Analysis

Unraveling the role of gut microbiota and immune cells in thyroid cancer and tumor drug resistance

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Abstract

The gut microbiota (GM) and immune cells (IC) are increasingly recognized as key players in cancer development and progression. This study aimed to explore the potential mediating role of IC in the causal relationship between GM and thyroid cancer (TC) using Mendelian randomization (MR) analysis. Data from genome-wide association studies (GWAS) encompassing 473 GM species, 731 IC types, and TC were utilized. MR analysis identified nine GM species with significant causal relationships to TC, mediated by 10 IC phenotypes such as "Switched Memory AC," "IgD-CD38dim AC," and "EM DN (CD4-CD8-) AC." These findings suggest a complex interplay where specific IC mediate the effects of GM on TC risk. Sensitivity analyses confirmed the robustness of these results, with no evidence of horizontal pleiotropy. This study high-lights potential mechanisms linking GM and IC to TC, offering insights that could inform GM-based immunotherapeutic strategies and IC-targeted interventions. However, further experimental research is needed to validate these causal pathways and better understand the underlying biological mechanisms.

Keywords Gut microbiota · Immune cells · Thyroid cancer · Mendelian randomization · Mediation analysis

1 Introduction

Since the 1990s, thyroid cancer (TC) has risen dramatically in incidence, ranking fifth in diagnosis and mortality among women diagnosed with cancer [1]. The overall prognosis of TC is generally favorable, however, 6–20% of patients will develop regional or distant metastases, immune system regulation has an impact on TC risk and progression [2]. The development and progression of cancer are influenced by complex molecular mechanisms that underpin tumor behavior and treatment responses. Key signaling pathways and oncoproteins are often implicated in these processes. For example, certain proteins involved in cell cycle regulation and microtubule stability can affect how cancer cells respond to chemotherapeutic agents, particularly in the context of drug resistance [3]. Additionally, pathways such as PI3K/AKT/mTOR have been highlighted for their role in regulating cancer cell growth, survival, and metabolic activity [4]. Recent findings suggest that modulating these pathways may provide adjunctive benefits to standard cancer therapies by enhancing the immune response and targeting tumor progression.

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The human gut microbiota (GM) is closely related to the host's metabolism, immunity, and health, playing an important role in the development of adaptive immune systems and the innate, which coordinate and maintain host-microbial symbiosis [5]. The GM modulates cancer immunotherapy and its immune-related adverse reactions. Fecal microbiota transplantation can improve the success rate of immunotherapy in cancer patients [6]. Immune cells (IC) are equally significant due to their central role in anti-tumor immunity and immune surveillance. They mediate responses that can either suppress or promote tumor growth, depending on the context and the interaction with external factors, including the GM. The rationale for targeting GM and IC in this study lies in their synergistic impact: GM-derived metabolites can modulate the activation and differentiation of IC, influencing the immune response against cancer. By focusing on these primary targets, the study aims to uncover the underlying mechanisms that could explain variability in tumor progression and treatment outcomes, potentially leading to innovative therapeutic strategies that enhance immune function through GM manipulation and IC-targeted interventions.

Cancer is a multifaceted disease characterized not only by uncontrolled cell proliferation but also by its interactions with the immune system and the surrounding tumor microenvironment (TME). The TME is composed of various immune cells, signaling molecules, and stromal components that collectively influence tumor growth and the response to therapy. The complex interplay between immune modulation and signaling pathways within the TME can affect both prognosis and therapeutic outcomes [7]. For instance, certain signaling pathways are known to contribute to immune evasion mechanisms and impact the effectiveness of immunotherapy [8]. Moreover, the identification of specific molecular signatures and biomarkers has shown promise in predicting cancer prognosis and tailoring treatment strategies to enhance patient response to therapy. These factors underscore the importance of understanding the dynamic interactions within the TME as a foundation for developing innovative therapeutic approaches.

Recent research on the thyroid-gut axis indicates that intestinal microbiota and its metabolites may influence the thyroid gland by affecting the uptake of intestinal trace elements and immune regulation, thereby enhancing our understanding of the pathogenesis and clinical treatment of thyroid diseases [9]. Beyond thyroid-specific effects, the broader role of GM and IC in cancer treatment outcomes has gained significant attention. The composition and diversity of the gut microbiota can modulate the host's immune response, which is critical for the effectiveness of immunotherapies such as immune checkpoint inhibitors. Certain bacterial species have been linked to better responses to these treatments, as they can promote the activation and infiltration of IC, including T cells, into the TME. Conversely, dysbiosis can impair immune activation, leading to reduced treatment efficacy and increased resistance to therapies. Understanding the interplay between GM, IC, and cancer treatments not only provides insight into patient variability in treatment outcomes but also supports the development of adjunctive therapies, such as probiotics or prebiotics, to enhance the success of cancer immunotherapy and reduce adverse effects.

The symbiotic intestinal microbiota inhabits the gastrointestinal tract, regulates the host's immune response and balance, and affects the metabolism of IC [10]. Dysbiosis of the intestinal microbiota can lead to autoimmune diseases and can be critical in regulating the immune system response [11]. The diversity and abundance of GMs are significantly reduced in individuals with TC [12]. Key mechanisms in GM-IC interactions include microbial metabolites like short-chain fatty acids (SCFAs), which enhance regulatory T cells (Tregs) and maintain immune tolerance, and microbial components like lipopolysaccharides (LPS), which activate immune pathways (e.g., NF-kB) through toll-like receptors (TLRs), modulating cytokine production. Dysbiosis may drive chronic inflammation, promoting tumor progression by creating a pro-inflammatory environment that aids immune evasion. These mechanisms suggest a therapeutic potential in targeting GM to regulate immune responses in TC.

