Pearls & Oy-sters: Whole-Genome Sequencing in Critically Ill Neurologic Patient Leads to Diagnosis With Treatment Implications

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Abstract

Many adult patients with a history of seizures and global developmental delay do not have an identified etiology for their epilepsy. Rapid whole-genome sequencing (rWGS) can be used to identify a genetic etiology in critically ill patients to provide actionable interventions. In this case, a 27-year-old patient with a history of epilepsy, global developmental delay, and intellectual disability presented with altered mental status and new abnormal movements. The patient acutely declined over the course of 24–48 hours of presentation, including nonconvulsive status epilepticus leading to intubation for airway protection, 2 episodes of ventricular tachycardia requiring synchronized cardioversion, and 1 episode of supraventricular tachycardia. The patient was found to be in metabolic crisis. Metabolic workup and rapid whole-genome sequencing were sent. Patient was treated with 10% dextrose in normal saline and a mitochondrial cocktail. She received treatment with ammonia scavengers and hemodialysis with resolution of metabolic crisis. rWGS found a homozygous pathogenic variant in TANGO2 and a de novo pathogenic variant in KCNQ1, ultimately leading to the creation of a metabolic emergency protocol and implantable cardioverter defibrillator placement. This case highlights the use of rWGS in an acutely ill patient leading to actionable interventions. It also highlights the utility and importance of genetic sequencing in reevaluation of adult neurologic patients.

Pearls

- TANGO2-related metabolic crisis can present at any age and can include lactic acidosis, hyperammonemia, hypoglycemia, life-threatening arrhythmias, rhabdomyolysis, and/or seizures.
- Whole-genome sequencing can provide new explanations for acute enigmatic presentation, altering management.

Oy-sters

- It is important to keep genetic metabolic epilepsies in the differential for patients with undiagnosed epilepsy syndromes without identifiable structural cause.
- Repeat or reevaluation of previous genetic testing can lead to new diagnoses with implications for management.
- Metabolic crisis can lead to life-threatening arrhythmias.

Case Report

A 27-year-old woman with diagnosis of probable focal epilepsy previously well controlled on levetiracetam, developmental delay (DD), intellectual disability (ID), episodic ataxia, and

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hypothyroidism presented to the emergency department (ED) with subacute functional decline and seizures. The day before presentation, she "slumped over" and was unable to ambulate or talk. During a neurology virtual visit, her activity, responsiveness, and mental status improved toward baseline. An increase in levetiracetam was recommended. On the day of presentation, she woke in her usual state of health but then experienced a second similar episode, prompting presentation to the ED.

On initial evaluation in the ED, she was obtunded, but mental status improved over the next 12 hours. Workup revealed elevated transaminases (aspartate transaminase 78 U/L [N = $\langle 34 \rangle$, alanine transaminase 114 U/L [N = 10–49]), urinalysis suggestive of infection (though culture was ultimately negative), and liver lesions on ultrasound. CT liver revealed several hypodense lesions, consistent with hemangiomas. She resumed ambulation, and speech improved toward baseline. Thus, her residual altered mental status was presumed to be postictal. Neurology recommended 2 g of levetiracetam bolus and increase in maintenance dosing, with outpatient followup. However, she became lethargic hours later and was admitted to an observation unit.

Over the next day, mental status worsened with marked lethargy. CT head, MRI brain, and continuous EEG were ordered. Neuroimaging was unrevealing (Figure 1A). EEG revealed multiple nonconvulsive seizures (Figure 1B), which improved with intravenous lorazepam. Venous blood gas revealed metabolic acidosis with lactate 5.5 mmol/L ($N =$ <2.2). She received multiple intravenous fluid boluses, was started on empiric antibiotics at meningitic dosing, and was transferred to the intensive care unit.

She was intubated for airway protection. After induction with rocuronium and etomidate, she developed supraventricular tachycardia, treated with metoprolol (Figure 2A). On day 2, she experienced episodes of pulseless ventricular tachycardia requiring synchronized cardioversion. Long QT was noted on electrocardiogram (Figure 2B). Multiple medical specialties were consulted, including genetics who were concerned for TANGO2-related disease, mitochondrial disorder, or other inborn error of metabolism (IEM). Metabolic workup including plasma amino acids, urine organic acids, acylcarnitine profile, creatine kinase (CK), ammonia, and rapid trio wholegenome sequencing (rWGS) was sent. Family history was noncontributory, and parents denied consanguinity.

