# scientific reports



## **Expression pattern of long non-OPEN coding RNAs in treatment-naïve and medicated schizophrenia patients**

**Kamran Javidi Aghdam<sup>1</sup>, Behzad Baradaran<sup>2</sup>, Shima Rahmani<sup>2</sup>, Fatemeh Manafzadeh<sup>1</sup>, Seyed Gholamreza Noor Azar<sup>3</sup>, ShahrokhAghayan<sup>4</sup>, Asghar Shayannia<sup>6</sup> & SoudehGhafouri-Fard<sup>5</sup>**

**Schizophrenia is a disabling mental disorder that affects 1% of people over their lifetime. The etiology and mechanism of schizophrenia are very complex, and many genes are involved in many different signaling pathways in the etiology of this disease. According to recent studies, one of the important mechanisms altered in this disorder is the regulation of immune system and the inflammation mechanism. In the present study, we evaluated the peripheral blood expression pattern of four lncRNAs and three protein-coding genes in the treatment- naïve patients, and medicated patients compared with sex and age-matched controls. In the medicated-patients, expression levels of**  *IFNG***,** *IL18RAP***,** *AC007278.2* **were significantly up-regulated (***P***<0.05); and the expression level of**  *IFNG-AS1-001* **was significantly down-regulated compared to healthy controls (***P***<0.05). However, levels of** *IL18R1***,** *AC007278.3* **and** *IFNG-AS1-003* **were not different between these groups. In the treatment-naïve patients,** *IFNG***,** *IL18R1***,** *IL18RAP***,** *IFNG-AS1-001***,** *AC007278.2***, and** *AC007278.3* **were significantly up-regulated compared to controls. On the other hand,** *IFNG-AS1-003* **was significantly down-regulated in the treatment-naïve patients compared to controls. Based on the Spearman correlation matrix, there was a significant correlation between genes in the treatment-naïve patients. We also showed the high sensitivity and specificity of** *IFNG-AS1-003***,** *IFNG***,** *IL18R1***, and** *AC007278.3* **in the identification of treatment-naïve patients from controls. The current study contributes further evidence to the understanding of the role of lncRNAs in the pathogenesis of schizophrenia. Future research is necessary to establish the validity of lncRNAs as peripheral markers for this condition.**

**Keywords** Schizophrenia, IFNG, IL18R, IL18RAP, IFNG-AS1-001

Schizophrenia is one of the most disabling mental disorders that affects about 1% of people over their lifetime<sup>1</sup>. The prevalence of this disease is approximately 0.3 to 0.7% worldwide. The disease usually occurs sporadically. Its symptoms usually appear in adulthood and early adolescence, leading to severe disability and high level of stress<sup>[1](#page-10-0)</sup>. This disorder is determined by the existence of various symptoms, including positive symptoms (hallucinations, delusions, unusual behaviors, unusual speech, abnormal thought, and movement disorder), negative symptoms (insensibility, lack of delight, attention impairment, and sociality withdrawal), and cognitive symptoms (imperfection in executive function and precision, and agnosia)<sup>2</sup>. The etiology of schizophrenia has not been completely identified<sup>[3](#page-10-2)</sup>. However, it is clear that mechanisms of schizophrenia are very complex, and many environmental and genetic factors play a pivotal role in causing this disease. There is massive evidence indicating that changes in gene expression in the immune system contribute to the pathogenesis of this disorder<sup>4[,5](#page-10-4)</sup>. Dysregulation of the immune system and its intricate interplay with the nervous system might play a part in the etiology and physiological mechanisms of schizophrenia<sup>[6](#page-10-5)</sup>. The reciprocal relationship between the immune system and the brain has sparked a rising curiosity regarding the involvement of the immune system

<sup>1</sup>Student Research Committee, School of Medicine, Shahroud University of Medical Sciences, Shahroud, Iran. <sup>2</sup>Immunology Research Center, Tabriz University of Medical Sciences, Tabriz, Iran. <sup>3</sup>Research Center of Psychiatry and Behavioral Sciences, Tabriz University of Medical Sciences, Tabriz, Iran. 4Sexual Health and Fertility Research Center, Shahroud University of Medical Sciences, Shahroud, Iran. 5Department of Medical Genetics, Shahid Beheshti University of Medical Sciences, Tehran, Iran. <sup>6</sup>Department of Medical Biotechnology, School of Medicine, Shahroud University of Medical Sciences, Shahroud, Iran. <sup>⊠</sup>email: a.shayannia@gmail.com; s.ghafourifard@sbmu.ac.ir

in neuropsychiatric disorders. Notably, the atypical blood lymphocyte parameters, such as the levels of total T lymphocytes and T helper cells, have specifically attracted attention $^7\!$  $^7\!$  $^7\!$ .

Numerous studies have shown that inflammation and immunity play significant roles in the development of symptoms associated with schizophrenia<sup>8,[9](#page-11-0)</sup>. These studies have provided compelling evidence that systemic inflammation can have a profound impact on the brain, resulting in alterations in mood, cognition, and behavior<sup>10</sup>.

Long non-coding RNAs are a subset of non-coding transcripts and have a length of more than 200 nucleotides<sup>11</sup>. Studies have demonstrated that some lncRNAs are effective in controlling the behavior of immune cells and immune responses<sup>12</sup>, including the differentiation and activity of T and B cells, macrophages, and NK cells. Several lncRNAs play pivotal roles in cell function and participate in the pathogenesis and development of various diseases such as cance[r13a](#page-11-4)nd especially neuropsychiatric disorders and neurodegenerative diseases such as Alzheimer's disease<sup>14</sup>, Parkinson's disease<sup>15</sup>, major depressive disorder<sup>16</sup>, autism spectrum disorders<sup>17</sup>and multiple sclerosis<sup>18</sup>.

