

RHEUMATOLOGY

Clinical science

Multiple serum biomarkers associate with mortality and interstitial lung disease progression in systemic sclerosis

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Abstract

Objectives: To investigate the prognostic utility of 28 serum biomarkers in systemic sclerosis (SSc), SSc-associated interstitial lung disease (SSc-ILD) and clinically relevant disease subgroups.

Methods: Participants with sera, high-resolution CT and lung function within 12 months of baseline were identified from the Australian Scleroderma Cohort Study. Baseline was the time of serum collection. Twenty-seven of the prespecified 28 serum biomarkers were analysed and biomarker associations with mortality and ILD progression were investigated in univariable and multivariable analyses, including within disease subgroups and combined with established risk factors for poorer prognosis in SSc.

Results: A total of 407 participants were identified, 252 (61.9%) with SSc-ILD. The median (interquartile range) follow-up after biomarker measurement was 6.31 (3.11–9.22) years. Sixteen biomarkers were associated with increased mortality. High levels of VCAM-1 were most strongly associated with mortality [hazard ratio (HR) 3.55; 95% CI 2.37–5.33; P < 0.001]. Five additional biomarkers had an HR >2: SP-D (2.28, 1.57–3.31; P < 0.001), E-selectin (2.19, 1.53–3.14; P < 0.001), IL-6 (2.15, 1.50–3.09; P < 0.001), MMP-3 (2.05, 1.42–2.95; P < 0.001) and ET-1 (2.03, 1.40–2.92; P < 0.001). Eleven biomarkers were independently associated with mortality following adjustment for sex, age and baseline forced vital capacity (FVC% predicted). Three biomarkers were associated with ILD progression at 1-year follow-up: CXCL4 (odds ratio 2.67, 1.46–4.88; P = 0.001), MMP-1 (2.56, 1.43–4.59; P = 0.002) and ET-1 (2.18, 1.24–3.83; P = 0.007).

Conclusion: Multiple biomarkers, especially VCAM-1, E-selectin, SP-D and CXCL4, provide prognostic utility beyond that of established risk factors for patients with SSc.

Keywords: scleroderma, prognosis, VCAM-1, E-selectin, SP-D, CXCL4, FVC.

Rheumatology key messages

- Multiple serum biomarkers provide prognostic utility beyond that of the previously established risk factors.
- VCAM-1 is strongly associated with mortality in patients with SSc.
- CXCL4 is associated with progression of SSc-associated interstitial lung disease in univariable analysis.

Introduction

SSc is a multisystem autoimmune disease characterized by tissue and organ fibrosis, vasculopathy and inappropriate inflammation [1]. Patients with SSc have >200-fold higher

mortality compared with healthy individuals. Disappointingly, despite improvements in management, overall survival has only marginally improved over the last few decades [2, 3].

Received: 16 August 2023. Accepted: 14 December 2023

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A major contributor to mortality in patients with SSc is interstitial lung disease (ILD), present in up to 80% of patients with SSc [3, 4]. Not all patients with SSc-associated ILD (SSc-ILD) will have clinically significant ILD and there is wide variation in the disease course for SSc-ILD [5]. Some patients may have stable disease over extended timeframes even without specific therapy, while others have inexorable progression despite optimal therapy, leading to respiratory failure and death. As potential therapeutic options for SSc improve but generally only attenuate progression of disease, accurate prognostication for the individual patient has never been more important.

Clinical and demographic prognostic risk factors for SSc-ILD include male sex, age and lower forced vital capacity (FVC) [6]. Alongside these, several serum biomarkers have been studied with some success [7]. However, to date, no biomarker or combination of biomarkers has been validated across multiple clinical cohorts to predict prognosis in SSc-ILD [8].

In our earlier work, we performed a comprehensive literature review of serum biomarkers in SSc-ILD and identified 28 biomarkers for further study and have evaluated these biomarkers for the identification of SSc-ILD [9, 10]. In the present study, our primary objective was to evaluate the prognostic utility of these 28 serum biomarkers in patients with SSc and SSc-ILD, alone and in combination with established clinical and demographic risk factors. Pre-specified subgroups including limited and dcSSc, patients with limited or extensive ILD at baseline, early disease and the presence of pulmonary hypertension at baseline, were identified for subgroup analysis.

Methods

Participants

The Australian Scleroderma Cohort Study (ASCS) is a national, multicentre, longitudinal observational cohort established in 2007 by the Australian Scleroderma Interest Group (ASIG). All Australian physicians are invited to refer SSc patients to 12 national screening centres. Patients fulfilling ACR/EULAR classification for SSc are eligible for enrolment [11]. Participants provide written consent at recruitment. Utilizing a standardized protocol, comprehensive deidentified data are recorded on consecutive patients at baseline and every 12 months including clinical assessment, investigations, treatment and outcomes, including vitality and lung transplantation. High resolution CT (HRCT) scan of the chest is performed when clinically indicated according to physician judgement. Disease extent on HRCT, defined according to the method of Goh et al. [12], is available on a subset of patients from prior studies. Selected centres have baseline and serial serum samples stored in a linked biobank. Clinical management remains as per the treating physician.

Study participants were identified through the ASCS. All patients with sera, HRCT to confirm presence or absence of ILD and lung function within 12 months of baseline at the time of study commencement (1 February 2017) were included. SSc-ILD was defined as SSc with radiologically confirmed ILD on HRCT. Patients with insufficient data to confirm ILD status or serum were excluded.

Our study complies with the Declaration of Helsinki. The Australian Scleroderma Cohort Study has human research ethics approval from all participating centres. Approval for this study was granted by the Research Ethics and Governance Office (protocol numbers X16-0311 & LNR/16/ RPAH/406 RPAH zone, SVHM HREC LRR 0121/20). Written informed consent was obtained from all participants.

