


# RIPK1 inhibition in malignant cells potentiates immunotherapy and radiotherapy outcome

Jonathan G. Pol <sup>a,b</sup>, Andrea Checcolia<sup>a,b,c</sup>, Manuela Lizarralde-Guerrero<sup>a,b,d</sup>, and Guido Kroemer<sup>a,b,e</sup>

<sup>a</sup>Centre de Recherche des Cordeliers, Equipe labellisée par la Ligue contre le cancer, Université de Paris Cité, Sorbonne Université, Inserm U1138, Institut Universitaire de France, Paris, France; <sup>b</sup>Metabolomics and Cell Biology Platforms, Institut Gustave Roussy, Villejuif, France; <sup>c</sup>Université Paris Sciences et Lettres (PSL), Paris, France; <sup>d</sup>Université Paris-Saclay, Faculté de Médecine, Kremlin-Bicêtre, France; <sup>e</sup>Department of Biology, Institut du Cancer Paris CARPEM, Hôpital Européen Georges Pompidou, AP-HP, Paris, France

## ABSTRACT

Apoptosis, necroptosis and pro-inflammatory NF- $\kappa$ B-dependent signaling are repressed by receptor-interacting serine/threonine-protein kinase 1 (RIPK1). A recent paper in *Immunity* describes a small molecule inducing the proteolytic degradation of RIPK1. In preclinical experiments, this RIPK1 inhibitor improved the anticancer efficacy of radiotherapy, immunotherapy (with PD-1 blockade) and radioimmunotherapy (with CTLA-4 blockade).

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## Main text

In a recent study, Mannion J et al. investigated the role of receptor-interacting serine/threonine-protein kinase 1 (RIPK1) in cancer biology, highlighting its dual function as both a driver of cell survival and an orchestrator of immunogenic cell death (ICD).<sup>1</sup> This builds upon a growing body of literature emphasizing the importance of ICD in enhancing anticancer immune responses, a concept that is increasingly seen as central to optimizing cancer therapies, particularly in the context of overcoming resistance to immunotherapy and radiotherapy.

ICD represents a critical mechanism in cancer immunosurveillance, wherein stressed or dying cancer cells release damage-associated molecular patterns (DAMPs) that stimulate an effective antitumor immune response.<sup>2,3</sup> In this process, various signaling pathways, including those regulated by RIPK1, play crucial roles in determining whether a cell death event will be immunogenic, thereby influencing the ability of the immune system to recognize and eradicate cancer cells.

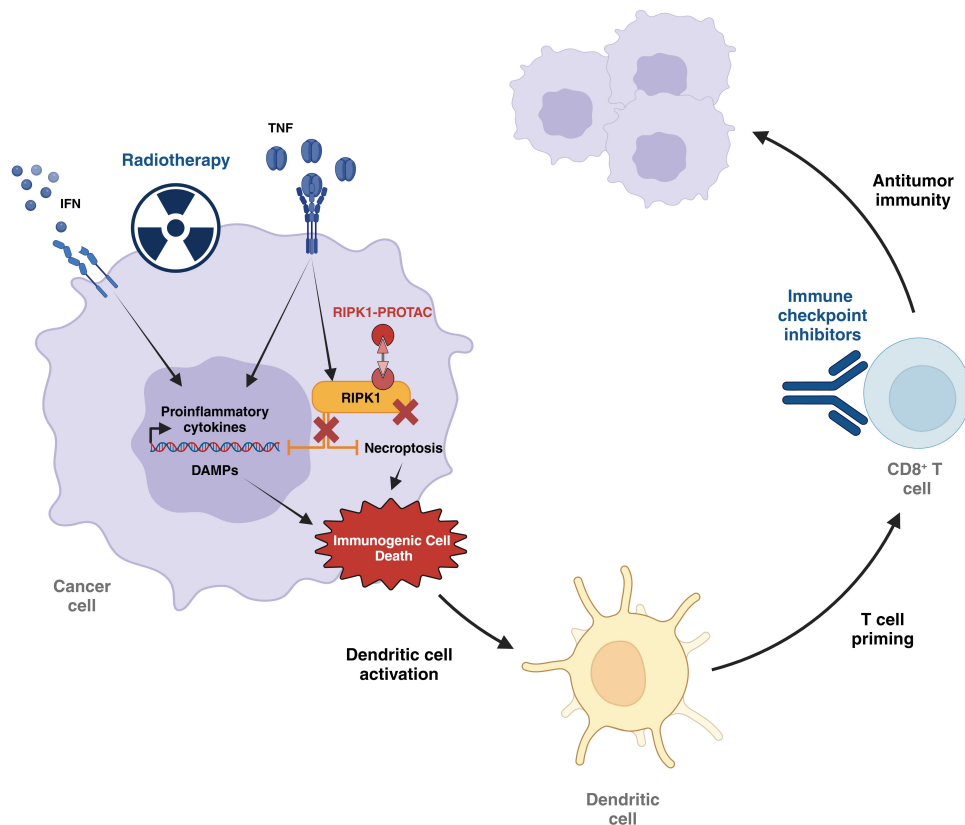
Mannion and colleagues extend these findings by focusing on the complex regulatory functions of RIPK1 in both cell death and survival, involving the NF- $\kappa$ B pathway, noting that cancer cells frequently exploit the scaffold function of RIPK1 to evade necroptosis, a form of regulated necrosis associated with robust immunogenicity.<sup>4</sup> By developing a small-molecule proteolysis-targeting chimera (PROTAC) that selectively degrades RIPK1, they have identified a strategy to circumvent this cancer defense mechanism, triggering necroptosis and promoting ICD.

