Check for updates

OPEN ACCESS

EDITED BY Yang Yang, Shanxi Agricultural University, China

REVIEWED BY Qinghua Yang, Northwest A & F University Hospital, China Hui Zhang, Shanxi Agricultural University, China

*CORRESPONDENCE Binjie Gan, ⊠ ganbinjie@163.com

RECEIVED 31 July 2024 ACCEPTED 10 October 2024 PUBLISHED 22 October 2024

CITATION

Ma S, Huang X, Zhao X, Liu L, Zhang L and Gan B (2024) Current status for utilization of cold resistance genes and strategies in wheat breeding program. *Front. Genet.* 15:1473717. doi: 10.3389/fgene.2024.1473717

COPYRIGHT

© 2024 Ma, Huang, Zhao, Liu, Zhang and Gan. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Current status for utilization of cold resistance genes and strategies in wheat breeding program

Shijie Ma, Xiaorong Huang, Xiaoqing Zhao, Lilong Liu, Li Zhang and Binjie Gan*

Crop Research Institute, Anhui Academy of Agricultural Sciences, Hefei, Anhui Province, China

Low temperature chilling is one of the major abiotic stresses affecting growth and yield of Triticum aestivum L. With global climate change, the risk of cold damage in wheat production has increased. In recent years, with the extensive research on wheat chilling resistance, especially the development of genetic engineering technology, the research on wheat chilling resistance has made great progress. This paper describes the mechanism of wheat cold damage, including cell membrane injury, cytoplasmic concentration increased as well as the imbalance of the ROS system. Mechanisms of cold resistance in wheat are summarised, including hormone signalling, transcription factor regulation, and the role of protective enzymes of the ROS system in cold resistanc. Functions of cloned wheat cold resistance genes are summarised, which will provide a reference for researchers to further understand and make use of cold resistance related genes in wheat. The current cold resistant breeding of wheat relies on the agronomic traits and observable indicators, molecular methods are lacked. A strategy for wheat cold-resistant breeding based on QTLs and gene technologies is proposed, with a view to breeding more coldresistant varieties of wheat with the deepening of the research.

KEYWORDS

Triticum aestivum L., cold resistance, gene, mechanism, breeding strategy

1 Introduction

Wheat (*Triticum aestivum L.*) is one of the world's top three grain crops. According to the FAO, around 200 million hectares of wheat will be planted globally with a production of about 787 million tonnes in 2023. Wheat is the calorie source for one-fifth of the global population (Pang et al., 2021), therefore, high and stable wheat production plays a crucial role in global food security. In recent decades, the likelihood of extreme weather on Earth has increased dramatically as greenhouse gases continue to be emitted (Jiang et al., 2022), low-temperature freezes occur from time to time, posing a significant threat to wheat production (Sutka, 2001; Vij and Tyagi, 2007). It occurs not only in the main temperate wheat-producing regions, but also in subtropical and Mediterranean climatic regions, covering the main wheat-producing countries such as China, the United States, Canada, Australia and some parts of South America. In the 25 years since 1990, the frequent occurrence of wheat freezes has caused severe losses over large areas in the main wheat-producing regions of central China, with the most severe reductions in yields of up to 30% (Yang and Yuan, 2014). In the central regions of the United States, statistics from 1955 to

2010 show that the yield of wheat is reduced by 210 kg per hectare due to low-temperature frosts (Holman et al., 2011). In the UK and eastern Australia, there has also been a marked increase in recorded frosts since the 1950s, causing some damage to wheat production (Whaley et al., 2004; Zheng et al., 2015). The incidence of frost damage results in millions of tonnes of food production being lost each year, with direct economic losses in excess of tens of billions of dollars. Wheat growth and development will be subjected to more cold stresses as the global climate continues to change, so improving varietal cold tolerance is very important in wheat breeding (Pang et al., 2021).

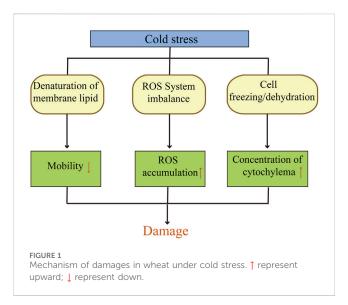
Wheat is a winter-habituated crop that must experience low temperatures to complete vernalisation (Fowler and Carles, 1979; Fowler et al., 1996), during inter-annual growth, they are exposed to the effects of multiple cold stress. The wheat is in the seedling stage before the winter, it will appear leaves dry, root damage or even dead seedlings phenomenon in case of a sudden drop in temperature and the yields will be also affected. After winter, if wheat is weakly resistant to cold the low-temperatures can damage plant tissues and inhibit growth and development (Subedi et al., 1998; Barton et al., 2014). During the period from nodulation to tasseling, wheat is in a period of parallel nutritive and reproductive growth, spikes can become hollow or underdeveloped as a result of frost or "spring cold" (Limin and Fowler, 2006), which will have a disastrous effect on wheat production. In summary, wheat is often exposed to low temperatures at critical stages of growth and development, and data shows that nearly 85% of the world's wheat area is affected by spring cold each year (Ferrante et al., 2021), it also shows the practical importance of research on cold resistance in wheat for production. The research on the function of plant cold resistance genes has been widely carried out with the deepening of research on the mechanism of plant cold resistance and plant genetic engineering. In this review, we will summarise the achievements in the mining and utilisation of cold-resistant genes in wheat and clarify the physiological and molecular mechanisms of cold resistance, and strategies for wheat cold resistance breeding will be provided.

2 Research progress of cold resistance mechanism in wheat

Low temperature affect the physiological and biochemical responses of plant cells seriously. Numerous studies have shown that the cold resistance of wheat is closely related to the mobility of cell membrane, reactive oxygen system and cellular osmoregulation. The mechanism of damages in wheat under cold stress is shown in Figure 1. The response to low-temperature involves multiple levels of regulation in wheat, including transcriptional regulation, hormonal regulation and signal transmission. These regulatory processes are quite complex and involve a large number of genes related to cold resistance.

2.1 Physiological mechanism of cold resistance in wheat

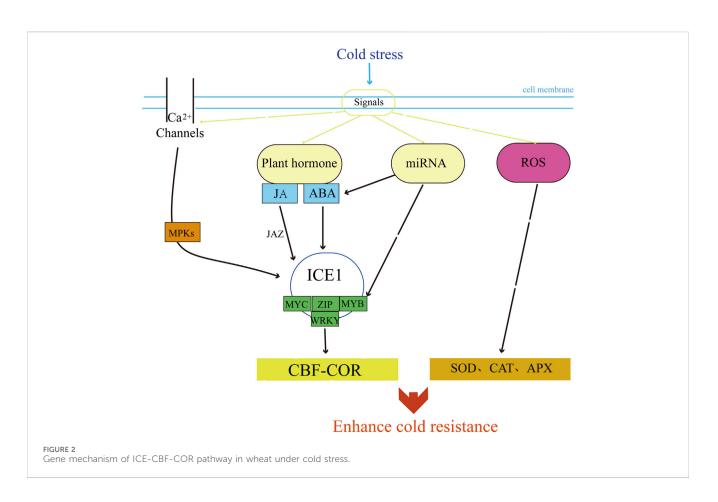
After suffering from cold damage, wheat cell tissues can be damaged or even die, mainly due to the destruction of cell



membrane structures (Lyons, 1973). The stability of cell membranes is determined by the composition of membrane lipid fatty acids. The stability of cell membrane is determined by the fatty acid composition of membrane lipids. Generally, a higher content of unsaturated fatty acids in membrane lipids correlates with an enhanced plant cold resistance. Therefore, fatty acid desaturase (FAD)plays a certain role in plant cold resistance (Ishizaki-Nishizawa et al., 1996; Kodama et al., 1995). The cell membrane also serves as a sensor and conduit for low-temperature signals, encompassing calcium signals, protein kinases, phosphatases, and transcription factors, all of which participate in the cold stress signaling pathway (Soleimani et al., 2022). Researchers indicated that the electrical conductivity method for determining the permeability of cell membranes of winter wheat leaves under indoor cold stress showed a significant positive correlation with the cold resistance of wheat in the field (Ju et al., 2012).