Definitions

Baseline was defined as the time of serum collection. Disease duration was defined as time from first non-Raynaud symptom until baseline. Percentage predicted values for forced vital capacity (FVC%) were reported using the *Quanjer* Global Lung Initiative 2012 predictive equation [13]. ILD extent was defined as per Goh *et al.* with 'limited disease' considered for <20% involvement on HRCT and 'extensive disease' for >20% involvement [12]. Disease subgroup (limited or diffuse) was defined by enrolling physician. In the absence of a widely accepted specific definition, early disease was defined as <5 years from first non-Raynaud's disease manifestation to biomarker collection.

Pulmonary arterial hypertension (PAH) was defined using right heart catheter-measured haemodynamic criteria applying 2019 ESC/ERS criteria. In a small proportion of participants where peripheral vascular resistance was not recorded the pre-existing definition of PAH was applied [14, 15]. Mortality was considered as all-cause mortality. ILD progression was defined as a relative decline in FVC of \geq 10% from baseline FVC [8]. Follow-up data were censored on 1 May 2019.

Serum samples and biomarker measurement

Serum samples were obtained from the ASCS biobanks and stored at -80°C until analysis. Sera with more than two freeze-thaw cycles were excluded. Twenty-eight biomarkers were selected by comprehensive literature review [9]. Equipment required to measure KL-6 was unavailable. The remaining 27 biomarkers were analysed at a single laboratory following manufacturer's protocols by magnetic Luminex (SP-D, MMP-1, MMP-3, MMP-7, MMP-12, TIMP-1, Ca15-3, periostin, CCL-2, IL-6, VEGF, IL-8, CXCL10, CXCL12, CXCL13, E-selectin, CXCL4, CCL-18, ICAM-1, VCAM-1), bead-based assay (TGF- β 1, - β 2, - β 3) and ELISA (amphiregulin, fibulin-1, LOXL-2, ET-1). Non-abbreviated biomarker labels and assay details are shown in Supplementary Table S1 (available at Rheumatology online). Any biomarkers with >25% of results below detectable range in initial analysis were re-tested with higher sensitivity assays. Individuals with undetectable biomarker values were imputed at half the lower limit of detection. Individual results above the quantitative threshold were imputed at maximum quantifiable value.

Statistical analysis

Continuous variables are reported as mean (s.D.) or median [interquartile range (IQR)] as appropriate, and categorical variables as an absolute number (relative frequency). *P*-values <0.05 were considered statistically significant.

Biomarker values were dichotomized with threshold values (in pg/ml) calculated using the Liu method of optimal cut-point estimation for a diagnostic test for the relevant outcome. Biomarker outcome associations were studied in univariable and multivariable analysis. In multivariable analysis, biomarkers were analysed with gender, age and baseline FVC%. The results of these analyses are expressed as a hazard ratio (HR) of outcome with 95% CI and P-value. All statistical analysis was performed using Stata statistical software (StataCorp. 2023. Stata Statistical Software: Release 15, College Station, TX: StataCorp LLC).

Results

Of 407 SSc participants identified, 252 (61.9%) had SSc-ILD. Patients with SSc-ILD had a median age of 58.7 years (IQR 50.6–66.7), 202 (80.2%) were female, mean FVC% was 87.5% (\pm 21.9%) with a median follow-up 6.50 years (IQR 2.9–9.2). Demographic and selected clinical features are detailed in Table 1. Participants with ILD were more likely to be non-Caucasian, have diffuse disease (39.7%), be anti-Scl70 positive (35.6%) and anti-centromere negative (19.6% positive), have a lower baseline FVC% and increased mortality (see Supplementary Fig. S1, available at *Rheumatology* online). HRCT data to establish ILD extent ('limited' or 'extensive' as defined in methods) was only available for 197/252 (78.2%) participants.

Mortality in entire SSc cohort

In univariable analysis of all participants, 16 biomarkers were associated with increased mortality (Table 2). High levels of VCAM-1 were most strongly associated with mortality (HR 3.55, 95% CI 2.37–5.33; P < 0.001). A further five biomarkers had a HR above 2.0, including SP-D (HR 2.28, 1.57–3.31; P < 0.001), E-selectin (HR 2.19, 1.53–3.14; P < 0.001), IL-6 (HR 2.15, 1.50–3.09; P < 0.001), MMP-3 (HR 2.05, 1.42–2.95; P < 0.001) and ET-1 (HR 2.03, 1.40–2.92; P < 0.001).

In multivariable analysis, combined with age, sex and baseline FVC%, 11 biomarkers remained significantly associated with increased mortality (Table 3). The strongest biomarker association with mortality was VCAM-1 (HR 2.85, 1.86– 4.38; P < 0.001).

Mortality in SSc-ILD and SSc no-ILD groups

Table 1. Participant characteristics

In SSc-ILD participants, 12 biomarkers were associated with increased mortality on univariable analysis (Table 2). VCAM-1 had the strongest association (HR 4.04, 2.37–6.90; P < 0.001). In multivariable analysis, seven biomarkers

remained significant after adjusting for age, sex and baseline FVC% (Table 3). VCAM-1 and E-selectin had the strongest associations with mortality. Interestingly, CXCL4, which was not found to be associated with mortality on multivariable analysis in the total cohort or the SSc no-ILD group, predicted mortality in this SSc-ILD cohort.

