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The histopathological correlates of lobar microbleeds in falsepositive cerebral amyloid angiopathy cases

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Abstract

Objective: A definite diagnosis of cerebral amyloid angiopathy (CAA), characterized by the accumulation of Amyloid- β in walls of cerebral small vessels, can only be obtained through pathological examination. A diagnosis of probable CAA during life relies on the presence of haemorrhagic markers, including lobar cerebral microbleeds (CMBs). The aim of this project was to study the histopathological correlates of lobar CMBs in false-positive CAA cases.

Methods: In three patients who met criteria for probable CAA during life, but showed no CAA upon neuropathological examination, lobar CMBs were counted on *ex vivo* 3T MRI and on *ex vivo* 7T MRI. Areas with lobar CMBs were next sampled and cut in serial sections, on which the CMBs were then identified.

Results: Collectively, there were 25 lobar CMBs on *in vivo* MRI and 22 on *ex vivo* 3T MRI of the analysed hemispheres. On *ex vivo* MRI we targeted 12 CMBs for sampling and definite histopathological correlates were retrieved for 9 of them of which seven were true CMBs. No CAA was found on any of the serial sections. The 'culprit vessels' associated with the true CMBs instead showed moderate to severe arteriolosclerosis. Furthermore, CMBs in false-positive CAA

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VP and SJvV contributed to the conception and design of the study; CAA, SJvV, MCZZ, and VP contributed to the acquisition and analysis of data; SJvV, JO, VP, MPF, AV, and SMG contributed to drafting the text or preparing the figures.

Potential conflicts of Interest

The authors declare that they have no conflict of interest.

cases tended to be located more often in the juxtacortical or subcortical white matter, than in the cortical ribbon.

Interpretation: These findings suggest that arteriolosclerosis can generate lobar CMBs and that more detailed investigations into the exact localization of CMBs with respect to the cortical ribbon could potentially aid the diagnosis of CAA during life.

Introduction

Cerebral amyloid angiopathy (CAA) is a condition pathologically defined by the accumulation of Amyloid- β (A β) in the walls of leptomeningeal and cortical vessels¹. Like many neurodegenerative pathologies, a definite diagnosis can only be made at autopsy. A diagnosis of probable CAA can be made during life using the Boston criteria with high sensitivity and specificity. These criteria rely on typical hemorrhagic neuroimaging markers of the disease, such as cortical superficial siderosis (cSS), lobar intracerebral hemorrhage (ICH), and lobar cerebral microbleeds (CMBs)². CMBs are defined as small round or ovoid hypointense foci, on T2*-weighted MRI sequences³. Patients >50 years of age, who present with clinical symptoms of CAA (i.e. symptomatic ICH, transient focal neurological episodes, subarachnoid hemorrhage, dementia/cognitive impairment) and with more than one strictly lobar CMB, meet criteria for a diagnosis of probable CAA during life². A small number of individuals with strictly lobar CMBs and a diagnosis of probable CAA during life, do not show significant A β -accumulation in the cerebral vessels upon standard neuropathological examination² (i.e. Vonsattel-grade 2 in full brain samples). The pathological substrate of lobar CMBs in these false-positive CAA cases is so far unknown. CAA can have a patchy distribution⁴ within the brain, so that it would be plausible to find it only in the vessels closely associated with the CMBs. CMBs have also been related to arteriolosclerosis in patients with hypertensive arteriopathy (HA)⁵. In these cases, the lesions are to be found mainly in deep brain regions, consistently with the thickening of the walls of the deep perforating vessels, which is associated with arteriolosclerosis⁶. However, arteriolosclerosis is not exclusively located in deep brain regions and often co-occurs with CAA⁷. CMBs are also a common finding in ageing⁸, Alzheimer's disease (AD)⁹, and other neurodegenerative diseases¹⁰, meaning that other pathologies could play a role.

A detailed analysis of such cases might give insight into factors (e.g. morphology or localization of the CMBs) that can aid discrimination during life. This is of particular clinical relevance, considering that both the presence (and degree) of CAA pathology¹¹ and the number of lobar CMBs have been related to risk of ICH^{12,13} and thus to the decision of administering anticoagulation.

The advent of MRI-guided targeted histopathological analysis in the past years^{14,15}, has handed the community better tools to find pathological correlates of neuroimaging markers, which would otherwise remain elusive in standard neuropathological examination. In the case of CMBs, these have been identified as accumulation of extravasated erythrocytes (when acute) or as gold-brown hemosiderin deposits (when older)¹⁵, often accompanied by iron positive macrophages.

In the present study we used *ex vivo* MRI-guided sampling and serial sectioning to unravel the underlying histopathology of lobar CMBs in individuals who fulfilled criteria for probable CAA during life but did not show the pathological signatures of CAA upon routine neuropathological examination at autopsy. Moreover, we explored whether the localization of strictly lobar CMBs and positron emission topography (PET) imaging during life could discriminate between definite CAA and false-positive CAA cases.