Mendelian randomization (MR) adheres to the Mendelian inheritance principle of "random assignment of genetic alleles" in order to infer the causal relations between exposure factors and research outcomes in observational studies [13]. The objective of this study was to identify the IC that mediate the causality between GM and TC. Therefore, we utilized MR analysis to deduce and dissect the causality between GM, IC, and TC.

2 Materials and methods

2.1 Study design

The procedural steps and methodology of MR in this study are detailed below (Fig. 1). Firstly, MR analysis was used to evaluate the causality of GM, TC and IC. Reverse analysis was performed to eliminate the risk of reverse causality. Following the three basic assumptions of MR analysis, the requirements for the selected SNPs are as follows:

1. They should be strongly related to exposure;



- 1. 2. The outcome should be affected only by exposure;
- 2. Confounding-related SNPs should be eliminated.

Secondly, MR analysis was used to assess the causality of IC on TC.

Finally, we explored the role of IC as a mediator in the pathways between GM and TC, and calculated the effect sizes (beta1, beta2, beta_all) and the proportions for each qualified mediator.

2.2 Data sources

The data for 473 GM traits comes from the NHGRI-EBI GWAS, and genome-wide association tests were performed on genetic variants among 5,959 Europeans. A total of 471 genetic taxa were identified [14]. Data on 731 ICs were derived from aggregated GWAS statistics of 3757 Europeans [15]. The 731 immunophenotypes comprised absolute cellularity (AC), morphological parameters, which include CDC and TBNK groups, median fluorescence intensity, and relative cellularity, including TBNK, Treg panel, T cells, bone marrow cells, natural killer cells, monocytes, and CDCs [11]. The data of TC were derived from Global Biobank Project Whole-Genome Genotyping GWAS, including 11,121 Hispanic or Latin American, 25,692 African unspecified, 326,915 East Asian, and 1,376,270 European individuals. The detailed data are shown in Table 1.

2.3 Selection of IVs

IVs were selected for subsequent analysis based on stringent criteria to ensure robust and reliable results. First, SNPs were chosen if they had a p-value of less than 5e-8 for their association with GM, IC, and TC, ensuring that only genome-wide significant variants were included. Second, to minimize the risk of linkage disequilibrium (LD) bias, SNPs were filtered using R software to exclude those with an R^2 value of less than 0.001 within a 10,000 kb range, which helped maintain the independence of the IVs. Third, SNPs that were strongly associated with the outcome (P < 5×10⁻⁵) were excluded to prevent potential confounding influences that could bias the causal inference. Fourth, the F-statistic was calculated for each SNP, and only those with an F-statistic greater than 10 (F > 10) were retained to ensure that the IVs were strong instruments with sufficient power to detect causal relationships.

2.4 MR analysis

Among GM, IC, and TC, inverse-variance weighting (IVW) was served as the methodology to evaluate the causality, along with four auxiliary analysis methods [16]. If P < 0.05 in the MR-IVW analysis, there is a causality between the two samples. Set the F > 10 to identify strong instrumental variables [17]. The P-value for the horizontal pleiotropy and heterogeneity

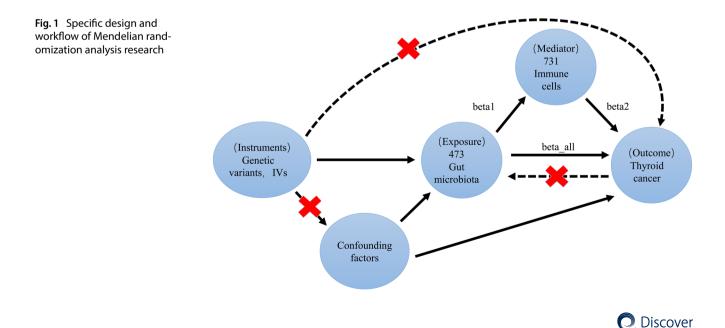


Table 1 Data sources

Analysis

Variable	ariable Phenotypes Cases/controls or sizes		Data source	Phenotypes code	Ancestry
Exposure	Gut microbiota	5959	GWAS Catalog	GCST90032172-GCST90032644	European
Mediator	Immune cells	3757	GWAS Catalog	GCST90001391-GCST90002121	European
Outcome	Thyroid cancer	1,620,354/119,644	GWAS Catalog	GCST90399737	Global

tests in MR should be greater than 0.05 to ensure reliability. According to these criteria, intestinal microbiota and IC with a positive causality, but no reverse causality with TC, were screened.

2.5 Analysis of intermediate MR method

Follow these steps to find potential IC in the GM-TC mediator pathway (Fig. 1):

We used MR analysis to identify IC causally affected by the GM and calculated the impact value (beta1).

Calculated the impact value (beta2) of the IC on TC.

Calculated the impact value (beta_all) of the identified GM on TC.

Calculated the mediation effect (beta12 = beta1 * beta2).

Calculated the direct effect (beta_dir = beta_all-beta12).