In the SSc no-ILD group, nine biomarkers were associated with mortality on univariable analysis (Table 2). The strongest associations were with CXCL12 (HR 4.23, 1.95–9.17; P < 0.001) and VCAM-1 (HR 3.99, 1.90–8.37; P < 0.001). Interestingly, ET-1 (3.42, 1.67–7.02; P = 0.001) had a stronger association with mortality in this group without ILD at baseline. CCL-18 was associated with mortality in this group but not in participants with ILD at baseline. In multivariable analysis (Table 3), the strongest association with mortality was again with CXCL12 followed by VCAM-1.

Subgroup analyses

We performed univariable and multivariable survival analyses in pre-specified subgroups including early disease, disease subtype (diffuse and limited), ILD extent (limited and extensive ILD on baseline HRCT) and the presence or absence of PAH. The univariable Cox regression analyses are presented in the Supplementary material (Supplementary Table S2, available at *Rheumatology* online). The multivariable analyses, adjusting for age, sex and baseline FVC%, are shown in Table 4.

Elevated VCAM-1 had the strongest association with increased mortality across all subgroups, with the exception of patients with extensive ILD at baseline and those with PAH. E-selectin also was associated with mortality across all subgroups except in participants with extensive ILD at baseline.

CXCL12 was associated with mortality in participants with early disease, limited disease, limited extent ILD and without PAH. In patients with more extensive ILD at baseline, SP-D, CCL-18, MMP-1 and Amphiregulin were associated with increased mortality.

Progression of lung disease in SSc-ILD

In the 252 participants with SSc-ILD, CXCL4 (HR 1.38, 1.2– 1.88; P = 0.039) was associated with ILD progression over the entire study period, on univariable analysis. This

Variable		SSc $(n = 407)$	SSc-ILD (<i>n</i> = 252)	SSc No-ILD $(n = 155)$	P-value
Age at biomarker collection		58.81 (50.51-66.40)	58.67 (50.56-66.69)	59.24 (49.46-65.51)	0.719
Disease duration, years		8.29 (3.38-16.98)	6.83 (2.52–15.63)	7.65 (2.43-15.58)	0.742
Follow-up period, years		6.31 (3.11–9.22)	6.50 (2.85-9.16)	6.09 (3.56-9.45)	0.699
Sex, $n(\%)$	Male	80 (19.7)	50 (19.8)	30 (19.4)	0.905
	Female	327 (80.3)	202 (80.2)	125 (80.6)	
Caucasian, $n(\%)$	No	45 (11.1)	37 (14.7)	8 (5.2)	0.003
	Yes	361 (88.9)	215 (85.3)	146 (94.8)	
Disease subtype, $n(\%)$	Limited	272 (66.8)	152 (60.3)	120 (77.4)	< 0.001
	Diffuse	135 (33.2)	100 (39.7)	35 (22.6)	
Antibody profile, $n(\%)$	ANA positive	386 (94.8)	242 (96.0)	144 (92.9)	0.166
	Centromere	124 (30.7)	49 (19.6)	75 (48.7)	< 0.001
	Scl-70	105 (26.1)	89 (35.6)	16 (10.5)	< 0.001
	RNA Polymerase III	49 (15.2)	31 (15.3)	18 (15.1)	0.972
FVC%	,	92.4 (22.60)	87.5 (21.88)	100.4 (21.5)	< 0.001
PAH, n (%)		70 (17.2)	42 (16.7)	28 (18.1)	0.717
Died during follow-up, n (%)		120 (29.5)	85 (33.7)	35 (22.6)	0.017
FVC% drop >10% during follow-up, <i>n</i> (%)		281 (72.6)	178 (75.4)		

Bold typeface used for statistically significant results ($P \le 0.05$).

SSc-ILD: SSc-associated interstitial lung disease; FVC%: forced vital capacity percentage predicted; PAH: pulmonary arterial hypertension.