Materials and Methods

Study design and case characteristics

The individuals included in the study, were part of an ongoing brain autopsy program that aims to evaluate the histopathology underlying MRI markers of CAA. The program was initiated in 2015 within the stroke research center at MGH and includes brain donations obtained from the MGH and outside hospitals¹⁶. Study approval was received from the MGH institutional review board (protocol number: 2021P001920), and informed consent was obtained from the next of kin or legal representative prior to autopsy according to the Declaration of Helsinki. Patients included in the study fulfilled the modified Boston criteria for probable CAA; they were > 55 years old at time of death, had multiple lobar CMBs or one lobar CMB and at least one area of cSS¹⁷. As per the clinical diagnostic criteria, possible alternative causes for lobar CMBs, such as antecedent head trauma or ischemic stroke, tumor of the central nervous system, vascular malformation, and vasculitis were excluded. After autopsy, one hemisphere underwent routine neuropathological examination by a board-approved neuropathologist, while the other and most intact hemisphere was fixed in 10% formalin for several weeks and underwent ex vivo 3T MRI scanning (Supplementary Figure 1). Then, the hemispheres were cut in 1 cm-thick coronal slabs and standardized histological examination was performed.

For this specific study, individuals who met criteria for probable CAA during life but showed no evidence of CAA upon standard histopathological examination (false-positive CAA cases) of their brains at autopsy were selected. Furthermore, cases with definite-CAA were chosen as a comparison group.

CMBs were detected on the *ex vivo* 3T MRI scan of the whole hemisphere. Additionally, for each false-positive case, the corresponding coronal slabs with CMBs were identified, based on anatomical landmarks (Figure 1). The selected tissue slabs were scanned again at higher resolution, using 7T MRI, which enabled a better localization of the CMBs. Finally, samples were taken from the presumed locations of the CMBs, they were embedded in paraffin, and serial sectioning was performed.

In vivo 3T MRI

During life, scans were performed within the frame of a single-center longitudinal study, that included probable and possible CAA patients from a memory-clinic cohort¹⁸. All three false-positive cases underwent the same 3T MRI scan protocol (Magnetom Prisma-Fit, Siemens Healthineers, Erlangen, Germany), using a 32-channel head coil. The protocol included: a 3D T1-weighted sequence (repetition time [TR] 2300–2510 ms; echo time

[TE] 1.69–2.98 ms; resolution $1 \times 1 \times 1$ mm), a 3D fluid-attenuated inversion recovery sequence (FLAIR, TR 5000–6000 ms; TE 356–455 ms; resolution $1 \times 1 \times 1$ mm), a susceptibility-weighted imaging sequence (SWI, TR 27–30 ms; TE 20 ms; slice thickness, 1.4 mm; in-plane resolution 0.9×0.9 mm), a 3D diffusion weighted imaging sequence (DWI, 60–64 directions; TR 8000–8040 ms; TE 82–84 ms; resolution $2 \times 2 \times 2$ mm; b-value, 700 s/mm²), and a T2-weighted sequence (TR 7500 ms; TE 84 ms; slice thickness, 2 mm; in-plane resolution 0.5×0.5 mm).

Ex vivo 3T MRI

The details of the whole-hemisphere *ex vivo* 3T MRI scanning procedure have been previously described^{16,20,21}. Briefly, hemispheres were vacuum sealed in a plastic bag containing periodate-lysine-paraformaldehyde, after eliminating air bubbles. Then, they were placed in the 32-channel head coil of a whole-body 3T MRI scanner (MAGNETOM Trio, Siemens Healthineers, Erlangen, Germany) and scanned overnight. The protocol included several sequences, including a T2-weighted turbo-spin echo (TSE) sequence (voxel size $0.5 \times 0.5 \times 0.5$ mm; TE 61 ms; TR 1800 ms; flip angle 150°; total scan time ~ 3 hours), and a gradient-echo fast low angle shot (FLASH) sequence (voxel size $0.5 \times 0.5 \times 0.5 \times 0.5$ mm; TE-1 4.49 ms; TE-2 11.02 ms; TR 20 ms; flip angle 10° , 20° , 30° ; receiver bandwidth 160 Hz/Px; 2 averages; total scan time ~ 2 hours), which was used in this study (Supplementary Figure 1).

Ex vivo 7T MRI

Individual formalin-fixed coronal tissue slabs of the false-positive CAA cases were scanned again with 7T MRI, in order to better identify the CMBs (Figure 1). Scans were acquired overnight on a whole-body 7T MRI scanner (MAGNETOM, Siemens Healthineers, Erlangen, Germany), with a custom built 32-channel head coil. The protocol included spoiled gradient echo T2*-weighted (FLASH) (voxel size $0.2 \times 0.2 \times 0.2$ mm; TE 8.67 ms; TR 20 ms; flip angle 20°; receiver bandwidth 180 Hz/Px total scan time ~ 1 hour) and T2-weighted (TSE) (voxel size $0.5 \times 0.5 \times 0.5$ mm; TE 54 ms; TR 1000 ms; flip angle 90°; receiver bandwidth 401 Hz/Px, total scan time ~ 1 hour) acquisitions.

