Check for updates

OPEN ACCESS

EDITED BY Tejeshwar Rao, University of Houston, United States

REVIEWED BY Salina Gairhe, National Institutes of Health (NIH), United States Chintan K Gandhi, The Pennsylvania State University, United States

*CORRESPONDENCE István Vadász istvan.vadasz@innere.med.uni-giessen.de

RECEIVED 23 September 2024 ACCEPTED 11 October 2024 PUBLISHED 22 October 2024

CITATION

Dada LA and Vadász I (2024) Editorial: Endocytic and trafficking events in acute lung injury and pulmonary inflammation. *Front. Immunol.* 15:1500369. doi: 10.3389/fimmu.2024.1500369

COPYRIGHT

© 2024 Dada and Vadász. This is an openaccess article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Editorial: Endocytic and trafficking events in acute lung injury and pulmonary inflammation

Laura A. Dada¹ and István Vadász^{2,3,4}*

¹Divison of Pulmonary and Critical Care Medicine, Feinberg School of Medicine, Northwestern University, Chicago, IL, United States, ²Department of Internal Medicine, Justus Liebig University, Universities of Giessen and Marburg Lung Center (UGMLC), German Center for Lung Research (DZL), Giessen, Germany, ³The Cardio-Pulmonary Institute (CPI), Giessen, Germany, ⁴Institute for Lung Health (ILH), Giessen, Germany

KEYWORDS

endocytosis, intracellular trafficking, inflammation, acute respiratory distress syndrome, acute lung injury, influenza A virus, SARS-CoV-2, COVID-19

Editorial on the Research Topic

Endocytic and trafficking events in acute lung injury and pulmonary inflammation

Lung inflammation and injury remain a major focus of research since the beginning of the coronavirus disease 2019 (COVID-19) pandemic. Various endocytic and intracellular trafficking events play a pivotal role in the integrity and physiological functions of the airblood barrier but also in the invasion of pathogens, such as, e.g. respiratory viruses. Hence, disturbances in endocytosis and trafficking across and within alveolar epithelial and endothelial cells are important drivers of alveolar-capillary barrier dysfunction during respiratory inflammation and failure. Moreover, interfering with these endocytic/trafficking processes may serve as rescue measures in life-threatening sequels of lung damage in the context of acute respiratory distress syndrome (ARDS). Of note, several recent studies have focused on better understanding of lung cell-specific signaling patterns and endocytic events, utilizing novel technological advances in complex model systems. This Research Topic consists of original research manuscripts that enhance our knowledge on the mechanisms regulating the endocytic machinery during infection, inflammation and damage of the lung, as well as review articles that summarize most recent advances on the field.

The review article of Kryvenko and Vadász covers the six major endocytic pathways (1), namely clathrin-, caveolae-, endophilin- and glycosylphosphatidyl inositol-anchored protein-mediated endocytosis, as well as, macropinocytosis and phagocytosis and the subsequent trafficking events that are involved in alveolar-capillary barrier (dys)function during acute lung injury (ALI) and ARDS. In particular, the role of these internalization events in viral and bacterial infections of the respiratory tract and in the regulation of various transporters and junctional molecules during formation and clearance of protein-rich alveolar edema is described in detail. This manuscript also focuses on major technological advances, which have facilitated better understanding of intracellular

trafficking events after endocytosis in various models and phases of ALI and describes potential novel therapeutic means, targeting these specific molecular events for patients with ARDS.

Most frequently, ARDS is a consequence of severe viral and/or bacterial pneumonia (2). Recently, the mechanisms of endocytosis of influenza A virus (IAV) and severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) by epithelial, and in particular by alveolar cells, have been intensively studied. An excellent review article in this Research Topic contributed by Hook and Bhattacharya covers important determinants of IAV deposition into the alveolar space, as well as the relevance of alveolar structure in the internalization process of IAV virions. The article also describes recent advances regarding the role of the alveolar glycocalyx and of the epithelial lining fluid (ELF) in IAV attachment and how post-endocytic events lead to failure of the alveolarepithelial barrier. Clearly, further research will be necessary to assess the potential therapeutic role of interfering with cellular IAV attachment during initial infection and viral spread.

