method of eliminating free chloride ions to reduce the formation of 2- and 3-MCPDE. Pre-washing with 10 % deionised water at 70 °C for 20 min resulted in a reduction of the final 2- and 3-MCPDE content by 19 and 22 %, respectively (109).

Gums, mainly a mixture of hydratable and non-hydratable phospholipids, are removed in the initial step of the refining process (75). The degumming step is generally applied in both physical and chemical refining by the addition of phosphoric acid to water. During refining, the degumming step can slightly reduce the 3-MCPDE content of crude and pre-refined palm oil by partly removing potential precursors such as diglycerides, monoglycerides, free chloride ions and polar compounds relative to triglycerides through their dissolution and removal with water and phosphoric acid (37,89). On the contrary, in another study, after the degumming step, the 3-MCPDE content of hot-pressed peanut oil was either not affected or it slightly increased from 0.13 to 0.14 mg/kg (37). The lower amount of phosphoric acid added during degumming increases the reduction of 3-MCPDE because an acidic medium induces the formation of 3-MCPDE and GE through a nucleophilic substitution pathway or the formation of intermediate acyloxonium ions (65,109). Although the degumming temperature was not reported by Sim *et al.* (38) as an effective factor in the formation of 3-MCPDE and GE, increasing the degumming temperature to 105 °C increases the solubility of phospholipids in the oil, leading to a decrease in degumming efficiency and, consequently, in a potentially higher 3-MCPDE formation.

Neutralisation or alkaline refining refers to the removal of free fatty acids by chemical saponification using an alkaline solution. Neutralisation, a step of chemical refining, contributes less to the formation of 3-MCPDE than bleaching and deodorisation (109). Franke *et al.* (89) reported an increase of 0.91 and 0.09 mg/kg in 3-MCPDE mass fractions of pre-refined palm oil and crude palm oil, respectively, which was still lower than the increase during deodorisation. On the contrary, chemical neutralisation of organic palm oil with 2 L of 33 % NaOH solution at 70 °C for 20 min and subsequent washing led to a 43 % decrease in 3-MCPDE mass fraction. However, the mitigation effect was not observed for GE, where neutralisation increased the GE mass fraction up to 29.4 mg/kg, while the GE mass fraction of physically refined (control) samples was 10.90 mg/kg (108). Neutralisation with sodium carbonate and sodium hydrogencarbonate had completely different effects on the GE mass fractions in crude palm oil. The GE was eliminated almost completely by neutralisation with sodium carbonate, while the amount of 3-MCPDE was reduced. When 1 mmol/kg sodium hydrogencarbonate was used, the total ester mass fraction was reduced to 1 mg/kg (5). In addition,

**Table 1.** Brief information about some studies discussing mitigation of 3-monochloropropane-1,2-diol (3-MCPD) and glycidyl ester (GE) during refining steps

Oil type	Operation/step	Condition	Effect	Reference
Crude palm oil	Prewashing (applied prior to degumming)	20 min at 70 °C, 10 % deionised water	At the end of the physical refining, 2- and 3-MCPDE ( $w=0.54$ and $1.04$ mg/kg, respectively) were 19 and 22 % lower when prewashing was applied	(108)
Crude palm oil	Neutralisation	20 min at 70 °C with 2 L of 33 % NaOH solution	$w(3\text{-MCPDE})$ was reduced by 43 % but $w(\text{GE})$ was doubled	(108)
Crude palm oil	Neutralisation (only the addition of alkali, refining step was not like industrial practice)	1 mmol/kg $\text{NaHCO}_3$	$w(3\text{-MCPDE})$ and $w(\text{GE})$ reduced 81 and 84 %, respectively	(5)
Crude palm oil	Neutralisation	KOH and NaOH were used (other conditions not given)	$w(3\text{-MCPDE})$ reduced 35 % and 45 % when NaOH and KOH were used, respectively	(20)
Pre-refined palm oil	Bleaching	$w(\text{Tonsil Optimum 214 FF bleaching earth})=0.8$ % was added to the oil (90 °C, up to 1 kPa) and removed by filtering after 20 min treatment	$w(3\text{-MCPDE}$ and related ester) was reduced from 6.06 to 2.48 mg/kg (59 % reduction)	(89)
Peanut oil (pressed)	Bleaching	The degummed oil was preheated to 95 °C and bleached by adding activated clay (activated by 3 % hydrochloric acid) up to 1 kPa and then filtered after 25 min treatment	$w(3\text{-MCPDE})$ was reduced by approx. 15–22 % after bleaching. Although DGF standard method C-VI 18 (10) of the German Society for Fat Science was used, GE results were not given	(37)
Crude soybean oil	Degumming, neutralisation, bleaching and deodorisation	Use of acid in the degumming stage and acid-activated clay	$w(3\text{-MCPDE})$ increased compared to crude oil	(109)
Crude palm oil	Degumming	Acid degumming (0.06 % of 85 % $\text{H}_3\text{PO}_4$ for 20 min at 90 °C) and water degumming (1 % water, 10 min at 90 °C)	Reduction ranging from 56 to 78 % for $w(3\text{-MCPDE})$ and 49–56 % for $w(\text{GE})$	(110)
Rice bran oil and palm oil	Bleaching	20 g oil was bleached with 200 mg activated earth at 110 °C and 10 kPa, after 20 min, cooled to 30 °C and filtered	$w(\text{GE})$ of the model oil spiked with different ester forms of glycidol was reduced effectively to >0.1 mg/kg	(72)
Crude palm oil	Bleaching	At 105 °C for 20 min with acid-activated bleaching earth (1, 2 and 3 %)	$w(3\text{-MCPDE})$ reduced as bleaching earth content increased	(38)
Crude palm oil	Bleaching	At 95 °C for 20 min with $w(\text{clay})=0.5$ %	The use of low Cl bleaching earth resulted in lower $w(\text{MCPD})$ than the use of high Cl bleaching earth	(111)
Crude palm oil	Bleaching	$w(\text{pure-Flo B80 natural bleaching earth})=10$ %	Bleaching reduced 93 % of $w(\text{GE})$ formed during deodorisation, while $w(3\text{-MCPDE})$ were unaffected	(29)
Palm oil	Deodorisation	10 g sample was heated to 200–290 °C for 45–360 min; 20 kg/t steam was purged	Despite the increase in deodorisation temperature to 270 °C, the 3-MCPDE+GE content in the samples increased. Deodorisation for 240–360 min at 290 °C resulted in a decrease in contaminant levels	(20)
Peanut oil (pressed)	Deodorisation	At 220–260 °C up to 3 h under 200 Pa with steam speed of 0.1 g/min	$w(3\text{-MCPDE}$ and related matter) were reduced after 2 h of deodorisation	(37)
Crude palm oil	Bleaching	$w(\text{zeolite-Fe})=1.5\text{--}15$ % and different bleaching temperature (60–140 °C)	The most effective bleaching procedure was achieved with a $w(\text{zeolite-Fe})=5$ % and a bleaching temperature of 80 °C for 30 min	(112)
Palm oil	Bleaching	Using activated carbon treated with a 2 M HCl solution	$w(3\text{-MCPD})$ 80 % reduction and $w(\text{GE})$ 97 % reduction	(113)
Palm oil	Bleaching	Using metal-organic frameworks (MOFs)	The MOFs, which are composed of 2,6-naphthalenedicarboxylic acid (Fe-MIL-88_BDC), exhibited the most effective adsorption capabilities	(114)
Palm oil	Bleaching	Using zeolite	The use of beta zeolites led to the most effective removal of 3-MCPDE (86 %) compared to other zeolites tested. Using a synergistic blend of beta zeolite and AC, the efficiency of removing 3-MCPDE was significantly improved to 94 %, resulting in a simultaneous reduction of 75 % in GE	(115)

Table 1. continued

Oil type	Operation/step	Condition	Effect	Reference
Coconut oil	Bleaching	At 105 °C with w(activated clay)=1 %. Bleaching was performed under vacuum. After 30 min, the slurry was allowed to cool (to 50 °C) and filtered	In crude coconut oil w(3-MCPDE) and w(GE) were 0.21 and 0.29 mg/kg, respectively. After bleaching, w(3-MCPDE) remained the same, but w(GE) was reduced to 0.19 mg/kg	(116)
Camellia oil	Degumming, bleaching and deodorisation	Degumming water addition at 2.97 %, bleaching earth (acid activated) 2.69 %, deodorisation temperature 230 °C, deodorisation duration 90 min	w(3-MCPDE) reduced by 76.9 %	(116)
Palm olein oil	Bleaching and deodorisation	Bleaching; 95 °C up to 1 h under 8 kPa using neutral and acid-activated bleaching earth Deodorisation (at 200–230 °C up to 1.5 h under 0.15–0.2 kPa)	Use of neutral bleaching earth and deodorisation at 230 °C led to w(MCPD) reduction of 69.91 %. The glycidol esters may be effectively reduced by up to 93.85 % with the use of acid-activated bleaching earth and deodorisation at a temperature of 230 °C, without the need for neutralisation	(117)
Palm oil	Deodorisation	At 250 °C up to 2 h using 100 g bleached oil	Potassium acetate reduced w(3-MCPDE) almost 99 % and w(GE) 40 %. Sodium hydrogencarbonate, sodium acetate and sodium carbonate were able to lower w(3-MCPDE) by more than 80 %	(118)
Palm oil	Deodorisation	Different temperatures (210–270 °C) and time (30–120 min)	After 30 min at 210 °C, the formation of 3-MCPDE was most pronounced under the mildest conditions	(119)
High oleic sunflower and rapeseed oil	Water washing, degumming, bleaching and deodorisation	Different deodorisation temperatures (220–260 °C), at 70 °C up to 20 min using 0.5 % citric acid for degumming, at 95 °C up to 20 min with 3 different bleaching earth (0.1–2 %)	Double refining successfully decreased w(GE) in high oleic (HO) sunflower and rapeseed oil by 70 and 94 %, respectively	(120)

Pudel *et al.* (20) reported that neutralisation caused a reduction in 3-MCPDE and related compounds when compared to crude palm oil. Moreover, when palm oil was neutralised with potassium hydroxide, a 45 % lower content of 3-MCPDE and related compounds was formed, while the reduction was 35 % when sodium hydroxide was used (20). The conflicting results of the reported effects of neutralisation on the content of 3-MCPDE and GE may be due to the use of washing before or after neutralisation, which reduces the concentration of precursors and ions.