Based on the functional roles in the regulation of immune responses, we selected four lncRNAs, namely *IFNG-AS1-001*, *IFNG-AS1-003*, *AC007278.2*, and *AC007278.3*; and three protein-coding genes, namely *IFNG*, *IL18R1*, and *IL18RAP* to assess their expression in peripheral blood of treatment-naive and medicated schizophrenia patients compared with matched healthy controls. *IFNG*has been suggested to partake in the pathogenesis of schizophrenia[19](#page-11-10),[20.](#page-11-11) Methylation of the *IFNG* locus is regulated by *IFNG-AS1-001*[21](#page-11-12). *IFNG-AS1-003* gene is also located on the same chromosome as the *IFNG*[22](#page-11-13), possibly contributing to regulation of this gene. Similarly, *IL-18*is involved in the pathologic events seen in this disorder<sup>[23](#page-11-14)</sup>. This cytokine exerts its effect through its receptor being encoded by *IL18R1* and *IL18RAPgenes<sup>24</sup>*. Two functionally related lncRNAs, namely *AC007278.2* and *AC007278.3* are located on chromosome 2 inside the introns of *IL18R1* and *IL18RAP*, respectively.

This study provides new insights into the altered expression of protein-coding and non-coding genes related to the immune system and proposes them as contributors in the pathogenesis of schizophrenia and as novel biomarkers for the diagnosis of schizophrenia.

### **Materials and methods**

#### **Study participants**

The present study was performed on 50 medicated and 25 treatment-naive schizophrenia patients, and 50 sex and age-matched healthy controls. Cases were recruited from Razi hospital, Tabriz, Iran. The fifth edition of the Diagnostic and Statistical Manual of Mental Disorders  $(DSM-V)^{25}$  was applied in the diagnostic process. The participants with alcohol drinking and substance abuse or cigarette smoking were excluded from the study. Healthy controls were selected from those were referred to the health centers of Tabriz University of Medical Sciences. The Mini-International Neuropsychiatric Interview was used for the assessment of healthy controls. Exclusion criteria were the existence of any systemic disorder, psychiatric condition, pregnancy, or a history of psychopathology in a first-degree biological relative. The study protocol was approved by the Shahroud University of Medical Sciences Ethical Committee (IR.SHMU.REC.1398.113). Informed consent forms were signed by all participants/or their guardians.

#### **RNA extraction and cDNA synthesis**

In the first step, 5 ml of the peripheral blood was gathered in K2-EDTA-containing tubes. The RNA of all samples was extracted by using the Hybrid-R blood RNA extraction Kit (Gene All, Seoul, Korea). The Quality of the extracted RNAs was verified by 1% agarose gel electrophoresis, and to eliminate any genomic DNA contamination, samples were treated with DNase I (Thermo scientific, Deutschland, Germany). The quantity of RNA was appraised by Nanodrop equipment (Thermo Scientific, MA, USA). The extracted RNA was subsequently converted to cDNA by using the High-Capacity cDNA Reverse Transcription FIRE Script RT cDNA Synthesis Kit (Solis Bio Dyne, Estonia).

#### **Primer design**

Primer design was accomplished using the NCBI Primer designing tool and verified through blasting in the nucleotide BLAST database to confirm specific binding to target sequences. The primer sequences are detailed in Table [1.](#page-2-0)

#### **qRT-PCR**

Expression levels of genes were assessed in all participants using the RealQ Plus 2 × PCR Master Mix Green with high ROX (Amplicon, Odense, Denmark). Cycling reactions were carried out in Step One Plus Real-Time PCR equipment (Applied Biosystems, Foster City, CA, USA). After evaluation of Ct values of *UBC* and *YWHAZ*, we used *UBC* gene as the reference gene based on its constant expression in the peripheral blood of schizophrenia patients. The stable expression of *UBC* was also confirmed by NormFinder software ([https://www.moma.dk/so](https://www.moma.dk/software/normfinder) [ftware/normfinder\)](https://www.moma.dk/software/normfinder).

#### *Statistical analysis*

Relative expression levels of genes were measured in all samples. The Ln [Efficiency^ΔΔCT] method considers the transcript levels of *UBC* as normalizer. The Shapiro-Wilk test was performed to evaluate the normality of the data. One-way ANOVA and Bonferroni's multiple comparisons test were used for comparison of expression data between study groups. Correlations between expressions of genes were valued by calculation of Spearman correlation coefficients. Data was analyzed using the GraphPad Prism 8.0.0 software. The diagnostic power of the transcript levels of genes was measured by depicting receiver operating characteristic (ROC) curves.

<span id="page-2-0"></span>

**Table 1**. Primers sequences.

#### **Results General data of patients and controls**

A total of 50 medicated patients, 25 treatment-naïve patients, and 50 healthy controls were recruited for the current case-control study. No significant difference was found between the age and sex ratios of cases and controls. Demographic data of the participants in the study are summarized in Table [2.](#page-3-0)

#### **Expression assays**

Relative expression of genes was compared between medicated patients, treatment-naïve patients, and healthy controls. Expression level of *IFNG-AS1-001* was significantly downregulated in medicated-patient (*P*<0.05); while expression levels of *IFNG*, *IL18RAP*, and *AC007278.2* were significantly upregulated in medicated patients compared to controls (*P*<0.05). Expression levels of *IFNG-AS1-003*, *AC007278.3* and *IL18R1* were not significantly different between this group of patients and healthy controls (Fig. [1\)](#page-4-0).

Expression levels of *IFNG*, *IFNG-AS1-001*, *IL18R1*, *IL18RAP*, *AC007278.2*, and *AC007278.3* were significantly upregulated in treatment-naïve patients compared to controls (*P*<0.0001). On the other hand, expression level of *IFNG-AS1-003* was significantly downregulated in treatment-naïve patients compared to controls (*P*<0.0001) (Fig. [1\)](#page-4-0).