The reactive oxygen species (ROS) accumulation and scavenging system in plants is crucial for maintaining the normal cellular functions. Generally, the accumulation and scavenging of ROS in this system is in dynamic balance. Persistent low temperatures can disrupt this balance, accelerating ROS accumulation and producing a large amount of free radicals, which in turn cause physiological disorders within cells (Zeng et al., 2011; Bhattacharjee, 2005). SOD, per-Oxidase (POD), CAT, and ascorbate per-Oxidase (APX) are important protective enzymes in wheat (Türkan et al., 2005). They can scavenge excess ROS and free radicals generated during cold stress, maintaining the normal growth of wheat (Demiral and Türkan, 2004).

When encountering low temperature cold damage, the loss of water molecules in plant cells increases the concentration of cell fluid, causing damage to the cells themselves. In order to avoid this kind of damage, the body will actively accumulate some soluble substances to increase the cell's ability to absorb water and maintain normal metabolic functions (Lalk and Dorffling, 1985). The soluble sugar content of wheat is positively correlated with cold resistance, and sucrose and fructose play an important role in the cold resistance of wheat (Kamata and Uemura, 2004; Zeng et al., 2011). The content of proline (Pro) is positively correlated with



the cold resistance of wheat. Many results show that when the temperature is low, the proline content in the wheat body rises, more water is aggregated on the protein, preventing the protein from deforming due to dehydration at low temperatures, thereby protecting wheat cells from damage (Bao et al., 2021; Chen et al., 2007).

2.2 Molecular mechanism of cold resistance in wheat

Plant cells can respond to low temperature stress by precisely regulating the expression of transcription factors and effector genes. These signaling pathways consist of transcription, translation, and post-transcriptional and post-translational regulatory factors that induce the expression of functional genes in response to cold temperatures. Within one of the signaling pathways is a ciselement known as the C-repeat element/dehydration-responsive element (CRT/DRE), to which C-repeat binding factors (CBF) can bind. This pathway is referred to as the CBF-COR signaling pathway. Studies on a variety of plants have shown that this pathway makes plants have cold resistance by regulating the expression of downstream cold-resistant proteins (Ding et al., 2019). CBF homologous genes in wheat and barley have also been identified (Mohseni et al., 2012; Skinner et al., 2005). ICE1 (CBF expression inducer) is a MYC-type bHLH transcription factor and is the most characteristic transcription activator of CBF genes so far (Chinnusamy et al., 2003; Lee et al., 2005). Overexpression of ICE1 can increase the expression of CBFs (Fursova et al., 2009; Agarwal et al., 2006; Juan et al., 2015). The expression of *ICE1* gene *TaICE41* in wheat is induced by cold stress (Badawi et al., 2008; Guo et al., 2019). The ICE-CBF-COR pathway plays an important regulatory role in the cold resistance pathway of wheat (Gong et al., 2020; Caccialupi et al., 2024) and is considered to be the main cold signal transduction pathway in wheat (Figure 2).

Plant hormones are crucial regulators in perceiving and transmitting various environmental signals and defense responses. Abscisic acid (ABA) and jasmonic acid (JA) are the most studied plant hormones in relation to wheat cold resistance. Lower temperatures can increase endogenous ABA levels in plants, participating in cold stress signal transduction and activating the expression of cold-resistance genes (Zhang et al., 2020; An et al., 2021; Liu et al., 2013). For instance, the cor gene family members such as cor6.6, cor15a, cor47, and cor78 are highly expressed due to increased ABA levels, thereby enhancing plant freeze tolerance (Hajela et al., 1990). Studies on wheat have found that endogenous ABA content increases during cold acclimation in winter wheat, with a rapid rise in ABA levels inducing the expression of stress-responsive genes and improving freeze resistance (Liu et al., 2013). Another plant hormone, JA, has also been recently discovered to play a regulatory role in plant cold stress responses. Cold stress activates the expression of JA biosynthesisrelated genes (AOC, AOS1, AOS2, and LOX2), increasing JA content (Hu et al., 2017; Du et al., 2013). Accumulated JA conjugates with isoleucine to form the active JA-Ile, which is perceived by the COI1-JAZ co-receptor and promotes the ubiquitination and degradation of the repressor protein JAZ via the 26S proteasome, thereby activating the ICE-CBF pathway and inducing the expression of certain cold-responsive genes, enhancing plant cold tolerance (Hu et al., 2013).

Excessive ROS production in plant cells under stress conditions is highly reactive and toxic to proteins, lipids, and nucleic acids. ROS in plants primarily exists in the form of free radicals and non-radicals, including hydrogen peroxide (H₂O₂), superoxide anion (O²⁻⁻), hydroxyl radical (OH⁻⁻), and singlet oxygen (1O₂), etc. Elevated ROS levels lead to various harmful cytological effects (Nadarajah, 2020), ultimately resulting in cell damage and death (Gill and Tuteja, 2010). On the other hand, the increase in ROS during stress periods is also considered a signal for the activation of stress response pathways (Baxter et al., 2014). ROS can also function as a signaling molecule, the overaccumulation of ROS at the initial stage of cold stress promotes the activation of antioxidant enzyme genes such as SOD, CAT, and APX to eliminate ROS toxicity. Notably, studies in wheat have demonstrated that the overexpression of the wheat TaCAT gene, which eliminates excessive H2O2, improves the freeze tolerance of transgenic plants (Matsumura et al., 2010).

While calcium (Ca²⁺) is known to enhance stress tolerance (Malko et al., 2023). In plants, low temperatures induce the accumulation of intracellular Ca2+ (Yang et al., 2010), and the calcium/calcineurin (Ca2+/CaM) regulated receptor kinase CLRK1/2 acts as a sensor for Ca²⁺/CaM under low temperatures. It interacts with and phosphorylates MEKK1, triggering the activation of downstream MPK4/6 and thereby positively regulating the cold response (Furuya et al., 2013). Cold stress activates MKK2, which in turn activates MPK4 and MPK6 (Ichimura et al., 2000). Additionally, cold stress activates protein kinases MPK3 and MPK6, phosphorylates ICE1 protein, and decreases its protein stability and transcriptional activity, thus negatively regulating the expression of CBF and the cold tolerance of plants (Li et al., 2017). Studies in wheat have shown that Ca²⁺ induces cold-responsive genes (WCOR413, WCOR410, WCOR14, and Wrab17) enhanced cold stress tolerance through maintained cellular redox homeostasis (Malko et al., 2023). Transformation with the Ta MPK6 gene significantly enhances the expression of the ICE-CBF-COR pathway genes in Arabidopsis, improving the plant's cold tolerance (Yu et al., 2023).