Testing revealed hyperammonemia (208 μ mol/L, N = <60) and elevated CK (1651 U/L, $N = 26-180$), suggesting metabolic crisis. Plasma amino acids, urine organic acids, and acylcarnitine profiles were consistent with liver dysfunction.

Figure 1 Representative MRI and EEG Findings Before and After Arrest

(A) Diffusion-weighted imaging (DWI) MRI sequence on day 1 of hospital admission showing no diffusion restriction and (B) initial EEG showing onset (red arrow) of a slowly evolving generalized electrographic seizure (low-frequency filter [LFF]: 1 Hz, high-frequency filter [HFF]: 70 Hz, notch: OFF, sensitivity: 10 μV/ mm, timebase: 15 mm/s). (C) DWI MRI sequence on day 6 of admission showing left posterior quadrant cortical diffusion restriction (white arrow) correlating to (D) left posterior quadrant focal seizures (black bounding boxes) in T3-T5, T5-O1, and P3-O1 seen on continuous EEG monitoring the following day (LFF: 1 Hz, HFF: 70 Hz, notch: OFF, sensitivity: 10 μV/mm, timebase: 15 mm/s).

Figure 2 Electrocardiogram (EKG) Findings Before and After Cardiac Arrest

B. Postarrest baseline

C. Routine follow-up post-ICD

(A) Preintervention EKG showing widened QRS complexes (arrows) and prolonged QT intervals with a QTc of 613 milliseconds (black bar). (B) Postarrest and intervention EKG with continued prolonged QTc of 601 milliseconds (black bar). (C) Surveillance EKG from approximately 1 year after initial presentation and following ICD placement. QTc now at 376 milliseconds (black bar). All EKGs taken at paper speed of 25 mm/s.

Ten percent dextrose in normal saline (D10NS) and a mitochondrial cocktail were started. She received a loading dose of ammonia scavenger Ammonul and a 3-hour run of hemodialysis for hyperammonemia. Ammonul infusion and D10NS were continued for 3 days, which resolved the hyperammonemia, hyperCKemia, and metabolic acidosis.

Sedation was weaned, but depressed mental status persisted. Repeat MRI brain showed restricted diffusion and mild pachymeningeal enhancement involving the left parietal and occipital lobes, radiographically concerning for mitochondrial encephalopathy, lactic acidosis, and stroke-like episodes (Figure 1C). Repeat continuous EEG on days 7–8 found 2 electrographic seizures at O1 (Figure 1D) treated with 2 separate 10 mg of phenytoin equivalents/kilogram fosphenytoin loads followed by maintenance dosing.

Due to testing laboratory delays, rWGS resulted after 12 days, revealing a homozygous pathogenic deletion of exons 3–9 in TANGO2 [(?_20041870)-(20074530_?)DEL 32.66 kb)], 1 inherited from each parent, and a de novo pathogenic variant in KCNQ1 (c.1766G>T, p.G589V) associated with autosomal dominant long QT syndrome 1 (MIM# 192500). Both TANGO2 and KCNQ1 confer significant risk for long QT; thus, she underwent implantable cardioverter defibrillator (ICD) placement (Figure 2C). She received a metabolic action plan and started supplementation of B complex vitamins. Nitrogen-scavenging medications were discontinued based on genomic results. After further improvement, she was discharged to inpatient rehabilitation and subsequently home.

Discussion

TANGO2-related metabolic encephalopathy and arrhythmias (TRMEA, MIM#: 616878) is an autosomal recessive condition first described in the literature in 2016 .¹ TRMEA was originally described in 9 families with episodic rhabdomyolysis, hypoglycemia, hyperammonemia, and susceptibility to tachyarrhythmias. The phenotype has expanded to include seizures, episodic ataxia, ID, DD, and hypothyroidism. The exact pathophysiology of TRMEA is unknown. Efforts to localize the protein have shown it to be involved in transport from the endoplasmic reticulum to the Golgi apparatus and functioning in the mitochondria, which may explain some features that are similar to a fatty acid oxidation disorder while in other ways acting similarly to a mitochondrial disorder.²

TRMEA typically presents in childhood, particularly with metabolic crises, but diagnosis varies from 8 months to 26 years. ³ Our patient's diagnosis at age 27 years underscores the need to consider TRMEA in adulthood. While identification of IEMs has improved with technological advancements and augmentation of knowledge regarding later-onset phenotypes, these syndromes are likely underdiagnosed. IEMs should be considered for patients with epilepsy who are critically ill, especially if the etiology of their epilepsy is unknown. A review of known IEMs identified 600 metabolic epilepsies, 70% which could be screened with standard metabolic testing, and more than 100 had significant treatment implications.⁴ While the ultimate trigger of the metabolic crisis in our patient cannot be determined, decreased caloric intake, stress, and possible infection may have contributed. Of note, despite usage immediately preceding supraventricular tachycardia, neither rocuronium nor etomidate are contraindicated from a metabolic standpoint.