Bleaching is the step in which pigments are removed using acid-activated or natural bleaching earth, which are adsorptive materials for pigments and phospholipids *via* van der Waals forces (75). Since bleaching clay adsorbs polar groups, numerous studies have reported a reduction of ester content during bleaching. Franke *et al.* (89) found a 59 % reduction of 3-MCPD. However, their analytical method at that time could not distinguish between 3-MCPDE and GE, so it was unclear which of the two was responsible for the reduction. Li *et al.* (37) conducted a separate analysis of 3-MCPDE and GE content. They reported a reduction of 15 to 22 % 3-MCPDE content after bleaching hot-pressed peanut oil with 3 % acid-activated bleaching earth. Another study focused on the effects of chemical refining steps on the physicochemical properties of coconut oil and the changes in 3-MCPDE and GE content of the samples (121). The GE mass fraction decreased from 0.29 to 0.21 mg/kg, which was statistically significant, however, the 3-MCPDE content was not significantly reduced. Sim *et al.* (38) investigated the effects of refining conditions on the 3-MCPDE and GE content of crude palm oil using an

optimisation approach of the response surface methodology. As the experimental design varied the amount of phosphoric acid, degumming temperature, bleaching earth amount and deodorisation temperature, the isolated effect of bleaching condition was not reported. However, linear modelling showed that a higher content of acid-activated bleaching earth decreased the 3-MCPDE mass fraction. Since the GE mass fraction varied between 0 and 0.64 mg/kg, a valid model for GE could not be determined. Furthermore, researchers indicated that the application of a modified (optimised) refining process enabled the extraction of refined palm oil with a lower 3-MCPDE mass fractions from 1.10 mg/kg (conventionally refined) to around 0.10 to 0.79 mg/kg, with a total reduction of 90 %. For mass fraction of 0.85 mg/kg GE, a reduction of 24.7 to 65.0 % was achieved with conventional refining (38). Some studies indicated a great potential of bleaching for the reduction of GE content in oil. The use of acid-activated bleaching earth was an undesirable practice because of the reported 3-MCPDE and GE propagation effect of chloride ion groups formed during acid activation of bleaching earth (111). However, Shimizu *et al.* (72) indicated that bleaching caused a significant reduction in the GE content in model oils spiked with different esterified forms of glycidol (palmitate, stearate, oleate, linoleate and linolenate). The mass fractions were reduced below 0.1 mg/kg regardless of the spike amounts. Studies have also indicated that bleaching was a crucial step in reducing GE mass fractions. Oey *et al.* (122) pointed out that bleaching reduced 93 % of the GE content formed during deodorisation, while the 3-MCPDE content remained unaffected. Anis *et al.* (112) used zeolite-Fe to improve the bleaching



process and decrease the chlorine concentration, and optimised bleaching settings by the use of response surface methodology (RSM). They also evaluated the properties of the bleached crude palm oil. Bleaching duration, zeolite-Fe content and bleaching temperature were evaluated as factors and subsequently adjusted. The results showed that the concentration of Fe in the modified zeolite-Fe increased significantly by 71.89 % compared to the natural zeolite. The mass fraction of zeolite-Fe, the duration of bleaching and the temperature of bleaching all had a remarkable effect on the adsorption of chlorine. The most effective bleaching process was achieved with a zeolite-Fe mass fraction of 5 % and a bleaching temperature of 80 °C for a duration of 30 min. The adsorption capacity of pretreated activated carbon for 3-MCPDE was evaluated in a batch system at different adsorption temperatures (112). Restiawaty *et al.* (113) found that the mass fraction of 3-MCPDE in the refined, bleached and deodorised palm oil was effectively reduced by 80 %, from 19 700 to below 4000 mg/kg, using activated carbon treated with a 2 M HCl solution. The application of the identical therapy led to a significant decrease in GE from 6680 to 200 mg/kg, which corresponds to a reduction of up to 97 % (113). In a study by Ahn *et al.* (114), metal-organic frameworks (MOFs) were used to selectively adsorb and remove 3-MCPD and glycidol, which are known to be potential genotoxic carcinogens. To increase the efficiency of adsorption, a series of Fe-MIL-88 compounds with various ligand types were employed, followed by further modification by successive carboxylation and deprotonation procedures. Of these ligands, the MOFs, consisting of 2,6-naphthalenedicarboxylic acid (Fe-MIL-88\_BDC), exhibited the most effective adsorption capabilities. Surface modification led to a significant increase in the efficiency of 3-MCPD and glycidol. Although it has lower porosity, the deprotonated Fe-MIL-88\_BDC showed the maximum adsorption capacity among the mentioned adsorbents. The researchers discovered that the significant increase in adsorption efficiency was a result of esterification with carboxyl groups on 3-MCPD and glycidol. The adsorption performance was found to be exclusively in the 3-MCPDE and GE forms (which are precursors of 3-MCPD and glycidol) in palm oil, without any change in the content of fat or minor components (114).

In another study, Restiawaty *et al.* (115) investigated the reduction of the 3-MCPDE and GE content in refined, bleached and deodorised palm oil by utilizing zeolite as an adsorbent. The adsorption was carried out in a batch reactor, varying the adsorbent percentage, temperature, zeolite type and the combination of zeolite and activated carbon. The results show that the optimum temperature for adsorption was 35 °C when 2 % zeolite was used. The use of beta zeolites led to the most effective removal of 3-MCPDE (86 %) compared to other zeolites tested. This can be attributed to their large pore volume, Si/Al ratio and overall acidity. By using a synergistic mixture of beta zeolite and activated carbon, the efficiency of 3-MCPDE removal was significantly improved to 94 %, resulting in a simultaneous reduction of GE by 75 % (115).

Deodorisation is the final step in which the main aim is to remove the odour-forming material from vegetable oil by distillation. As the previous neutralisation step is not used in physical refining, the free fatty acid content is also removed at this point. Significant amount of GE and 3-MCPDE formed during deodorisation as higher temperatures (>220 °C) are removed in this step. An earlier study indicated that the content of 3-MCPDE and a related compound (GE) decreased at extreme temperatures (290 °C) during deodorisation for 240–360 min, although it increased 10 times when deodorized at 270 °C. It was also reported that the amount of contaminants increased with time up to 250 °C, particularly the amount of GE. This effect was reversed at temperatures above 270 °C, and at 290 °C the amount of contaminants decreased with time, probably due to thermal degradation reactions or distillation (20). A similar effect of thermal degradation was also found in another study where 3-MCPDE and related matter content in hot-pressed peanut oil was reduced after deodorisation at 220–260 °C for 2 h (37). Since deodorisation accelerates the formation of 3-MCPDE and GE, some optimisation studies have been carried out to limit or minimise the formation of 3-MCPDE and GE during deodorisation while meeting the industrial demand for refined oil. The optimisation studies aim to find the factors and their interactions that have an effect on the formation of 3-MCPDE and GE during processes. Studies suggest that temperature is the most significant factor in the formation of 3-MCPDE and GE during deodorisation, and that duration is another important factor as it represents effective total thermal load of the oil (110). Sim *et al.* (38) investigated the effect of optimised refining conditions including phosphoric acid amount, degumming temperature, bleaching earth amount and deodorisation temperature (240, 250 and 260 °C) and reported that the application of a lower temperature was important to limit the formation of 3-MCPDE and GE during deodorisation. During the optimisation of deodorisation parameters, while temperature was the most effective factor for 3-MCPDE and GE formation, the provoking effect of temperature on GE formation was not observed at high steam stripping rates. This result was associated with an increased mass transfer rate of precursors such as diglycerides and GE at high steam stripping rates (92). Optimisation studies generally provided valid results and approaches only for the used laboratory-scale equipment and experimental conditions discussed. Positive results were observed in the reduction of 3-MCPDE and GE in the final product by using the optimised refining conditions at pilot and industrial scale (38). Camellia oil is a type of oil that has a high 3-MCPDE content due to the deodorisation process applied at high temperatures, however, certain physical refining steps were optimised to minimise the 3-MCPDE content (116). A 76.9 % reduction of 3-MCPDE content was achieved compared to commercial samples. The study suggests degumming at 2.97 % moisture, degumming temperature of 50.5 °C, activated clay amount of 2.69 %, deodorisation temperature of 230 °C and deodorisation time of 90 min.

Elmas and Akpınar Bayazit (117) investigated the optimal process parameters to minimise the formation of 3-MCPD

and GE during palm oil refining. They have varied refining parameters such as bleaching agent and deodorisation temperature, and obtained a reduction of 3-MCPD by 69.91 % with neutral bleaching earth and deodorisation at 230 °C, and a reduction of GE by 93.85 % with acid-activated bleaching earth and deodorisation at 230 °C without neutralisation.

Studies have also investigated the reduction of 3-MCPDE and GE by the use of adsorbents, metals, salts and amino acids before and during deodorisation (118). Potassium acetate reduced the 3-MCPDE content by almost 99 % and the GE content by 40 %. However, it led to an increase in free fatty acid content in the refined oil. The other compounds used caused a small decrease of just 1–30 % of 3-MCPDE and 6–49 % of GE. In the same study, other salts such as sodium hydrogen carbonate, sodium acetate and sodium carbonate were able to reduce 3-MCPDE by more than 80 %. However, they caused an increase in GE and changes of both the colour and FFA content of the refined, bleached and deodorised palm oil. Similar results were obtained when adsorbents, metals or amino acids were used, indicating their effectiveness in removing a specific pollutant, either 3-MCPDE or GE.

Furthermore, Tivanello *et al.* (119) investigated the deodorisation of palm oil at different temperatures (210, 230, 250 and 270 °C) and durations (30, 60, 90 and 120 min), and concluded that the formation of 3-MCPDE was most pronounced under the mildest conditions (30 min at 210 °C), with values ranging from 1.91 to 2.70 mg/kg. Notably, the study did not find a significant correlation between the 3-MCPDE content and the physicochemical changes in the deodorised palm oil (119).

Mayayo *et al.* (120) carried out studies with the aim of developing strategies to reduce the formation of contaminants, especially GE and 2- and 3-MCPDE in organic oils. The focus was on sunflower oil with high oleic content and rapeseed oil, and the research looked specifically at the refining process to achieve this reduction. The study compared several techniques, including twofold refining with a high or low deodorisation temperature, water washing and increasing amounts of bleaching earth, with a single physical refining treatment (control treatment). The results showed that the twofold refining successfully reduced the GE content in sunflower oil with high oleic content and rapeseed oil by 70 and 94 % respectively, compared to the control treatment. The second degumming and bleaching steps were very important for the removal of GE contamination. Both 2- and 3-MCPDE were formed during the temperature ramping phase before deodorisation and their content remained constant throughout the refining process. Increasing the amount of bleaching earth by a factor of two led to a 48 % reduction in the content of 2- and 3-MCPDE in the sunflower oil with high oleic content. However, no reduction was observed in rapeseed oil (120).