2.6 Sensitive MR analysis

In our research, three MR methods were served as sensitivity analysis, namely MR-Egger, MR-PRESSO, and leave-one-out. We assessed heterogeneity and horizontal pleiotropy by calculating Cochran's Q statistic and the MR-Egger regression intercept, respectively. Horizontal pleiotropy in IVs is addressed using MR-PRESSO [18]. Evidence of pleiotropy would invalidate the causality. We used the leave-one-out method to thoroughly investigate the effect of single SNP on IVW and used funnel plots to evaluate potential biases in the study results, ensuring robustness [19].

The MR study relied on R software (version 4.3.3) with the software packages: "Variant Annotation" (version 1.48.1), "foreach" (version 1.5.2), "Two Sample MR" (version 0.5.10), "data.table" (version 1.15.4), "ggplot2" (version 3.5.1), and "gwasglue" (version 0.0.0.90).

3 Results

3.1 IVs for exposure

The IVs needed for GM, IC, and TC studies were screened using the aforementioned method. The number of SNPs in GM, IC, and TC we studied ranged from 6 to 22, 12 to 31, and 22, respectively. Additionally, the F-statistic of SNPs screened in this study was at least 15. These data are detailed in Supplementary Material Table V1-V20.

3.2 Causality between GM and TC

After MR analysis, GM species with a causality (P < 0.05) with TC were screened among 473 GM. Ultimately, 9 GM species were found to have a causal association with TC, with no reverse causality identified (Fig. 2). Using IVW as the primary method, specific GM species demonstrated notable odds ratios (OR), indicating their strength of association with TC. For example, s_Veillonella rogosae had an OR of 1.1871, which implies that an increase in its abundance is associated with an approximately 18.7% higher risk of TC. Similarly, s_Bacteroides stercoris had an OR of 1.6441, suggesting a 64.4% increased risk of TC, highlighting its strong potential role in tumorigenesis. The highest effect was seen with f_Mycobacteriaceae (OR = 2.1736), indicating that its presence could more than double the risk of TC (a 117.4% increase), pointing to a particularly significant influence.



These effect sizes illustrate the varying degree of impact that different GM species may have on TC. The OR values reflect the potential risk modulation of TC as mediated by GM, providing insight into which species might be prioritized for further research. This highlights the importance of specific GM species not just in terms of their presence but also their quantitative effect on TC risk. The summary results of the MR analysis, including detailed ORs for all GM species, are provided in Fig. 3 and the Supplementary Materials (Figures S1-9, Tables T1-9, U1-9, and V1-9).

3.3 Causality between IC and TC

After conducting MR analysis, we screened 731 IC to identify those causally related to TC based on GM (P < 0.05), ultimately identifying 10 types that were causally related to TC (Fig. 4). When IVW serves as the primary method to evaluate

Fig. 2 MR analysis OR and P value of 9 gut microbiota on thyroid cancer

Exposures_Outcome	Used_SNPS	S OR (95% CI) P·	-value
s_Veillonella rogosae on Thyroid cancer		1	
MR Egger	13	1.0417 (0.7668 - 1.4151) 0.	.7988
Weighted median	13	1.0956 (0.8924 - 1.3452) 0.	.3830
Inverse variance weighted	13	····· 1.1871 (1.0202 − 1.3813) 0.	.0265
Simple mode	13	1.1176 (0.8400 - 1.4870) 0.	.4599
Weighted mode	13	1.1027 (0.8544 - 1.4233) 0.	.4670
s Massiliomicrobiota on Thyroid cancer			
MR Egger	10	1.2616 (0.8719 - 1.8255) 0.	.2527
Weighted median	10	. ,	.1333
Inverse variance weighted	10		.0434
Simple mode	10		.3391
Weighted mode	10		.3066
s Phocea massiliensis on Thyroid cancer			
MR Egger	22	→ 1.3196 (0.7563 - 2.3025) 0.	.3404
Weighted median	22		.0565
Inverse variance weighted	22		.0066
Simple mode	22		.0766
Weighted mode	22		.0967
g_Saccharomonospora on Thyroid cancer			.0307
MR Egger	15	2 6820 (1 2206 - 5 4142) 0	.0164
	15		
Weighted median			.1333
Inverse variance weighted	15		.0400
Simple mode	15		.3692
Weighted mode	15	→ 1.3087 (0.7571 – 2.2622) 0.	.3517
s_Bacteroides stercoris on Thyroid cancer			
MR Egger	18		.1982
Weighted median	18	· · · · · · · · · · · · · · · · · · ·	.0886
Inverse variance weighted	18		.0393
Simple mode	18	,	.3180
Weighted mode	18	→ 1.7357 (0.6477 – 4.6514) 0.	.2882
f_Acetobacteraceae on Thyroid cancer			
MR Egger	20		.9133
Weighted median	20	• 1.4222 (1.0744 – 1.8826) 0.	.0138
Inverse variance weighted	20	······ 1.4005 (1.1328 − 1.7315) 0.	.0019
Simple mode	20	· · · · · · · · · · · · · · · · · · ·	.1260
Weighted mode	20	· · · · · · · · · · · · · · · · · · ·	.1430
g_UNC496MF on Thyroid cancer			
MR Egger	17	• 1.1812 (0.7012 – 1.9897) 0.	.5408
Weighted median	17	1.2175 (0.9722 - 1.5246) 0.	.0864
Inverse variance weighted	17	····· 1.2121 (1.0245 − 1.4340) 0.	.0250
Simple mode	17	→ → → → → → → → → →	.2686
Weighted mode	17	→ 1.2569 (0.8561 − 1.8453) 0.	.2602
s_Ruminococcus C on Thyroid cancer			
MR Egger	8	→ 0.8133 (0.2109 - 3.1364) 0.	.7742
Weighted median	8	·····→ 1.4900 (0.8169 – 2.7177) 0.	.1934
Inverse variance weighted	8	·→ 1.7637 (1.0881 – 2.8586) 0.	.0213
Simple mode	8	→ 1.5819 (0.5775 - 4.3333) 0.	.4020
Weighted mode	8	· · · · · · · · · · · · · · · · · · ·	.4834
f Mycobacteriaceae on Thyroid cancer			
MR Egger	6	→ 2.3562 (0.2102 - 26.4088) 0.	.5253
Weighted median	6		.1209
Inverse variance weighted	6		.0062
Simple mode	6		.3481
Weighted mode	6		.3994
theighted mode	•		
		0 1 2	