Their research reveals that degradation of RIPK1 not only sensitizes cancer cells to treatment-induced tumor necrosis factor (TNF) and interferons but also potentiates the

immunostimulatory effects of radiotherapy, anti-PDCD1 (programmed cell death 1, best known as PD-1)-based immunotherapy, and anti-CTLA4-based radioimmunotherapy (**Figure 1**). This finding echoes previous reports showing that necroptotic cancer cells can enhance CD8<sup>+</sup> T cell cross-priming and elicit durable antitumor immunity.<sup>5,6</sup>

However, at least at first glance, the role of RIPK1 in modulating the immunogenicity of necroptosis appears controversial. A prior report suggested that RIPK1 activation is required for necroptosis to be immunogenic because knockout of the *Ripk1* gene reduced the capacity of polyinosinic-polycytidylic acid (poly I:C)-treated cells (which undergo necroptosis) to induced immune responses against the model antigen ovalbumin and to elicit protective immune responses in vaccination experiments.<sup>5</sup> In sharp contrast, PROTAC-induced degradation of RIPK1 enhances the immunogenicity of cell death.<sup>4</sup> Future studies must investigate whether the knockout of the *Ripk1* gene and the PROTAC-induced degradation of RIPK1 protein have exactly the same cell biological consequences or whether one of the two methods for inhibiting RIPK1 may have off-target effects explaining these discrepant conclusions. Alternatively, the duration of RIPK1 inhibition might affect the system. Indeed, acute pharmacological removal of RIPK1 may have rather distinct effects than knockout of the gene, because the latter can lead to long-term adaptation of the cells, as this has been suggested by Mannion et al.<sup>4</sup>

The work by Mannion et al. aligns with emerging approaches aimed at exploiting immunogenic forms of cell death, such as necroptosis and ferroptosis, to boost



**Figure 1.** RIPK1-PROTAC improves the efficacy of radio- and immune-therapy. RIPK1 inhibits RIPK3/MLKL-triggered necroptosis and the production of proinflammatory factors. RIPK1-PROTAC degrades RIPK1 which sensitizes cancer cells treated by radiotherapy and/or immune checkpoint inhibitors (anti-PD-1, anti-CTLA-4) to immunogenic necroptotic cell death. The related emission/exposure of DAMPs accompanied by the secretion of inflammatory cytokines (e.g. chemokines, IFNs, TNFs) favor the recruitment and activation of dendritic cells and the subsequent priming of tumor-specific cytotoxic CD8<sup>+</sup> T cells. DAMP, damage-associated molecular pattern; IFN, interferon; PROTAC, proteolysis-targeting chimera; TNF, tumor-necrosis factor.

antitumor immune responses.<sup>7–9</sup> Along this way, Zhang J et al. recently described a synthetic compound named necroside-1 (NC1). NC1 operates as an inducer of TNF-independent necrosis accompanied with the release of DAMPs, underscoring the therapeutic potential of pharmacologically triggering necrotic cell death to enhance the immunogenicity of cancer cells.<sup>10</sup>

This work highlights not only the versatility of RIPK1 as a therapeutic target but also the far-reaching implications of RIPK1 degradation in driving lasting antitumor immunity. By disrupting cancer cells' reliance on RIPK1 for survival and immune modulation, the PROTAC approach amplifies the inflammatory cascade associated with DAMP release, enhances dendritic cell recruitment, and strengthens CD8<sup>+</sup> T cell priming, all of which are critical to effective antitumor responses. Mannion et al.'s findings provide a significant contribution to the field, bridging molecular targeting and immune modulation to potentiate current treatment modalities and present a promising pathway for translational cancer therapies that synergize with existing therapeutic frameworks. This could be transformative in clinical contexts where treatment-resistant malignancies necessitate innovative and multi-faceted therapeutic strategies.