MicroRNA (miRNA) is a class of non-coding small RNA molecules derived from the genomes of eukaryotic organisms, mostly ranging from 21 to 25 nt in length. Their main mode of action is post-transcriptional regulation of target gene expression, which also plays an important role in plant stress response (Naya et al., 2014; Tiwari et al., 2020). Using deep sequencing and bioinformatics prediction methods, miRNAs responding to cold stress have been identified in various crops and model species (Lv et al., 2010; Cao et al., 2014; Sun et al., 2015). In wheat, overexpression of miR399 can positively regulate the protein levels of ICE1 and the expression of key genes *CORs* in the CBF signaling pathway of plants (Peng et al., 2021).

3 Application of cold resistant genes and germplasm innovation in wheat breeding

At present, many studies have explained the physiological and molecular mechanisms of cold resistance in wheat from different

perspectives, and the research on the genes related to cold resistance in wheat has also progressed rapidly. Cold resistant genes are a class of induced genes that can be activated and expressed in large quantities to produce corresponding cold-resistant substances only at specific periods and conditions. Cold resistance is a complex trait involving many physiological mechanisms, which is the result of the combined action of multiple genes. With the rapid development of molecular biology technology and its wide application in the field of plant breeding, several quantitative trait loci (QTL) related to cold resistance have been mapped by linkage analysis.

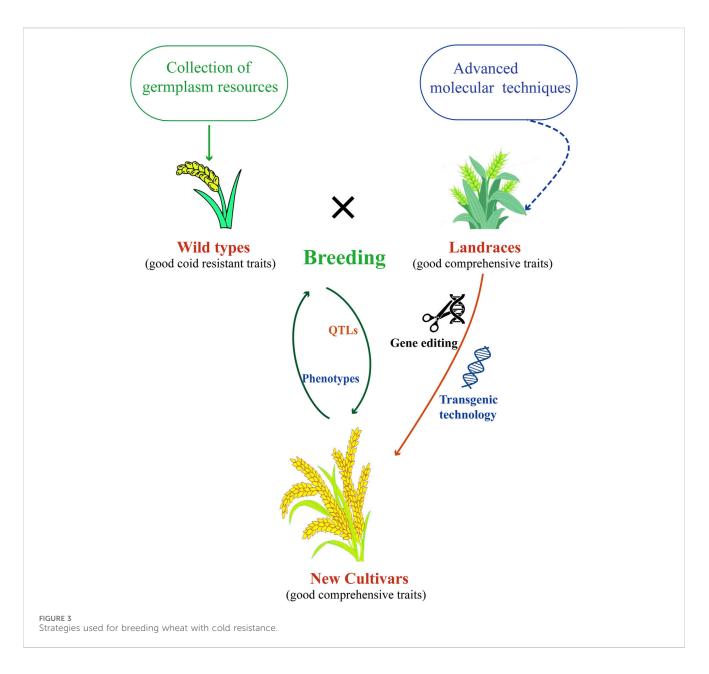
Many studies have confirmed that due to the huge genetic background of wheat, wheat cold resistance is controlled and played a role by multiple minor genes. Researchers have mapped allelic loci related to wheat cold resistance on more than 10 pairs of chromosomes such as 1B, 1D, 2B, 2D, 4D, 5A, 5D, and 7A of wheat. The DH population of wheat was used to map QTL for the semi lethal temperature, an index related to cold resistance traits of wheat, and found a total of five major QTL loci controlling the semi lethal temperature, which were distributed on chromosomes 2A, 5A, 1D and 6D of wheat (Båga et al., 2007). Some studies have also found that there are some alleles on chromosomes 5A and 5D that regulate the cold resistance of wheat. At present, QTL mapping loci cover almost all wheat chromosomes, among which 5A and 5D chromosomes are most closely related to cold resistance in wheat (Sutka, 1994; Limin et al.,. 1997), As well as a major-effect QTL related to frost tolerance was reported located on chromosomes 4A recently (Bolouri et al., 2023). The vrnl FRL segment of 5A chromosome in wheat regulates cold resistance in wheat. In common wheat, the major loci controlling cold resistance (FR-1 and FR-2) have been mapped to the long arm of chromosome 5 (Galiba et al., 1995; Tóth et al., 2003). FR-2 coincides with the CBFs gene cluster in wheat and barley (Miller et al., 2006; Francia et al., 2007) and directly induces downstream cor/lea gene expression during cold adaptation (Takumi et al., 2008). A large number of deletions in the CBF cluster of *fr-b2* significantly reduced the cold tolerance of tetraploid and hexaploid wheat (Pearce et al., 2013). A recent study reported that two genes named Wcr-3 and Wcr-4 control cold resistance in wheat, loci on 2B and 2D chromosomes (Lei et al., 2023).

In order to further explore the molecular mechanism of winter wheat response to low temperature stress, several cold resistance genes in wheat were transferred into model plants and proved to affect its cold tolerance. Genes with cold resistance function were cloned from wheat as shown in Table 1. Studies have shown that overexpression of TabZIP1 reduces the MDA content and electrical conductivity of Arabidopsis at 4°C, and improves the cold tolerance of Arabidopsis (Lv et al., 2018); Overexpression of TaMYB56-B transgenic Arabidopsis has a higher survival rate at 4°C, and the expression of CBF3 and COR15a was strongly induced (Zhang et al., 2012). A recent study showed that overexpression of TaMYB4 enhanced the freezing tolerance of transgenic Arabidopsis, AtCBF1, AtCBF2, AtCBF3, AtCOR15A, AtCOR47, AtKIN1 and AtRD29A in transgenc lines was significantly upregulated (Tian et al., 2023). TabZIP6 was found as a negative regulator of cold resistance in wheat. The cold tolerance of Arabidopsis thaliana

TABLE I Summary of cold resistance genes in wheat.			
Resistance mechanism	Protein types	Cloned genes	References
Increase cell membrane fluidity	FAD (fatty acid desaturase)	TaFAD2.8,TaFAB2.15	Hajiahmadi et al. (2020)
Eliminate effects of ROS	CAT catalase	TaCAT	Matsumura et al. (2010), Ghorbel et al. (2023)
	Glucose-6-phosphate dehydrogenase	TaG6PDH	Liu et al. (2019)
	6-phosphogluconate dehydrogenase	Ta6PGDH	Liu et al. (2019)
Increase the content of soluble sugar	FBA (Fructose-1, 6-bisphosphate aldolase)	Ta FBA-A10	Zeng et al. (2011)
	TPS (trehalose 6-phosphate synthase)	TaTPS11	Peng et al. (2022), Lu et al. (2024)
Transcription factor	МҮВ	ТаМҮВ56-В, ТаМҮВ4	Zhang et al. (2012), Tian et al. (2023)
	МҮС	TaMYC2	Wang et al. (2022)
	bZIP	TabZIP1, TabZIP6	Lv et al. (2018), Cai et al. (2018)
	NAC	TaNAC2a,TANAC4a,TaNAC6,TaNAC7,TaNAC13,TaNTL5	Lv et al. (2018)
Cold regulated gene	LEA (late embriogenesis abundant protein)	Wap27,Pvlea218,Wrab17	Takumi et al. (2008)
	Cor protein	Wcr3,Wcr4,Wcr719,Wcor80,Wcor14,Wcor72	Soltész et al. (2013), Lei et al. (2023)
	Wheat cold acclimation protien	WCS120, WCS19	Limin et al. (1997)
	ICE	TaICE41	Badawi et al. (2008)
Hormone regulation	SAMT (salicylic acid methyltransferase)	TaSAMT1	Chu et al. (2024)
Others	MAPK (Mitogen-activated protein kinase)	ТаМРКЗ, ТаМРК6	Yu et al. (2023), Wang et al. (2024)