MRI analysis

On *in vivo* and *ex vivo* MRI, CMBs were defined as round/ovoid foci < 10 mm, that appear hypointense on T2*-weighted imaging, according to established criteria³. *Ex vivo* T2*-weighted imaging is susceptible to air bubbles, which remain trapped on the surface of the brain and in the sulci and can mimic CMBs. Therefore, the T2-weighted images were used to discriminate between actual CMBs and the artefacts. CMBs were visually assessed by one expert observer (V.P., 7 years of experience) and consensus was then achieved with another expert (S.J.v.V., 10 years of experience). Each CMB was identified, when possible, in all imaging modalities (*in vivo* and *ex vivo* 3T MRI, *ex vivo* 7T MRI), guided by anatomical landmarks (Table 2). Furthermore, a third observer blinded to clinical information and pathological diagnosis (M.C.Z.Z., a neuroradiologist with 9 years of experience) classified the lobar CMBs on the *ex vivo* 3T MRI of the three false-positive CAA cases and in the three true-positive CAA cases as either located strictly in the cortex, in the juxtacortical WM or in the subcortical WM.

PET Scans and analysis

PET scans were acquired on a Siemens/CTI (Knoxville, TN) ECAT HR+ (3-D mode, 63 image planes, 15.2-cm axial field of view, 5.6-mm transaxial resolution, 2.4-mm slice interval) at MGH. Immediately before the scan, 8.5 to 15 mCi of [11C]-Pittsburgh compound B was injected as a bolus followed by a 60- to 70-minute dynamic acquisition in 69 frames, as previously published²². The [11C]- PiB was prepared directly at MGH.

PET data were reconstructed, corrected for attenuation, and inspected to verify adequate count statistics and absence of significant head motion. Data processing was performed using FreeSurfer (version 6.0), as previously described²³. The images were aligned to T1-weighted MR images and measurements were made in regions of interest (ROIs) defined by the Desikan-Killian atlas²⁴. The specific binding of PiB was calculated using the Logan graphical method as distribution volume ratio (DVR), on data collected 40 to 60 minutes after injection, adopting the cerebellar cortex as a reference region²⁵. We chose to evaluate the DVRs of a composite ROI consisting of the frontal, lateral temporal/parietal, and retrosplenial (FLR) cortex, because previous studies demonstrated an increased PiB retention in these brain areas in both AD and CAA²⁶. To this aim, we adopted a cutoff score (FLR DVR 1.08), that had demonstrated good sensitivity to presence of Aβ^{23,27}.

Histopathology

Samples were taken from predefined standard brain areas of the frontal, parietal, temporal and occipital cortex and from the BG area in all cases, as previously described²⁸. All tissue blocks were placed in thin cassettes and embedded in paraffin. Six µm-thick sections were cut with a microtome and stained using hematoxylin & eosin (H&E), following standard histology protocols. Brightfield immunohistochemistry against A β (mouse, Agilent Technologies, Santa Clara, CA; 1:200, Cat# M0872) was performed on adjacent sections. Following a standardized immunohistochemistry protocol, sections were deparaffinized and rehydrated through xylene and graded ethanol series. Next, endogenous peroxidase was blocked with 3% H₂O₂ (20 min), followed by antigen retrieval using formic acid treatment (for A β , 5 min). Tissue was then blocked with normal horse or goat serum (1 hour) and incubated overnight with the primary antibody at 4°C. On the next day, the biotinylated mouse or rabbit secondary antibody was applied (1 hour), followed by a mixture of avidin (A) and biotinylated HRP (B) (Vectastain ABC kit, Vector laboratories, 30 min) and 3,3'-Diaminobenzidine (DAB, Vector laboratories). Negative controls were included by omitting the primary antibodies and showed no immunopositivity, while the presence of Aβ-plaques served as a positive control.

Further samples were taken from the three false-positive CAA cases, targeting several CMBs. These samples had the same dimensions as the ones described above to fit a tissue cassette. They were also embedded in paraffin and cut in serial sections at a microtome. Sections at the estimated lesion location were collected and each fifth section was stained with H&E and then matched with the *ex vivo* MRI scans to verify whether the targeted CMB (or another correlate for the hypointensity seen on MRI) was visible. If this was the case, immunohistochemistry was performed on directly adjacent sections for A β , following the protocol described above, and for smooth muscle actin (SMA) (mouse,

Agilent Technologies, Santa Clara, CA; 1:250, Cat# M0851), using heated citrate buffer (pH 6.0) for the antigen retrieval step. Moreover, adjacent sections were stained for Perls' Prussian blue using a standard protocol, to identify iron deposits. Briefly, sections were incubated with a 1:1 mixture of 5% hydrochloric acid and 5% potassium ferrocyanide (30 min), and counterstained with filtered neutral red (1 min).