This case demonstrates the utility of rWGS in a critically ill adult patient. Others have documented clear clinical and financial benefit to using this testing. 5 However, utility of genomic testing is still dependent on information provided to the interpreting laboratory. While TRMEA was highly suspected based on clinical presentation and could have been identified with single gene sequencing or gene panel, targeted testing would have missed the de novo variant in KCNQ1, another risk factor of arrhythmias or death. Without genetic diagnosis, this patient may not have undergone ICD placement (identification of TANGO2 variants alone does not usually prompt ICD placement, but identification of a second significant arrhythmogenic predisposition provided additional evidence of necessity) or implementation of a metabolic action plan and supplementation of B complex vitamins. In addition, this diagnosis has screening protocols for long-term management, which altered her ongoing care.

This case highlights the importance of genetic testing and reevaluation in adults with epilepsy. The patient previously completed chromosomal microarray (CMA) in 2010 that was reportedly negative. However, a 6-exon deletion in a gene of unknown significance then may not have been reported. Using today's technology, the homozygous deletion would be reportable, but evolution in testing will continue, and repeat testing remains important. Genetic testing panels now include multiple common genes associated with epilepsy (including TANGO2) and can detect multiexon deletions on Next-Generation Sequencing platforms. As of May 2022, TANGO2 is included on 6 commonly ordered epilepsy panels and other types of panels including hypoglycemia and rhabdomyolysis/ metabolic myopathies.

Genetic testing in adults with epilepsy leads to diagnosis in 10.9%–23% of patients tested.⁶⁻⁸ Diagnostic yield of genetic testing in adults increases with decreasing age at onset of epilepsy, infancy being highest (29.5%–38.6%).⁸⁻¹⁰ ID (14%–16%) or DD $(36.4%)$ also increase diagnostic yield.^{6,8,10} Diagnostic yield in pediatric studies for epilepsy starting less than 2–3 years of age is similarly elevated at 15.3%–38%, with highest yield in patients with epileptic encephalopathies.⁹⁻¹² Genetic diagnoses lead to actionable changes in management in $17% - 30%$ of patients.^{7,11} Our patient's dual diagnoses of TRMEA and long QT syndrome 1 led to ICD placement, mitochondrial cocktail prescription, and implementation of a metabolic emergency protocol plan.

Given lower cost, CMA is usually considered first in patients with dysmorphisms, multiple congenital anomalies, autism, DD, and/or ID. Patients who do not fit these criteria may benefit instead from targeted gene panels before more comprehensive testing.¹³ In addition, reevaluation of previously completed genetic testing is important and can lead to diagnosis for 4.8%–23.4% of patients with previously nondiagnostic tests.^{9,12} Subsequent genomic sequencing can lead to a genetic diagnosis in 32.1% of patients with previously negative testing.¹³ Therefore, previous negative testing does not eliminate the possibility of a genetic condition and must be considered in the differential diagnosis, especially when the clinical phenotype is consistent.

Conclusion

Our case highlights the utility of rWGS in an acutely ill patient leading to diagnosis with actionable interventions including metabolic emergency protocol, mitochondrial cocktail prescription, and ICD placement. Genetic testing in adults with epilepsy can lead to diagnoses more often in patients with younger age at onset, ID, or DD. Genetic diagnosis of epilepsy should remain on the differential in the appropriate clinical context, and genetic testing should be reevaluated because ongoing discoveries can lead to diagnoses with actionable interventions.