#### **ROC curve analysis**

In the present study, we evaluated the diagnostic power of transcript quantities of *IFNG*, *IL18R1*, *IL18RAP*, *IFNG-AS1-003*, *AC007278.2*, and *AC007278*.3 in identifying between the treatment-naïve patients and controls by depicting ROC curve (Table [3\)](#page-5-0).

Based on the area under cover (AUC) value, *INFG-AS1-003*, *IFNG*, *IL18R1*, and *AC007278.3* had powerful diagnostic power (AUC=0.90, 0.9, 0.87, and 0.80, *P*<0.0001) (Fig. [2](#page-5-1)).

Additionally, we assessed the diagnostic power of all differentially expressed genes in *3* in identifying between treatment-naïve patients and controls (Fig. [3\)](#page-6-0). Transcript levels of these genes could separate these groups with AUC = 0.93, sensitivity = 0.96 and specifiicity = 0.73.

#### *Correlation matrix analysis*

The analysis of the Spearman correlation matrix among genes in all subjects allows us to examine the connections between genes, as evidenced by their correlation coefficients and corresponding p-values. Significant positive and negative correlations between genes were observed in all three groups of medicated patients, treatmentnaïve patients, and healthy controls, which indicated their expression relationship in the disease (Figs. [4,](#page-6-1) [5](#page-7-0) and [6](#page-7-1)).

Finally, we assessed correlation between expression of genes and clinicopathological scores (Tables [4](#page-8-0) and [5](#page-8-1)). In the medicated patients, we found positive correlations between expression of *IL18RAP* and BPRS, expression of *IFNG-AS1-001* and both PANNS and negative symptoms score, and expression of *AC007278.2* and PANNS. Moreover, inverse correlations were detected between CRP levels and expression levels of both *IFNG-AS1-001* and *AC007278.3*.

Among treatment-naïve patients, expression of *IFNG-AS1-001* was inversely correlated with BPRS and positive symptoms.

<span id="page-3-0"></span>

**Table 2**. Demographic data of patients and controls.

#### **Discussion**

After intensive research, it is now clear that a gene or protein cannot explain such a complex disease as schizophrenia. Schizophrenia is a multifactorial disease in which a large number of genes and different cellular signaling pathways are involved in its initiation and development. One of the important pathways involved in the development of schizophrenia is the inflammatory pathwa[y26](#page-11-17). Cytokines have a critical role in initiating and maintaining immune responses. They can easily cross the blood-brain barrier and act as a major mediator between the brain and the immune system<sup>27</sup>. Many studies showed that there is a significant alteration in the levels of inflammatory cytokines in the blood of schizophrenia patients compared to healthy control[s28](#page-11-19).

The aim of this study was to evaluate the expression level of *IFNG*, *IL18R1*, *IL18RAP*, and *IFNG-AS1-001*, *IFNG-AS1-003*, *AC007278.2*, and *AC007278.3* in treatment-naïve schizophrenic patients who were in the acute

<span id="page-4-0"></span>

Control Medicated Treatment-naive patient patients

**Figure 1**. Expressions levels of genes in treatment-naïve and medicated patients compared to healthy controls. Expression levels of genes were calculated using the (Efficiency^-ΔΔCT) method. ns: not significant, *P*<0.05(\*), *P*<0.01(\*\*), *P*<0.001(\*\*\*), *P*≤0.0001(\*\*\*\*).

**Scientific Reports** | (2024) 14:27654 | https://doi.org/10.1038/s41598-024-78220-w 5

 $\bf{0}$ 

<span id="page-5-0"></span>

**Table 3**. Result of ROC curve analysis between treatment-naïve patients and controls.

<span id="page-5-1"></span>

**Figure 2**. The diagnostic power of transcript quantities of *IFNG*, *IL18R1*, *IFNG-AS1-003*, and *AC007278.3* in identifying between treatment-naïve patients and controls.

phase of the disease as well as patients who were in the remission phase of the disease (medicated) compared to matched controls.

Interleukin 18 (IL-18) is one of the multifunctional cytokines that is structurally similar to the IL-1 family and is one of the main factors inducing the secretion of IFNG from T-helper cells<sup>29</sup>. IL-18 has a role in many psychiatric disorders. IL-18 signaling has been found to be interrupted in the central amygdala in an animal psychiatric disorders. IL-10 signaling has been found to be measured in the modulation of model of post-traumatic stress and alcohol use disorder<sup>[30](#page-11-21)</sup>. This cytokine has a possible role in the modulation of the hypothalamic–pituitary–adrenal axis and might mediate the CNS dependent impacts on the susceptibility to related disorders<sup>31</sup>. Moreover, in many neurodegenerative and inflammatory diseases, the expression of IL-18 receptor (IL18R) and also the expression of *IL18*were increased[32.](#page-11-23) The IL18R has two subunits, including IL18R1 (IL18α) and IL18RAP or (IL18ß)[24](#page-11-15). Two functionally related lncRNAs, namely *AC007278.2* and

<span id="page-6-0"></span>

**Figure 3**. The diagnostic power of combination of transcript quantities of *IFNG*, *IL18R1*, *IFNG-AS1-003*, and *AC007278.3* in identifying between treatment-naïve patients and controls.

<span id="page-6-1"></span>

**Figure 4**. Spearman correlation between genes in treatment-naive patients.

*AC007278.3* are located on chromosome 2 inside the introns of *IL18R1* and *IL18RAP*, respectively. IL-18 is secreted by macrophage-like cells and plays a pivotal role in the response of helper  $T$  cells<sup>33</sup>. IL18R is widely present in neurons. Thus, the IL-18 that is synthesized in the CNS, can affect neuronal growth, differentiation, and apoptosis<sup>34</sup>. This interleukin exerts its inflammatory role by increasing IFNG production from T cells and NK cells. Expression level of *IL18*in the blood of patients with schizophrenia has been shown to be higher than control[s23](#page-11-14). This finding has also been confirmed by Szabo et al., the serum level of IL18 significantly increased in schizophrenia patients compared to  $HCs^{35}$  $HCs^{35}$  $HCs^{35}$ .