Biomarkers	SSc- Total (n = 407) Threshold (pg/mL)	HR (95% CI), P-value	SSc-ILD $(n = 252)$ Threshold (pg/mL)	HR (95% CI), P-value	SSc no- ILD (n = 155) Threshold (pg/mL)	HR (95% CI), <i>P</i> -value
VCAM-1	1 176 536.8	3.55 (2.37–5.33), <i>P</i> < 0.001	1103174.7	4.04 (2.37–6.90), <i>P</i> < 0.001	1203068.7	3.99 (1.90–8.37), <i>P</i> < 0.001
SP-D	33 023.6	2.28 (1.57–3.31), <i>P</i> < 0.001	32894.5	2.06 (1.29–3.27), $P = 0.002$	22099	2.85 (1.37–5.95), $P = 0.005$
E-selectin	40 267.0	2.19 (1.53–3.14), <i>P</i> < 0.001	40267.0	2.22 (1.44–3.43), <i>P</i> < 0.001	37989.1	2.06 (1.05–4.04), $P = 0.036$
IL-6	4.3	2.15 (1.50–3.09), <i>P</i> < 0.001	4.4	2.08 $(1.36-3.19), P = 0.001$	3.55	2.04 (1.04–3.99), $P = 0.037$
MMP-3	14 226.4	2.05 (1.42–2.95), <i>P</i> < 0.001	14526.5	2.36 (1.52–3.68), <i>P</i> < 0.001	NS	NS
ET-1	11.8	2.03 (1.40–2.92), <i>P</i> < 0.001	10.6	1.67 (1.08–2.58), $P = 0.022$	13.0	3.42 (1.67–7.02), <i>P</i> = 0.001
MMP-7	2069.5	1.99 (1.39–2.85), <i>P</i> < 0.001	2281.7	1.92 (1.25–2.96), <i>P</i> = 0.003	1753.1	2.22 (1.13–4.34), <i>P</i> = 0.020
CXCL12	482.6	1.90 (1.32–2.73), $P = 0.001$	485.7	2.18 (1.41–3.35), <i>P</i> < 0.001	682.55	4.23 (1.95–9.17), <i>P</i> < 0.001
CXCL13	81.0	1.88 (1.31–2.70), $P = 0.001$	80.5	1.85 (1.20–2.86), $P = 0.005$	96.6	2.01 (1.03–3.92), <i>P</i> = 0.040
MMP-1	2256.5	1.68 (1.17–2.42), <i>P</i> = 0.005	2285.5	1.70 (1.10–2.62), <i>P</i> = 0.016	NS	NS
CCL-18	85 515.0	1.61 (1.12–2.30), $P = 0.009$	NS	NS	81596.766	2.03 (1.04–3.97), <i>P</i> = 0.037
CA15-3	49.6	1.56 (1.09–2.24), $P = 0.015$	51.0	1.67 (1.09–2.58), $P = 0.019$	NS	NS
CCL-2	534.4	1.56 (1.08–2.24), $P = 0.018$	NS	NS	NS	NS
VEGF	99.4	1.56 (1.07–2.27), $P = 0.021$	NS	NS	NS	NS
Periostin	173 525.6	1.55 (1.08–2.22), $P = 0.017$	174842.5	1.70 (1.11–2.61), $P = 0.016$	NS	NS
CXCL/IL-8	19.8	1.47 (1.02–2.10), $P = 0.036$	NS	NS	NS	NS
Age ^a		1.06 (1.04–1.08), <i>P</i> < 0.001		1.06 (1.04–1.08), <i>P</i> < 0.001		1.06 (1.03–1.10), <i>P</i> < 0.001
Sex (male)		2.11 (1.43–3.11), <i>P</i> < 0.001		2.62 (1.67–4.12), <i>P</i> < 0.001		1.27 (0.58 - 2.81), P = 0.55
FVC% baselin	e ^b	0.98 (0.97–0.99), <i>P</i> < 0.001		0.98 (0.97–0.99), <i>P</i> < 0.001		0.99 (0.97 - 1.00), P = 1.27

Table 2. Biomarkers associated with mortality in univariable analysis

Bold typeface used for statistically significant results ($P \le 0.05$).

^a Per year increase.

^b Per percentage point increase. SSc-ILD: SSc-associated interstitial lung disease; SSc no-ILD: SSC, no interstitial lung disease; FVC%: forced vital capacity percentage predicted; NS: not significant.

Table 3. Biomarkers associated with mortality in multivariable analysis combined with gender, age and baseline	FVC%

Biomarker	SSc-Total $(n = 407)$	SSc-ILD $(n=252)$	SSc no-ILD $(n = 155)$
	HR (95% CI), <i>P</i> -value	HR (95% CI), <i>P</i> -value	HR (95% CI), <i>P</i> -value
VCAM-1	2.85 (1.86–4.38), <i>P</i> < 0.001	3.11 (1.77–5.46), <i>P</i> < 0.001	3.61 (1.66–7.86), <i>P</i> = 0.001
E-selectin	2.20 (1.51–3.20), <i>P</i> < 0.001	2.29 (1.46–3.61), <i>P</i> < 0.001	2.03 (1.03–4.03), $P = 0.042$
SP-D	1.94 $(1.33-2.83)$, $P = 0.001$	1.81 (1.13–2.89), $P = 0.014$	2.59 $(1.24-5.43)$, $P = 0.012$
ET-1	1.82 $(1.25-2.65)$, $P = 0.002$	NS	3.11 (1.51–6.42), $P = 0.002$
CXCL12	1.75 $(1.21-2.54)$, $P = 0.003$	1.89 (1.20–2.95), $P = 0.006$	4.58 (1.98–10.58), <i>P</i> < 0.001
CCL-18	1.66 $(1.15-2.39), P = 0.006$	NS	2.01 $(1.03-3.95)$, $P = 0.042$
IL-6	1.65 $(1.12-2.42), P = 0.010$	NS	2.02 (1.01–4.02), $P = 0.046$
MMP-3	1.65 (1.12–2.42), $P = 0.010$	1.79 (1.12–2.86), $P = 0.015$	NS
MMP-7	1.62 $(1.12-2.35)$, $P = 0.010$	NS	2.05 (1.03–4.08), $P = 0.040$
Ca15-3	1.56 $(1.08-2.24), P = 0.018$	1.71 (1.10–2.65), $P = 0.017$	NS
MMP-1	1.54(1.06-2.22), P = 0.023	NS	NS
CXCL4	NS	1.87 (1.20–2.92), <i>P</i> = 0.005	NS

Bold typeface used for statistically significant results ($P \le 0.05$). SSc-ILD, SSc-associated interstitial lung disease; SSc no-ILD, SSc, no interstitial lung disease; NS, not significant; FVC%, forced vital capacity percentage predicted.

association did not remain significant when adjusted for sex, age and baseline FVC%.

Three biomarkers were associated with ILD progression at 1 year: CXCL4 [odds ratio (OR) 2.67, 1.46–4.88, 1.67; P = 0.001], MMP-1 (OR 2.56, 1.43–4.59; P = 0.002) and ET-1 (OR 2.18, 1.24–3.83; P = 0.007). The relationship of biomarkers and ILD progression in the pre-specified subgroups are shown in Table 5. E-selectin was negatively associated with ILD progression in participants with limited disease (HR 0.68, 0.47–1.00; P = 0.049) and PAH (HR 0.42, 0.20–0.90; P = 0.025).