the causality of IC on TC in the MR analysis, Switched Memory AC (OR = 1.075), CD25 on CD39+CD4+(OR = 1.033), IgD-CD38dim AC (OR = 1.0876), Effector Memory Double Negative (EMDN) (CD4-CD8-) AC (OR = 1.105), B cell %lymphocyte (OR = 1.0537), CD28 + DN (CD4-CD8-) AC (OR = 1.1009), CCR2 on CD14 + CD16 + monocyte (OR = 1.024), SSC-A on granulocyte, the expression of CD11c on myeloid Dendritic Cells (DC) (OR = 1.0568), and CD11b on Mo MDSC (OR = 1.0508) were correlated with TC. The detailed results of all MR analyses are shown in Fig. 5 and Supplementary Figures (S21-30, T21-30, U21-30, and V10-19).

3.4 Causal relationship between GM and IC

When IVW serves as the primary method to evaluate the causality of GM on IC in the MR analysis, s_Veillonella rogosae on Switched Memory AC (OR = 1.2306), s_Veillonella rogosae on IgD- CD38dim AC (OR = 1.2288), s_Massiliomicrobiota on B cell % lymphocyte (OR = 1.1992), s_Massiliomicrobiota on SSC-A on granulocyte (OR = 1.2347), s_Phocea massiliensis on CCR2 on CD14 + CD16 + monocyte (OR = 1.3946), and g_Saccharomonospora on EM DN (CD4-CD8-) AC (OR = 1.5444). The above results revealed that GM had a positive correlation with IC. Detailed results of all MR analyses are shown in Fig. 6 and Supplementary Figures (S10-20, T10-20, U10-20, and V20).

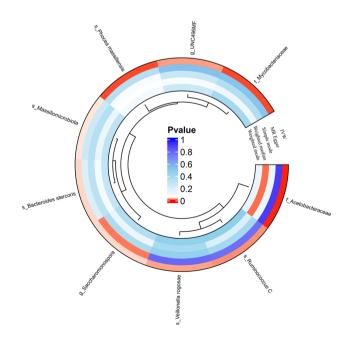
3.5 Analysis of potential IC mediation

MR analysis revealed that 10 IC mediated the causality between 9 GM and TC. Notably, 'EM DN (CD4-CD8-) AC' exhibited the highest mediation ratio (beta_P of 15.9%) in the pathway between g_Saccharomonospora and TC, suggesting that this immune cell type may play a particularly significant role in influencing tumor progression. The strong mediation by 'Switched Memory AC' (beta_P of 8.7%) and 'IgD-CD38dim AC' (beta_P of 10.1%) in the relationship between s_Veillonella rogosae and TC highlights the potential importance of adaptive immune memory responses in modulating cancer outcomes. These cell types are known for their roles in sustaining long-term immune surveillance and response, which could explain their notable mediation effects.

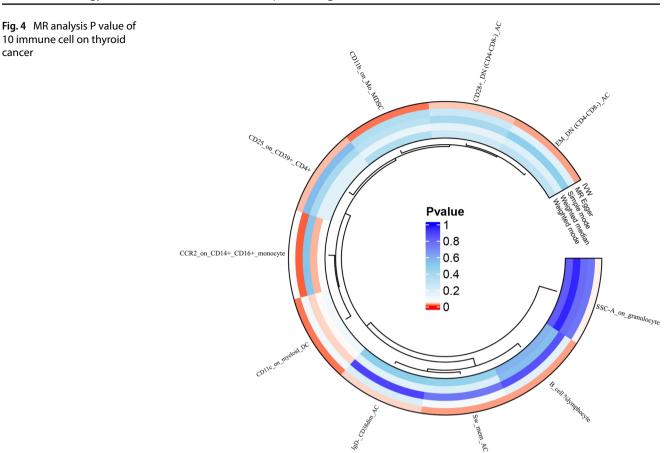
In contrast, 'CCR2 on CD14 + CD16 + monocyte' had a lower mediation ratio (beta_P of 2.2%) in the pathway between s_Phocea massiliensis and TC, possibly reflecting a more specialized or context-dependent role of this monocyte subtype in the immune response to tumorigenesis. The involvement of 'CD28 + DN (CD4-CD8-) AC' (beta_P of 9.7%) in mediating the effect of s_Bacteroides stercoris on TC may indicate a crucial role in immune regulation, given the dual functions of CD28 in co-stimulatory signaling for T cells.