Luo et al. have shown that the concentration of IL-18 in the serum of patients is not significantly altered between treatment-naïve and medicated patients, and the concentration level of IL-18 is increased in both groups compared to healthy controls<sup>36</sup>. Thus, one can infer that antipsychotic treatment does not change the concentration of IL-18. This finding was also verified in our study.

<span id="page-7-0"></span>

**Figure 5**. Spearman correlation between genes in medicated patients.

<span id="page-7-1"></span>

**Figure 6**. Spearman correlation between genes in healthy controls.

In the present study, we also evaluated the expression level of IL-18 receptor subunits, namely *IL18R1* and *IL18RAP*. The findings from the earlier research indicate that the binding of IL-18 to IL18R1 induces a sequence of processes that activates multiple signaling pathways, such as NF-kB and MAPK pathways, resulting in the generation of inflammatory cytokines and chemokines<sup>37</sup>. Therefore, based on previous studies, there is a direct relationship between the level of IL-18 and IL18R, IL18RAP. In the present study, we showed that the expression levels of *IL18R1* and *IL18RAP* were higher in both treatment-naive and medicated groups compared to healthy controls. We did not observe any significant expression changes between medicated and treatment-naive patients, which indicates that the use of antipsychotic drugs does not have a significant effect on the expression of IL-18 receptor subunits. In the ROC curve analysis of IL18R1, the sensitivity of 84% and the specificity of 90% indicated that it can be used as a suitable diagnostic biomarker to identify treatment-naïve patients from healthy with a expression cutoff of 8.4, which is statistically significant.

Another subunit of the IL18 receptor is IL18RAP. The expression of *IL18RAP* was significantly increased in treatment-naïve and medicated patients.

In line with our results, Xu et al. have analyzed GWAS and replicated the results in an independent cohort of schizophrenia patients. They showed association signals within *IL18R1* and *IL18RAP* genes, with the most significant marker being *IL18R1*rs1035130. They have also reported altered levels of IL-18 and IL18R1 in schizophrenia patients compared with controls<sup>38</sup>.

We suggest that by increasing the expression of IL-18 receptor subunits, the expression of IL-18 also increases, which causes an increase in the amount of IFNG. IL-18 and IFNG are closely related cytokines that play important roles in the immune response. IL-18 is known to enhance the production of IFNG, and both

<span id="page-8-0"></span>

**Table 4**. Spearman correlation matrix analysis between genes and schizophrenia assessments criteria in the medicated patients.

<span id="page-8-1"></span>

**Table 5**. Spearman correlation matrix analysis between genes and schizophrenia assessments criteria in treatment-naive patients.

cytokines can synergistically amplify immune responses<sup>39</sup>. In Spearman's correlation matrix analysis, there was no significant relationship between the expression of *IL18R1* and *IL18RAP* in any of study subgroups.

We found positive correlation between expression of *IL18RAP* and BPRS in the medicated patients. In a previous study, elevation of serum concentration of IL-18 and a certain polymorphisms within *IL-18*gene have been reported to be positively associated with the PANSS general psychopathology subscore and the PANSS total scor[e40](#page-11-31). However, there was no data about correlation between expression of IL-18 receptor subunits and mentioned scores.

IFNG is a soluble cytokine of the type II class of IFNs<sup>41</sup>. It is a key activator of macrophages and inducer of MHC ΙΙ molecule expression[42.](#page-11-33) Abnormal expression of *IFNG*has been associated with numerous autoinflammatory and autoimmune diseases<sup>43</sup>. This cytokine is synthesized by T helper cells (particularly, Th1 cells), cytotoxic T cells, macrophages, mucosal epithelial cells, and NK cells. Moreover, it acts as a crucial autocrine signal for professional antigen-presenting cells (APCs) in early innate immune responses and a key paracrine signal in the adaptive immune response. Notably, expression of this molecule is induced by a number of cytokines, including  $IL-18$  and type I IFN<sup>44</sup>. It is a cytokine of T helper  $1^{45}$  and regulates the presentation of antigens and the division and differentiation of lymphocytes. Meta-analyses of cytokine changes in schizophrenia have shown that IFNG can be a marker of disease diagnosis. The IFNG secretion is controlled by APCs, IL12 and IL1[846](#page-11-37). When the IFNG ligand binds to the receptor, the receptor is dimerized and causes JAK1 and JAK2 to come close to each other, leading to the phosphorylation and activation of STAT1. Activated STAT1 causes activation of STAT4. Activated STAT4 goes to the nucleus and increases the expression of the *IFNG*gene[47](#page-11-38). Figure [7](#page-9-0) shows the relationships between IL-18, IFNG, JAK1, JAK2, STAT1, and STAT4.

<span id="page-9-0"></span>

**Figure 7**. The relationships between IL18, IFNG, JAK1, JAK2, STAT1, and STAT4. When the IFNG ligand binds to the receptor, the receptor is dimerized. Then, JAK1 and JAK2 become close to each other. This leads to the phosphorylation and activation of STAT1. Activated STAT1 causes activation of STAT4. This cascade of events leads to up-regulation of IFNG.

*IFNG*has been shown to be significantly decreased in the medicated schizophrenia patients compared to healthy controls, while its expression has been up-regulated in the treatment-naïve patients<sup>48</sup>. Another study has shown that the expression level of this gene is higher in schizophrenia patients compared with controls<sup>[49](#page-11-40)</sup>. In the medicated patients, we found positive correlations between expression of *IFNG-AS1-001* and both PANNS and negative symptoms score, and expression of *AC007278.2* and PANNS. Moreover, inverse correlations were detected between CRP levels and expression levels of both *IFNG-AS1-001* and *AC007278.3*. However, among treatment-naïve patients, expression of *IFNG-AS1-001* was inversely correlated with BPRS and positive symptoms. Thus, it can be inferred that medication affects correlation between mentioned genes and clinicopathological scores.