#### *Effects of the oil modification on the reduction of 3-MCPDE and GE*

In addition to the refining processes, several modifications are used to produce different fat and oil products to

alter the physical properties of the fats and oils to meet the requirements of the intended use. The processes that offer the possibility of modifying fats and oils are fractionation, winterisation, interesterification and hydrogenation (7,123). Fractionation is a common method applied to palm oil that often refers to drying; however, a wet or solvent fractionation can also be used. Wet fractionation, typically applied to cottonseed products, uses hexane or acetone to separate different components in the miscella stage. Kyselka *et al.* (98) investigated the reduction of 3-MCPDE and GE content in palm oil during hydrogenation and wet fractionation. By wet fractionation with acetone palm oil was fractionated into four different fractions, namely super stearin, hard stearin, super olein and top olein. The results show that the distribution of 3-MCPDE and GE in the fractions tends to be more than a reduction. The 3-MCPDE and GE were highly concentrated in the top olein phase at 16.16 and 9.2 mg/kg, respectively. Super olein contained 5.01 mg/kg 3-MCPDE and 1.08 mg/kg GE. Super stearin and hard stearin contained lower amounts of 3-MCPDE and GE. The results showed that 3-MCPDE and GE tend to remain in the liquid fractions of palm oil during fractionation. During the complete hydrogenation of the palm oil sample, the mass fraction of 3-MCPDE and GE was reduced from 4.72 to 2.17 mg/kg and from 1.07 to 0.62 mg/kg, respectively. Although deodorisation applied after complete hydrogenation caused a slight increase in the contents of both contaminants, the reductions during hydrogenation were notable. Another significant decrease in glycidyl ester content was observed after bleaching of fully hydrogenated palm oil, where GE content was not detected, while 3-MCPDE content was not affected by the bleaching (98).

Enzymatic approaches have been shown to be a promising tool for reducing 3-MCPD in the extraction of palm oil, serving as an alternative to mechanical extraction. Modification of the fatty acid arrangement in the final product can be customised by the development of lipases used in the enzymatic production of structural lipids to benefit specific initial characteristics of materials, such as the location of the glycerol backbone and fatty acids. In 2010, Bornscheuer and Hessler (124) investigated the potential of using enzymes to remove 3-MCPDE and GE from refined oils. They were the pioneers in uncovering the fact that 3-MCPDE can be converted into the harmless by-product glycerol by enzymatic cascade reactions. The enzyme cascade consisting of an epoxide hydrolase from *Agrobacterium radiobacter* AD1 and a halohydrin dehalogenase from *Arthrobacter* sp. AD2 was used to fully convert 3-MCPD into glycerol. This conversion took place at a lower reaction temperature of 30 °C in a thermomixer (124). Bel-Rhliid *et al.* (125) conducted another enzymatic treatment with 1242 and 0.3358 mg/kg of racemic 3-MCPD in the presence of oxygen. At pH=8.2 and in the presence of glucose, a mass fraction of 1242 mg/kg of racemic 3-MCPD was converted at an ideal rate of 68 %. In contrast, without glucose, a mass fraction of 0.3358 mg/kg of 3-MCPD degraded by 73 % after 72 h (125). In addition, Tsai *et al.* (126) used modified

QuEChERS (quick, easy, cheap, effective, rugged and safe) and lipase hydrolysis as pretreatment methods for sample material to detect 3-MCPDE in edible oils. Acetonitrile and salts are used in QuEChERS, a common sample preparation method for the extraction of highly polar compounds such as 3-MCPD. Their approach showed superior precision and accuracy compared to the widely used official method (AOCS Cd 29c-13), as they presented a rapid and accurate analytical method to quantify the amount of 3-MCPDE in edible oils (126). In addition, Miyazaki and Koyama (127) conducted a study that used *Candida cylindracea* in an indirect enzymatic technique to identify 3-MCPDE. This method involved the removal of sodium bromide during the hydrolysis. Recovery rates of 3-MCPD from concentrated sardine oil were found to be good, ranging from 91 to 109 % at a mass fraction of 20 mg/kg (127). In addition, Chai *et al.* (128) investigated the effect of tallow by enzymatic hydrolysis and the addition of NaCl on the production of 3-MCPDE. In the preparation of beef flavour, they found that the specific conditions of enzymatic hydrolysis (80 % tallow (*m/V*), 47.5 °C, pH=7 and 9.5 h) had a notable effect on the formation of 3-MCPD mono- and diesters. There were no appreciable amounts of 3-MCPD monoesters detected, while the amount of 3-MCPD diesters at all temperatures depended strongly on the content of NaCl and the time of its addition (either before or after the thermal reaction). The reason for this is that tallow has a very low melting point of 47.5 °C. At temperatures above 50 °C, the lipase becomes practically inactive (128). According to all published studies, the decrease in the synthesis of 3-MCPDE can be attributed to the elimination of the associated precursors before deodorisation. This is because several studies have shown that most 3-MCPDE are formed during the deodorisation of oil. Research into the exact processes behind the formation of 3-MCPD is still ongoing. In order to effectively reduce and eliminate 3-MCPD from vegetable oils by enzymatic hydrolysis, it is crucial to understand the link between the potential of the enzyme and its contribution to the process.

Deep fat frying is known as a cooking method that potentially triggers the formation of 3-MCPDE and GE due to the prolonged exposure of the oil to high temperature. The content of both contaminants was reduced after three days of frying; however, 3-MCPDE and GE formation in rosemary oleoresin and *tert*-butylhydroquinone (TBHQ) decreased to a larger extent in refined, bleached and deodorised palm oil fortified with 200 mg/kg natural (rosemary and sage extract) and synthetic (butylated hydroxy toluene (BHT), butylated hydroxy anisole (BHA) and TBHQ) antioxidants, from 3.2 to 1 mg/kg and from 5.2 to 3.5 mg/kg, respectively (14). The reduction of 3-MCPDE and GE was associated with the reducing effect of antioxidants on the intermediate radical acyloxonium shortly before the nucleophilic attack of chloride ions during the 3-MCPDE production pathway (14,129).

As the theory about the inhibitory effect of antioxidants on the formation of 3-MCPDE and GE was established, the potential of natural antioxidants to replace unsafe conventional

synthetic antioxidants was investigated. Therefore, sesame oil was used to inhibit the formation of GE in corn oil, palm oil and rice bran oil during the deodorisation. Sesame oil contains a considerable mass fraction of sesamin, sesamol and sesamol lignans (6.5–17.3 mg/kg), and tocopherols (56.9–99.3 mg/kg, mainly  $\alpha$ -tocopherol), which all have antioxidant activity. When the addition of sesame oil was increased from 10 to 50 %, the inhibition of GE formation increased from 4.29 to 18.86 % in corn oil, from 16.82 to 27.39 % in palm oil, and from 23.64 to 29.89 % in rice bran oil (130).

A recent study investigated the inhibitory effect of seven antioxidants, including quercetin, astaxanthin, TBHQ, propyl gallate, L-ascorbyl palmitate, puerarin and  $\alpha$ -tocopherol, on the formation of GE in rice bran oil during the exposure to high temperature (131). Among the seven antioxidants, propyl gallate, astaxanthin and L-ascorbyl palmitate with different antioxidant mechanisms had a better inhibitory effect on GE formation. The study also reported that after high temperature application at 240 °C for 1.5 h, the inhibition rates decreased due to probable degradation of the antioxidants. In particular, the epoxy structure of GE would compromise their stability at high temperatures, leading to thermal degradation to MAG *via* a ring-opened process. The increasing trend in the inhibition rates of the antioxidants was attributed to the combined suppressive effect of antioxidants on GE formation and thermal degradation of GE during high temperature application.

## DEVELOPMENT OF METHODS FOR THE EXTRACTION AND DETERMINATION OF 3-MCPDE AND GE

Chromatographic techniques are widely used today in studies for the detection of chloropropanols or their esters in food matrices. Apart from these methods, electrochemical and optical methods are also used. Each technique offers different mechanisms for qualitative or quantitative analysis. Intrinsic limitations such as molecular structure, low volatility and low molecular mass hinder the direct gas chromatographic (GC) analysis of chloropropanols. Researchers addressed these challenges with a pretreatment step involving derivatisation. This technique converts chloropropanols into more volatile derivatives, minimising unwanted interactions with other sample components during preparation and GC analysis. Derivatisation has enabled the development of different analytical methods for the determination of chloropropanols, including GC (132), GC-MS (133) and HPLC-FLD (134). When analysing chloropropanol, the choice of derivatisation reagent is crucial. Phenylboronic acid (PBA) (135), butaneboronic acid (BBA) (136,137), heptafluorobutyrylimidazole (HFBI) and heptafluorobutyric acid (HFBA) (138) are common reagents for the detection of chloropropanol. However, the innate structural properties of chloropropanols can lead to adverse interactions with the GC system, which can affect sensitivity and cause poor peak shape. It is also problematic to identify targets based on retention times alone.

GC-MS offers superior sensitivity for the determination of chloropropanols and achieves LODs below 3–5 µg/kg for 3-MCPD in various food products (139). The Association of Official Analytical Chemists (AOAC) has established a GC-MS method for the analysis of 3-MCPD in food with an LOD of 0.005 mg/kg (140). HPLC-FLD is an alternative, especially because of its high sensitivity. Prior to HPLC-FLD analysis, Hu *et al.* (141) developed a derivatisation method using periodate oxidation for the detection of 3-MCPD in vegetable oil and water, and achieved an LOD of 0.36 ng/mL. This method is based on the oxidation cleavage of 3-MCPD with sodium periodate to generate chloroacetaldehyde. Subsequent fluorescence derivatisation with adenine enables HPLC analysis with FLD detection. Compared to GC-MS, HPLC-FLD has a significantly lower LOD (by approx. ten orders of magnitude) and eliminates the need for complex pretreatment, extraction or enrichment steps. Notably, in a relevant study, an average recovery of 95.36 % and a relative standard deviation (RSD) of less than 3.44 % was found for repeated measurements, highlighting the excellent accuracy and sensitivity of the method. Beyond derivatisation, researchers have explored alternative pretreatment methods for the detection of chloropropanols. These include molecularly imprinted polymer membrane extraction coupled with GC-MS (142) and the combination of ultra-high performance liquid chromatography (UHPLC) with microwave-assisted derivatisation (MAD) followed by HPLC-UV detection (143). These approaches offer versatility and can be used for a wide range of samples. They also achieve sufficient sensitivity for detection in the mg/kg or µg/kg range. However, there are also limitations. The techniques often require expensive equipment and time-consuming procedures. Furthermore, the presence of nucleophilic compounds in the extract can react with derivatisation reagents, resulting in increased background noise, compromised selectivity and reduced abundance of target ions. Notably, HFBI, a popular derivatisation reagent, is very sensitive to moisture, which can hinder consistent derivatisation.