TABLE 1 Summary of cold resistance genes in wheat.

overexpressing TabZIP6 at 4 °C was significantly decreased, and the expression of multiple genes in the CBF-COR cold signaling pathway was downregulated (Cai et al., 2018). Wheat TaICE41 and TaICE87 are homologous to Arabidopsis AtICE1 genes (Badawi et al., 2008). TaCBF14 and TaCBF15 play a role in cold tolerance of spring barley, and transgenic TaCBF14 and TaCBF15 genes improve the cold tolerance of spring barley (Soltész et al., 2013). Researchers found that TaMYC2 interacted with TaICE41, activated the downstream CBF-COR signaling pathway, and positively regulated the freezing resistance of Arabidopsis thaliana (Wang et al., 2022). TaMPK3 is also involved in regulating the ICE-CBF-COR cold resistance module through its interaction with TaICE41, improving freezing tolerance in wheat (Wang et al., 2024). TaFAD2.8 genes high expression in response to temperature stress (Hajiahmadi et al., 2020). Transgenic TaMPK6 gene significantly reduced the production and accumulation of ROS in Arabidopsis plants. The expression levels of genes encoding superoxide dismutase (SOD) and catalase (CAT) and the enzyme activities of SOD and CAT were significantly increased. The CAT genes were upregulated by cold stress (Ghorbel et al., 2023). Overexpression of TaCAT increased the CAT activity of transgenic rice, eliminated excessive accumulation of H₂O₂, and improved the cold resistance of transgenic rice (Matsumura et al., 2010). The soluble sugar content of winter wheat seedlings was significantly accumulated (Yu et al., 2008), and the expression of genes encoding key enzymes in glucose metabolism was significantly upregulated. The ectopic expression of wheat TaFBA-A10 gene was showed in Arabidopsis thaliana could increase the content of soluble sugar by affecting the rate of glycolysis and Calvin cycle, thereby improving the cold resistance of Arabidopsis thaliana (Zeng et al., 2011); ectopic expression of TaTPS11, a gene encoding wheat trehalose-6-phosphate synthase, in Arabidopsis can increase the frost resistance of Arabidopsis by catalyzing the expression level of SnRK1 in sucrose (Peng et al., 2022), and downregulates the expression of this gene with significantly lower cold resistance than Arabidopsis wild-types (Lu et al., 2024). Overexpressing the key enzyme genes TaG6PDH and Ta6PGDH in the PPP pathway of wheat could improve its cold resistance by scavenging ROS generated at low temperature (Liu et al., 2019; Tian et al., 2021). The mechanism of action of some cold-resistant genes in wheat depends on hormone signal transduction. Lv found that exogenous MeJA treatment can significantly increase the expression of transcription factors (TabZIP1, TaWABI5, TaMYB80, TaNAC2, TaWRKY80) in winter wheat under low temperature stress, thereby regulating the CBF signaling pathway to improve cold resistance (Lv et al., 2017; Lv et al., 2018). Chu identified a SA methyltransferase, TaSAMT1 that converts SA to methyl SA (MeSA) and confers cold tolerance



in wheat (Chu et al., 2024). For another example, under low temperature, exogenous ABA increased the expression of bZIP1, NAC2 and several key enzyme genes of sugar metabolism in wheat (Lv et al., 2018; Liu et al., 2013).

4 Breeding strategies for cold resistant wheat

Breeding cold-resistant wheat varieties to protect wheat from low-temperature damage and maintain high and stable yields is of significant importance in production. However, breeding for cold resistance in wheat is still at the level of phenotypic selection, the cloned genes were also only functionally characterized in model plants. To make more rational and effective use of cold resistance genes, the following cold resistance breeding strategies for wheat are proposed (Figure 3).

4.1 Screening for suitable wheat resources based on cold tolerance traits

Agronomic traits are key factors in wheat cultivars breeding (Tshikunde et al., 2019). Try harder to collect, identify, and study the cold resistance of various germplasm resources (commercial varieties, breeding lines, local varieties, wild relatives, and distant species) is very necessary, and clarify their genetic characteristics and traits of resistance. Only by collecting enough germplasm resources can we explore new diversity of cold resistance genes for cloning, functional verification and aggregation.

There are a number of methods for evaluating and characterising cold resistance criteria in wheat (Valluru et al., 2012; Ou and Wang, 2019). To develop an accurate, fast and scalable method for identifying cold resistance indexes of wheat, so as to judge the cold resistance of breeding materials and germplasm resources through relevant indexes in the field.

4.2 QTLs associated with cold resistance

QTLs have great potential for accelerating traditional breeding processes (Jha et al., 2017). Fine mapping and cloning of cold resistance genes/QTLs, studying the mechanism of cold resistance in wheat, and analyzing the regulatory network. Effectively use the existing cold-resistant genes with identification functions, use polymerization hybridization and backcross breeding, combined with molecular marker-assisted selection and generation-adding breeding techniques, accumulate different cold-resistant genes to high-yield varieties with excellent comprehensive traits, and create new materials for high-yield and multi-resistance breeding. On this basis, different hybrid combinations were combined by conventional breeding methods to cultivate new varieties with high yield and durable multi-resistance. Construct various genetic populations and natural populations, and use current wheat genome sequencing and molecular marker chips to carry out the mapping of cold resistance gene QTLs. Develop simple, accurate, and reliable molecular markers and detection technology systems that can be applied in practical breeding.

4.3 Functional gene research and new technology applications

In-depth study of key functional genes is a method to improve the yield and quality of crops such as wheat. Genome editing methods, such as CRISPR/Cas9, allowed to manipulate wheat genome to improve agronomic traits (Kim et al., 2018). Transgenic and genome editing was carried out to directly create new varieties of high-yield and high-quality cold-resistant wheat on the basis of the original excellent large varieties. Some technologies in non-biological fields also canbe applied to cold resistance in wheat, such as nanotechnology (Venzhik et al., 2024).