IL-18 and IFNG, two closely related cytokines, have a significant impact on the immune response as they often interact with each other. Both IL-18 and IFNG are classified as pro-inflammatory cytokines and play crucial roles in regulating immune responses, particularly in the context of defending the host against infections and managing the inflammatory response. One of the key aspects of their interaction is that IL-18 can stimulate the production of IFNG from various immune cells, such as T cells and NK cells. In response, IFNG can further enhance the expression of IL-18 receptors on immune cells. This creates a positive feedback loop within the immune response, where IL-18 and IFNG mutually reinforce each other's effects. This interaction between IL-18 and IFNG serves to amplify the inflammatory response, leading to a more robust immune defense against pathogens. By working together, IL-18 and IFNG contribute to the coordination and regulation of immune responses, ultimately aiding in the protection of the host organism<sup>[50](#page-11-41)</sup>. Based on previous studies, the elevated *IL18R1* and *IL18RAP* expression can positively up-regulate the expression level of *IFNG*. In the present study, we showed that the expression level of *IFNG* in treatment-naïve patients significantly increased, and on the other hand, the level of *IFNG* expression in medicated patients compared decreased compared to controls. Based on the ROC curve analysis, *IFNG* had the highest AUC among the genes, at 0.94, and sensitivity of 0.8, specificity of 0.9 which significantly indicate its diagnostic ability  $(P < 0.0001)$ .

*IFNG-AS1*, which is an intergenic lncRNAs, is located on the same chromosome as the *IFNG* gene, and is known as *Tmevpg1*; *NEST<sup>[51](#page-11-42)</sup>*. *IFNG-AS1*has a number of variants<sup>52</sup>, and in the present study, we investigated the expression level of two variants, including *IFNG-AS-001* and *IFNG-AS-003*. The expression level of the IFNG-AS1-001 variant decreased twofold in the medicated patients compared with controls, while its expression level was higher in the treatment-naïve patients. Most notably, while the expression level of the *IFNG-AS-003* gene was similar between the medicated patients and controls, its expression was significantly lower in the treatmentnaïve patients compared to controls.

Expression level of the *IFNG-AS1-001* gene was decreased in the medicated patients compared with controls. *IFNG-AS1-001* contributes to the methylation of the *IFNG*locus and decreases its expression by binding to WDR5, a scaffolding protein in the H3K4 methyltransferase complex<sup>21</sup>. Thus, down-regulation of *IFNG-AS1-001* is expected to result in the up-regulation of IFNG. *IFNG-AS1-003* gene is also located on the same chromosome as the *IFNG*[22.](#page-11-13) The observed downregulation of the *IFNG-AS1-003*variant in the current study is consistent with the previous studies that were conducted on other diseases such as Hashimoto's disease[53](#page-11-44). In fact, *AC007278.2* and *IFNG-AS1-001* have been shown to be up-regulated in the relapsing phase in multiple sclerosis patients, while *IFNG-AS1-003has* been up-regulated in the remitting phase compared with relapsing phase<sup>52</sup>.

Expression levels of *AC007278.2* and *AC007278.3* were higher in the treatment-naïve patient compared to the controls. These lncRNAs are directly related with the increase of IL18R1 and IL18RAP genes. These genes are located on the positive strand of chromosome 2q12, and inside the *IL18R1*gene intron and play an important role in the expression and differentiation of T helper1 cells<sup>54</sup>. It has been disclosed that the expression level of *AC007278.2* and *AC007278.3* lncRNAs is aligned with the *IL18R1* and *IL18RAP* gene expression. Upregulation of the mentioned lncRNAs has been correlated with over-expression of *IL18R1* and *IL18RAP* genes<sup>52</sup>. The increase in the expression of *IL18R1* and *IL18RA*genes causes the activation of STAT4 and IL-2, as well as activation of histone acetyltransferase and DNA methylase complex. Finally, IL18R1 and IL18RAP cause the differentiation of T helper1 cells, and this differentiation causes the production of pro-inflammatory cytokines from T helper1 cells, which is one of the main causes of autoimmune diseases<sup>55</sup>. Therefore, it is likely that the expression level of *AC007278.2* and *AC007278.3* is directly related to the level of expression of genes coding for the IL-18 receptor protein, and the use of antipsychotic drugs may reduce their levels. Based on the ROC curve analysis, *AC007278.3* had the sensitivity of 84% and the specificity of 82% with AUC=80%, thus it can be used as a suitable diagnostic biomarker to identify treatment-naïve patients from controls with an expression cutoff of 4.66.

Finally, assessment of pairwise correlation in the patients group showed that there was a significant direct relationship between genes. In both medicated and treatment-naïve patients, *IFNG-AS1-001* showed significantly strong positive correlations with *AC007278.2*, *AC007278.3*, and *IL18RAP*. Moreover, *AC007278.2* had a significant correlation with *IFNG-AS1-001*, *AC007278.3* and *IL18RAP* in both groups. However, the correlations between *AC007278.2*/*IFNG* and *AC007278.3*/ *IFNG* were only significant in the treatment-naïve patients. On the other hand, the correlation between and *IFNG* and *IFNG-AS1-003* was only significant among medicated patients. Thus, it can be inferred from the correlation data that medication affects the correlation between *IFNG* and other genes.