Histopathological Image Analysis

Stained sections were scanned using a Hamamatsu NanoZoomer Digital Pathology (NDP)whole slide scanner (C9600–12, Hamamatsu Photonics KK, Japan), with a 20x objective. Obtained high-resolution (457 nm/pixel; 55579 dpi) digital images were visualized using the NDP.view2 software (version 2.8.24). The samples taken from the predefined standard brain areas were scored for cortical and leptomeningeal CAA severity, which was assessed on A β -stained sections, using a 4-point scale; absent (0), scant A β deposition (1), some circumferential A β (2), and widespread circumferential A β (3), following proposed consensus criteria²⁹. Parenchymal A β -plaque severity was evaluated using a 4-point scale; absent (0), mild (1), moderate (2), severe (3). CAA severity and A β -plaques were assessed by two independent raters (S.J.v.V. and V.P.), followed by a consensus rating to obtain a final score. Moreover, arteriolosclerosis in the juxtacortical WM of the sections and in the BG was assessed according to a 4-point scale (Grades 0–3)⁶, assigning to the section the highest grade reached by at least three arteries. Two independent raters (V.P. and C.A.A.) evaluated the sections and reached consensus to obtain a final score.

The H&E sections collected when targeting the CMBs were matched to the *ex vivo* MRIs and visually inspected. The positive retrieval of a CMB was defined as presence of erythrocyte extravasation (indicative of a recent hemorrhage) or blood-breakdown products (such as hemosiderin, indicative of subacute or old hemorrhages). Because vasculopathies (such as microaneurysms, or vessel calcifications) have been previously reported as CMB-mimics¹⁵, these were also scored. Positivity for iron was addressed on adjacent sections stained for Perls' Prussian blue. When possible, the 'culprit vessel' of the hemorrhage was identified if extravasation from it or near the CMB could be observed. When necessary, the blocks were further cut to completely retrieve the CMB or the culprit vessel. When a CMB was found, adjacent A β -stained sections, were inspected for presence and grade of CAA at a whole section level, at the site of rupture, as well as up- and downstream of the culprit vessel. Similarly, arteriolosclerosis was assessed on adjacent HE-stained sections. Moreover, the integrity of smooth muscle cells in the culprit vessel was examined on the adjacent SMA-stained sections and recorded as either complete circumferential or not. Observations were carried out by V.P. and discussed with S.J.v.V.

Statistical Analysis

The difference in ratio of the sum of CMBs in the juxtacortical plus subcortical WM and total CMBs, for false-positive vs true-positive CAA cases was calculated using a two-tailed Mann Whitney-U test. Statistical analysis was performed in R, version 3.6.0 (R Foundation for Statistical Computing, Vienna, Austria; www.R-project.org) and significance was set at p < 0.05.

Results

False-positive CAA cases

Three individuals who met criteria for probable CAA during life but showed no evidence of CAA upon standard histopathological examination of their brains at autopsy, were selected for the present study (Table 1a). During life, these three patients all presented with cognitive complaints to the Memory Disorders Division of the Massachusetts General Hospital (MGH) and were enrolled in a single-center longitudinal memory-clinic cohort, as previously described in detail elsewhere¹⁸. They underwent a neurological clinical evaluation, neuropsychological examination, 3 tesla (T) research MRI (on which CMBs were detected), and a Pittsburgh Compound-B (PiB) positron emission tomography (PET) scan. Case 1 was an 87-year-old Caucasian female with 16 years of education, who presented with increasing memory loss, that begun approximately six years earlier. Brain MRI showed three lobar CMBs, periventricular white matter hyperintensities (WMH) and moderate burden of MRI-visible perivascular spaces (PVS) in the centrum semiovale (CSO) and basal ganglia (BG). The second patient (case 2) was an 80-year-old Caucasian male, with 19 years of education, who first presented with cognitive complaints that begun two to three years earlier. Baseline MRI showed lobar CMBs, two intragyral hemorrhages¹⁹, lobar lacunes and WMH in the posterior periventricular region and CSO. Severe bilateral hippocampal atrophy could also be observed. The third patient (case 3) was a 79-year-old female, who had 18 years of education and presented with cognitive impairment beginning two years earlier. On the brain MRI performed on the same day of the first visit, two lobar CMBs, mild WMH, moderate burden of MRI-visible PVS in the CSO and a lacune in the caudate nucleus were seen. With the exception of the third patient, who had an ocular transient ischemic attach after initial evaluation, no further neurological symptoms were documented.

True-positive CAA cases

To assess differences in the localization of lobar CMBs between false-positive and truepositive CAA cases, we included three additional individuals from the same pathological cohort, who had comparable radiological characteristics (i.e. low number of CMBs). All cases fulfilled criteria for probable CAA during life, which was confirmed as definite CAA upon standard histopathological examination. Case 4 was a male who presented at age 68 at MGH with his second hemorrhagic stroke and deceased one year later. Case 5 was an 80-year-old female patient, who was evaluated at MGH because of memory impairment and diagnosed with probable CAA and dementia. She deceased at age 84. Case 6 was an 85-year-old male, who presented to an outside hospital, deceased the same year, and for which no further clinical information was available (Supplementary Table 1).