References

Agarwal, M., Hao, Y., Kapoor, A., Dong, C. H., Fujii, H., Zheng, X., et al. (2006). A R2R3 type MYB transcription factor is involved in the cold regulation of *CBF* genes and in acquired freezing tolerance. *J. Biol. Chem.* 281, 37636–37645. doi:10.1074/jbc.M605895200

An, J. P., Wang, X. F., Zhang, X. W., You, C. X., and Hao, Y. J. (2021). Apple B-box protein BBX37 regulates jasmonic acid mediated cold tolerance through the JAZ-BBX37-ICE1-CBF pathway and undergoes MIEL1-mediated ubiquitination and degradation. *New Phytol.* 229 (5), 2707–2729. doi:10.1111/nph.17050

Badawi, M., Reddy, Y. V., Agharbaoui, Z., Tominaga, Y., Danyluk, J., Sarhan, F., et al. (2008). Structure and functional analysis of wheat *ICE* (inducer of *CBF* expression) genes. *Plant Cell Physiol.* 49 (8), 1237–1249. doi:10.1093/pcp/pcn100

Båga, M., Chodaparambil, S. V., Limin, A. E., Pecar, M., Fowler, D. B., and Chibbar, R. N. (2007). Identification of quantitative trait loci and associated candidate genes for low-temperature tolerance in cold-hardy winter wheat. *Funct. and Integr. Genomics* 7 (1), 53–68. doi:10.1007/s10142-006-0030-7

Bao, Y., Yang, N., Meng, J., Wang, D., Fu, L., Wang, J., et al. (2021). Adaptability of winter wheat Dongnongdongmai 1 (*Triticum aestivum L.*) to overwintering in alpine regions. *Plant Biol.* 23 (3), 445–455. doi:10.1111/plb.13200

Barton, D. A., Cantrill, L. C., Law, A. M., Phillips, C. G., Sutton, B. G., and Overall, R. L. (2014). Chilling to zero degrees disrupts pollen formation but not meiotic microtubule arrays in *Triticum aestivumL. Plant Cell and Environ.* 37 (12), 2781–2794. doi:10.1111/pce.12358

Baxter, A., Mittler, R., and Suzuki, N. (2014). ROS as key players in plant stress signalling. J. Exp. Bot. 65 (5), 1229–1240. doi:10.1093/jxb/ert375

Bhattacharjee, S. (2005). Reactive oxygen species and oxidative burst: roles in stress, senescence and signal transduction in plants. *Currnt Sci.* 89 (7). doi:10.1073/pnas. 0506897102

Author contributions

SM: Conceptualization, Investigation, Software, Writing-original draft. XH: Writing-review and editing. XZ: Methodology, Writing-review and editing. LL: Software, Writing-review and editing. LZ: Methodology, Software, Writing-review and editing. BG: Conceptualization, Investigation, Writing-review and editing, Writing-original draft.

Funding

The author(s) declare that financial support was received for the research, authorship, and/or publication of this article. The work was supported by Open Project of Key Laboratory of Crop Quality improvement of Anhui Province (2024ZW005) and Joint research project on Wheat Breeding of Anhui Province (2021–2025).

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Bolouri, P., Haliloğlu, K., Mohammadi, S. A., Türkoğlu, A., İlhan, E., Niedbała, G., et al. (2023). Identification of novel QTLs associated with frost tolerance in winter wheat (*Triticum aestivum L.*). *Plants (Basel)* 12 (8), 1641. doi:10.3390/plants12081641

Caccialupi, G., Milc, J., Caradonia, F., Nasar, M. F., and Francia, E. (2023). The *triticeae CBF* gene cluster-to frost resistance and beyond. *Cells* 12 (22), 2606. doi:10. 3390/cells12222606

Cai, W., Yang, Y., Wang, W., Guo, G., Liu, W., and Bi, C. (2018). Overexpression of a wheat (*Triticum aestivum L.*) bZIP transcription factor gene, *TabZIP6*, decreased the freezing tolerance of transgenic Arabidopsis seedlings by down-regulating the expression of *CBFs. Plant physiology Biochem.* 124, 100–111. doi:10.1016/j.plaphy.2018.01.008

Cao, X., Wu, Z., Jiang, F., Zhou, R., and Yang, Z. (2014). Identification of chilling stress-responsive tomato microRNAs and their target genes by high-throughput sequencing and degradome analysis. *BMC Genomics* 15 (1), 1130. doi:10.1186/1471-2164-15-1130

Chen, X., Li, J., Ma, L., and Zhang, F. (2007). Effect of low temperature stress on change of free proline content in the leaves of spring and winter wheat. *Xinjiang Agric. Sci.* 4 (5), 553–556. doi:10.3969/j.issn.1001-4330.2007.05.002

Chinnusamy, V., Ohta, M., Kanrar, S., Lee, B. H., Hong, X., Agarwal, M., et al. (2003). ICE1: a regulator of cold-induced transcriptome and freezing tolerance in Arabidopsis. *Genes and Dev.* 17 (8), 1043–1054. doi:10.1101/gad.1077503

Chu, W., Chang, S., Lin, J., Zhang, C., Li, J., Liu, X., et al. (2024). Methyltransferase TaSAMT1 mediates wheat freezing tolerance by integrating brassinosteroid and salicylic acid signaling. *Plant Cell* 36 (7), 2607–2628. doi:10.1093/plcell/koae100

Demiral, T., and Türkan, I. (2004). Does exogenous glycinebetaine affect antioxidative system of rice seedlings under NaCl treatment? *J. Plant Physiology* 161 (10), 1089–1100. doi:10.1016/j.jplph.2004.03.009

Ding, Y., Shi, Y., and Yang, S. (2019). Advances and challenges in uncovering cold tolerance regulatory mechanisms in plants. *New Phytol.* 222 (4), 1690–1704. doi:10. 1111/nph.15696

Du, H., Liu, H., and Xiong, L. (2013). Endogenous auxin and jasmonic acid levels are differentially modulated by abiotic stresses in rice. *Front. Plant Sci.* 4 (397), 397. doi:10. 3389/fpls.2013.00397

Ferrante, A., Cullis, B. R., Smith, A. B., and Able, J. A. (2021). A multi-environment trial analysis of frost susceptibility in wheat and barley under Australian frost-prone field conditions. *Front. Plant Sci.* 12, 722637. doi:10.3389/fpls.2021.722637

Fowler, D. B., and Carles, R. J. (1979). Growth, development, and cold tolerence of fall-acclimated cereal grains. *Crop Sci.* 19 (6), 915–922. doi:10.2135/cropsci1979. 0011183X001900060040x

Fowler, D. B., Chauvin, L. P., Limin, A. E., and Sarhan, F. (1996). The regulatory role of vernalization in the expression of low-temperature-induced genes in wheat and rye. *Theor. and Appl. Genet.* 93 (4), 554–559. doi:10.1007/BF00417947

Francia, E., Barabaschi, D., Tondelli, A., Laidò, G., Rizza, F., Stanca, A. M., et al. (2007). Fine mapping of a HvCBF gene cluster at the frost resistance locus Fr-H2 in barley. *Theor. and Appl. Genet.* 115 (8), 1083–1091. doi:10.1007/s00122-007-0634-x

Fursova, O. V., Pogorelko, G. V., and Tarasov, V. A. (2009). Identification of *ICE2*, a gene involved in cold acclimation which determines freezing tolerance in *Arabidopsis thaliana*. *Gene* 429 (1-2), 98–103. doi:10.1016/j.gene.2008.10.016

Furuya, T., Matsuoka, D., and Nanmori, T. (2013). Phosphorylation of Arabidopsis thaliana MEKK1 via $Ca^{(2+)}$ signaling as a part of the cold stress response. J. Plant Res. 126 (6), 833–840. doi:10.1007/s10265-013-0576-0