#### **Conclusion**

In summary, this study showed changes in the expression of a number of lncRNA genes and related proteincoding genes in the blood of patients with schizophrenia. Notably, *IFNG*, *IFNG-AS1-003*, *IL18R1*, and *AC007278.3* genes were found to be suitable biomarkers in this disorder. In order to confirm the biomarker power of genes and gene expression changes in the acute phase and remission of the disease, more studies in a larger statistical population are suggested. Our study had some limitations including lack of assessment of expression of mRNA coding genes at protein level and lack of functional studies.

#### **Data availability**

The datasets generated and/or analysed during the current study are available in the NCBI repository: IFNG (NC\_000012.12), IL18R (NC\_058098.1), IL18RAP (NC\_000002.12), IFNG-AS1(NC\_000012.12).

Received: 10 July 2024; Accepted: 29 October 2024 Published online: 12 November 2024

#### **References**

- <span id="page-10-0"></span>1. McCutcheon, R. A., Marques, T. R. & Howes, O. D. Schizophrenia—an overview. *JAMA Psychiatry*. **77** (2), 201–210 (2020).
- <span id="page-10-1"></span>2. Correll, C. U. & Schooler, N. R. Negative symptoms in Schizophrenia: a review and clinical guide for Recognition, Assessment, and treatment. *Neuropsychiatr Dis. Treat.* **16**, 519–534 (2020). PubMed PMID: 32110026. Pubmed Central PMCID: PMC7041437. Epub 2020/02/29. eng.
- <span id="page-10-2"></span>3. Kahn, R. S. On the origins of schizophrenia. *Am. J. Psychiatry*. **177** (4), 291–297 (2020).
- <span id="page-10-3"></span>4. Ermakov, E. A., Melamud, M. M., Buneva, V. N. & Ivanova, S. A. Immune System abnormalities in Schizophrenia: an integrative view and translational perspectives. *Front. Psychiatry.* **13***,* 880568 (2022). PubMed PMID: 35546942. Pubmed Central PMCID: PMC9082498. Epub 2022/05/14. eng.
- <span id="page-10-4"></span>5. Müller, N. & Schwarz, M. J. Immune System and Schizophrenia. *Curr. Immunol. Rev.* **6** (3), 213–220 (2010). PubMed PMID: 21057585. Pubmed Central PMCID: PMC2971548. Epub 2010/11/09. eng.
- <span id="page-10-5"></span>6. Pouget, J. G. The emerging immunogenetic architecture of schizophrenia. *Schizophr. Bull.* **44** (5), 993–1004 (2018).
- <span id="page-10-6"></span>7. Ermakov, E. A., Melamud, M. M., Buneva, V. N. & Ivanova, S. A. Immune system abnormalities in schizophrenia: an integrative view and translational perspectives. *Front. Psychiatry*. **13**, 880568 (2022).
- <span id="page-10-7"></span>8. Müller, N., Weidinger, E., Leitner, B. & Schwarz, M. J. The role of inflammation in schizophrenia. *Front. Neurosci.* **9**, 372 (2015). PubMed PMID: 26539073. Pubmed Central PMCID: PMC4612505. Epub 2015/11/06. eng.
- <span id="page-11-0"></span>9. Vallée, A. Neuroinflammation in Schizophrenia: the key role of the WNT/β-Catenin pathway. *Int. J. Mol. Sci.* **23** (5). [https://doi.or](https://doi.org/10.3390/ijms23052810) [g/10.3390/ijms23052810](https://doi.org/10.3390/ijms23052810) (2022). PubMed PMID: 35269952. Pubmed Central PMCID: PMC8910888. Epub 2022/03/11. eng.
- <span id="page-11-1"></span>10. Birnbaum, R. & Weinberger, D. R. A genetics perspective on the role of the (neuro) immune system in schizophrenia. *Schizophr. Res.* **217**, 105–113 (2020).
- <span id="page-11-2"></span>11. Bridges, M. C., Daulagala, A. C. & Kourtidis, A. LNCcation: lncRNA localization and function. *J. Cell Biol.* **220** (2), e202009045 (2021).
- <span id="page-11-3"></span>12. Peltier, D. C., Roberts, A. & Reddy, P. LNCing RNA to immunity. *Trends Immunol.* **43** (6), 478–495 (2022).
- <span id="page-11-4"></span>13. Jiang, M-C., Ni, J-J., Cui, W-Y., Wang, B-Y. & Zhuo, W. Emerging roles of lncRNA in cancer and therapeutic opportunities. *Am. J. cancer Res.* **9** (7), 1354 (2019).
- <span id="page-11-5"></span>14. Ma, N., Tie, C., Yu, B., Zhang, W. & Wan, J. Identifying lncRNA–miRNA–mRNA networks to investigate Alzheimer's disease pathogenesis and therapy strategy. *Aging (Albany NY)*. **12** (3), 2897 (2020).
- <span id="page-11-6"></span>15. Cai, L-J. et al. LncRNA MALAT1 facilitates inflammasome activation via epigenetic suppression of Nrf2 in Parkinson's disease. *Mol. Brain*. **13** (1), 1–15 (2020).
- <span id="page-11-7"></span>16. Cui, X. et al. Long non-coding RNA: potential diagnostic and therapeutic biomarker for major depressive disorder. *Med. Sci. Monitor: Int. Med. J. Experimental Clin. Res.* **22**, 5240 (2016).
- <span id="page-11-8"></span>17. Parikshak, N. N. et al. Genome-wide changes in lncRNA, splicing, and regional gene expression patterns in autism. *Nature*. **540** (7633), 423–427 (2016).
- <span id="page-11-9"></span>18. Teimuri, S. et al. Integrative analysis of lncRNAs in Th17 cell lineage to discover new potential biomarkers and therapeutic targets in autoimmune diseases. *Mol. Therapy-Nucleic Acids*. **12**, 393–404 (2018).
- <span id="page-11-10"></span>19. Arolt, V. et al. Production of interferon-gamma in families with multiple occurrence of schizophrenia. *Psychiatry Res.* **66** (2–3), 145–152 (1997). PubMed PMID: 9075278. Epub 1997/02/07. eng.
- <span id="page-11-11"></span>20. Paul-Samojedny, M. et al. Association Study of Interferon Gamma (IFN-γ)+874T/A gene polymorphism in patients with paranoid Schizophrenia. *J. Mol. Neurosci.* **43**(3), 309–315 (2011).
- <span id="page-11-12"></span>21. Collier, S. P., Henderson, M. A., Tossberg, J. T. & Aune, T. M. Regulation of the Th1 genomic locus from Ifng through Tmevpg1 by T-bet. *J. Immunol.* **193** (8), 3959–3965 (2014).
- <span id="page-11-13"></span>22. Peng, H. et al. Elevated expression of the long noncoding RNA IFNG-AS1 in the peripheral blood from patients with rheumatoid arthritis. *J. Immunol. Res.* **2020**, 1–8 (2020).
- <span id="page-11-14"></span>23. Syed, A. A. S., He, L., Shi, Y. & Mahmood, S. Elevated levels of IL-18 associated with schizophrenia and first episode psychosis: a systematic review and meta‐analysis. *Early. Interv. Psychiat.* **15** (4), 896–905 (2021).
- <span id="page-11-15"></span>24. Kaplanski, G. Interleukin-18: Biological properties and role in disease pathogenesis. *Immunol. Rev.* **281** (1), 138–153 (2018).
- <span id="page-11-16"></span>25. American Psychiatric Association D & American Psychiatric Association, D. *Diagnostic and Statistical Manual of Mental Disorders: DSM-5* (American psychiatric association Washington, DC, 2013).