General Neuropathology Findings

No evidence of severe (i.e. grade >2) cortical or leptomeningeal CAA was found in any of the three false-positive CAA cases upon routine neuropathological analysis. Only a few sections from the predefined standard areas of the donated hemisphere showed mild CAA, in the form of grade 1 leptomeningeal (occipital cortex in case 1 and all areas in case 2) or

cortical CAA (parietal cortex in case 3), or grade 2 leptomeningeal CAA (parietal cortex in case 3). Moderate arteriolosclerosis was found in all three cases (Table 1b).

Neuropathology of MRI-observed CMBs

Across the three false-positive CAA cases, 25 MRI-observed CMBs were found on *in vivo* MRI, 15 of which were located in the post-mortem hemisphere that was included in our study. On the *ex vivo* 3T MRI scans, a total of 22 CMBs could be identified on the donated hemispheres. Nine of these were precisely localized on 7T MRI, 2 additional smaller ones were seen only on 7T MRI and one CMB was not searched for at high resolution, because the localization was easy to determine using 3T MRI (Figure 2). These 12 7T MRI-observed CMBs were targeted for histopathological analysis and a correlate was found for 9 of them (Table 2): 7/9 were hemosiderin deposits, indicating older microhemorrhages, or erythrocytes, indicating a more recent hemorrhage. 2/9 proved to be mimics on histopathology (one was a microaneurysm and one a calcified vessel) (Table 2). Two additional CMBs were identified only on histopathology and had no MRI correlate, also retrospectively. Five CMBs and the calcification were positive for iron.

Neuropathology of vessels associated with CMBs

The culprit vessel was identified for all histopathological CMBs. A total of 34 H&E-stained sections, and their adjacent sections stained for A β , iron and SMA, were examined in detail. In six sections, two CMBs were located on the same section. No CAA was found on any of the serial sections stained for A β (Figure 3), at the site of rupture of the culprit vessel, or up-/downstream of it. Conversely, 20/34 sections displayed moderate-to-severe arteriolosclerosis. In 8/9 CMBs and in 2/2 CMB-mimics (Figure 4) the culprit vessels had evidence of moderate-to-severe arteriolosclerosis at the rupture site (or site of most severe calcification or aneurysmal dilation), characterized by loss of smooth muscle cells with fewer nuclei visible within the wall, thickening of the vessel's wall, and dysmorphic changes, associated with partial occlusion. The degree of arteriolosclerosis remained similar also when the vessel was traceable in adjacent sections (up- or downstream). A non-circumferential ring of SMA was visible only in two of the culprit vessels.

Location of CMBs in false-positive CAA cases

It was noticed that the CMBs of false-positive CAA cases were mostly located in the juxtacortical or subcortical WM (7/9 pathologically confirmed CMBs) (Figure 5). This observation prompted a systematic comparison between the location of MRI-observed CMBs in false-positive and definite CAA cases, which was conducted on the 3T *ex vivo* MRI. The CMBs were classified as being located in the cortex, juxtacortical WM, or subcortical WM. The ratio between the sum of CMBs located in the juxtacortical WM and subcortical WM, to the total number of CMBs tended to be higher in the false-positive CAA cases (Median = 0, 5) compared to the true-positive CAA cases (Median = 0; W = 9, p = 0.076).

PiB-PET findings

Case 1 and 3 had a negative PiB PET scan (FLR DVR of 1.06 and 0.92 respectively). Case 2 had a positive scan (FLR DVR of 1.70), corresponding to the presence of parenchymal A β plaques on histopathology in this case (Figure 6).

Discussion

Lobar CMBs are a typical marker of CAA, but they occur also in absence of this pathology. In three individuals who met Boston criteria for a clinical diagnosis of probable CAA during life but proved to have no CAA in their brains at autopsy, we performed *ex vivo* MRI-guided histological serial sectioning of lobar CMBs, with the aim of identifying the neuropathological correlates of these lesions. We found that the majority of MRI-observed CMBs had extravasation of erythrocytes (suggesting an acute hemorrhagic event) or blood-breakdown products (e.g. hemosiderin, revealing older hemorrhages) as pathological correlates. Two of 9 were CMB-mimics (one microaneurysm and one calcification), which is consistent with the previously described heterogeneity of the pathological correlates of lobar CMBs.³⁰ Calcifications have been observed on *in vivo* imaging of small vessel disease patients³¹ and in hereditary Dutch-type CAA, where they were regarded as a marker of advanced pathology³².

No A β -depositions were found in any of the culprit vessels related to the CMBs or in any other vessels of the targeted sections, confirming that indeed CAA did not play a role in these lobar CMBs. Although it has been previously observed that the severity of CSVD-burden on MRI was lower in CAA cases without symptomatic ICH, compared to those with ICH, no differences have been reported in the number of CMBs on MRI between these two groups.³³ Furthermore, the positive predictive value of probable CAA according to the modified Boston criteria was previously shown to be high (87.5%)³⁴ in CAA patients without symptomatic ICH that came through a hospital-based memory clinic. Hence, the misclassification was likely not attributable to the absence of symptomatic ICH in these false-positive CAA cases. As per study design, inclusion in the MGH pathological cohort was based on the modified Boston criteria for the diagnosis of probable CAA². However, it is important to emphasize that cases 1 and 3 only had 2–3 lobar CMBs on in vivo MRI, and that in clinical practice they would possibly be considered as equivocal for a diagnosis of probable CAA.