Galiba, G., Quarrie, S. A., Sutka, J., Morgounov, A., and Snape, J. W. (1995). RFLP mapping of the vernalization (*Vrn1*) and frost resistance (*Fr1*) genes on chromosome 5A of wheat. *Theor. and Appl. Genet.* 90 (7-8), 1174–1179. doi:10.1007/BF00222940

Ghorbel, M., Zribi, I., Haddaji, N., Siddiqui, A. J., Bouali, N., and Brini, F. (2023). Genome-wide identification and expression analysis of catalase gene families in *triticeae*. *Plants (Basel)* 13 (1), 11. doi:10.3390/plants13010011

Gill, S. S., and Tuteja, N. (2010). Reactive oxygen species and antioxidant machinery in abiotic stress tolerance in crop plants. *Plant Physiology Biochem.* 48 (12), 909–930. doi:10.1016/j.plaphy.2010.08.016

Gong, Z., Xiong, L., Shi, H., Yang, S., Herrera-Estrella, L. R., Xu, G., et al. (2020). Plant abiotic stress response and nutrient use efficiency. *Sci. China. Life Sci.* 63 (5), 635–674. doi:10.1007/s11427-020-1683-x

Guo, J., Ren, Y., Tang, Z., Shi, W., and Zhou, M. (2019). Characterization and expression profiling of the *ICE-CBF-COR* genes in wheat. *PeerJ* 7 (1–2), e8190. doi:10. 7717/peerj.8190

Hajela, R. K., Horvath, D. P., Gilmour, S. J., and Thomashow, M. F. (1990). Molecular cloning and expression of cor (Cold-Regulated) genes in *Arabidopsis thaliana*. *Plant Physiol*. 93 (3), 1246–1252. doi:10.1104/pp.93.3.1246

Hajiahmadi, Z., Abedi, A., Wei, H., Sun, W., Ruan, H., Zhuge, Q., et al. (2020). Identification, evolution, expression, and docking studies of fatty acid desaturase genes in wheat (*Triticum aestivum L.*). *BMC Genomics* 21 (1), 778. doi:10.1186/s12864-020-07199-1

Holman, J. D., Schlegel, A. J., Thompson, C. R., and Lingenfelser, J. E. (2011). Influence of precipitation, temperature, and 56 years on winter wheat yields in western Kansas. *Crop Manag.* 10, 1–10. doi:10.1094/CM-2011-1229-01-RS

Hu, Y., Jiang, L., Wang, F., and Yu, D. (2013). Jasmonate regulates the inducer of cbf expression-C-repeat binding factor/DRE binding factor1 cascade and freezing tolerance in Arabidopsis. *Plant Cell* 25 (8), 2907–2924. doi:10.1105/tpc.113.112631

Hu, Y., Jiang, Y., Han, X., Wang, H., Pan, J., and Yu, D. (2017). Jasmonate regulates leaf senescence and tolerance to cold stress: crosstalk with other phytohormones. *J. Exp. Bot.* 68 (6), 1361–1369. doi:10.1093/jxb/erx004

Ichimura, K., Mizoguchi, T., Yoshida, R., Yuasa, T., and Shinozaki, K. (2000). Various abiotic stresses rapidly activate Arabidopsis MAP kinases ATMPK4 and ATMPK6. *Plant J.* 24 (5), 655–665. doi:10.1046/j.1365-313x.2000.00913.x

Ishizaki-Nishizawa, O., Fujii, T., Azuma, M., Sekiguchi, K., Murata, N., Ohtani, T., et al. (1996). Low-temperature resistance of higher plants is significantly enhanced by a nonspecific cyanobacterial desaturase. *Nat. Biotechnol.* 14 (8), 1003–1006. doi:10.1038/ nbt0896-1003

Jha, U. C., Bohra, A., and Jha, R. (2017). Breeding approaches and genomics technologies to increase crop yield under low-temperature stress. *Plant Cell Rep.* 36 (1), 1–35. doi:10.1007/s00299-016-2073-0

Jiang, T., Zhai, J. Q., Luo, Y., Su, B. D., Chao, Q. C., Wang, Y. J., et al. (2022). Understandings of assessment reports on climate change impacts, adaptation and vulnerability:progress from IPCC AR5 to IPCC AR6. *Trans. Atmos. Sci.* 45 (4), 502–511. doi:10.13878/j.cnki.dqkxxb.20220529013

Ju, W., Yang, C. F., Zhang, S. H., Tian, J. C., Hai, Y., and Yang, X. J. (2012). Mapping QTL for cell membrane permeability of leaf treated by low temperature in winter wheat. *Crop J.* 38 (7), 1247–1252. doi:10.3724/SP.J.1006.2012.01247

Juan, J. X., Yu, X. H., Jiang, X. M., Gao, Z., Zhang, Y., Li, W., et al. (2015). Agrobacterium-mediated transformation of tomato with the *ICE1* transcription factor gene. *Genet. and Mol. Res.* 14 (1), 597–608. doi:10.4238/2015.January.30.1 Kamata, T., and Uemura, M. (2004). Solute accumulation in heat seedlings during cold acclimation: contribution to increased freezing tolerance. *Cryoletters* 25 (5), 311–322. doi:10.1163/1568540042781748

Kim, D., Alptekin, B., and Budak, H. (2018). CRISPR/Cas9 genome editing in wheat. Funct. and Integr. Genomics 18, 31-41. doi:10.1007/s10142-017-0572-x

Kodama, H., Horiguchi, G., Nishiuchi, T., Nishimura, M., and Iba, K. (1995). Fatty acid desaturation during chilling acclimation is one of the factors involved in conferring low-temperature tolerance to young tobacco leaves. *Plant Physiol.* 107 (4), 1177–1185. doi:10.1104/pp.107.4.1177

Lalk, I., and Dorffling, K. (1985). Hardening, abscisic acid, proline and freezing resistance in two winter wheat varieties. *Physiol. Plant.* 63 (3), 287–292. doi:10.1111/j. 1399-3054.1985.tb04267.x

Lee, B. H., Henderson, D. A., and Zhu, J. K. (2005). The Arabidopsis cold-responsive transcriptome and its regulation by ICE1. *Plant Cell* 17 (11), 3155–3175. doi:10.1105/tpc.105.035568

Lei, C., Li, M., Chen, Z., He, W., Liu, B., Liu, S., et al. (2023). Molecular mapping of two novel cold resistance genes in common wheat by 660K SNP array. *Mol. Breed.* 43 (12), 83. doi:10.1007/s11032-023-01425-w

Li, H., Ding, Y., Shi, Y., Zhang, X., Zhang, S., Gong, Z., et al. (2017). MPK3- and MPK6-mediated ICE1 phosphorylation negatively regulates ICE1 stability and freezing tolerance in Arabidopsis. *Dev. Cell* 43 (5), 630–642. doi:10.1016/j.devcel.2017.09.025

Limin, A. E., Danyluk, J., Chauvin, L. P., Fowler, D. B., and Sarhan, F. (1997). Chromosome mapping of low-temperature induced *Wcs120* family genes and regulation of cold-tolerance expression in wheat. *Mol. General Genet.* 253 (6), 720–727. doi:10.1007/s004380050376