- <span id="page-11-17"></span>26. Müller, N. Inflammation in schizophrenia: pathogenetic aspects and therapeutic considerations. *Schizophr. Bull.* **44** (5), 973–982 (2018).
- <span id="page-11-18"></span>27. Upthegrove, R., Khandaker, G.M. Cytokines, Oxidative Stress and Cellular Markers of Inflammation in Schizophrenia. In: Khandaker, G., Meyer, U., Jones, P. (eds) *Neuroinflammation and Schizophrenia*. Current Topics in Behavioral Neurosciences, vol 44. Springer, Cham. [https://doi.org/10.1007/7854\\_2018\\_88](https://doi.org/10.1007/7854_2018_88) (2019)
- <span id="page-11-19"></span>28. Lee, E. E., Hong, S., Martin, A. S., Eyler, L. T. & Jeste, D. V. Inflammation in schizophrenia: cytokine levels and their relationships to demographic and clinical variables. *Am. J. Geriatric Psychiatry*. **25** (1), 50–61 (2017).
- <span id="page-11-20"></span>29. Yasuda, K., Nakanishi, K. & Tsutsui, H. Interleukin-18 in health and disease. *Int. J. Mol. Sci.* **20** (3), 649 (2019).
- <span id="page-11-21"></span>30. Borgonetti, V. et al. IL-18 signaling in the Rat Central Amygdala is disrupted in a Comorbid Model of post-traumatic stress and Alcohol Use Disorder. *Cells.* **12** (15). <https://doi.org/10.3390/cells12151943>(2023). PubMed PMID: 37566022. Pubmed Central PMCID: PMC10416956. Epub 2023/08/11. eng.
- <span id="page-11-22"></span>31. Sugama, S. & Conti, B. Interleukin-18 and stress. *Brain Res. Rev.* **58**(1), 85–95 (2008).
- <span id="page-11-23"></span>32. Piancone, F., La Rosa, F., Marventano, I., Saresella, M. & Clerici, M. The role of the inflammasome in neurodegenerative diseases. *Molecules*. **26** (4), 953 (2021).
- <span id="page-11-24"></span>33. Fang, D. et al. Differential regulation of transcription factor T-bet induction during NK cell development and T helper-1 cell differentiation. *Immunity*. **55** (4), 639–655 (2022). e7.
- <span id="page-11-25"></span>34. Wu, D. et al. Interleukin-18 from neurons and microglia mediates depressive behaviors in mice with post-stroke depression. *Brain. Behav. Immun.* **88**, 411–420 (2020).
- <span id="page-11-26"></span>35. Szabo, A. et al. Increased circulating IL-18 levels in severe mental disorders indicate systemic inflammasome activation. *Brain. Behav. Immun.* **99**, 299–306 (2022).
- <span id="page-11-27"></span>36. Luo, Y., He, H., Zhang, J., Ou, Y. & Fan, N. Changes in serum TNF-α, IL-18, and IL-6 concentrations in patients with chronic schizophrenia at admission and at discharge. *Compr. Psychiatr.* **90**, 82–87 (2019). PubMed PMID: 30782515. Epub 2019/02/21. eng.
- <span id="page-11-28"></span>37. Rex, D. et al. A comprehensive pathway map of IL-18-mediated signalling. *J. cell. Communication Signal.* **14** (2), 257–266 (2020).
- <span id="page-11-29"></span>38. Xu, Y. et al. Potential involvement of the interleukin-18 pathway in schizophrenia. *J. Psychiatr. Res.* **74**, 10–6 (2016).
- <span id="page-11-30"></span>39. Meda Spaccamela, V. et al. High levels of IL-18 and IFN-γ in chronically inflamed tissue in chronic granulomatous disease. *Front. Immunol.* **10**, 2236 (2019).
- <span id="page-11-31"></span>40. Zhang, X. Y. et al. Serum IL-18 level, clinical symptoms and IL-18-607A/C polymorphism among chronic patients with schizophrenia in a Chinese Han population. *Psychoneuroendocrinology* **68**, 140–147 (2016).
- <span id="page-11-32"></span>41. Burke, J. D. & Young, H. A. (eds) *IFN-γ: A Cytokine at the Right time, is in the Right Place. Seminars in Immunology* (Elsevier, 2019).
- <span id="page-11-33"></span>42. Axelrod, M. L., Cook, R. S., Johnson, D. B. & Balko, J. M. Biological consequences of MHC-II expression by tumor cells in cancer. *Clin. Cancer Res.* **25** (8), 2392–2402 (2019).
- <span id="page-11-34"></span>43. Matta, B., Song, S., Li, D. & Barnes, B. J. Interferon regulatory factor signaling in autoimmune disease. *Cytokine*. **98**, 15–26 (2017).
- <span id="page-11-36"></span><span id="page-11-35"></span>44. Mirlekar, B. & Pylayeva-Gupta, Y. IL-12 family cytokines in cancer and immunotherapy. *Cancers*. **13** (2), 167 (2021). 45. Paibomesai, M. A. *Epigenetic Influences on Bovine T-helper 1 and T-helper 2 Cytokines (interferon-gamma and Interleukin-4) in high*
- *and low Immune Responders around the* (University of Guelph, 2017).
- <span id="page-11-37"></span>46. Zhu, Q. & Kanneganti, T-D. Cutting edge: distinct regulatory mechanisms control proinflammatory cytokines IL-18 and IL-1β. *J. Immunol.* **198** (11), 4210–4215 (2017).
- <span id="page-11-38"></span>47. Philips, R. L. et al. The JAK-STAT pathway at 30: much learned, much more to do. *Cell*. **185** (21), 3857–3876 (2022).
- <span id="page-11-39"></span>48. Ghafelehbashi, H., Pahlevan Kakhki, M., Kular, L., Moghbelinejad, S. & Ghafelehbashi, S. Decreased expression of IFNG-AS 1, IFNG and IL‐1B inflammatory genes in Medicated Schizophrenia and bipolar patients. *Scand. J. Immunol.* **86** (6), 479–485 (2017).
- <span id="page-11-40"></span>49. Lorestani, R., Boozarpour, S., Alijanpour, S. & Ahangar, L. Evaluation of IFN-γ and T-bet expression levels as possible molecular markers of Schizophrenia. *J. Cell. Mol. Res.* **11** (2), 82–89 (2020).
- <span id="page-11-41"></span>50. Nakanishi, K. Unique action of interleukin-18 on T cells and other immune cells. *Front. Immunol.* **9**, 358681 (2018).
- <span id="page-11-42"></span>51. Lodde, V. et al. Long noncoding RNAs and circular RNAs in autoimmune diseases. *Biomolecules*. **10** (7), 1044 (2020).
- <span id="page-11-43"></span>52. Hosseini, A. et al. LncRNAs associated with multiple sclerosis expressed in the Th1 cell lineage. *J. Cell. Physiol.* **234** (12), 22153– 22162 (2019).
- <span id="page-11-44"></span>53. Peng, H. et al. The long noncoding RNA IFNG-AS1 promotes T helper type 1 cells response in patients with Hashimoto's thyroiditis. *Sci. Rep.* **5** (1), 17702 (2015).
- <span id="page-12-0"></span>54. You, Y. et al. Integrated transcriptome profiling revealed that elevated long non-coding RNA-AC007278. 2 expression repressed CCR7 transcription in systemic lupus erythematosus. *Front. Immunol.* **12**, 615859 (2021).
- <span id="page-12-1"></span>55. Hedl, M., Zheng, S. & Abraham, C. The IL18RAP region disease polymorphism decreases IL-18RAP/IL-18R1/IL-1R1 expression and signaling through innate receptor–initiated pathways. *J. Immunol.* **192**(12), 5924–5932 (2014).