The varying mediation ratios observed suggest that some IC, such as 'EM DN (CD4-CD8-) AC' and 'Switched Memory AC,' may have broader and more impactful roles in the interplay between GM and TC, potentially due to their involvement in maintaining immune homeostasis and response modulation. Meanwhile, mediators like 'CD11b on Mo MDSC'

Fig. 3 MR analysis P value of 9 gut microbiota on thyroid cancer



cancer



(beta_P of 7.0%) and 'CD11c on myeloid DC' (beta_P of 5.6%) underscore the influence of myeloid lineage cells in immune suppression and tumor immune escape. These differences emphasize the complexity of the immune response and the varied influence of different IC in mediating GM's effects on TC, warranting further experimental validation to uncover the mechanisms driving these interactions. These data are detailed in Table 2.

3.6 MR sensitivity analysis

This MR study found no evidence to support the existence of horizontal pleiotropy, as indicated by the MR-Egger and MR-PRESSO. Leave-one-out study revealed that independent SNPs didn't significantly influence the overall effect in the MR analyses of GM-TC and TC-GM (Supplementary Material Figure U1-U30). Overall, the findings of the MR study were confirmed to be robust through the conducted sensitivity analyses.

4 Discussion

Our research identified that 9 specific GM species (s_Veillonella rogosae, s_Massiliomicrobiota, s_Phocea massiliensis, g Saccharomonospora, s Bacteroides stercoris, f Acetobacteraceae, g UNC496MF, s Ruminococcus C, and f Mycobacteriaceae) were causally related to TC and might be mediated by 10 IC phenotypes (EM DN (CD4-CD8-) AC, Switched Memory AC, B cell % lymphocyte, IgD-CD38dim AC, CD28+(DN) (CD4-CD8-) AC, CD25 on CD39+CD4+cells, CD11c on myeloid DC, CCR2 on CD14+CD16+monocytes, SSC-A on granulocytes, and CD11b on Mo MDSC). These findings elucidate potential immune-mediated mechanisms between GM and TC, providing new insights into GM-based immunotherapeutic strategies and IC-targeted interventions for TC. This study highlights the complex interplay between specific gut microbiota and immune cells, emphasizing the potential for targeted approaches that leverage these interactions to modulate the immune environment and improve therapeutic outcomes in thyroid cancer.



(2024) 15:683

Fig. 5	MR analysis OR and P
value	of 10 immune cell on
thyroi	d cancer

Exposures_Outcome	Used_SNPS		OR (95% CI)	P-value
Sw mem AC on Thyroid cancer		1		
MR Egger	18	⊨=-1	1.0890 (0.9917 - 1.1958)	0.0930
Weighted median	18	H	1.0593 (0.9704 - 1.1564)	0.1977
Inverse variance weighted	18) -	1.0750 (1.0092 - 1.1450)	0.0249
	18		1.0217 (0.8938 - 1.1680)	0.7565
•	18		1.0402 (0.9402 - 1.1509)	
IgD- CD38dim AC on Thyroid cancer				
	16		1.0963 (0.9596 - 1.2523)	0 1974
	16		1.0853 (0.9663 - 1.2189)	
5	16		. ,	
5	16		1.0876 (1.0042 - 1.1781) 1.0114 (0.8561 - 1.1948)	
•			(/	
0	16		1.0480 (0.9217 - 1.1917)	0.4852
EM DN (CD4-CD8-) AC on Thyroid cancer				
	15	₩ ₩ ₩₩	1.1290 (0.9564 - 1.3326)	
5	15		1.0845 (0.9621 - 1.2224)	
	15	i+•-1	1.1050 (1.0119 - 1.2066)	0.0262
Simple mode	15		1.0628 (0.9031 - 1.2508)	0.4754
Weighted mode	15	∺ •	1.0743 (0.9423 - 1.2248)	0.3019
B cell %lymphocyte on Thyroid cancer				
MR Egger	29		1.0580 (0.9787 - 1.1437)	0.1674
Weighted median	29	Here .	1.0214 (0.9540 - 1.0934)	0.5434
-	29		1.0537 (1.0064 - 1.1032)	
-	29	⊢∎1	1.0100 (0.9066 - 1.1252)	
•	29		1.0232 (0.9443 - 1.1087)	
CD28+ DN (CD4-CD8-) AC on Thyroid cancer	20		1.0202 (0.0440 1.1007)	0.0100
	12		1.0971 (0.9432 - 1.2761)	0.2572
		H	, , ,	
5	12	+	1.0952 (0.9668 - 1.2406)	
0	12	H- -1	1.1009 (1.0063 - 1.2044)	
	12		1.0936 (0.8998 - 1.3291)	
Weighted mode	12	i ∔ ∎1	1.0957 (0.9462 - 1.2688)	0.2477
CD25 on CD39+ CD4+ on Thyroid cancer				
MR Egger	19	-	1.0137 (0.9701 - 1.0593)	0.5515
Weighted median	19	(= 1	1.0293 (0.9874 - 1.0729)	0.1733
Inverse variance weighted	19	-	1.0330 (1.0027 - 1.0643)	0.0326
Simple mode	19	H=-1	1.0351 (0.9593 - 1.1168)	0.3862
Weighted mode	19	in the second se	1.0270 (0.9896 - 1.0658)	0.1760
CCR2 on CD14+ CD16+ monocyte on Thyroid cancer			. ,	
	31	-	1.0407 (1.0113 - 1.0710)	0.0106
	31	-	1.0389 (1.0036 - 1.0754)	
0	31		1.0240 (1.0001 - 1.0485)	
	31		1.0174 (0.9653 - 1.0723)	
•	31			
-	51		1.0306 (1.0001 - 1.0619)	0.0565
SSC-A on granulocyte on Thyroid cancer	o.(
	24	HH	1.0105 (0.9458 - 1.0796)	
0	24	H i	0.9983 (0.9385 - 1.0618)	
Inverse variance weighted	24	=·	1.0446 (1.0007 - 1.0905)	0.0466
Simple mode	24	+	1.0136 (0.9202 - 1.1164)	0.7869
Weighted mode	24	H i I	0.9936 (0.9363 - 1.0543)	0.8328
CD11c on myeloid DC on Thyroid cancer				
MR Egger	25	i	1.0576 (0.9917 - 1.1280)	0.1016
	25	H=H	1.0710 (1.0004 - 1.1466)	
	25) 	1.0568 (1.0104 - 1.1054)	
	25		1.1365 (1.0132 - 1.2748)	
	25		1.0617 (0.9908 - 1.1377)	
*	20		1.0017 (0.0000 1.1077)	0.1020
CD11b on Mo MDSC on Thyroid cancer	45		4 0005 (0 0000 4 4050)	0.0505
	15	Heri	1.0335 (0.9666 - 1.1050)	
	15	h=-1	1.0435 (0.9855 - 1.1050)	
•		N=+	4 0500 (4 0006 4 0027)	0.0152
Inverse variance weighted	15)=-1	1.0508 (1.0096 - 1.0937)	
Inverse variance weighted	15 15	µ== - - - -	1.0508 (1.0096 - 1.0937) 1.0559 (0.9526 - 1.1705) 1.0395 (0.9600 - 1.1256)	0.3181