Limin, A. E., and Fowler, D. B. (2006). Low-temperature tolerance and genetic potential in wheat (*Triticum aestivum L.*): response to photoperiod, vernalization, and plant development. *Planta* 224 (2), 360–366. doi:10.1007/s00425-006-0219-y

Liu, L., Cang, J., Yu, J., Wang, X., Huang, R., Wang, J., et al. (2013). Effects of exogenous abscisic acid on carbohydrate metabolism and the expression levels of correlative key enzymes in winter wheat under low temperature. *Biosci. Biotechnol. Biochem.* 77 (3), 516–525. doi:10.1271/bbb.120752

Liu, X., Fu, L., Qin, P., Sun, Y., Liu, J., and Wang, X. (2019). Overexpression of the wheat *trehalose 6-phosphate synthase 11* gene enhances cold tolerance in *Arabidopsis thaliana*. *Gene* 710, 210–217. doi:10.1016/j.gene.2019.06.006

Lu, X., Zhang, F., Zhang, C., Li, G., Du, Y., Zhao, C., et al. (2024). TaTPS11 enhances wheat cold resistance by regulating source-sink factor. *Plant physiology Biochem.* 211, 108695. doi:10.1016/j.plaphy.2024.108695

Lv, D. K., Bai, X., Li, Y., Ding, X. D., Ge, Y., Cai, H., et al. (2010). Profiling of coldstress-responsive miRNAs in rice by microarrays. *Gene* 459 (1-2), 39–47. doi:10.1016/j. gene.2010.03.011

Lv, Y., Cang, J., Lu, Q., Yang, N., Feng, M. F., Meng, D. Y., et al. (2017). Bioinformatic analysis and effect of low temperature and ABA on *TabZIP1* expression pattern of winter wheat dongnongdongmai 1. *J. Triticeae Crops* 37 (1), 22–29. doi:10.7606/j.issn. 1009-1041.2017.01.04

Lv, Y., Song, C. H., Lu, Q. W., Tian, Y., Li, H. D., Zhang, D., et al. (2018). The expression characteristics of transcription factors regulated by exogenous ABA in winter wheat (*Triticum aestivum*) under cold stress. *Russ. J. Plant Physiology* 65 (6), 842-848. doi:10.1134/s1021443718060134

Lyons, J. M. (1973). Chilling injury in plants. Annu. Rev. Plant Physiology 24 (1), 445-466. doi:10.1146/annurev.pp.24.060173.002305

Malko, M. M., Peng, X., Gao, X., Cai, J., Zhou, Q., Wang, X., et al. (2023). Effect of exogenous calcium on tolerance of winter wheat to cold stress during stem elongation stage. *Plants (Basel).* 12 (21), 3784. doi:10.3390/plants12213784

Matsumura, T., Tabayashi, N., Kamagata, Y., Souma, C., and Saruyama, H. (2010). Wheat catalase expressed in transgenic rice can improve tolerance against low temperature stress. *Physiol. Plant.* 116 (3), 317–327. doi:10.1034/j.1399-3054.2002. 1160306.x

Miller, A. K., Galiba, G., and Dubcovsky, J. (2006). A cluster of 11 CBF transcription factors is located at the frost tolerance locus Fr-Am2 in Triticum monococcum. Mol. General Genet. 275 (2), 193–203. doi:10.1007/s00438-005-0076-6

Mohseni, S., Che, H., Djillali, Z., Dumont, E., Nankeu, J., and Danyluk, J. (2012). Wheat CBF gene family: identification of polymorphisms in the *CBF* coding sequence. *Genome* 55 (12), 865–881. doi:10.1139/gen-2012-0112

Nadarajah, K. K. (2020). ROS homeostasis in abiotic stress tolerance in plants. Int. J. Mol. Sci. 21 (15), 5208. doi:10.3390/ijms21155208

Naya, L., Paul, S., Oswaldo, V.-L., Mendoza-Soto, A. B., Nova-Franco, B., Sosa-Valencia, G., et al. (2014). Regulation of copper homeostasis and biotic interactions by MicroRNA 398b in common bean. *Plos One* 9, e84416. doi:10.1371/journal.pone. 0084416

Ou, X. Q., and Wang, Y. (2019). Preliminary study on wheat breeding for late spring coldness tolerance in south of Huanghuai region. *J. Triticeae Crops* 39, 560–566. doi:10. 7606/j.issn.1009-1041.2019.05.07

Pang, Y., Wu, Y., Liu, C., Li, W., St Amand, P., Bernardo, A., et al. (2021). Highresolution genome-wide association study and genomic prediction for disease resistance and cold tolerance in wheat. Theor. Appl. Genet. 134 (9), 2857-2873. doi:10.1007/s00122-021-03863-6

Pearce, S., Zhu, J., Boldizsár, Á., Vágújfalvi, A., Burke, A., Garland-Campbell, K., et al. (2013). Large deletions in the *CBF* gene cluster at the *Fr-B2* locus are associated with reduced frost tolerance in wheat. *Theor. Appl. Genet.* 126 (11), 2683–2697. doi:10.1007/s00122-013-2165-y

Peng, K., Tian, Y., Cang, J., Yu, J., Wang, D., He, F., et al. (2022). Overexpression of *TaFBA-A10* from winter wheat enhances freezing tolerance in *Arabidopsis thaliana*. *J. Plant Growth Regul.* 41 (1), 314–326. doi:10.1007/s00344-021-10304-7

Peng, K., Tian, Y., Sun, X., Song, C., Ren, Z., Bao, Y., et al. (2021). tae-miR399-UBC24 module enhances freezing tolerance in winter wheat via a CBF signaling pathway. J. Agric. food Chem. 69 (45), 13398–13415. doi:10.1021/acs.jafc.1c04316

Skinner, J. S., von Zitzewitz, J., Szucs, P., Marquez-Cedillo, L., Filichkin, T., Amundsen, K., et al. (2005). Structural, functional, and phylogenetic characterization of a large *CBF* gene family in barley. *Plant Mol. Biol.* 59 (4), 533–551. doi:10.1007/s11103-005-2498-2

Soleimani, B., Lehnert, H., Babben, S., Keilwagen, J., Koch, M., Arana-Ceballos, F. A., et al. (2022). Genome wide association study of frost tolerance in wheat. *Sci. Rep.* 12 (1), 5275. doi:10.1038/s41598-022-08706-y

Soltész, A., Smedley, M., Vashegyi, I., Galiba, G., Harwood, W., and Vágújfalvi, A. (2013). Transgenic barley lines prove the involvement of *TaCBF14* and *TaCBF15* in the cold acclimation process and in frost tolerance. *J. Exp. Bot.* 64 (7), 1849–1862. doi:10. 1093/jxb/ert050

Subedi, K. D., Gregory, P. J., Summerfield, R. J., and Gooding, M. J. (1998). Cold temperatures and boron deficiency caused grain set failure in spring wheat (*Triticum aestivum L.*). *Field Crops Res.* 57 (3), 277–288. doi:10.1016/S0378-4290(97)00148-2

Sun, X., Fan, G., Su, L., Wang, W., Liang, Z., Li, S., et al. (2015). Identification of coldinducible microRNAs in grapevine. *Front. plant Sci.* 6, 595. doi:10.3389/fpls.2015.00595