## Disclosure statement

J.G.P. is the inventor of patents covering the diagnosis, prognosis, and treatment of cancers, including patents licensed to Turnstone Biologics

and Therafast Bio. GK has been holding research contracts with Daiichi Sankyo, Eleor, Kaleido, Lytix Pharma, PharmaMar, Osasuna Therapeutics, Samsara Therapeutics, Sanofi, Sutro, Tollys, and Vascage. GK is on the Board of Directors of the Bristol Myers Squibb Foundation France. GK is a scientific co-founder of everImmune, Osasuna Therapeutics, Samsara Therapeutics and Therafast Bio. GK is in the scientific advisory boards of Hevolution, Institut Servier, Longevity Vision Funds and Rejuveron Life Sciences. GK is the inventor of patents covering therapeutic targeting of aging, cancer, cystic fibrosis and metabolic disorders. GK's wife, Laurence Zitvogel, has held research contracts with Glaxo Smyth Kline, Incyte, Lytix, Kaleido, Innovate Pharma, Daiichi Sankyo, Pilege, Merus, Transgene, 9 m, Tusk and Roche, was on the on the Board of Directors of Transgene, is a cofounder of everImmune, and holds patents covering the treatment of cancer and the therapeutic manipulation of the microbiota. GK's brother, Romano Kroemer, was an employee of Sanofi and now consults for Boehringer-Ingelheim. The funders had no role in the writing of the manuscript.

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## ORCID

Jonathan G. Pol  <http://orcid.org/0000-0002-8355-7562>

## Data availability statement

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## References

- Mannion J, Gifford V, Bellenie B, Fernando W, Ramos Garcia L, Wilson R, John SW, Udainiya S, Patin EC, Tiu C, et al. A RIPK1-specific PROTAC degrader achieves potent antitumor activity by enhancing immunogenic cell death. *Immunity*. 2024;57(7):1514–1532.e15. doi:10.1016/j.immuni.2024.04.025.
- Kroemer G, Chan TA, Eggermont AMM, Galluzzi L. Immunosurveillance in clinical cancer management. *CA A Cancer J Clinicians*. 2024;74(2):187–202. doi:10.3322/caac.21818.
- Galluzzi L, Guilbaud E, Schmidt D, Kroemer G, Marincola FM. Targeting immunogenic cell stress and death for cancer therapy. *Nat Rev Drug Discov*. 2024;23(6):445–460. doi:10.1038/s41573-024-00920-9.
- Yang H, Ma Y, Chen G, Zhou H, Yamazaki T, Klein C, Pietrocola F, Vacchelli E, Souquere S, Sauvat A, et al. Contribution of RIP3 and MLKL to immunogenic cell death signaling in cancer chemotherapy. *Oncoimmunology*. 2016;5(6):e1149673. doi:10.1080/2162402X.2016.1149673.
- Yatim N, Jusforgues-Saklani H, Orozco S, Schulz O, Barreira da Silva R, Reise, Green DR, Reis e Sousa C, Oberst A, Albert ML. RIPK1 and nf-κB signaling in dying cells determines cross-priming of CD8 + T cells. *Science*. 2015;350(6258):328–334. doi:10.1126/science.aad0395.
- Aaes TL, Kaczmarek A, Delvaeye T, De Craene B, De Koker S, Heyndrickx L, Delrue I, Taminau J, Wiernicki B, De Groot P, et al. Vaccination with necroptotic cancer cells induces efficient anti-tumor immunity. *Cell Rep*. 2016;15(2):274–287. doi:10.1016/j.celrep.2016.03.037.
- Meier P, Legrand AJ, Adam D, Silke J. Immunogenic cell death in cancer: targeting necroptosis to induce antitumour immunity. *Nat Rev Cancer*. 2024;24(5):299–315. doi:10.1038/s41568-024-00674-x.
- Catanzaro E, Demuyneck R, Naessens F, Galluzzi L, Krysko DV. Immunogenicity of ferroptosis in cancer: a matter of context? *Trends In Cancer*. 2024;10(5):407–416. doi:10.1016/j.trecan.2024.01.013.
- Kayagaki N, Webster JD, Newton K. Control of cell death in health and disease. *Annu Rev Pathol Mech Dis*. 2024;19(1):157–180. doi:10.1146/annurev-pathmechdis-051022-014433.
- Zhang J, Trojel-Hansen C, Wang J, Zhang Z, Wang X, Qiao Y, Jiao H, Michaud M, Kepp O, Jäättelä M, et al. Necrocidin 1 mediates necrotic cell death and immunogenic response in human cancer cells. *Cell Death Dis*. 2023;14(4):238. doi:10.1038/s41419-023-05740-0.