Instead of CAA, all the identified culprit vessels had a moderate-to-severe degree of arteriolosclerosis, suggesting that this pathology might play a role not only in deep CMBs, but also in lobar CMBs. Moderate-to-severe arteriolosclerosis has indeed also been observed in the cortex and juxtacortical WM⁶. In fact, a co-occurrence of arteriolosclerosis and CAA was determined in 42% of cases with a lobar ICH³⁵. In line with these previous observations, arteriolosclerosis is a frequent finding across our CAA pathological cohort. The co-occurrence of the two small vessel pathologies suggests shared pathophysiological pathways leading to hemorrhages, possibly through a process including vessel remodelling.^{14,36} Recent work from our group, using histopathological analysis of MRI-targeted CMBs in cases with advanced CAA has shown that cortical CMBs are the

result of vascular remodeling, including hyaline thickening of the vessel wall, loss of smooth muscle cells, and loss of A β from affected vessels.^{14,28} While the exact mechanisms that trigger vessel remodeling and rupture in the context of advanced CAA remain unclear, it is possible that a comparable sequence of events may be implicated in the context of advanced arteriolosclerosis.

Relationships between CMBs and other neurodegenerative and neuroinflammatory pathologies have also been hypothesized^{10,37}, though they have not been directly confirmed in neuropathology. Therefore, it cannot be excluded that the investigation of further pathological hallmarks could inform us on the genesis of these lesions. The contribution of globular glial tauopathy to hemorrhage, which was found in case 1, appears to be unlikely^{38,39}.

Because of the clinical relevance that CAA burden has in predicting ICH risk during life, we explored possible characteristics that could aid the differentiation of false-positive CAA from true-positive CAA cases. Previous observations¹⁵ and a recent study on the validation of the Boston criteria V2.0 in a community-dwelling cohort (Zanon Zotin et al., under review)⁴⁰ have pointed out that lobar CMBs in definite CAA cases are observed mainly within the cortical ribbon, suggesting potential discriminative power between definite and non-CAA cases. This is in line with the fact that CAA is a cortical pathology, which only very rarely affects subcortical areas. In our false-positive CAA cases, the CMBs were more often seen in the juxtacortical area (including the U-fibers) and in the subcortical WM. Furthermore, a trend towards higher ratio between juxtacortical/subcortical CMBs and total number of lobar CMBs was observed in the false-positive versus the true-positive CAA cases, suggesting that the localization of the CMBs might aid in the differentiation between pathologies, if replicated in future work. The use of PiB-PET imaging might further aid the differentiation between true- and false-positive CAA cases, as two out of three cases in this study had indeed a negative PiB-PET scan during life. That said, previous studies have suggested PiB-PET might not be sensitive enough to detect mild CAA⁴¹ and the co-occurrence of A β -plaques in the brain parenchyma may result in a positive scan, as was true for one of our cases⁴². As such, further research is needed to help determine the added value of PET imaging to aid diagnosis of CAA during life.

For a few of the CMBs seen on MRI, a histopathological correlate could not be retrieved, most likely because a mismatch between MRI and histopathology occurred, even though it cannot be excluded that pathologies we did not detect (e.g. thrombi) underlined some of the MRI-observed CMBs. Nonetheless, the combination of *ex vivo* 7T MRI and histopathology represents a strength of our study, which allowed to first localize and then characterize these minute lesions, which would have been missed on standard pathological examination. This exploratory study, based on a series of pathological cases, has some limitations. Firstly, only three false-positive CAA cases were available. Future case series should include a greater number of subjects, which will possibly disclose more heterogeneity within the pathological correlates of the lobar CMBs observed on MRI. At present, the qualitative nature of the study, together with its small sample size, hinders an analysis that accounts for the co-dependence by subject. Secondly, the cross-sectional nature of histopathological examination does not allow to infer the sequence of events that resulted in microhemorrhage.

Thirdly, further pathological hallmarks of disease in the proximity of the lesions were not assessed. Systemic and local neuroinflammation and blood-brain-barrier leakage might for example play a role in the genesis of CMBs, as it has been suspected in multiple sclerosis⁴³ and shown in CAA⁴⁴. Future analyses should include the observation of reactive astrocytes and activated microglia.