Sutka, J. (1994). "Genetic control of frost tolerance in wheat (*Triticum aestivum L*)," in *Breeding fodder crops for marginal conditions. Developments in plant breeding.* Editors O. A. Rognli, E. Solberg, and I. Schjelderup (Dordrecht: Springer). doi:10.1007/978-94-011-0966-6_42

Sutka, J. (2001). Genes for frost resistance in wheat. *Euphytica* 119 (1-2), 169–177. doi:10.1023/a:1017520720183

Takumi, S., Shimamura, C., and Kobayashi, F. (2008). Increased freezing tolerance through up-regulation of downstream genes via the wheat *CBF* gene in transgenic tobacco. *Plant Physiology Biochem.* 46 (2), 205–211. doi:10.1016/j.plaphy.2007.10.019

Tian, Y., Peng, K., Bao, Y., Zhang, D., Meng, J., Wang, D., et al. (2021). *Glucose-6-phosphate dehydrogenase* and 6-phosphogluconate dehydrogenase genes of winter wheat enhance the cold tolerance of transgenic Arabidopsis. *Plant physiology Biochem.* 161, 86–97. doi:10.1016/j.plaphy.2021.02.005

Tian, Y., Peng, K., Ma, X., Ren, Z., Lou, G., Jiang, Y., et al. (2023). Overexpression of *TaMYB4* confers freezing tolerance in *Arabidopsis thaliana*. *Int. J. Mol. Sci.* 24 (13), 11090. doi:10.3390/ijms241311090

Tiwari, B., Habermann, K., Arif, M. A., Weil, H. L., Garcia-Molina, A., Kleine, T., et al. (2020). Identification of small RNAs during cold acclimation in *Arabidopsis thaliana*. *BMC Plant Biol.* 20 (1), 298. doi:10.1186/s12870-020-02511-3

Tóth, B., Galiba, G., Fehér, E., Sutka, J., and Snape, J. W. (2003). Mapping genes affecting flowering time and frost resistance on chromosome 5B of wheat. *Theor. Appl. Genet.* 107 (3), 509–514. doi:10.1007/s00122-003-1275-3

Tshikunde, N. M., Mashilo, J., Shimelis, H., and Odindo, A. (2019). Agronomic and physiological traits, and associated quantitative trait loci (QTL) affecting yield response

in wheat (Triticum aestivum L.): a review. Front. plant Sci. 10, 1428. doi:10.3389/fpls. 2019.01428

Türkan, I., Bor, M., Izdemir, F., and Koca, H. (2005). Differential responses of lipid peroxidation and antioxidants in the leaves of drought-tolerant P. acutifolius Gray and drought-sensitive P. vulgaris L. subjected to polyethylene glycol mediated water stress. *Plant Sci.* 168 (1), 223–231. doi:10.1016/j.plantsci.2004.07.032

Valluru, R., Link, J., and Claupein, W. (2012). Consequences of early chilling stress in two Triticum species: plastic responses and adaptive significance. *Plant Biol. Stuttg. Ger.* 14 (4), 641–651. doi:10.1111/j.1438-8677.2011.00540.x

Venzhik, Y., Deryabin, A., and Zhukova, K. (2024). Au-based nanoparticles enhance low temperature tolerance in wheat by regulating some physiological parameters and gene expression. *Plants (Basel)* 13 (9), 1261. doi:10.3390/plants13091261

Vij, S., and Tyagi, A. K. (2007). Emerging trends in the functional genomics of the abiotic stress response in crop plants. *Plant Biotechnol. J.* 5 (3), 361–380. doi:10.1111/j. 1467-7652.2007.00239.x

Wang, R., Yu, M., Xia, J., Xing, J., Fan, X., Xu, Q., et al. (2022). Overexpression of *TaMYC2* confers freeze tolerance by ICE-CBF-COR module in *Arabidopsis thaliana*. *Front. plant Sci.* 13, 1042889. doi:10.3389/fpls.2022.1042889

Wang, R., Yu, M., Zhao, X., Xia, J., Cang, J., and Zhang, D. (2024). Overexpression of *TaMPK3* enhances freezing tolerance by increasing the expression of *ICE-CBF-COR* related genes in the *Arabidopsis thaliana*. *Funct. plant Biol.* 51, FP23144. doi:10.1071/FP23144

Whaley, J. M., Kirby, E. J. M., Spink, J. H., Foulkes, M. J., and Sparkes, D. L. (2004). Frost damage to winter wheat in the UK: the effect of plant population density. *Eur. J. Agron.* 21 (1), 105–115. doi:10.1016/S1161-0301(03)00090-X

Yang, C., and Yuan, P. (2014). The occurrence of wheat frost damage and preventive measures in the China Huanghuai region. *Mod. Agric. Sci. Technol.* 2 (2). doi:10.3969/j. issn.1007-5739.2014.02.060

Yang, T., Chaudhuri, S., Yang, L., Du, L., and Poovaiah, B. W. (2010). A calcium/ calmodulin-regulated member of the receptor-like kinase family confers cold tolerance in plants. *J. Biol. Chem.* 285 (10), 7119–7126. doi:10.1074/jbc.M109.035659

Yu, J., Zhang, L., Cui, H., Zhang, Y. X., Cang, J., Hao, Z. B., et al. (2008). Physiological and biochemical characteristics of dongnongdongmai 1. *Crop J.* 34 (11), 2019–2025. doi:10.3724/SP.J.1006.2008.02019

Yu, M. M., Wang, R., Xia, J. Q., Li, C., Xu, Q. H., Cang, J., et al. (2023). JA-induced TaMPK6 enhanced the freeze tolerance of *Arabidopsis thaliana* through regulation of ICE-CBF-COR module and antioxidant enzyme system. *Plant Sci.* 329, 111621. doi:10. 1016/j.plantsci.2023.111621

Zeng, Y., Yu, J., Cang, J., Liu, L., Mu, Y., Wang, J., et al. (2011). Detection of sugar accumulation and expression levels of correlative key enzymes in winter wheat (*Triticum aestivum*) at low temperatures. *J. Agric. Chem. Soc. Jpn.* 75 (4), 681–687. doi:10.1271/bbb.100813

Zhang, L., Zhao, G., Xia, C., Jia, J., Liu, X., and Kong, X. (2012). Overexpression of a wheat MYB transcription factor gene *TaMYB56-B* enhances tolerances to freezing and salt stresses in transgenic Arabidopsis. *Gene* 505 (1), 100–107. doi:10.1016/j.gene.2012. 05.033

Zhang, N., Li, L., Zhang, L., Li, J., Fang, Y., Zhao, L., et al. (2020). Abscisic acid enhances tolerance to spring freeze stress and regulates the expression of ascorbate–glutathione biosynthesis-related genes and stress-responsive genes in common wheat. *Mol. Breed.* 40 (11), 108–118. doi:10.1007/s11032-020-01187-9

Zheng, B., Chapman, S. C., Christopher, J. T., Frederiks, T. M., and Chenu, K. (2015). Frost trends and their estimated impact on yield in the Australian wheatbelt. *Procedia Environ. Sci.* 29, 171–172. doi:10.1016/j.proenv.2015.07.244