Electrochemical approaches represent an alternative strategy for the detection of 3-MCPD. Sun *et al.* (144) developed a sensor based on a glassy carbon electrode with deposited gold nanoparticles (AuNP/GCE) and coated with a molecularly imprinted polymer film. This sensor showed high sensitivity (LOD of  $3.8 \cdot 10^{-18}$  mol/L) and excellent recovery rates (95.0–106.4 %) for the detection of 3-MCPD in soy sauce, demonstrating strong adsorption and selectivity. However, the complex manufacturing process and limited reusability of the sensor limit its practical application (144). Yuan *et al.* (145) proposed a method in which hemoglobin (Hb) is immobilised on magnetic molecularly imprinted nanoparticles. This approach uses the reducibility of Hb for the detection of 3-MCPD and achieves an LOD of 0.25 mg/L. The method relies on the electrochemically generated reduction peak current at  $-0.236$  V, which is further reduced by 3-MCPD through Hb catalysis. Higher 3-MCPD amounts lead to a corresponding increase in the intensity of the reduction peak. While this method offers

acceptable repeatability (RSD of 2.9 %), limitations include the need for nitrogen supplementation in the electrolyte and the short shelf-life of the modified electrode (145).

Fang *et al.* (146) introduced a fluorescence-based detection method for 3-MCPD in soy sauce using a molecularly imprinted polymer on a paper substrate. This method eliminates the need for pretreatment and enables direct sample analysis. With an LOD of 0.6 ng/mL, recovery rates between 97.2 and 105.3 % and an RSD below 5.6 %, it surpasses the accuracy of the HPLC-FLD approach. Notably, the HPLC-FLD method suffers from potentially incomplete conversion during the complex pretreatment steps, which include oxidation cleavage and derivatisation, as evidenced by the reported conversion rate of 98.96 %. However, the fluorescence method has its own limitations, including the fragility of the paper substrate and the lengthy surface modification. In addition to the methods explored here, several other techniques, such as UV-Vis spectroscopy, have been developed for the detection of chloropropanols, each offering unique advantages and disadvantages (147).

Traditional indirect methods remain the mainstay for the determination of chloropropanols and chloropropanol esters. This dominance can be attributed to several factors: affordability and accessibility of consumables and instruments, ease of use and repeatability, and satisfactory analytical performance (accuracy, precision and sensitivity). These methods typically involve a sequence of steps: alkaline or acid-catalysed cleavage, solid-phase extraction (SPE), derivatisation with PBA or HFBI, and final detection by GC-MS or GC-MS/MS. Future advances are likely to refine this core approach. Direct methods, while potentially valuable, are less commonly used due to the high cost of instrumentation and extensive standard requirements. Among indirect methods, lipase-catalysed cleavage, advocated by the AOCS, is a promising alternative with several advantages. However, further research and validation for different sample types are required for wider application. Solid-phase extraction (SPE) offers potential for improvement by minimising eluent volume and toxicity. Ideally, specialised SPE materials and columns tailored for efficient chloropropanol extraction should be developed. Non-derivatisation methods are promising replacements for derivatisation-based approaches as they offer less manipulation and lower consumption of resources. However, overcoming the challenges of chromatographic separation of chloropropanols remains critical. Rapid screening methods would benefit from simplified material preparation and streamlined determination procedures.

Custodio-Mendoza *et al.* (43) developed and validated two new indirect approaches for total 3-MCPD. 3-MCPD was released from 3-MCPDE by both alkaline and acidic transesterification processes. For the first time, these reactions were accelerated with ultrasound. The overall content of 3-MCPD was determined using an ultrasound-assisted dispersive liquid-liquid microextraction method and subsequently analysed by GC-MS. Both methods were optimised, with the emphasis



on variables such as sample size, reagent quantity, temperature and agitation technique. The results showed that the alkaline approach achieved a lower limit of quantification, while the acidic method showed good accuracy and precision. Both approaches showed similar performance in terms of analytical properties. The alkaline method was shown to require fewer different types of aqueous solutions, lower amounts of organic liquids, less waste, less reaction time and lower temperatures. Ultrasound greatly shortens response times, resulting in faster, more environmentally friendly and more economical processes (43).

In conclusion, traditional indirect methods currently dominate the analysis of chloropropanols. While existing techniques are being further developed and new approaches explored, further research is needed to overcome limitations and improve overall analytical capabilities.

## CONCLUSIONS

3-Monochloropropane-1,2-diol (3-MCPD) and glycidyl esters (GE) are process contaminants found in vegetable oils and/or oil-containing food as a result of processing. While mono- and diglycerides and chloride ions are effective in the formation of these contaminants, high temperature and time are also important factors that increase their formation and quantity. Mono- and diglycerides formed during the breakdown of triglycerides can turn into glycidyl esters with genotoxic and carcinogenic properties at high temperatures and 3-MCPD esters with carcinogenic properties in the presence of chloride ions in the environment. According to the report published by EFSA in 2018, the tolerable daily intake of 3-MCPD was set at 2 µg/kg body mass. These contaminants are even found in different foods such as coffee, smoked foods and baby food. For this reason, the amount of 3-MCPD found in many foods and the reasons for its formation have gained attention in recent years. Existing studies to reduce or avoid these contaminants, improvement of process conditions, reduction of salt content of foods, reduction of mono- and diglyceride content in raw materials, deactivation of lipase enzyme, improvement of the growth and storage conditions of raw material, low temperature applications during refining, *etc.* are addressing these issues. Due to the negative impact of these toxic contaminants on consumer health, it is important to investigate them in more detail and carry out the necessary studies to reduce/prevent their formation and implement this into industrial processes. The scope and performance of direct techniques will be greatly enhanced by the commercial availability of a solution with internal standards and mixed chloropropanol ester standards. Finally, as a quick screening method, less complicated material preparation and a quicker decision process are needed for alternative approaches. Future studies on chloropropanols, their esters and GE could focus on creating more practical experimental models to understand how they are formed, developing simpler sample preparation techniques, improving test methods with lower limits of detection, and finding more effective and economical ways to eliminate them.

## CONFLICT OF INTEREST


All authors declare that they have no conflicts of interest.


## AUTHORS' CONTRIBUTION

K. Yıldız was responsible for the design of the study, search and interpretation of literature, and preparation of the manuscript. O. Özdikicierler and P. Günç Ergönül were responsible for the study design and drafting the final manuscript. O. Özdikicierler and P. Günç Ergönül gave valuable assistance in the major revision of the manuscript. All authors reviewed the final manuscript before submission.

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## REFERENCES

- Nas S, Gökalg HY, Ünsal M. Vegetable oil technology. Pamukkale: Denizli Pamukkale University Faculty of Engineering Textbooks; 2001 (in Turkish).
- Custodio-Mendoza JA, Carro AM, Lage-Yusty MA, Herrero A, Valente IM, Rodrigues JA, Lorenzo RA. Occurrence and exposure of 3-monochloropropanediol diesters in edible oils and oil-based foodstuffs from the Spanish market. *Food Chem.* 2019;270:214–22. <https://doi.org/10.1016/j.foodchem.2018.07.100>
- Arisseto AP, Vicente E, Furlani RPZ, Toledo MCF. Estimate of dietary intake of chloropropanols (3-MCPD and 1,3-DCP) and health risk assessment. *Ciênc Tecnol Aliment.* 2013; 33(suppl 1):125–33.
- Dunford NT. Oil- and oilseed-based bioactive compounds and their health effects. In: Nutritionally enhanced edible oil and oilseed processing. New York, NY, USA: AOCS Publishing; 2004. pp. 1–30.
- Freudenstein A, Weking J, Matthäus B. Influence of precursors on the formation of 3-MCPD and glycidyl esters in a model oil under simulated deodorization conditions. *Eur J Lipid Sci Technol.* 2013;115:286–94. <https://doi.org/10.1002/ejlt.201200226>
- Hrncirik K, van Duijn G. An initial study on the formation of 3-MCPD esters during oil refining. *Eur J Lipid Sci Technol.* 2011;113:374–9. <https://doi.org/10.1002/ejlt.201000317>
- Shahidi F. editor. *Bailey's industrial oil and fat products.* London, UK: John Wiley & Sons Inc; 2005.
- Deniz Şirinyıldız D, Aydın E, Öztürk Y, Avcı T, Yıldırım A, Yorulmaz A. The level of 3-MCPD fatty acid esters in vegetable oils and margarines collected from Turkish market. *J Food.* 2014;44(3):491–7. <https://doi.org/10.15237/gida.gd19039>