Discussion

This study describes the largest panel of biomarkers measured concurrently in a national, well-phenotyped cohort of patients with SSc. All the biomarkers studied had been previously demonstrated to have associations with potentially clinically meaningful outcomes. In this study, we analysed the biomarkers collectively and with established risk factors to determine which biomarker individually contributes most powerfully to the outcomes of interest. Accurate prognostication may help inform management strategies for screening, treatment interventions and intensity.

The most striking association in this study was between increased mortality and elevated levels of VCAM-1. VCAM-1 demonstrated a strong association with mortality in participants both with and without ILD, maintained when accounting for established risk factors for worse prognosis, age, gender and FVC%. This relationship remained strong for participants both with and without ILD at baseline as well as in clinically relevant subgroups, except participants with

Biomarkers associate with mortality and ILD progression in systemic sclerosis

Table 4. Biomarkers associated with mortality in multivariable subgroup analysis

Subgroup		Biomarker	Adjusted HR (95% CI), P-value
Early disease $(n = 136)$		VCAM-1	4.99 (1.85–13.44), <i>P</i> = 0.001
		E-selectin	3.95(1.78-8.77), P = 0.001
		CCL-2	3.60 $(1.55-8.35)$, $P = 0.003$
		ET-1	3.14(1.08-9.13, P=0.035)
		CXCL12	3.08 (1.40–6.78), $P = 0.005$
		CCL-18	2.82 (1.24–6.45), $P = 0.014$
		MMP-1	2.73 $(1.28-5.81), P = 0.009$
		TIMP-1	2.58 (1.16–5.72), $P = 0.020$
		CXCL/IL-8	2.31 (1.07–5.02), $P = 0.034$
Disease subtype	Diffuse disease, $n = 135$	TIMP-1	3.59(1.77-7.29), P < 0.001
71	,	VCAM-1	2.99 (1.41–6.36), $P = 0.004$
		E-selectin	2.87 (1.43–5.75), $P = 0.003$
		Ca15-3	2.42 (1.24–4.75), $P = 0.010$
		SP-D	2.41 (1.20–4.82), $P = 0.013$
		CCL-18	2.19 (1.14–4.21), $P = 0.019$
		CXCL4	2.08 (1.10–3.93), $P = 0.024$
		MMP-1	2.01 (1.03–3.92), $P = 0.042$
	Limited disease, $n = 272$	VCAM-1	2.89 $(1.71 - 4.89), P < 0.001$
		MMP-3	2.42 $(1.52 - 3.86), P < 0.001$
		IL-6	2.25 $(1.41 - 3.60)$, $P = 0.001$
		ET-1	2.10 (1.31–3.35), $P = 0.002$
		MMP-7	2.04 $(1.27-3.26), P = 0.003$
		E-selectin	2.01 $(1.26-3.19), P = 0.003$
		SP-D	1.96 $(1.21 - 3.19), P = 0.006$
		CXCL12	1.78 (1.14–2.79), $P = 0.011$
		CXCL13	1.66 $(1.05-2.64), P = 0.031$
ILD extent	ILD extent $< 20\%$, $n = 111$	VCAM-1	2.79(1.16-6.71), P = 0.022
		E-selectin	2.41 (1.16–5.03), $P = 0.019$
		CXCL12	2.40 $(1.14-5.06)$, $P = 0.021$
		IL-6	2.27 (1.06–4.86), $P = 0.034$
	ILD extent >20%, $n = 86$	SP-D	2.58 $(1.39-4.76)$, $P = 0.003$
		CCL-18	2.48 (1.29–4.77), $P = 0.007$
		MMP-1	2.25 $(1.18-4.29), P = 0.014$
		Amphiregulin	2.16 $(1.12-4.19)$, $P = 0.022$
PAH ^a	Yes, $n = 70$	SP-D	2.64 (1.27–5.47), $P = 0.009$
		E-selectin	2.30 (1.18–4.48), $P = 0.014$
	No, <i>n</i> = 337	VCAM-1	2.86 (1.66–4.90), <i>P</i> < 0.001
		CCL-18	1.86 (1.17–2.97), <i>P</i> = 0.009
		E-selectin	1.81 (1.12–2.91), <i>P</i> = 0.015
		MMP-3	1.75 $(1.07-2.84), P = 0.024$
		CXCL12	1.75 $(1.08-2.81), P = 0.022$
		CXCL4	1.59 $(1.00-2.54), P = 0.050$

Bold typeface used for statistically significant results ($P \le 0.05$).

^a PAH was not investigated for with right heart catheter study in every patient in this cohort. ILD: interstitial lung disease; PAH: pulmonary arterial hypertension.

extensive ILD and PAH. This would suggest that elevated VCAM-1 may be an important prognostic marker in patients with SSc in general. The weaker association in participants with extensive ILD and PAH was interesting as these subgroups of patients tend to have higher mortality. Possible explanations for this finding include that VCAM-1 may be a marker of progression independent of ILD and PAH disease manifestations, or that VCAM-1 may predict progression prior to these manifestations which, when established, progress in association with other biomarkers.

VCAM-1 is a 90-kDa glycoprotein predominantly expressed in endothelial cells with expression upregulated by pro-inflammatory cytokines such as TNF- α . It is a transmembrane protein involved in the adhesion and trans-endothelial migration of leucocytes. In some diseases, VCAM-1 is also expressed on tissue macrophages, dendritic cells and other cell types potentially relevant to SSc pathophysiology [16]. Elevated VCAM-1 has previously been associated with mortality in SSc and clinical progression in SSc [17] and idiopathic pulmonary fibrosis [18].