The GM is crucial for the progression, differentiation, as well as maturation of the human immune system and has a disproportionate impact on triggering thyroid autoimmune diseases [20]. In vitro studies using mature monocytes treated with antiretroviral therapy have shown that HIV + CD14 + CD16 + monocytes are the first to migrate [21]. Our study suggests that the IC phenotype "CCR2 on CD14 + CD16 + " may mediate the positive effects of "Phocea massiliensis" on TC. Whole genome expression analysis using RNA sequencing showed that Ess2 deficiency altered the expression of immune-related genes and Myc target genes in CD4 single-positive thymocytes [22]. Our findings indicated that the IC phenotype "EM DN (CD4-CD8-) AC" mediates the positive effects of "s_Phocea massiliensis" on TC. CD4 + CD25 + Tregs have significant impact on preventing immune attacks and exert immune surveillance by modulating



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Analysis

J.6 MR analysis OR and P	Exposures_Outcome	Used_SNPS		OR (95% CI)	P−va
lue of 9 gut microbiota on	s_Veillonella rogosae on Sw mem AC				
3	MR Egger	19		1.2687 (0.9672 - 1.6641)	0.103
immune cells	Weighted median	19		1.2802 (1.0146 - 1.6153)	0.037
	Inverse variance weighted	19		1.2306 (1.0621 - 1.4258)	0.005
	Simple mode	19		0.9532 (0.6487 - 1.4005)	
	Weighted mode	19		1.2975 (1.0163 - 1.6565)	
	-	19		1.2975 (1.0105 - 1.0505)	0.001
	s_Veillonella rogosae on IgD- CD38dim AC	10		1 0010 (0 0015 1 0015)	0.075
	MR Egger	19		1.2912 (0.9915 - 1.6815)	
	Weighted median	19		1.2389 (0.9852 - 1.5579)	0.066
	Inverse variance weighted	19		1.2288 (1.0646 - 1.4182)	0.00
	Simple mode	19	·	1.2379 (0.8910 - 1.7200)	0.21
	Weighted mode	19)— • —•	1.2639 (1.0165 - 1.5713)	0.04
	s Massiliomicrobiota on B cell %lymphocyte				
	MR Egger	14		1.2656 (0.9266 - 1.7287)	0.16
	Weighted median	14		1.1591 (0.9189 - 1.4623)	0.21
	Inverse variance weighted	14		1.1992 (1.0290 - 1.3974)	
	-				
	Simple mode	14		1.0948 (0.7752 - 1.5463)	
	Weighted mode	14		1.1207 (0.8000 - 1.5699)	0.51
	s_Massiliomicrobiota on SSC-A on granulocyte				
	MR Egger	14		1.1686 (0.8323 - 1.6408)	0.38
	Weighted median	14		1.2230 (0.9631 - 1.5530)	0.09
	Inverse variance weighted	14		1.2347 (1.0456 - 1.4581)	
	Simple mode	14		1.2021 (0.8829 - 1.6367)	0.26
	Weighted mode	14		1.2281 (0.9741 - 1.5484)	
	s_Phocea massiliensis on CCR2 on CD14+ CD16+				0.10
	-			> 1 2407 (0 7474 - 0 5000)	0.20
	MR Egger	23		→ 1.3407 (0.7171 - 2.5066)	0.36
	Weighted median	23		→ 1.3651 (0.8758 - 2.1278)	
	Inverse variance weighted	23		→ 1.3946 (1.0299 - 1.8886)	0.03
	Simple mode	23		→ 1.5774 (0.8616 - 2.8880)	0.15
	Weighted mode	23		→ 1.3319 (0.7825 - 2.2669)	0.30
	g Saccharomonospora on EM DN (CD4-CD8-) AC				
	MR Egger	17		→ 2.0030 (0.6938 - 5.7827)	0.21
	Weighted median	17		→ 1.2742 (0.8087 - 2.0078)	
	Inverse variance weighted	17		→ 1.5444 (1.1161 - 2.1371)	
		17		, , ,	
	Simple mode			→ 0.9943 (0.4811 - 2.0549)	0.98
	Weighted mode	17		→ 1.0542 (0.5497 - 2.0217)	0.87
	s_Bacteroides stercoris on CD28+ DN (CD4-CD8-)				
	MR Egger	22		→ 1.0232 (0.4086 - 2.5625)	0.96