9. Destailats F, Craft BD, Sandoz L, Nagy K. Formation mechanisms of monochloropropanediol (MCPD) fatty acid diesters in refined palm (*Elaeis guineensis*) oil and related fractions. *Food Addit Contam Part A Chem Anal Control Expo Risk Assess.* 2012;29:29–37.  
<https://doi.org/10.1080/19440049.2011.633493>
10. Hernandez EM, Kamal-Eldin A. *Processing and nutrition of fats and oils.* London, UK: John Wiley & Sons Inc; 2013.
11. MacMahon S, Begley TH, Diachenko GW. Occurrence of 3-MCPD and glycidyl esters in edible oils in the United States. *Food Addit Contam Part A Chem Anal Control Expo Risk Assess.* 2013;30:2081–92.  
<https://doi.org/10.1080/19440049.2013.840805>
12. Destailats F, Angers P. On the mechanisms of cyclic and bicyclic fatty acid monomer formation in heated edible oils. *Eur J Lipid Sci Technol.* 2005;107:767–72.  
<https://doi.org/10.1002/ejlt.200501159>
13. Günc Ergönül P, Gödeli T. Presence of 3-monochloropropane-1,2-diol (3-MCPD) and their esters in foods, mechanisms of their occurrence and detection method. *Akademik Gıda.* 2013;11(2):102–9 (in Turkish).
14. Wong YH, Goh KM, Nyam KL, Nehdi IA, Sbihi HM, Tan CP. Effects of natural and synthetic antioxidants on changes in 3-MCPD esters and glycidyl ester in palm olein during deep-fat frying. *Food Control.* 2018;96:488–93.  
<https://doi.org/10.1016/j.foodcont.2018.10.006>
15. Turan S, Solak R, Keskin Ş. Formation and determination of monochloropropanediol esters in foods. *Akademik Gıda.* 2018;16(2):210–7 (in Turkish).  
<https://doi.org/10.24323/akademik-gida.449866>
16. Albuquerque TG, Costa HS, Silva MA, Oliveira MBPP. Are chloropropanols and glycidyl fatty acid esters a matter of concern in palm oil? *Trends Food Sci Technol.* 2020;105:494–514.  
<https://doi.org/10.1016/j.tifs.2019.01.005>
17. Kamikata K, Vicente E, Ariseto-Bragotto AP, Miguel AMR de O, Milani RF, Tfouni SAV. Occurrence of 3-MCPD, 2-MCPD and glycidyl esters in extra virgin olive oils, olive oils and oil blends and correlation with identity and quality parameters. *Food Control.* 2019;95:135–41.  
<https://doi.org/10.1016/j.foodcont.2018.07.051>
18. Ermacora A, Hrnčirik K. Influence of oil composition on the formation of fatty acid esters of 2-chloropropane-1,3-diol (2-MCPD) and 3-chloropropane-1,2-diol (3-MCPD) under conditions simulating oil refining. *Food Chem.* 2014;161:383–9.  
<https://doi.org/10.1016/j.foodchem.2014.03.130>
19. Karl H, Merkle S, Kuhlmann J, Fritsche J. Development of analytical methods for the determination of free and ester bound 2-, 3-MCPD, and esterified glycidol in fishery products. *Eur J Lipid Sci Technol.* 2016;118:406–17.  
<https://doi.org/10.1002/ejlt.201400573>
20. Pudiel F, Benecke P, Fehling P, Freudenstein A, Matthäus B, Schwaf A. On the necessity of edible oil refining and possible sources of 3-MCPD and glycidyl esters. *Eur J Lipid Sci Technol.* 2011;113:368–73.  
<https://doi.org/10.1002/ejlt.201000460>
21. Cheng WW, Liu GQ, Wang LQ, Liu ZS. Glycidyl fatty acid esters in refined edible oils: a review on formation, occurrence, analysis, and elimination methods. *Compr Rev Food Sci Food Saf.* 2017;16:263–81.  
<https://doi.org/10.1111/1541-4337.12251>
22. Demydova A, Levchuk I, Yevlash V, Aksanova O, Foschan A. Effect of oxidation products and other precursors on the content of 3-MCPD esters and glycidol esters in deodorised sunflower oil. *Food Sci Tech.* 2022;16(1):50–7.  
<https://doi.org/10.15673/fst.v16i1.2284>
23. Jędrkiewicz R, Kupska M, Glowacz A, Gromadzka J, Namieśnik J. 3-MCPD: A worldwide problem of food chemistry. *Crit Rev Food Sci Nutr.* 2016;56:2268–77.  
<https://doi.org/10.1080/10408398.2013.829414>
24. Seefelder W, Varga N, Studer A, Williamson G, Scanlan FP, Stadler RH. Esters of 3-chloro-1,2-propanediol (3-MCPD) in vegetable oils: Significance in the formation of 3-MCPD. *Food Addit Contam Part A Chem Anal Control Expo Risk Assess.* 2008;25:391–400.  
<https://doi.org/10.1080/02652030701385241>
25. EFSA Panel on Contaminants in the Food Chain (CONTAM), Knutsen HK, Alexander J, Barregård L, Bignami M, Brüschweiler B, et al. Update of the risk assessment on 3-monochloropropanediol and its fatty acid esters. *EFSA J.* 2018;16(1):e05083.  
<https://doi.org/10.2903/j.efsa.2018.5083>
26. Commission Regulation (EC) No 2018/290 of 26 February 2018 amending Regulation (EC) No 1881/2006 as regards maximum levels of glycidyl fatty acid esters in vegetable oils and fats, infant formula, follow-on formula and foods for special medical purposes intended for infants and young children. *OJ L.* 2018;55:27–9.
27. Davidek J, Velíšek J, Kubelka V, Janíček G, Šimicová Z. Glycerol chlorohydrins and their esters as products of the hydrolysis of tripalmitin, tristearin and triolein with hydrochloric acid. *Z Lebensm Unters Forch.* 1980;171(1):14–17.  
<https://doi.org/10.1007/BF01044410>
28. Stadler RH. Monochloropropane-1,2-diol esters (MCPDES) and glycidyl esters (GEs): An update. *Curr Opin Food Sci.* 2015;6:12–8.  
<https://doi.org/10.1016/j.cofs.2015.11.008>
29. Zelinková Z, Svejková B, Velíšek M, Doležal M. Fatty acid esters of 3-chloropropane-1,2-diol in edible oils. *Food Addit Contam.* 2006;23:1290–8.  
<https://doi.org/10.1080/02652030600887628>
30. Wöhrlin F, Fry H, Lahrssen-Wiederholt M, Preiß-Weigert A. Occurrence of fatty acid esters of 3-MCPD, 2-MCPD and

- glycidol in infant formula. *Food Addit Contam Part A Chem Anal Control Expo Risk Assess.* 2015;32:1810–22.  
<https://doi.org/10.1080/19440049.2015.1071497>
31. Beekman JK, Granvogl M, Macmahon S. Analysis and occurrence of MCPD and glycidyl esters in infant formulas and other complex food matrices. In: Granvogl M, MacMahon S, editors. *Food-borne toxicants: Formation, analysis, and toxicology*, vol. 1306. ACS Symposium Series. Washington DC, USA: American Chemical Society; 2019. pp. 67–90.  
<https://doi.org/10.1021/bk-2019-1306.ch005>
  32. Arisseto AP, Silva WC, Marcolino PFC, Scaranelo GR, Berbari SAG, Miguel AMRO, Vicente E. Influence of potato cultivar, frying oil and sample pre-treatments on the contamination of French fries by 3-monochloropropane-1,2-diol fatty acid esters. *Food Res Int.* 2019;124:43–8.  
<https://doi.org/10.1016/j.foodres.2018.10.070>
  33. Abraham K, Appel KE, Berger-Preiss E, Apel E, Gerling S, Mielke H, *et al.* Relative oral bioavailability of 3-MCPD from 3-MCPD fatty acid esters in rats. *Arch Toxicol.* 2013;87:649–59.  
<https://doi.org/10.1007/s00204-012-0970-8>
  34. Andres S, Appel KE, Lampen A. Toxicology, occurrence and risk characterisation of the chloropropanols in food: 2-Monochloro-1,3-propanediol, 1,3-dichloro-2-propanol and 2,3-dichloro-1-propanol. *Food Chem Toxicol.* 2013;58:467–78.  
<https://doi.org/10.1016/j.fct.2013.05.024>
  35. Joint FAO/WHO Expert Committee on Food Additives (JECFA). Evaluation of certain food additives and contaminants. Fifty-seventh report of the Joint FAO/WHO Expert Committee on Food Additives, WHO Technical Report Series 909. Geneva, Switzerland: Food and Agriculture Organization of the United Nations and World Health Organization (FAO/WHO); 2002. Available from: <https://iris.who.int/handle/10665/42601>.
  36. Lee BQ, Khor SM. 3-Chloropropane-1,2-diol (3-MCPD) in soy sauce: A review on the formation, reduction, and detection of this potential carcinogen. *Compr Rev Food Sci Food Saf.* 2015;14:48–66.  
<https://doi.org/10.1111/1541-4337.12120>
  37. Li C, Li L, Jia H, Wang Y, Shen M, Nie S, Xie M. Formation and reduction of 3-monochloropropane-1,2-diol esters in peanut oil during physical refining. *Food Chem.* 2016;199:605–11.  
<https://doi.org/10.1016/j.foodchem.2015.12.015>
  38. Sim BI, Khor YP, Lai OM, Yeoh CB, Wang Y, Liu Y, *et al.* Mitigation of 3-MCPD esters and glycidyl esters during the physical refining process of palm oil by micro and macro laboratory scale refining. *Food Chem.* 2020;328:127147.  
<https://doi.org/10.1016/j.foodchem.2020.127147>
  39. IARC Working Group on the Evaluation of Carcinogenic Risks to Humans. Some chemicals present in industrial and consumer products, food and drinking-water, vol. 101. IARC Monographs. Lyon, France; 2013. pp. 9–549.
  40. Some industrial chemicals, vol. 77. IARC Monographs on the evaluation of carcinogenic risk of chemicals to humans. Lyon, France; 2000. pp. 469–86.
  41. Cheng W, Liu G, Wang X, Han L. Adsorption removal of glycidyl esters from palm oil and oil model solution by using acid-washed oil palm wood-based activated carbon: Kinetic and mechanism study. *J Agric Food Chem.* 2017;65:9753–62.  
<https://doi.org/10.1021/acs.jafc.7b03121>
  42. Oey SB, Fogliano V, Fels-Klerx HJ, Leeuwen SPJ. Mitigation strategies for the reduction of 2- and 3-MCPD esters and glycidyl esters in the vegetable oil processing industry. *Compr Rev Food Sci Food Saf.* 2019;18:349–61.  
<https://doi.org/10.1111/1541-4337.12415>
  43. Custodio-Mendoza JA, Arribas García A, Lorenzo Ferreira RA, Carro Díaz AM. Comparative analysis of ultrasound-accelerated alkaline and acid hydrolysis for the indirect determination of 3-monochloropropane-1,2-diol fatty acid esters. *J Food Compos Anal.* 2024;125.  
<https://doi.org/10.1016/j.jfca.2023.105764>
  44. Liu M, Gao BY, Qin F, Wu PP, Shi HM, Luo W *et al.* Acute oral toxicity of 3-MCPD mono- and di-palmitic esters in Swiss mice and their cytotoxicity in NRK-52E rat kidney cells. *Food Chem Toxicol.* 2012;50:3785–91.  
<https://doi.org/10.1016/j.fct.2012.07.038>
  45. Onami S, Cho YM, Toyoda T, Mizuta Y, Yoshida M, Nishikawa A, Ogawa K. A 13-week repeated dose study of three 3-monochloropropane-1,2-diol fatty acid esters in F344 rats. *Arch Toxicol.* 2014;88(4):871–80.  
<https://doi.org/10.1007/s00204-013-1190-6>
  46. Sawada S, Oberemm A, Buhrke T, Meckert C, Rozycki C, Braeuning A, Lampen A. Proteomic analysis of 3-MCPD and 3-MCPD dipalmitate toxicity in rat testis. *Food Chem Toxicol.* 2015;83:84–92.  
<https://doi.org/10.1016/j.fct.2015.06.002>
  47. IARC. Some Chemicals Present in Industrial and Consumer Products, Food and Drinking-Water. Vol 101; 2012
  48. Cho WS, Han BS, Nam KT, Park K, Choi M, Kim SH, Jeong J, Jang DD. Carcinogenicity study of 3-monochloropropane-1,2-diol in Sprague-Dawley rats. *Food Chem Toxicol.* 2008;46:3172–7.  
<https://doi.org/10.1016/j.fct.2008.07.003>
  49. EFSA. Risks for human health related to the presence of 3- and 2-monochloropropanediol (MCPD), and their fatty acid esters, and glycidyl fatty acid esters in food. *EFSA J.* 2016;14.  
<https://doi.org/10.2903/j.efsa.2016.4426>
  50. Samaras VG, Giri A, Zelinkova Z, Karasek L, Buttinger G, Wenzl T. Analytical method for the trace determination of esterified 3- and 2-monochloropropanediol and glycidyl fatty acid esters in various food matrices. *J Chromatogr A.*