Two additional biomarkers, E-selectin and SP-D, were consistently associated with mortality in our study. E-selectin was associated with mortality on univariable and multivariable analysis in all subgroups, except for participants with extensive ILD. E-selectin is a cell adhesion molecule expressed solely by endothelial cells. Levels have been found to be high in patients with SSc [19]. E-selectin has been correlated with disease activity in SSc and systemic organ involvement, particularly renal crisis [20, 21].

SP-D similarly was associated with mortality including in all subgroups except in early disease, limited extent ILD and in patients without PAH. SP-D may predict progression in participants who have already developed major internal organ manifestations.

SP-D is a pulmonary surfactant lipoprotein and elevated serum levels have been found to reflect the extent of damage to Table 5. Biomarkers associated with ILD progression in univariable subgroup analysis

Subgroup			Biomarker: HR (95% CI), P-value
Early disease	<i>n</i> = 87		MMP-3: 1.89 (1.08–3.33), <i>P</i> = 0.027
			CCL-18: $1.79 (1.07 - 3.00), P = 0.028$
Disease subtype		Diffuse disease, $n = 100$	E-selectin: 2.18 (1.19–4.01), P = 0.012
71			ICAM-1: 2.03 (1.23–3.36), P = 0.006
			LOXL2: 1.83 $(1.14-2.94), P = 0.012$
		Limited disease, $n = 152$	CXCL4: 1.84 (1.12–3.01), P = 0.016
			Ca15-3: 1.51 (1.02–2.23), $P = 0.038$
			E-selectin: 0.68 (0.47–1.00), P = 0.049
			ICAM-1: 0.64 (0.44–0.93), P = 0.019
ILD extent		ILD extent <20%, $n = 111$	MMP-7: 2.06 $(1.20-3.54)$, $P = 0.009$
		ILD extent $>20\%$, $n = 86$	CXCL4: 1.92 (1.13–3.28), $P = 0.016$
			ET-1: 1.83 (1.06–3.13), <i>P</i> = 0.029
			Ca15-3: 1.76 (1.03–3.00), $P = 0.039$
			CCL-18: 1.72 (1.02–2.89), $P = 0.040$
PAH ^a		Yes, $n = 42$	CXCL4: 3.97 $(1.21-13.05)$, $P = 0.023$
			MMP-1: 2.01 $(1.00-4.03)$, $P = 0.050$
			E-selectin: $0.42 (0.20-0.90), P = 0.025$
		No, <i>n</i> = 210	N/A

Bold typeface used for statistically significant results ($P \le 0.05$).

^a PAH was not investigated for with right heart catheter study in every patient in this cohort. ILD: interstitial lung disease; PAH: pulmonary arterial hypertension.

the capillary/epithelial barrier in ILD [22]. SP-D levels have been found to be elevated in patients with SSc-ILD [23] and may reflect the severity of SSc-ILD [24–27]. Levels of SP-D have been found to be higher in patients with SSc-ILD compared with those without ILD [10, 24, 28]. In a large prospective cohort, SP-D values combined with anti-Scl-70 (topoisomerase I) antibodies were used to accurately identify patients with SSc-ILD [28] but SP-D levels were not correlated with the severity of lung disease, its progression or mortality. Our data suggests there is an important correlation between mortality and elevated SP-D levels.

Other biomarker associations appear to be more specific for certain participant subgroups. MMP-3 was only associated with increased mortality in participants with ILD at baseline and in those with limited disease. Amphiregulin was only associated with mortality in those with extensive ILD at baseline. The association between CXCL12 and mortality was particularly strong in participants without ILD at baseline, where it was the strongest single biomarker association. Elevated levels of CXCL12 (or elevated cellular expression of the corresponding receptor CXCR4) have been found in previous studies of patients with SSc, but there are no previous reports of a link to mortality or of disproportionate expression in particular disease subgroups [29-31]. Genetic polymorphisms in the SDF1 gene that would be predicted to result in increased levels of CXCL12 have been found to be more common in SSc patients with PAH and/or digital ulcers [32]. ET-1 was particularly associated with mortality in participants with early and limited disease.

The associations between biomarkers and mortality were strongest in participants with early disease. This may in part link to the natural history of the disease where patients with longer established disease are less likely to progress. Additionally, in patients with more severe ILD, lower FVC% is a powerful clinical predictor, and so it is difficult to demonstrate any additional prognostic benefit beyond FVC% in this cohort. We have demonstrated that multiple serum biomarkers improve prognostic accuracy beyond that of established risk factors. This is particularly relevant in this group of patients with early disease—knowing early which patients are likely to do worse could influence treatment decisions and hopefully help better target therapies to those patients who are most likely to benefit.

Collectively our data suggests that multiple biomarkers could add to established risk factors in better predicting mortality in patients with a broad spectrum of SSc. It may be a that a combination of biomarkers including some that are useful in all patients combined with others more specific to the individual phenotype will provide the best performance.

Our study demonstrated potentially important associations between CXCL4 and ILD disease progression, as defined by decline in FVC%. CXCL4 was associated with progression in participants with ILD at baseline on univariable analysis. CXCL4 is released predominantly from activated platelets and has a chemotactic activity on neutrophils, monocytes and fibroblasts. It also stimulates pro-fibrotic cytokines and inhibits the expression of antifibrotic IFN-y [33]. A proteome-wide study in SSc found that plasmacytoid dendritic cells predominantly secrete CXCL4 [34]. CXCL4 levels were markedly elevated in SSc patients and highly correlated with skin fibrosis, ILD and PAH [34]. Elevated CXCL4 levels have also been shown to be associated with a more rapid decline in gas transfer and decreased levels were predictive of improved pulmonary function during immunosuppressive therapy [34]. CXCL4 overexpression in SSc has a direct mechanistic link to upregulated IFN-1 pathways by acting as both a chaperone and adjuvant in breaking immune tolerance to self-DNA [35]. The findings in our study contribute to and support this emerging literature.