Conclusion

This exploratory study aimed to investigate the pathological substrate of MRI-observed lobar CMBs in false-positive CAA cases. We confirmed that vascular A β accumulation was not present at the lesion site. Instead, the combination of ultra-high-resolution *ex vivo* MRI and serial sectioning gave insight in the likely role that arteriolosclerosis has in this context. Moreover, the observations made in this study, combined with those from a recent larger community-based study (Zanon Zotin et al., *under review*)⁴⁰, suggest that the localization of CMBs strictly within the cortical ribbon rather than in the juxtacortical or subcortical WM, could represent an additional discriminative diagnostic factor during life. These observations are qualitative in nature and need to be confirmed in additional studies, ideally in population-based studies not subject to the selection bias present in this autopsy cohort, which is a highly selected group of clinical CAA patients.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

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Data Availability

Source data can be made available to collaborators upon reasonable request.

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FIGURE 1:

Study design. During life, the participants of this study underwent a research 3T MRI scan (A), which included a susceptibility weighted imaging (SWI) sequence, that allowed to identify cerebral microbleeds (CMBs) as hypointense round structures (inset). After death, the most intact hemisphere was donated to our research program, fixed in formalin, before undergoing ex vivo 3T MRI (B), including a spoiled gradient echo T2*-weighted (FLASH) on which hypointense round/oval lesions were identified as CMBs (inset). The hemisphere was afterwards cut in 1 cm thick coronal slabs (C) and samples were taken from standardized lobar regions (frontal (F), temporal (T), parietal (P), occipital (O) lobe) and from the basal ganglia (BG). These samples were embedded in paraffin and adjacent 6 µm-thick sections were cut and stained for standard histopathology, which involved Hematoxylin and Eosin (HE) and immunohistochemistry against Amyloid- β (A β), two examples displayed are from the frontal and parietal lobe (D)). Additionally, the tissue slabs that contained CMBs in the false-positive CAA cases were identified according to anatomical landmarks (black box in C) and scanned with an ultra-high resolution 7T MRI protocol that encompassed a FLASH sequence, on which the location of the CMBs was confirmed (E). Guided by the MRI, samples were cut to fit a standard tissue cassette, they were embedded in paraffin and underwent complete serial sectioning. Each 5th section was stained for HE and when a CMB correlate was found, adjacent sections were subjected

to immunohistochemistry against $A\beta$, smooth muscle actin (SMA), and stained for Perls' Prussian Blue to detect iron (F). When the presumed culprit vessel of the lesion was identified, it was traced, and adjacent sections were also stained for the above. In the here represented CMB, hemosiderin deposits (arrow in F, HE-enlargement) and iron (arrow in F, iron-enlargement) were observed predominantly within the occluded vessel, which likely represents an example of end-stage vessel remodeling. Neither $A\beta$ (F, $A\beta$ -enlargement), nor smooth muscle cells (F, SMA-enlargement) were seen along the course of the vessel.



FIGURE 2:

Flow chart exemplifying the number of lobar cerebral microbleeds (CMBs) analyzed in the three false-positive CAA cases.



FIGURE 3:

Examples of cerebral microbleeds (CMBs) identified on MRI and their pathological correlates. (A) displays an example of a coronal view of the susceptibility weighted imaging (SWI) scan of the *in vivo* 3T MRI of case 2, on which a CMB was identified (inset). The *ex vivo* 3T MRI (B), confirmed the presence of this first CMB (inset with continued line) and allowed to detect a second one (inset with broken line). On the *ex vivo* 7T MRI of the tissue slab (C) the localization of both CMBs was confirmed (the second CMB was visible on another plane) and a third CMB was observed in proximity to the first (arrow, inset with continued line). This area was sampled and cut in serial sections to retrieve both CMBs (D, E). Hemosiderin deposits (arrows in D and E, HE-enlargements) and iron (D and E, iron-enlargements) were seen at different depths as correlates for both CMBs. Moreover, the culprit vessels for each CMB were traced and showed thickening and loss of structure of the vessel's wall and partial occlusion of the lumen, consistent with grade 3 arteriolosclerosis at the rupture site. There were no A β deposits in any of the vessels seen on the sections (D, E on Amyloid β). One of the culprit vessels had circumferential smooth muscle cells (E, SMA), while the other did not (D, SMA).



FIGURE 4:

Examples of cerebral microbleed (CMB) – mimics. Two of the CMBs identified on MRI proved to be CMB-mimics upon histopathological analysis. The histological correlate of the first, which was observed on *ex vivo* 3T MRI in case 1 (A, inset), and *ex vivo* 7T MRI (not shown), was a partially occluded (arrow) microaneurysm with hemosiderin deposits within the vessel wall (B, HE), but no iron deposits (B, iron). Vascular A β was absent (B, Amyloid β). The smooth muscle cell layer was largely intact (not shown). The second CMB-mimic was found in case 3, also on *ex vivo* 3T (inset, C) and 7T MRI (inset, D). At the corresponding location on serial sectioning (E), we observed a calcified vessel (E, HE), which stained positively for iron (E, iron), had no A β deposits within the vessel wall (E, Amyloid β), nor circumferential smooth muscle cells (E, SMA).