- 2016;1466:136–47.  
<https://doi.org/10.1016/j.chroma.2016.08.071>
51. Knutsen HK, Alexander J, Barregård L, Bignami M, Brüschweiler B, Ceccatelli S, *et al.* Update of the risk assessment on 3-monochloropropane diol and its fatty acid esters. *EFSA J.* 2018;16.  
<https://doi.org/10.2903/j.efsa.2018.5083>
  52. Schilter B, Scholz G, Seefelder W. Fatty acid esters of chloropropanols and related compounds in food: Toxicological aspects. *Eur J Lipid Sci Technol.* 2011;113:309–13.  
<https://doi.org/10.1002/ejlt.201000311>
  53. Crews C, Chiodini A, Granvogl M, Hamlet C, Hrnčirik K, Kuhlmann J, *et al.* Analytical approaches for MCPD esters and glycidyl esters in food and biological samples: A review and future perspectives. *Food Addit Contam – Part A.* 2013;30:11–45.  
<https://doi.org/10.1080/19440049.2012.720385>
  54. Kaze N, Watanabe Y, Sato H, Murota K, Kotaniguchi M, Yamamoto H, *et al.* Estimation of the intestinal absorption and metabolism behaviors of 2- and 3-Monochloropropanediol esters. *Lipids.* 2016;51:913–22.  
<https://doi.org/10.1007/s11745-016-4143-z>
  55. Liu PW, Li CI, Huang KC, Liu CS, Chen HL, Lee CC, *et al.* 3-MCPD and glycidol coexposure induces systemic toxicity and synergistic nephrotoxicity via NLRP3 inflammasome activation, necroptosis, and autophagic cell death. *J Hazard Mater.* 2021;405.  
<https://doi.org/10.1016/j.jhazmat.2020.124241>
  56. Lynch BS, Bryant DW, Hook GJ, Nestmann ER, Munro IC. Carcinogenicity of monochloro-1,2-propanediol ( $\alpha$ -chlorohydrin, 3-MCPD). *Int J Toxicol.* 1998;17:47–76.
  57. Bakhiya N, Abraham K, Gürtler R, Appel KE, Lampen A. Toxicological assessment of 3-chloropropane-1,2-diol and glycidol fatty acid esters in food. *Mol Nutr Food Res.* 2011;55:509–21.  
<https://doi.org/10.1002/mnfr.201000550>
  58. Buhrke T, Frenzeli F, Kuhlmann J, Lampen A. 2-Chloro-1,3-propanediol (2-MCPD) and its fatty acid esters: cytotoxicity, metabolism, and transport by human intestinal Caco-2 cells. *Arch Toxicol.* 2015;89:2243–51.  
<https://doi.org/10.1007/s00204-014-1395-3>
  59. Evaluation of certain contaminants in food: Eighty-third report of the Joint FAO/WHO Expert Committee on Food Additives. WHO Technical Report Series; 1002. Geneva, Switzerland: World Health Organization & Joint FAO/WHO Expert Committee on Food Additives; 2017. Available from: <https://iris.who.int/handle/10665/254893>.
  60. Huang G, Xue J, Sun X, Wang J, Yu L. Necroptosis in 3-chloro-1, 2-propanediol (3-MCPD)-dipalmitate-induced acute kidney injury in vivo and its repression by miR-223-3p. *Toxicology.* 2018;406–407:33–43.  
<https://doi.org/10.1016/j.tox.2018.05.015>
  61. Wakabayashi K, Kurata Y, Harada T, Tamaki Y, Nishiyama N, Kasamatsu T. Species differences in toxicokinetic parameters of glycidol after a single dose of glycidol or glycidol linoleate in rats and monkeys. *J Toxicol Sci.* 2012;37:691–8.
  62. Aasa J, Törnqvist M, Abramsson-Zetterberg L. Measurement of micronuclei and internal dose in mice demonstrates that 3-monochloropropane-1,2-diol (3-MCPD) has no genotoxic potency in vivo. *Food Chem Toxicol.* 2017;109:414–20.  
<https://doi.org/10.1016/j.fct.2017.09.019>
  63. Aasa J, Vryonidis E, Abramsson-Zetterberg L, Törnqvist M. Internal doses of glycidol in children and estimation of associated cancer risk. *Toxics.* 2019;7.  
<https://doi.org/10.3390/toxics7010007>
  64. Commission Regulation (EU) amending Regulation (EC) No 1881/2006 as regards maximum levels of 3-monochloropropanediol (3-MCPD), 3-MCPD fatty acid esters and glycidyl fatty acid esters in certain foods. *OJ L.* 2020;131/2. Available from: <https://eur-lex.europa.eu/legal-content/EN/TXT/PDF/?uri=CELEX:32020R1322&qid=1654409203078&from=EN>.
  65. Hamlet CG, Asuncion L, Velíšek J, Doležal M, Zelinková Z, Crews C. Formation and occurrence of esters of 3-chloropropane-1,2-diol (3-CPD) in foods: What we know and what we assume. *Eur J Lipid Sci Technol.* 2011;113:279–303.  
<https://doi.org/10.1002/ejlt.201000480>
  66. Hamlet CG, Sadd PA, Crews C, Velíšek J, Baxter DE. Occurrence of 3-chloro-propane-1,2-diol (3-MCPD) and related compounds in foods: A review. *Food Addit Contam.* 2002;19:619–31.  
<https://doi.org/10.1080/02652030210132391>
  67. Rahn AKK, Yaylayan VA. What do we know about the molecular mechanism of 3-MCPD ester formation? *Eur J Lipid Sci Technol.* 2011;113:323–9.  
<https://doi.org/10.1002/ejlt.201000310>
  68. Craft BD, Chiodini A, Garst J, Granvogl M. Fatty acid esters of monochloropropanediol (MCPD) and glycidol in refined edible oils. *Food Addit Contam – Part A.* 2013;30:46–51.  
<https://doi.org/10.1080/19440049.2012.709196>
  69. Zhang X, Gao B, Qin F, Shi H, Jiang Y, Xu X, *et al.* Free radical mediated formation of 3-monochloropropanediol (3-MCPD) fatty acid diesters. *J Agric Food Chem.* 2013;61:2548–55.  
<https://doi.org/10.1021/jf305252q>
  70. Ramli MR, Siew WL, Ibrahim NA, Kuntom A, Raznim RA. Other factors to consider in the formation of chloropropanediol fatty esters in oil processes. *Food Addit Contam Part A Chem Anal Control Expo Risk Assess.* 2015;32:817–24.  
<https://doi.org/10.1080/19440049.2015.1032368>
  71. Sevindirici G. 3-MCPD and GE risk in refined vegetable oils: structure, formation mechanism, legal regulations and mitigation techniques. *J Food.* 2018:886–95.  
<https://doi.org/10.15237/gjida.gd18053>