Our results should be interpreted in the context of the patient population. Although representative of SSc populations seen in clinical practice, the relatively high proportion of long-established and limited disease may have influenced the results in ways for which we were unable to control. PAH, an important contributor to increased mortality, was defined accurately by strict criteria but not all patients were systematically investigated for its presence. This was a retrospective study and biospecimens were collected at variable times in relation to definitions of disease subgroups. No clinically correlated externally validated values for the biomarkers are available so the thresholds calculated in this study are specific to this cohort and the techniques described. Participant characteristics and subgroups were defined at baseline and not assessed during follow-up. This study did not look at cause of death which could be particularly important in those with strong biomarker signals. In addition to the variables already described, anti-Scl70 has also been associated with progression of SSc-ILD [34]. In *post hoc* analysis (data not shown but available on request), adding anti-Scl70 status to multivariable analysis had no significant effect on the serum biomarker associations.

The strengths of our study include the unique combination of a large, well-phenotyped cohort of patients with comprehensive datasets and long follow-up together with the largest multiple biomarker panel investigated in SSc research to date. Further work could legitimately focus on certain subgroups in more detail, such as those with early diffuse disease or in those with already more extensive ILD at baseline. It would be useful to see how biomarkers can be best combined together and with clinical variables to improve prognostication. External validation of our findings is required. The strong associations identified in this study between identified biomarkers and important prognostic outcomes may support ongoing research into disease pathogenesis and management.

Conclusion

We have demonstrated that multiple biomarkers, especially VCAM-1, E-selectin, SP-D and CXCL4, provide prognostic utility beyond that of established risk factors for patients with SSc. These findings provide a framework for future investigation.

Supplementary material

Supplementary material is available at Rheumatology online.

Data availability

The data underlying this article cannot be shared publicly to preserve the privacy of individuals that participated in the study. The data will be shared on reasonable request to the corresponding author.

Funding

M.J.S.P. is supported by the Brian Eaton Memorial PhD Scholarship administered by Lung Foundation Australia and The Thoracic Society of Australia & New Zealand. T.J.K. is supported by NHMRC Project Grant GNT1162767. The Australian Scleroderma Interest Group and the Australian Scleroderma Cohort Study are supported by Janssen (Actelion), Scleroderma Australia, Scleroderma Victoria, Arthritis Australia, Musculoskeletal Australia, the Australian Rheumatology Association, St Vincent's Hospital Melbourne Information Technology Department, the Scleroderma Clinical Trials Consortium, Boehringer-Ingelheim and Bayer. Lung Foundation Australia facilitated the Australian IPF Registry with educational grants from Foundation partners Roche Products, Pty Ltd and Boehringer Ingelheim. Institutional and in-kind support was provided by the Department of Allergy and Clinical Immunology and

Department of Respiratory at the Royal Prince Alfred Hospital, the University of Sydney and the NHMRC Centre of Research Excellence in Pulmonary Fibrosis, Australia, which is funded by the NHMRC, Lung Foundation Australia and anonymous philanthropy.

Disclosure statement: T.J.C. reports grants or contracts from Boehringer Ingelheim, Roche, Biogen and Three Lakes Foundation; consulting fees from, and participation on a Data Safety Monitoring Board or Advisory Board for, Boehringer Ingelheim, Roche and Bristol Myers Squibb; payment or honoraria from Boehringer Ingelheim; and is the cochair of Australian ILD Registry. All other authors declare no conflicts of interests.

References

- 1. Gabrielli A, Avvedimento EV, Krieg T. Scleroderma. N Engl J Med 2009;360:1989–2003.
- Elhai M, Meune C, Avouac J, Kahan A, Allanore Y. Trends in mortality in patients with systemic sclerosis over 40 years: a systematic review and meta-analysis of cohort studies. Rheumatology (Oxford) 2012;51:1017–26.
- 3. Steen VD, Medsger TA. Changes in causes of death in systemic sclerosis, 1972-2002. Ann Rheum Dis 2007;66:940–4.
- 4. Elhai M, Meune C, Boubaya M *et al.*; EUSTAR Group. Mapping and predicting mortality from systemic sclerosis. Ann Rheum Dis 2017;76:1897–905.
- Vonk MC, Walker UA, Volkmann ER *et al.* Natural variability in the disease course of SSc-ILD: implications for treatment. Eur Respir Rev 2021;30:200340.
- Khanna D, Tashkin DP, Denton CP *et al*. Etiology, risk factors, and biomarkers in systemic sclerosis with interstitial lung disease. Am J Respir Crit Care Med 2020;201:650–60.
- Wermuth PJ, Piera-Velazquez S, Rosenbloom J, Jimenez SA. Existing and novel biomarkers for precision medicine in systemic sclerosis. Nat Rev Rheumatol 2018;14:421–32.
- Distler O, Assassi S, Cottin V *et al*. Predictors of progression in systemic sclerosis patients with interstitial lung disease. Eur Respir J 2020;55:1902026.
- Jee AS, Sahhar J, Youssef P *et al.* Review: serum biomarkers in idiopathic pulmonary fibrosis and systemic sclerosis associated interstitial lung disease—frontiers and horizons. Pharmacol Ther 2019; 202:40–52.
- Jee AS, Stewart I, Youssef P et al.; Australian Scleroderma Cohort Study, Australian Scleroderma Interest Group, Australian Idiopathic Pulmonary Fibrosis Registry, and associated investigators. A Composite Serum Biomarker Index for the Diagnosis of Systemic Sclerosis-Associated Interstitial Lung Disease: a Multicenter, Observational Cohort Study. Arthritis Rheumatol 2023;75:1424–33.
- van den Hoogen F, Khanna D, Fransen J et al. 2013 classification criteria for systemic sclerosis: an American College of Rheumatology/European League against Rheumatism collaborative initiative. Arthritis Rheum 2013;65:2737–47.
- 12. Goh NS, Desai SR, Veeraraghavan S *et al.* Interstitial lung disease in systemic sclerosis: a simple staging system. Am J Respir Crit Care Med 2008;177:1248–54.
- 13. Quanjer PH, Stanojevic S, Cole TJ *et al.*; ERS Global Lung Function Initiative. Multi-ethnic reference values for spirometry for the 3-95-yr age range: the global lung function 2012 equations. Eur Respir J 2012;40:1324–43.
- Simonneau G, Montani D, Celermajer DS *et al*. Haemodynamic definitions and updated clinical classification of pulmonary hypertension. Eur Respir J 2019;53:1801913.
- 15. Hatano S, Strasser T; World Health Organization. Primary pulmonary hypertension: report on a WHO meeting. Geneva: World Health Organization, 1975.