A key function of the alveolar-capillary barrier is providing adequate gas exchange that requires a relatively "dry" alveolar compartment where the epithelium is covered by a thin ELF layer (3). In contrast, during ARDS the alveolar space is flooded by protein-rich edema fluid, persistence of which is associated with worse outcomes (2). The research article of Alberro-Brage et al. describes a novel mechanism by which IAV in cultured epithelial cells and in three-dimensional precision cut lung slices impairs alveolar uptake and thus averts removal of excess protein content from the alveolar space in vivo. The authors establish the central involvement of matrix metalloprotease-14 as a negative regulator of megalin, a major receptor for albumin internalization in the alveolar epithelium (4, 5), which promotes shedding of the megalin ectodomain from the cell surface (6), thereby impairing its function. Therefore, inhibition of metalloprotease-14 may rescue alveolar protein clearance in the setting of ALI/ARDS.

In case of SARS-CoV-2, infection occurs when the spike protein of the virus that contains the receptor binding domain (RBD) binds to the host cell receptor angiotensin-converting enzyme 2 (ACE2) (7). Of note, patients with chronic lung diseases are more susceptible for cellular SARS-CoV-2 entry (8) in part as a consequence of elevated ACE2 expression levels (9). The original article of Chen et al. demonstrates that hypercapnia (an elevated level of CO₂ in blood and tissues), which is often observed in patients with chronic lung diseases, increases ACE2 protein expression and viral infection in cultured human bronchial epithelial cells and in murine airway epithelium. Of note, similar effects were observed when bronchial epithelial cells were exposed to extracts of cigarette smoke, the most common cause of chronic lung disease. Moreover, both hypercapnia and cigarette smoke extract increased total cellular and lipid raft-associated cholesterol, which was found to be relevant for viral entry and assembly. Further to this notion, inhibition of cholesterol synthesis and depletion of cellular cholesterol reduced ACE2 expression and SARS-CoV-2 infection, revealing a novel and pharmacologically targetable mechanism that may contribute to the poor clinical outcomes of smokers and patients with advanced lung disease with hypercapnia upon COVID-19.

As compared to other etiologies of viral pneumonia, COVID-19 presents with higher levels of vasculopathy, which may lead to an array of complications including thrombotic events and pulmonary edema due to loss of alveolar-capillary barrier function (10, 11). Romero et al. characterized the pathways that are involved in SARS-CoV-2-triggered vasculopathy and identified a potential novel therapeutic strategy to ameliorate endothelial dysfunction during and post COVID-19. The authors found that activity of the epithelial sodium channel (ENaC), which is also expressed in the endothelium (12), is markedly inhibited by SARS-CoV-2 RBD. RBD also increased oxidative stress and production of tissue factor, causing barrier dysfunction and coagulopathy. Importantly, these effects could be restored by a TNF-derived peptide, called TIP that directly binds ENaC. TIP also limited pneumococcal infection, which was stimulated by SARS-CoV-2. Thus, TIP might represent a potent therapeutic mean, as it appears to stabilize/activate ENaC in both the alveolar endothelium and epithelium (13, 14) and may limit bacterial infections secondary to COVID-19.

Air-blood barrier integrity is also critically dependent on pulmonary surfactant (PS) that covers the alveolar epithelium, thereby reducing surface tension, and also plays an important role in innate immunity of the lung (15). PS is synthetized in the lamellar bodies (LB) of alveolar type 2 (AT2) cells and consists of phospholipids, cholesterol and four specific proteins, whereas some of the lipids that are not synthesized in the LB are transported to it via the Golgi apparatus (16, 17). While it is clear that PS alterations are associated with impaired barrier function (18, 19), the mechanisms that are at play remain less understood. Novel bioimaging techniques, described in detail in the review article by Garcia et al., which combine microscopy and spectroscopy, enable precise temporo-spatial depiction of LB structure and function during maturation, PS secretion and recycling in two- and threedimensional cell culture systems. This is particularly remarkable, as better understanding of the implicated impairments e.g., during synthesis and transport to the alveolar surface may lead to therapeutic options in PS alterations associated with lung diseases.

During the inflammatory phase of ARDS and mechanical ventilation, an extracellular release of adenosine triphosphate (ATP) is observed that drives alveolar-capillary barrier dysfunction and thus, lung edema formation, in part through activation of purinergic, G-protein-coupled P2Y receptors (20, 21). In their manuscript, Kargarpour et al. convincingly demonstrate that in the setting of lipopolysaccharide (LPS)-induced ALI in mice, which triggers an acute recruitment of neutrophils to the lung; the purinergic P2Y2 receptors drive inflammation. Furthermore, neutralizing extracellular ATP, blocking P2Y2 and genetically deleting the receptor in neutrophils reduced neutrophil recruitment and rescued the inflammatory phenotype, which may open new avenues for therapy in the treatment of ARDS.