	Weighted median	22		→ 1.3192 (0.6916 - 2.5164)	0.40
	Inverse variance weighted	22		→ 1.6549 (1.0678 - 2.5648)	0.02
	Simple mode	22		→ 1.5044 (0.5572 - 4.0620)	0.42
	Weighted mode	22		→ 1.2915 (0.5927 - 2.8146)	0.52
	f_Acetobacteraceae on CD11c on myeloid DC	<u></u>		1.2010 (0.0021 2.0140)	0.01
		25		> 1 2680 (0 6680 - 2 4070)	0.47
	MR Egger	25		→ 1.2680 (0.6680 - 2.4070)	
	Weighted median	25		1.1577 (0.8055 - 1.6641)	
	Inverse variance weighted	25		- 1.4048 (1.0485 - 1.8822)	0.02
	Simple mode	25		→ 1.2855 (0.7495 - 2.2049)	
	Weighted mode	25	H	→ 1.2417 (0.7971 - 1.9342)	0.34
	g_UNC496MF on CD25 on CD39+ CD4+				
	MR Egger	19		→ 1.1656 (0.5537 - 2.4539)	0.69
	Weighted median	19		1.3370 (0.9620 - 1.8582)	
	-	19		1.3158 (1.0384 - 1.6672)	0.08
	Inverse variance weighted				
	Simple mode	19		→ 1.2751 (0.7279 - 2.2335)	
	Weighted mode	19		→ 1.3509 (0.8270 - 2.2066)	0.24
	s_Ruminococcus C on CD11b on Mo MDSC				
	MR Egger	11	·	→ 4.2669 (1.0852 - 16.7766) 0.06
	Weighted median	11		→ 1.5023 (0.5714 - 3.9499)	0.40
	Inverse variance weighted	11		→ 2.2367 (1.0699 - 4.6762)	
	Simple mode	11		→ 3.2803 (0.6747 - 15.9491	
	Weighted mode	11			
	-	11		→ 2.1047 (0.6215 - 7.1272)	0.25
	f_Mycobacteriaceae on B cell %lymphocyte				
	MR Egger	12		→ 3.1486 (1.0803 - 9.1763)	
	Weighted median	12	-	→ 1.6963 (0.9330 - 3.0841)	0.08
	Inverse variance weighted	12		→ 1.5814 (1.0164 - 2.4605)	0.04
	Simple mode	12		→ 1.8160 (0.6988 - 4.7189)	
	Weighted mode	12	1	→ 1.9280 (0.9170 - 4.0533)	
					U. I I

the function of antigen-presenting cells [23]. Our study found that the phenotype "CD25 on CD39+CD4+" mediates the positive effect of g_UNC496MF on TC.

MDSCs are cells that suppress anti-tumor immunity, including CD11b + Gr1 + Iy6 PMN-MDSCs and CD11b + Gr1 + Iy6 Mo-MDSCs [24]. Our study found that the IC phenotype "CD11b on Mo MDSC" mediates the positive influence of "s_Ruminococcus C" on TC.

The GM regulates the efficacy of cancer immunotherapy and its immune-related adverse reactions. Fecal microbiota transplantation or dietary intervention may be clinically employed to enhance the success rate of immunotherapy in cancer patients [6]. Consuming a diet that triggers inflammation has been found to be associated with higher levels of



Gut microbiota	Immune cells	beta all	beta dir	beta 1	beta 12	beta P (%)	beta 2
s_Veillonella rogosae	Sw mem AC	0.172	0.157	0.208	0.015	8.7	0.072
s_Veillonella rogosae	lgD- CD38dim AC	0.172	0.154	0.206	0.017	10.1	0.084
s_Massiliomicrobiota	SSC-A on granulocyte	0.154	0.145	0.211	0.009	6.0	0.044
s_Massiliomicrobiota	B cell %lymphocyte	0.154	0.145	0.182	0.009	6.2	0.052
s_Phocea massiliensis	CCR2 on CD14+CD16+monocyte	0.355	0.347	0.333	0.008	2.2	0.024
g_Saccharomonospora	EM DN (CD4-CD8-) AC	0.274	0.23	0.435	0.043	15.9	0.1
s_Bacteroides stercoris	CD28+DN (CD4-CD8-) AC	0.497	0.449	0.504	0.048	9.7	0.106
f_Acetobacteraceae	CD11c on myeloid DC	0.337	0.318	0.34	0.019	5.6	0.055
g_UNC496MF	CD25 on CD39 + CD4 +	0.192	0.183	0.274	0.009	4.6	0.033
s_Ruminococcus C	CD11b on Mo MDSC	0.567	0.527	0.805	0.04	7.0	0.05
f_Mycobacteriaceae	B cell %lymphocyte	0.776	0.752	0.458	0.024	3.1	0.052