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Disclosure

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References

- 1. Lalani SR, Liu P, Rosenfeld JA, et al. Recurrent muscle weakness with rhabdomyolysis, metabolic crises, and cardiac arrhythmia due to bi-allelic TANGO2 mutations. Am J Hum Genet. 2016;98(2):347-357. doi:10.1016/ j.ajhg.2015.12.008
- 2. Milev MP, Saint-Dic D, Zardoui K, et al. The phenotype associated with variants in TANGO2 may be explained by a dual role of the protein in ER-to-Golgi transport and at the mitochondria. J Inherit Metab Dis. 2021;44(2):426-437. doi:10.1002/ jimd.12312
- Dines JN, Golden-Grant K, LaCroix A, et al. TANGO2: expanding the clinical phenotype and spectrum of pathogenic variants. Genet Med. 2019;21(3):601-607. doi: 10.1038/s41436-018-0137-y
- 4. Tumiene B, Ferreira CR, van Karnebeek CDM. 2022 Overview of metabolic epilepsies. Genes. 2022;13(3):508. doi:10.3390/genes13030508
- 5. Dimmock D, Caylor S, Waldman B, et al. Project Baby Bear: rapid precision care incorporating rWGS in 5 California children's hospitals demonstrates improved clinical outcomes and reduced costs of care. Am J Hum Genet. 2021;108(7): 1231-1238. doi:10.1016/j.ajhg.2021.05.008
- 6. Borlot F, de Almeida BI, Combe SL, Andrade DM, Filloux FM, Myers KA. Clinical utility of multigene panel testing in adults with epilepsy and intellectual disability. Epilepsia. 2019;60(8):1661-1669. doi:10.1111/epi.16273
- 7. Johannesen KM, Nikanorova N, Marjanovic D, et al. Utility of genetic testing for therapeutic decision-making in adults with epilepsy. Epilepsia. 2020;61(6):1234-1239. doi:10.1111/epi.16533
- 8. McKnight D, Bristow SL, Truty RM, et al. Multigene panel testing in a large cohort of adults with epilepsy: diagnostic yield and clinically actionable genetic findings. Neurol Genet. 2022;8(1):e650. doi:10.1212/NXG.0000000000000650
- 9. Balciuniene J, DeChene ET, Akgumus G, et al. Use of a dynamic genetic testing approach for childhood-onset epilepsy. JAMA Netw Open. 2019;2(4):e192129. doi: 10.1001/jamanetworkopen.2019.2129
- 10. Lee J, Lee C, Ki CS, Lee J. Determining the best candidates for next-generation sequencing-based gene panel for evaluation of early-onset epilepsy. Mol Genet Genomic Med. 2020;8(9):e1376. doi:10.1002/mgg3.1376
- 11. Costain G, Cordeiro D, Matviychuk D, Mercimek-Andrews S. Clinical application of targeted next-generation sequencing panels and whole exome sequencing in childhood epilepsy. Neuroscience. 2019;418:291-310. doi:10.1016/j.neuroscience.2019.08.016
- 12. Salinas V, Martínez N, Maturo JP, et al. Clinical next generation sequencing in developmental and epileptic encephalopathies: diagnostic relevance of data re-analysis and variants re-interpretation. Eur J Med Genet. 2021;64(12):104363. doi:10.1016/j.ejmg.2021.104363
- 13. Sánchez Fernández I, Loddenkemper T, Gaínza-Lein M, Sheidley BR, Poduri A. Diagnostic yield of genetic tests in epilepsy: a meta-analysis and cost-effectiveness study. Neurology. 2019;92(5):e418-e428. doi:10.1212/WNL.0000000000006850

CORRECTIONS

CSF Findings in Relation to Clinical Characteristics, Subtype, and Disease Course in Patients With Guillain-Barré Syndrome

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In the Research Article "CSF Findings in Relation to Clinical Characteristics, Subtype, and Disease Course in Patients With Guillain-Barré Syndrome" by Al-Hekem et al.,¹ author Gerardo Gutiérrez-Gutiérrez's affiliations should be listed as follows: Department of Neurology, Hospital Universitario Infanta Sofía, Paseo de Europa, 34, 28702, San Sebastián de los Reyes, Spain, and Universidad Europea de Madrid. The authors regret the omission.

Reference

Al-Hakem H, Doets AY, Stino AM, et al. CSF findings in relation to clinical characteristics, subtype, and disease course in patients with Guillain-Barré syndrome. Neurology. 2023; 100(23):e2386-e2397.

Determining Vascular Risk Factors for Dementia and Dementia Risk Prediction Across Midto Later Life

The Framingham Heart Study

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In the Research Article "Determining Vascular Risk Factors for Dementia and Dementia Risk Prediction Across Mid- to Later Life: The Framingham Heart Study" by McGrath et al.,¹ the URL linking to the supplemental material should be [links.lww.com/](http://links.lww.com/WNL/B977) [WNL/B977.](http://links.lww.com/WNL/B977) The publisher regrets the error.

Reference

1. McGrath ER, Beiser AS, O'Donnell A, et al. Determining vascular risk factors for dementia and dementia risk prediction across mid- to later life: the Framingham Heart study. Neurology. 2022;99(2):e142-e153.