Endogenous glucocorticoids limit inflammation in ARDS, however systemic inflammation-associated glucocorticoid resistance may lessen these effects (22). Mahida et al. study the role of 11 β -hydroxysteroid dehydrogenase type-1 (HSD-1), which converts inactive cortisone to active cortisol, in both alveolar macrophages in broncho-alveolar lavage samples from patients with sepsis-associated ARDS, in mice in LPS-induced ALI, as well as in a murine polymicrobial sepsis model (cecal ligation and puncture). The work establishes that HSD-1 activity of alveolar macrophages is markedly reduced in these scenarios, leading to decreased macrophage efferocytosis (phagocytosis of apoptotic cells), which may contribute to worse outcomes and thus, identifies HSD-1 as a potential therapeutic target in patients with sepsis and sepsis-associated ARDS.

Collectively, the articles published in this Research Topic highlight the pivotal role of alveolar-capillary endocytic and trafficking events in the pathogenesis and potential therapy of pulmonary inflammation and injury. Further studies to explore the therapeutic potential of these pathways are warranted.

Author contributions

LD: Writing – original draft, Writing – review & editing. IV: Writing – original draft, Writing – review & editing.

Funding

The author(s) declare that financial support was received for the research, authorship, and/or publication of this article. This work was supported in part by grants from the National Institutes of

Health (R01HL147070, P01AG049665 and P01HL154998 to LD) and the German Federal Ministry of Education and Research (German Center for Lung Research (DZL/ALI 1.5, 3.3 and 3.4)), the von Behring Röntgen Foundation (Project 66-LV07), the German Research Foundation (DFG/KFO309, project ID: 284237345; P5; The Cardio-Pulmonary Institute (EXC 2026, project ID: 390649896) (to IV).

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

The author(s) declared that they were an editorial board member of Frontiers, at the time of submission. This had no impact on the peer review process and the final decision.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

1. Thottacherry JJ, Sathe M, Prabhakara C, Mayor S. Spoiled for choice: diverse endocytic pathways function at the cell surface. *Annu Rev Cell Dev Biol.* (2019) 35:55–84. doi: 10.1146/annurev-cellbio-100617-062710

2. Matthay MA, Zemans RL, Zimmerman GA, Arabi YM, Beitler JR, Mercat A, et al. Acute respiratory distress syndrome. *Nat Rev Dis Primers*. (2019) 5:18. doi: 10.1038/ s41572-019-0069-0

3. Herold S, Gabrielli NM, Vadasz I. Novel concepts of acute lung injury and alveolar-capillary barrier dysfunction. *Am J Physiol Lung Cell Mol Physiol.* (2013) 305: L665–81. doi: 10.1152/ajplung.00232.2013

4. Buchackert Y, Rummel S, Vohwinkel CU, Gabrielli NM, Grzesik BA, Mayer K, et al. Megalin mediates transepithelial albumin clearance from the alveolar space of intact rabbit lungs. *J Physiol*. (2012) 590:5167–81. doi: 10.1113/tjp.2012.590.issue-20

5. Vohwinkel CU, Buchackert Y, Al-Tamari HM, Mazzocchi LC, Eltzschig HK, Mayer K, et al. Restoration of megalin-mediated clearance of alveolar protein as a novel therapeutic approach for acute lung injury. *Am J Respir Cell Mol Biol.* (2017) 57:589–602. doi: 10.1165/rcmb.2016-0358OC

6. Mazzocchi LC, Vohwinkel CU, Mayer K, Herold S, Morty RE, Seeger W, et al. TGF-beta inhibits alveolar protein transport by promoting shedding, regulated intramembrane proteolysis, and transcriptional downregulation of megalin. *Am J Physiol Lung Cell Mol Physiol.* (2017) 313:L807–24. doi: 10.1152/ajplung.00569.2016