#### **Acknowledgements**

This study was founded by the Shahroud University of Medical Science, Iran. The present study was supported by Shahroud University of medical sciences as a MSc Thesis. We hereby acknowledge the research deputy for grant No 9870

#### **Research involving human participants and/or animals**

The study was carried out according to the principles of the 1964 Declaration of Helsinki. Each enrolled subject provided a written informed consent, and all study data were obtained and elaborated in accordance with our institutional ethical committee regulations. The study protocol was approved by the Shahroud University of Medical Sciences Ethical Committee (IR.SHMU.REC.1398.113).

#### **Informed consent**

Informed consent was obtained from patients.

#### **Consent to participate**

Not applicable.

#### **Author contributions**

Kamran Javidi and Shima Rahmani performed the majority of experiments and data analysis. Faezeh Mehdizadeh, Fatemeh Manafzadeh, Seyed Gholamreza Noor Azar and Shahrokh Aghayan contributed to the experiments and interpreted the results. Faezeh Mehdizadeh wrote the manuscript and helped with the experiments. Behzad Baradaran and Soudeh Ghafouri-Fard revised the manuscript critically for important intellectual content. Asghar Shayannia designed and conducted the project.

#### **Declarations**

#### **Consent for publication**

Not applicable.

#### **Competing interests**

The authors declare no competing interests.

#### **Additional information**

**Supplementary Information** The online version contains supplementary material available at [https://doi.org/1](https://doi.org/10.1038/s41598-024-78220-w) [0.1038/s41598-024-78220-w](https://doi.org/10.1038/s41598-024-78220-w).

**Correspondence** and requests for materials should be addressed to A.S. or S.G.-F.

**Reprints and permissions information** is available at www.nature.com/reprints.

**Publisher's note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

**Open Access** This article is licensed under a Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International License, which permits any non-commercial use, sharing, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if you modified the licensed material. You do not have permission under this licence to share adapted material derived from this article or parts of it. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit [http://creativecommo](http://creativecommons.org/licenses/by-nc-nd/4.0/) [ns.org/licenses/by-nc-nd/4.0/.](http://creativecommons.org/licenses/by-nc-nd/4.0/)

© The Author(s) 2024