72. Shimizu M, Moriwaki J, Shiiba D, Nohara H, Kudo N, Katsuragi Y. Elimination of glycidyl palmitate in diolein by treatment with activated bleaching earth. *J. Oleo Sci.* 2012;61:23–8.
73. Lucas-Torres C, Pérez Á, Cabañas B, Moreno A. Study by <sup>31</sup>P NMR spectroscopy of the triacylglycerol degradation processes in olive oil with different heat-transfer mechanisms. *Food Chem.* 2014;165:21–8.  
<https://doi.org/10.1016/j.foodchem.2014.05.092>
74. Matthäus B, Pudel F. Mitigation of 3-MCPD and glycidyl esters within the production chain of vegetable oils especially palm oil. *Lipid Technol.* 2013;25:151–5.  
<https://doi.org/10.1002/lite.201300288>
75. Shahidi F, Zhong Y. Lipid Oxidation: Measurement Methods. In: *Bailey's Industrial Oil and Fat Products*. London, UK; John Wiley&Sons Inc; 2005. pp. 357–85.  
<https://doi.org/10.1002/047167849X.bio050>
76. Dijkstra AJ, Segers JC. Production and Refining of Oils and Fats. In: *The Lipid Handbook with CD-ROM*. CRC Press; 2007.
77. Aniołowska M, Kita A. The effect of type of oil and degree of degradation on glycidyl esters content during the frying of french fries. *J Am Oil Chem Soc.* 2015;92:1621–31.  
<https://doi.org/10.1007/s11746-015-2715-3>
78. Cheng W, Liu G, Liu X. Formation of glycidyl fatty acid esters both in real edible oils during laboratory-scale refining and in chemical model during high temperature exposure. *J Agric Food Chem.* 2016;64:5919–27.  
<https://doi.org/10.1021/acs.jafc.6b01520>
79. Masukawa Y, Shiro H, Nakamura S, Kondo N, Jin N, Suzuki N, *et al.* A new analytical method for the quantification of glycidol fatty acid esters in edible oils. *J Oleo Sci.* 2010;59:81–8.
80. Destailats F, Craft BD, Dubois M, Nagy K. Glycidyl esters in refined palm (*Elaeis guineensis*) oil and related fractions. Part I: Formation mechanism. *Food Chem.* 2012;131:1391–8.  
<https://doi.org/10.1016/j.foodchem.2011.10.006>
81. Vicente E, Arisseto AP, Furlani RPZ, Monteiro V, Gonçalves LM, Pereira ALD, *et al.* Levels of 3-monochloropropane-1,2-diol (3-MCPD) in selected processed foods from the Brazilian market. *Food Res Int.* 2015;77:310–4.  
<https://doi.org/10.1016/j.foodres.2015.03.035>
82. Šmidrkal J, Tesařová M, Hrádková I, Berčíková M, Adamčíková A, Filip V. Mechanism of formation of 3-chloropropane-1,2-diol (3-MCPD) esters under conditions of the vegetable oil refining. *Food Chem.* 2016;211:124–9.  
<https://doi.org/10.1016/j.foodchem.2016.05.039>
83. Merkle S, Ostermeyer U, Rohn S, Karl H, Fritsche J. Mitigation strategies for ester bound 2-/3-MCPD and esterified glycidol in pre-fried breaded and frozen fish products. *Food Chem.* 2018;245:196–204.  
<https://doi.org/10.1016/j.foodchem.2017.10.076>
84. Tiong SH, Saparin N, Teh HF, Theresa LMN, Zain MZM, Neoh BK, *et al.* Natural organochlorines as precursors of 3-monochloropropanediol esters in vegetable oils. *J Agric Food Chem.* 2018;66:999–1007.  
<https://doi.org/10.1021/acs.jafc.7b04995>
85. Nagy K, Sandoz L, Craft BD, Destailats F. Mass-defect filtering of isotope signatures to reveal the source of chlorinated palm oil contaminants. *Food Addit Contam – Part A.* 2011;28:1492–500.  
<https://doi.org/10.1080/19440049.2011.618467>
86. Collison M. Current status of MCPD and glycidol analysis in oils. In: *104th AOCS Annual Meeting and Expo*. Montreal, Quebec, Canada; 2013.
87. Clemens R, Hayes AW, Sundram K, Pressman P. Palm oil and threats to a critically important food source: The chloropropanols – Caution, controversy, and correction. *Toxicol Res App.* 2017;1.  
<https://doi.org/10.1177/2397847317699844>
88. de Albuquerque TG, Beatriz M, Oliveira PP, Costa HS. 3-MCPD occurrence in vegetable oils: impact on human nutrition and future challenges. *EC Nutr.* 2018;17:455–69.
89. Franke K, Strijowski U, Fleck G, Pudel F. Influence of chemical refining process and oil type on bound 3-chloro-1,2-propanediol contents in palm oil and rapeseed oil. *LWT – Food Sci Technol.* 2009;42:1751–4.  
<https://doi.org/10.1016/j.lwt.2009.05.021>
90. Zhao Y, Zhang Y, Zhang Z, Liu J, Wang YL *et al.* Formation of 3-MCPD fatty acid esters from monostearoyl glycerol and the thermal stability of 3-MCPD monoesters. *J Agric Food Chem.* 2016;64:8918–26.  
<https://doi.org/10.1021/acs.jafc.6b04048>
91. Weißhaar R. 3-MCPD-esters in edible fats and oils – A new and worldwide problem. *Eur J Lipid Sci Technol.* 2008;110:671–2.  
<https://doi.org/10.1002/ejlt.200800154>
92. Özdikicierler O, Yemişçiöğlü F, Saygın Gümüşkesen A. Effects of process parameters on 3-MCPD and glycidyl ester formation during steam distillation of olive oil and olive pomace oil. *Eur Food Res Technol.* 2016;242:805–13.  
<https://doi.org/10.1007/s00217-015-2587-7>
93. Wong YH, Muhamad H, Abas F, Lai OM, Nyam KL, Tan CP. Effects of temperature and NaCl on the formation of 3-MCPD esters and glycidyl esters in refined, bleached and deodorized palm olein during deep-fat frying of potato chips. *Food Chem.* 2017;219:126–30.  
<https://doi.org/10.1016/j.foodchem.2016.09.130>
94. Codex Alimentarius Commission Committee on Food Additives and Contaminants. Proposed draft code of practice for the reduction of 3-monochloropropane-1,2-diol esters (3-MCPDE) and glycidyl esters (GE) in refined oils and products made with refined oils, especially infant formula. In: *Joint FAO/WHO Food Standards Programme*. Utrecht, The Netherlands; 2018.

95. Matthäus B, Pudel F, Fehling P, Vosmann K, Freudenstein A. Strategies for the reduction of 3-MCPD esters and related compounds in vegetable oils. *Eur J Lipid Sci Technol.* 2011;113(3):380–6.  
<https://doi.org/10.1002/ejlt.201000300>
96. FEDIOL. MCPD esters and glycidyl esters. Bruxelles, Belgium: FEDIOL AISBL – The EU Vegetable Oil and Protein Meal Industry; 2015. Available from: <https://www.fediol.eu/data/FEDIOL%20Review%20of%20Mitigation%20Measures%20MCPD%20Esters%20and%20Glycidyl%20Esters%20-%2024%20June%202015.pdf>.
97. Craft BD, Nagy K, Sandoz L, Destailats F. Factors impacting the formation of monochloropropanediol (MCPD) fatty acid diesters during palm (*Elaeis guineensis*) oil production. *Food Addit Contam Part A Chem Anal Control Expo Risk Assess.* 2012;29:354–61.  
<https://doi.org/10.1080/19440049.2011.639034>
98. Kyselka J, Matějková K, Šmidrkal J, Bercikova M, Pesek E, Belkova B *et al.* Elimination of 3-MCPD fatty acid esters and glycidyl esters during palm oil hydrogenation and wet fractionation. *Eur Food Res Technol.* 2018;244:1887–95.  
<https://doi.org/10.1007/s00217-018-3101-9>
99. Craft BD, Nagy K, Seefelder W, Dubois M, Destailats F. Glycidyl esters in refined palm (*Elaeis guineensis*) oil and related fractions. Part II: Practical recommendations for effective mitigation. *Food Chem.* 2012;132:73–9.  
<https://doi.org/10.1016/j.foodchem.2011.10.034>
100. Sulin SN, Mokhtar MN, Mohammed MAP, Baharuddin AS. Review on palm oil contaminants related to 3-monochloropropane-1, 2-diol (3-MCPD) and glycidyl esters (GE). *Food Res.* 2020;4:11–8.  
[https://doi.org/10.26656/fr.2017.4\(56\).051](https://doi.org/10.26656/fr.2017.4(56).051)
101. Mou Y, Sun L, Geng Y, Xie Y, Chen F, Xiao J, *et al.* Chloropropanols and their esters in foods: Exposure, formation and mitigation strategies. *Food Chem Adv.* 2023;3.  
<https://doi.org/10.1016/j.focha.2023.100446>
102. Santiago JK, Silva WC, Capristo MF, Ferreira MC, Ferrari RA, Vicente E, *et al.* Organic, conventional and sustainable palm oil (RSPO): Formation of 2- and 3-MCPD esters and glycidyl esters and influence of aqueous washing on their reduction. *Food Res Int.* 2021;140.  
<https://doi.org/10.1016/j.foodres.2020.109998>
103. Syed Putra SS, Basirun WJ, Elgharbawy AAM, Hayyan M, Abdulmonem WA, Aljohani ASM, *et al.* 3-Monochloropropane-1,2-diol (3-MCPD): a review on properties, occurrence, mechanism of formation, toxicity, analytical approach and mitigation strategy. *J Food Meas Charact.* 2023;17:3592–615.  
<https://doi.org/10.1007/s11694-023-01883-y>
104. Yung YL, Lakshmanan S, Chu CM, Tham HJ, Kumaresan S. Optimization of water washing for mitigation of 3-monochloropropane 1,2 diol ester in palm oil physical refining process. *Food Addit Contam Part A.* 2023;40:1541–50.  
<https://doi.org/10.1080/19440049.2023.2283873>
105. Ramli MR, Tarmizi AHA, Hammid ANA, Razak RAA, Kuntom A, Lin SW, *et al.* Preliminary large scale mitigation of 3-Monochloropropane-1,2-diol (3-MCPD) esters and glycidyl esters in palm oil. *J Oleo Sci.* 2020;69:815–24.  
<https://doi.org/10.5650/jos.ess20021>
106. Nagy K, Redeuil K, Lahrichi S, Nicolas M. Removal of organochlorines from vegetable oils and its benefits in preventing formation of monochloropropanediol diesters. *Food Addit Contam Part A Chem Anal Control Expo Risk Assess.* 2019;36:712–21.  
<https://doi.org/10.1080/19440049.2019.1588999>
107. Kopas PM, Kopas G. Cosmeceutical formulation containing palm oils. US patent US8524292B2. 2013.
108. Oey SB, van der Fels-Klerx HJ, Fogliano V, van Leeuwen SPJ. Chemical refining methods effectively mitigate 2-MCPD esters, 3-MCPD esters, and glycidyl esters formation in refined vegetable oils. *Food Res Int.* 2022;156.  
<https://doi.org/10.1016/j.foodres.2022.111137>
109. Xie D, Zhou H, Jiang X. Effect of chemical refining on the levels of bioactive components and hazardous substances in soybean oil. *J Food Meas Charact.* 2019;13:1423–30.  
<https://doi.org/10.1007/s11694-019-00058-y>
110. Hew KS, Khor YP, Tan TB, Yusoff MM, Lai OM, Asis AJ, *et al.* Mitigation of 3-monochloropropane-1,2-diol esters and glycidyl esters in refined palm oil: A new and optimized approach. *LWT – Food Sci Technol.* 2021;139.  
<https://doi.org/10.1016/j.lwt.2020.110612>
111. Theurillat X, Nicolas M, Redeuil K, Nagy K. Synergistic effects during physical refining of palm oil to mitigate the formation of monochloropropanediols. *Eur J Lipid Sci Technol.* 2020;122.  
<https://doi.org/10.1002/ejlt.202000071>
112. Anis U, Millati R, Hidayat C. Optimization of crude palm (*Elaeis guineensis*) oil bleaching using zeolite-Fe by response surface methodology. *agriTECH.* 2022;42:23.  
<https://doi.org/10.22146/agritech.48114>
113. Restiawaty E, Maulana A, Umi Culsum NT, Aslan C, Suendo V, Nishiyama N, *et al.* The removal of 3-monochloropropane-1,2-diol ester and glycidyl ester from refined-bleached and deodorized palm oil using activated carbon. *RSC Adv.* 2021;11:16500–9.  
<https://doi.org/10.1039/d1ra00704a>
114. Ahn Y, Choi S, Kwak SY. Remarkable effect of deprotonation on adsorption of 3-MCPD and glycidol on carboxylated Fe-MIL-88s. *J Environ Chem Eng.* 2020;8.  
<https://doi.org/10.1016/j.jece.2020.104456>
115. Restiawaty E, Rida FM, Maulana A, Culsum NTU, Saputera WH, Widiatmoko P, *et al.* Adsorption of 3-monochloropropane-1,2-diol ester and glycidyl ester from refined bleached deodorized palm oil using zeolite-based adsor-