7. Hoffmann M, Kleine-Weber H, Schroeder S, Kruger N, Herrler T, Erichsen S, et al. SARS-coV-2 cell entry depends on ACE2 and TMPRSS2 and is blocked by a clinically proven protease inhibitor. *Cell.* (2020) 181:271–280 e8. doi: 10.1016/j.cell.2020.02.052

8. Bui LT, Winters NI, Chung MI, Joseph C, Gutierrez AJ, Habermann AC, et al. Chronic lung diseases are associated with gene expression programs favoring SARS-CoV-2 entry and severity. *Nat Commun.* (2021) 12:4314. doi: 10.1038/s41467-021-24467-0

9. Jacobs M, Van Eeckhoutte HP, Wijnant SRA, Janssens W, Joos GF, Brusselle GG, et al. Increased expression of ACE2, the SARS-CoV-2 entry receptor, in alveolar and bronchial epithelium of smokers and COPD subjects. *Eur Respir J.* (2020) 56:2002378. doi: 10.1183/13993003.02378-2020

10. Birnhuber A, Fliesser E, Gorkiewicz G, Zacharias M, Seeliger B, David S, et al. Between inflammation and thrombosis: endothelial cells in COVID-19. *Eur Respir J.* (2021) 58:2100377. doi: 10.1183/13993003.00377-2021

11. Flaumenhaft R, Enjyoji K, Schmaier AA. Vasculopathy in COVID-19. Blood. (2022) 140:222–35. doi: 10.1182/blood.2021012250

12. Czikora I, Alli AA, Sridhar S, Matthay MA, Pillich H, Hudel M, et al. Epithelial sodium channel-alpha mediates the protective effect of the TNF-derived TIP peptide in pneumolysin-induced endothelial barrier dysfunction. *Front Immunol.* (2017) 8:842. doi: 10.3389/fimmu.2017.00842

13. Czikora I, Alli A, Bao HF, Kaftan D, Sridhar S, Apell HJ, et al. A novel tumor necrosis factor-mediated mechanism of direct epithelial sodium channel activation. *Am J Respir Crit Care Med.* (2014) 190:522–32. doi: 10.1164/rccm.201405-0833OC

 Vadasz I, Schermuly RT, Ghofrani HA, Rummel S, Wehner S, Muhldorfer I, et al. The lectin-like domain of tumor necrosis factor-alpha improves alveolar fluid balance in injured isolated rabbit lungs. Crit Care Med. (2008) 36:1543–50. doi: 10.1097/CCM.0b013e31816f485e

15. Perez-Gil J. A recipe for a good clinical pulmonary surfactant. *BioMed J.* (2022) 45:615–28. doi: 10.1016/j.bj.2022.03.001

16. Agassandian M, Mallampalli RK. Surfactant phospholipid metabolism. *Biochim Biophys Acta*. (2013) 1831:612-25. doi: 10.1016/j.bbalip.2012.09.010

17. Olmeda B, Martinez-Calle M, Perez-Gil J. Pulmonary surfactant metabolism in the alveolar airspace: Biogenesis, extracellular conversions, recycling. *Ann Anat.* (2017) 209:78–92. doi: 10.1016/j.aanat.2016.09.008

18. Gunther A, Siebert C, Schmidt R, Ziegler S, Grimminger F, Yabut M, et al. Surfactant alterations in severe pneumonia, acute respiratory distress syndrome, and cardiogenic lung edema. *Am J Respir Crit Care Med.* (1996) 153:176–84. doi: 10.1164/ ajrccm.153.1.8542113

19. Wang S, Li Z, Wang X, Zhang S, Gao P, Shi Z. The role of pulmonary surfactants in the treatment of acute respiratory distress syndrome in COVID-19. *Front Pharmacol.* (2021) 12:698905. doi: 10.3389/fphar.2021.698905

20. Idzko M, Ferrari D, Eltzschig HK. Nucleotide signalling during inflammation. *Nature*. (2014) 509:310–7. doi: 10.1038/nature13085

21. Rich PB, Douillet CD, Mahler SA, Husain SA, Boucher RC. Adenosine triphosphate is released during injurious mechanical ventilation and contributes to lung edema. *J Trauma*. (2003) 55:290–7. doi: 10.1097/01.TA.0000078882.11919.AF

22. Meduri GU, Yates CR. Systemic inflammation-associated glucocorticoid resistance and outcome of ARDS. Ann N Y Acad Sci. (2004) 1024:24–53. doi: 10.1196/annals.1321.004