Table 2 Mediating effect on thyroid cancer

beta all: Total effect of gut microbiota on thyroid cancer

beta dir: Direct effects of gut microbiota on thyroid cancer

beta 1: Effects of gut microbiota on immune cells

beta 12: Mediating effect of gut microbiota on thyroid cancer

beta P: Percentage of mediating effects of gut microbiota on thyroid cancer

beta 2: Effects of immune cells on thyroid cancer

"Veillonella rogosae" in individuals with inflammatory bowel disease who are in remission, which suggests that the food we choose to eat can impact the composition of the GM and the level of inflammation experienced by these patients [25]. According to our analysis results, the IC phenotypes " Switched Memory AC" and "IgD-CD38dim AC" acted as mediators, both of which may mediate the positive correlation between "s_Veillonella rogosae" and TC. Enhancing the effectiveness of immunotherapy for TC by maintaining a favorable microbiota profile can enhance the clinical outcomes and the quality of survival sufferers with TC [26]. These findings underscore the potential for integrating gut microbiota modulation into clinical practice as an adjunctive approach to existing cancer therapies. Personalized dietary interventions, probiotics, or fecal microbiota transplantation could be tailored to individual patients to optimize their microbiota composition, thus potentially improving immune response and reducing adverse reactions. Additionally, understanding the specific immune cell phenotypes involved could aid in the development of targeted immunotherapies that harness the body's natural defenses more effectively. This approach opens new avenues for precision medicine, where treatment strategies are adapted based on a patient's microbiome and immune cell profile to enhance therapeutic success and minimize side effects.

Immune elimination and escape may partly rely on bacteria to shape immunity by mediating host immune regulation, and the mutual regulation of host-microbiome provides a novel therapeutic strategy to enhance the efficacy of anti-cancer treatment [27]. Dietary regulation of the intestinal microbiota directly influences the microbial metabolites produced in the intestinal mucosa and their impact on IC [28]. However, it is important to note that not all GM or IC species demonstrated significant associations with thyroid cancer in our MR analysis. This could be due to several factors, such as insufficient statistical power for certain less abundant microbial or immune cell types, potential confounding variables not accounted for, or the complex interplay of genetic and environmental factors that may mask specific relationships. Future studies should explore these null results in greater detail to understand if they indicate true non-associations or are influenced by limitations in data or methodology.

In the adaptive immune system, thyroid hormone may activate T lymphocytes through multiple potential mechanisms, including mediation of NF- κ B signaling pathways, as well as β -adrenergic receptors, resulting in increased T lymphocyte activation [29]. Our results suggest that "SSC-A on granulocyte" and "B cell % lymphocyte" as IC phenotypes may mediate s_Massiliomicrobiota, while "B cell % lymphocyte" may mediate f_Mycobacteriaceae, all exhibiting positive mediating effects on TC. CD11c+CD8+T cells, when activated, can potentially stop the development of autoimmune colitis through adoptive transfer; meanwhile, in specific viral and cancer models, they function as immune effectors, enhancing immune potential [30].

The intestinal microbiota is considered to be an important factor affecting thyroid homeostasis, and low abundance of Faecalibacterium may lead to GM dysbiosis before or after the development of TC [31]. The GM can interact with the

host's colon epithelial cells and IC by releasing a variety of metabolites, thereby regulating the development of colorectal cancer [32].

5 Limitation

This study utilized a large GWAS dataset, encompassing summary data from 473 GM, 731 IC, and TC. The MR analysis method not only identifies statistically significant findings but also guarantees strong statistical efficacy. Secondly, through MR analysis, this study identified 10 IC phenotypes as mediating factors in the causality from 9 types of GM to TC. However, this study has several limitations. The causality between GM, IC, and TC determined by MR analysis may be influenced by potential confounders, including environmental factors such as diet, lifestyle, and microbiome exposure, as well as genetic background, which add complexity to these relationships. Additionally, while MR analysis helps mitigate some biases present in observational studies, it cannot fully account for all unmeasured confounding variables or the intricacies of indirect pathways in the GM-IC-TC axis. The use of GWAS summary data limits the ability to explore interactions at an individual level and may introduce biases related to population-specific genetic structures. To address these challenges, future research should include experimental studies such as cell culture and animal models to validate the causal pathways suggested by MR analysis and provide deeper insight into the biological interactions among GM, IC, and thyroid cancer. Such approaches would enhance the understanding of these complex interactions and help identify potential confounding factors more precisely.

6 Conclusion

This Mendelian randomization study provided an in-depth exploration of the causal relationships between GM, IC, and TC. We identified nine GM species that demonstrated a causal association with TC, mediated by ten distinct IC phenotypes, including "Switched Memory AC," "IgD-CD38dim AC," and "CD28+DN (CD4-CD8-)." These findings illuminate the complex interplay between GM and IC in influencing tumor progression and suggest potential avenues for GM-based immunotherapies and IC-targeted treatments. While this study advances our understanding of the GM-IC-TC axis, further experimental validation is necessary. Future research should focus on in vitro and in vivo studies to corroborate these mediating effects and unravel the underlying molecular mechanisms. Such work could bridge the gap between statistical associations and biological causality, enabling the development of innovative therapeutic interventions.

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Declarations

Ethics approval and consent to participate Not applicable.

Consent for publication Not applicable.

Competing interests The authors declare no competing interests.

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