- bents. *J Am Oil Chem Soc.* 2024;101(9):893–904.  
<https://doi.org/10.1002/aocs.12838>
116. Zhang L, Wu P, Xiang X, Yang D, Wang L, Hu Z. Optimization of physical refining process of camellia oil for reduction of 3-monochloropropane-1,2-diol (3-MCPD) ester formation using response surface methodology on a laboratory scale. *Molecules.* 2023;28:3616.  
<https://doi.org/10.3390/molecules28083616>
  117. Elmas A, Akpınar Bayızit A. The impact of process conditions on the formation of 3-MCPD and glycidol esters in palm oil. *Bursa Uludağ Üniv Ziraat Fak Derg.* 2021;35(2):333–54.
  118. Tivanello RG, Capristo MF, Leme FM, Ferrari RA, Sampaio KA, Ariseto AP *et al.* Mitigation studies based on the contribution of chlorides and acids to the formation of 3-MCPD, 2-MCPD, and glycidyl esters in palm oil. *ACS Food Sci Technol.* 2021;1:1190–7.  
<https://doi.org/10.1021/acfoodscitech.1c00084>
  119. Tivanello R, Capristo M, Vicente E, Ferrari R, Sampaio K, Ariseto A. Effects of deodorization temperature and time on the formation of 3-MCPD, 2-MCPD, and glycidyl esters and physicochemical changes of palm oil. *J Food Sci.* 2020;85:2255–2260.  
<https://doi.org/10.1111/1750-3841.15304>
  120. Mayayo C, Oey SB, van der Fels-Klerx HJ, van Leeuwen SPJ. Mitigation methods during physical refining for the reduction of 2- and 3-MCPD esters and glycidyl esters in (organic) high oleic sunflower and rapeseed oils. *Food Control.* 2024;162:110480.  
<https://doi.org/10.1016/j.foodcont.2024.110480>
  121. Liu R, Guo X, Cheng M, Zheng L, Gong M, Chang M, *et al.* Effects of chemical refinement on the quality of coconut oil. *J Food Sci Technol.* 2019;56:3109–16.  
<https://doi.org/10.1007/s13197-019-03810-w>
  122. Oey SB, van der Fels-Klerx HJ, Fogliano V, van Leeuwen SPJ. Effective physical refining for the mitigation of processing contaminants in palm oil at pilot scale. *Food Res Int.* 2020; 138.  
<https://doi.org/10.1016/j.foodres.2020.109748>
  123. Bockisch M. *Fats and Oils Handbook.* Bockisch M, editor. Hamburg, Germany: AOCS Press; 1998.  
<https://doi.org/10.1016/C2015-0-02417-0>
  124. Bornscheuer UT, Hessler M. Enzymatic removal of 3-monochloro-1,2-propanediol (3-MCPD) and its esters from oils. *Eur J Lipid Sci Technol.* 2010;112:552–6.  
<https://doi.org/10.1002/ejlt.200900245>
  125. Bel-Rhaid R, Talmon JP, Fay LB, Juillierat MA. Biodegradation of 3-chloro-1,2-propanediol with *Saccharomyces cerevisiae*. *J Agric Food Chem.* 2004;52:6165–9.  
<https://doi.org/10.1021/jf048980k>
  126. Tsai HY, Hsu JN, Fang CJ, Su NW. Determination of glycidyl esters and 3-MCPD esters in edible oils by sample pretreatment with the combination of lipase hydrolysis and modified QuEChERS for GC-MS analysis. *J Food Drug Anal.* 2021;29:153–67.  
<https://doi.org/10.38212/2224-6614.3202>
  127. Miyazaki K, Koyama K. An improved enzymatic indirect method for simultaneous determinations of 3-MCPD esters and glycidyl esters in fish oils. *J Oleo Sci.* 2017;66:1085–93.  
<https://doi.org/10.5650/jos.ess17082>
  128. Chai Q, Hayat K, Karangwa E, Duhoranimana E, Zhang X, Xia S, *et al.* Investigating the optimum conditions for minimized 3-chloropropane-1,2-diol esters content and improved sensory attributes during savory beef flavor preparation. *Food Chem.* 2018;243:96–102.  
<https://doi.org/10.1016/j.foodchem.2017.09.113>
  129. Aniołowska M, Kita A. The effect of frying on glycidyl esters content in palm oil. *Food Chem.* 2016;203:95–103.  
<https://doi.org/10.1016/j.foodchem.2016.02.028>
  130. Han L, Li J, Wang S, Cheng W, Ma L, Liu G, *et al.* Sesame oil inhibits the formation of glycidyl ester during deodorization. *Int J Food Prop.* 2021;24:505–16.  
<https://doi.org/10.1080/10942912.2021.1900236>
  131. Wang X, Cheng W, Wang X, Wang Q, El-Maksoud AAA, Wang M, *et al.* Inhibition effects of typical antioxidants on the formation of glycidyl esters in rice oil and chemical model during high temperature exposure. *LWT – Food Sci Technol.* 2022;166:113794.  
<https://doi.org/10.1016/j.lwt.2022.113794>
  132. Plantinga WJ, Van Toorn WG, van der Stegen GHD. Determination of 3-chloropropane-1,2-diol in liquid hydrolysed vegetable proteins by capillary gas chromatography with flame ionization detection. *J Chromatogr A.* 1991;555: 311–4.  
[https://doi.org/10.1016/S0021-9673\(01\)87196-X](https://doi.org/10.1016/S0021-9673(01)87196-X)
  133. Mezouari S, Liu WY, Pace G, Hartman TG. Development and validation of an improved method for the determination of chloropropanols in paperboard food packaging by GC-MS. *Food Addit Contam Part A.* 2015;32(5):768–78.  
<https://doi.org/10.1080/19440049.2015.1010606>
  134. He B, Wang L, Li M. A biosensor for direct bioelectrocatalysis detection of 3-MCPD in soy sauce using Cyt-c incorporated in Au@AgNSs/FeMOF nanocomposite. *J Iran Chem Soc.* 2021;18:151–8.  
<https://doi.org/10.1007/s13738-020-02011-z>
  135. Huang M, Jiang G, He B, Zhou Q, Fu W, Wu Y. Determination of 3-chloropropane-1,2-diol in liquid hydrolyzed vegetable proteins and soy sauce by solid-phase microextraction and gas chromatography/mass spectrometry. *Anal Sci.* 2005;21:1343–7.  
<https://doi.org/10.2116/analsci.21.1343>
  136. Rétho C, Blanchard F. Determination of 3-chloropropane-1,2-diol as its 1,3-dioxolane derivative at the  $\mu\text{g kg}^{-1}$

- level: Application to a wide range of foods. *Food Addit Contam.* 2005;22:1189–97.  
<https://doi.org/10.1080/02652030500197680>
137. Cao X, Song G, Gao Y, Zhao J, Zhang M, Wu W, *et al.* A novel derivatization method coupled with GC–MS for the simultaneous determination of chloropropanols. *Chromatographia.* 2009;70:661–4.  
<https://doi.org/10.1365/s10337-009-1203-z>
138. Abu-El-Haj S, Bogusz MJ, Ibrahim Z, Hassan H, Al Tufail M. Rapid and simple determination of chloropropanols (3-MCPD and 1,3-DCP) in food products using isotope dilution GC–MS. *Food Control.* 2007;18:81–90.  
<https://doi.org/10.1016/j.foodcont.2005.08.014>
139. Sun C, Wu N, Kou S, Wu H, Liu Y, Pei A *et al.* Occurrence, formation mechanism, detection methods, and removal approaches for chloropropanols and their esters in food: An updated systematic review. *Food Chem X.* 2023;17:100529.  
<https://doi.org/10.1016/j.fochx.2022.100529>
140. Brereton P, Kelly J, Crews C, Honour S, Wood R, Davies A *et al.* Determination of 3-chloro-1,2-propanediol in foods and food ingredients by gas chromatography with mass spectrometric detection: Collaborative study. *J AOAC Int.* 2001;84:455–65.  
<https://doi.org/10.1093/jaoac/84.2.455>
141. Hu Z, Cheng P, Guo M, Zhang W, Qi Y. A novel approach of periodate oxidation coupled with HPLC–FLD for the quantitative determination of 3-chloro-1,2-propanediol in water and vegetable oil. *J Agric Food Chem.* 2013;61:6614–21.  
<https://doi.org/10.1021/jf400167f>
142. Qiu X, Xu XY, Liang Y, Guo H. The molecularly imprinted polymer supported by anodic alumina oxide nanotubes membrane for efficient recognition of chloropropanols in vegetable oils. *Food Chem.* 2018;258:295–300.  
<https://doi.org/10.1016/j.foodchem.2018.03.071>
143. Chung HY, Kumar Ponnusamy V, Jen JF. Determination of 3-chloropropanediol in soy sauce samples by liquid phase extraction coupled with microwave-assisted derivatization and high performance liquid chromatography-ultra-violet detection. *Int J Eng Res Sci.* 2018;4:54–61.
144. Sun X, Zhang L, Zhang H, Qian H, Zhang Y, Tang L, *et al.* Development and application of 3-chloro-1,2-propanediol electrochemical sensor based on a polyaminothiophenol modified molecularly imprinted film. *J Agric Food Chem.* 2014;62:4552–7.  
<https://doi.org/10.1021/jf4055159>
145. Yuan Y, Wang J, Ni X, Cao Y. A biosensor based on hemoglobin immobilized with magnetic molecularly imprinted nanoparticles and modified on a magnetic electrode for direct electrochemical determination of 3-chloro-1,2-propanediol. *J Electroanal Chem.* 2019; 834:233–40.  
<https://doi.org/10.1016/j.jelechem.2018.12.034>
146. Fang M, Zhou L, Zhang H, Liu L, Gong ZY. A molecularly imprinted polymers/carbon dots-grafted paper sensor for 3-monochloropropane-1,2-diol determination. *Food Chem.* 2019;274:156–61.  
<https://doi.org/10.1016/j.foodchem.2018.08.133>
147. Martin AA, Fodjo EK, Marc GBI, Albert T, Kong C. Simple and rapid detection of free 3-monochloropropane-1,2-diol based on cysteine modified silver nanoparticles. *Food Chem.* 2021;338:127787.  
<https://doi.org/10.1016/j.foodchem.2020.127787>