FIGURE 5:

Example of cerebral microbleeds (CMB) identified in the subcortical and juxtacortical white matter (WM). (A) coronal view of the spoiled gradient echo T2*-weighted (FLASH) sequence at *ex vivo* 3T MRI of case 3, which shows a lobar subcortical CMB in the WM (arrow) and a juxtacortical CMB (inset). The juxtacortical CMB was found in the histopathological analysis (B) where hemosiderin deposits (B, HE) and iron (B, iron) were observed at different depths in serial sections. The presumed culprit vessel was negative for both A β (arrow in B, Amyloid β -enlargement) and smooth muscle cells (arrow in B, SMA-enlargement).



FIGURE 6:

The figure displays the SUVR values of the Pittsburgh Compound-B PET in form of a heat map in the three false-positive CAA cases.

TABLE 1a.

Characteristics of the false-positive CAA cases and MRI findings. Cortical superficial siderosis, WMH, lacunes, intragyral bleeds, PVS and infarcts were assessed on the last *in vivo* scan before death, according to the STRIVE criteria³ and proposed differentiation of WMH-patterns⁴⁵. The latter include the multi-spot WMH pattern, which have recently been included as a supporting feature for the diagnosis of probable CAA according to the Boston criteria V2.0. Key: Basal ganglia (BG); Body-mass index (BMI); Cerebral Amyloid Angiopathy (CAA); Cerebral microbleeds (CMBs); Cerebral microinfarcts (CMIs); Centrum semiovale (CSO); Perivascular spaces (PVS); Intracerebral hemorrhage (ICH); white matter hyperintensities (WMH).

Demographic information	Case 1	Case 2	Case 3	
CAA	Probable CAA	Probable CAA	Probable CAA	
Age at death	88	88 85		
Sex	Female	Male	Female	
Hypertension	+	+	+	
Diabetes mellitus II	+	-	+	
Hyperlipidemia	+	+	+	
BMI	25.2	25.8	58	
Stroke/TIA	-	-	+	
MRI findings	Case 1	Case 2	Case 3	
MRI – death interval (months)	17	30	38	
No. lobar CMBs in vivo (total)	3	20	2	
No. lobar CMBs in vivo/ex vivo (relevant hemisphere)	2/5	12/13	1/4	
No. lobar CMBs ex vivo subcortical + juxtacortical/total	2/5	8/13	2/4	
No. CMBs targeted for histopathology	2 7		3	
No. correlates found on histopathology	2	2 4		
No. of extra CMBs found on histopathology	1 0		1	
Cortical Superficial Siderosis	No	No	No	
WMH	Multi-spot WMH	Confluent WMH	Multi-spot WMH	
Lacunes	1	4	3	
Intragyral bleeds	0	2	0	
ICH	0	0	0	
MRI-visible PVS CSO	3	1	0	
MRI-visible PVS BG	1	2	0	
Ischemic macroinfarcts	0	1	1	

TABLE 2.

Histopathological findings of the MRI-observed CMBs and of their culprit vessel. CAA and arteriolosclerosis score were assessed at a whole section level, at the site of rupture of the culprit vessel, as well as up- and downstream of it. Key: not available (N/A); smooth muscle cells (SMC).

MB	Detection on <i>in vivo</i> 3T MRI	Detection on ex vivo 3T MRI	Detection on <i>ex vivo</i> 7T MRI	Histological correlate	Location (histology)	Iron	CAA score (whole section/ culprit vessel at site of rupture/ culprit vessel down- and upstream)	Arteriolo- sclerosis score (whole section/ culprit vessel at site of rupture/ culprit vessel down- and upstream)	SMC
Case 1 – 1	+	+	Not scanned	Hemosiderin/ Erythrocytes	Subcortical	-	0/0/0	1/1/1	+
Case 1 – 2	-	+	+	Microaneurysm	Juxtacortical	-	0/0/0	2/3/3	+
Case 1 – 3	-	-	-	Hemosiderin/ Erythrocytes	Subcortical	-	0/0/0	2/3/3	+
Case 2 – 1	-	+	+	No correlate found	N/A	N/A		N/A	N/A
Case 2 – 2	-	+	+	No correlate found	N/A	N/A		N/A	N/A
Case 2 – 3	-	+	+	No correlate found	N/A	N/A		N/A	N/A
Case 2 – 4	+	+	+	Hemosiderin	Cortical	+	0/0/0	3/3/3	-
Case 2 – 5	+	+	+	Hemosiderin	Juxtacortical	+	0/0/0	3/3/3	-
Case 2 – 6	-	-	+	Hemosiderin	Juxtacortical	+	0/0/0	3/3/3	+
Case 2 – 7	+	+	+	Hemosiderin	Cortical	+	0/0/0	3/3/3	+
Case 3 – 1	-	+	+	Hemosiderin	Juxtacortical	+	0/0/0	2/2/2	+
Case 3 – 2	-	+	+	Hemosiderin	Subcortical	-	0/0/0	1/3/3	+
Case 3 – 3	-	-	-	Hemosiderin	Subcortical	-	0/0/0	1/3/3	N/A
Case 3 –	-	-	+	Calcification	Subcortical	+	0/0/0	1/3/3	+