- Kong DH, Kim YK, Kim MR, Jang JH, Lee S. Emerging Roles of Vascular Cell Adhesion Molecule-1 (VCAM-1) in immunological disorders and cancer. Int J Mol Sci 2018;19:1057.
- Denton CP, Bickerstaff MC, Shiwen X *et al.* Serial circulating adhesion molecule levels reflect disease severity in systemic sclerosis. Br J Rheumatol 1995;34:1048–54.
- Richards TJ, Kaminski N, Baribaud F *et al.* Peripheral blood proteins predict mortality in idiopathic pulmonary fibrosis. Am J Respir Crit Care Med 2012;185:67–76.
- Hasegawa M, Asano Y, Endo H *et al.* Serum adhesion molecule levels as prognostic markers in patients with early systemic sclerosis: a multicentre, prospective, observational study. PLoS One 2014;9:e88150.
- Gruschwitz MS, Hornstein OP, von Den Driesch P. Correlation of soluble adhesion molecules in the peripheral blood of scleroderma patients with their in situ expression and with disease activity. Arthritis Rheum 1995;38:184–9.
- Kuryliszyn-Moskal A, Klimiuk PA, Sierakowski S. Soluble adhesion molecules (sVCAM-1, sE-selectin), vascular endothelial growth factor (VEGF) and endothelin-1 in patients with systemic sclerosis: relationship to organ systemic involvement. Clin Rheumatol 2005;24:111–6.
- Guiot J, Moermans C, Henket M, Corhay JL, Louis R. Blood Biomarkers in Idiopathic Pulmonary Fibrosis. Lung 2017; 195:273–80.
- 23. Kuroki Y, Takahashi H, Chiba H, Akino T. Surfactant proteins A and D: disease markers. Biochim Biophys Acta 1998; 1408:334–45.
- Asano Y, Ihn H, Yamane K et al. Clinical significance of surfactant protein D as a serum marker for evaluating pulmonary fibrosis in patients with systemic sclerosis. Arthritis Rheum 2001;44:1363–9.
- 25. Yanaba K, Hasegawa M, Takehara K, Sato S. Comparative study of serum surfactant protein-D and KL-6 concentrations in patients with systemic sclerosis as markers for monitoring the activity of pulmonary fibrosis. J Rheumatol 2004;31:1112–20.

- Hant FN, Ludwicka-Bradley A, Wang HJ et al.; Scleroderma Lung Study Research Group. Surfactant protein D and KL-6 as serum biomarkers of interstitial lung disease in patients with scleroderma. J Rheumatol 2009;36:773–80.
- Elhaj M, Charles J, Pedroza C et al. Can serum surfactant protein D or CC-chemokine ligand 18 predict outcome of interstitial lung disease in patients with early systemic sclerosis? J Rheumatol 2013;40:1114–20.
- Elhai M, Hoffmann-Vold AM, Avouac J *et al*. Performance of candidate serum biomarkers for systemic sclerosis-associated interstitial lung disease. Arthritis Rheumatol 2019;71:972–82.
- Liakouli V, Cipriani P, Marrelli A *et al.* Angiogenic cytokines and growth factors in systemic sclerosis. Autoimmun Rev 2011; 10:590–4.
- Tourkina E, Bonner M, Oates J *et al.* Altered monocyte and fibrocyte phenotype and function in scleroderma interstitial lung disease: reversal by caveolin-1 scaffolding domain peptide. Fibrogenesis Tissue Repair 2011;4:15.
- Ikawa T, Miyagawa T, Fukui Y *et al.* Association of serum CXCL12 levels with arthropathy in patients with systemic sclerosis. Int J Rheum Dis 2021;24:260–7.
- Manetti M, Liakouli V, Fatini C et al. Association between a stromal cell-derived factor 1 (SDF-1/CXCL12) gene polymorphism and microvascular disease in systemic sclerosis. Ann Rheum Dis 2009;68:408–11.
- Utsunomiya A, Oyama N, Hasegawa M. Potential biomarkers in systemic sclerosis: a literature review and update. J Clin Med 2020;9:3388.
- van Bon L, Affandi AJ, Broen J *et al*. Proteome-wide analysis and CXCL4 as a biomarker in systemic sclerosis. N Engl J Med 2014; 370:433–43.
- Lande R, Lee EY, Palazzo R *et al.* CXCL4 assembles DNA into liquid crystalline complexes to amplify TLR9-mediated interferon-alpha production in systemic sclerosis. Nat Commun 2019; 10:1731.

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https://doi.org/10.1093/rheumatology/keae110 Original Article