ORIGINAL ARTICLE [OPEN ACCESS](https://doi.org/10.1111/mec.17578)

Evolution of Size-Fecundity Relationship in Medaka Fish From Different Latitudes

Shingo Fujimoto^{1,2} | Bayu [K. A](https://orcid.org/0000-0001-8659-5698). Sumarto^{[2](#page-0-1)} | Iki Murase² | Daniel F. Mo[kod](https://orcid.org/0000-0002-5184-6554)ongan² | Taijun Myosho³ | Mitsuharu Yagi^{[4](#page-0-3)} | Satoshi Ansai^{[5,6](#page-0-4)} | Jun Kitano⁵ | Satoshi Takeda⁷ | Kazunori Yamahira^{[2](#page-0-1)} |

¹Integrated Technology Center, University of the Ryukyus, Okinawa, Japan | ²Tropical Biosphere Research Center, University of the Ryukyus, Okinawa, Japan | ³Laboratory of Molecular Reproductive Biology, Institute for Environmental Sciences, University of Shizuoka, Shizuoka, Japan | ⁴Graduate School of Fisheries and Environmental Sciences, Nagasaki University, Nagasaki, Japan | 5Department of Genomics and Evolutionary Biology, Ecological Genetics Laboratory, National Institute of Genetics, Mishima, Japan | ⁶Ushimado Marine Institute, Okayama University, Setouchi, Japan | 7Research Center for Marine Biology, Graduate School of Life Sciences, Tohoku University, Aomori, Japan

Correspondence: Shingo Fujimoto [\(fujimoto.s@outlook.com](mailto:fujimoto.s@outlook.com))

Received: 17 May 2024 | **Revised:** 26 September 2024 | **Accepted:** 9 October 2024

Handling Editor: Andrew DeWoody

Funding: This work was supported by Japan Society for the Promotion of Science (13J04092, 19K16232, 20570019, 23570031, 24K09610), University of the Ryukyus, Spatiotemporal Genomics Project. National Institute of Genetics, Cooperative Research Program (2013-A35).

Keywords: fecundity selection | life history adaptation | operational sex ratio | *Oryzias latipes* species complex | sexual selection

ABSTRACT

In most fishes, the number of offspring increases with maternal body size. Although this size-fecundity relationship often varies among species as a result of the coevolution of life-history traits, the genetic basis of such size-fecundity relationships remains unclear. We explored the genetic basis underlying this size-fecundity relationship in two small medaka species, *Oryzias latipes* and *O. sakaizumii*. Our findings showed that *O. sakaizumii* has a higher fecundity than *O. latipes*, and quantitative trait locus analysis using interspecific F_2 hybrids showed that chromosome 23 is linked to the size-fecundity relationship. In particular, the genes *igf1* and *lep-b* in this region are known to be associated with life-history traits, including somatic growth, gonad maturation, and progeny numbers in various taxa. Because *O. sakaizumii* is distributed at higher latitudes and has a shorter spawning season than *O. latipes* in the wild, we propose that the relatively high fecundity observed in *O. sakaizumii* is an adaptation to high latitudes. We also discuss the potential ecological ramifications associated with the evolution of increased fecundity in this species.

1 | Introduction

In numerous taxa, the number of offspring per reproductive event, thats is, fecundity, increases with larger body size in a population, for example, reptiles (Shine [2005\)](#page-12-0), insects (Berger, Walters, and Gotthard [2008](#page-10-0); Green and Extavour [2014](#page-11-0)), and fishes (Barneche et al. [2018](#page-10-1)). This relationship between size and fecundity has been shown to vary markedly among species (Bunger et al. [2005](#page-10-2); Orgogozo, Broman, and Stern [2006;](#page-12-1) Barneche et al. [2018](#page-10-1); Healy et al. [2019;](#page-11-1) Álvarez-Noriega et al. [2023](#page-10-3)). From an ontogenetic perspective, the amount of energy invested in reproduction changes throughout the development of an individual and is influenced by factors such as age and body size (Stearns [1992](#page-12-2); Kooijman [2010](#page-11-2); White et al. [2022](#page-13-0)). For example, ontogenetic changes in investment for reproduction are associated with physiological trade-offs in energy allocation between survival, growth, and reproduction (Kooijman [2010;](#page-11-2) Simmons, Lupold, and Fitzpatrick [2017;](#page-12-3) Healy et al. [2019;](#page-11-1) White et al. [2022\)](#page-13-0), all of which are subject to adaptive evolution in relation to life-history traits (Stearns [1992;](#page-12-2) Simmons, Lupold, and Fitzpatrick [2017](#page-12-3); White et al. [2022\)](#page-13-0). Hence, the ecological factors and selection pressures shaping

This is an open access article under the terms of the [Creative Commons Attribution](http://creativecommons.org/licenses/by/4.0/) License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

^{© 2024} The Author(s). *Molecular Ecology* published by John Wiley & Sons Ltd.

the size-fecundity relationships within species have been extensively studied (Pianka [1960;](#page-12-4) Healy et al. [2019](#page-11-1); White et al. [2022](#page-13-0)) to understand the coevolution of life-history traits between species (Charnov, Turner, and Winemiller [2001](#page-10-4); White et al. [2022;](#page-13-0) Álvarez-Noriega et al. [2023](#page-10-3)).

Investigations into the genetic mechanisms that affect fecundity have focused primarily on intraspecific variation in domesticated animals that have been reared to maximize agricultural productivity (see review by Bunger et al. [2005](#page-10-2)). For example, both quantitative trait locus (QTL) analysis and genome-wide association studies (GWAS) have been employed to elucidate the genetic basis of fecundity in chickens (Wright et al. [2010](#page-13-1); Wolc et al. [2014;](#page-13-2) Zhao et al. [2021](#page-13-3)), quails (Minvielle et al. [2005\)](#page-11-3), and salmonids (Sauvage et al. [2012;](#page-12-5) D'Ambrosio et al. [2020\)](#page-10-5). Some genetic analyses have targeted wild populations; for example, the heritability of fecundity has been examined in the great tit, *Parus major* using genome-wide nuclear genetic markers (Santure et al. [2013](#page-12-6)). In these studies, the heritability of fecundity ranged from low to moderate (0.14–0.42), as estimated by genome-wide nuclear genetic markers (Santure et al. [2013;](#page-12-6) Wolc et al. [2014;](#page-13-2) D'Ambrosio et al. [2020\)](#page-10-5). Single nucleotide polymorphisms (SNPs) contained in a single quantitative trait locus (QTL) explain only 1%–3% of the total genetic variance (e.g., D'Ambrosio et al. [2020\)](#page-10-5). Although several candidate genes influencing intraspecific variation in fecundity have been identified (Wright et al. [2010](#page-13-1); Swanson and Dantzer [2014;](#page-12-7) D'Ambrosio et al. [2020;](#page-10-5) Zhao et al. [2021\)](#page-13-3), it remains unclear whether these genes can also explain interspecific variation in size-fecundity relationships.

To explore the genetic basis of interspecific variation in sizefecundity relationships, we focused on medaka, or ricefish, in genus *Oryzias*, which is widely used as an animal model in genetics and molecular biology (Iwamatsu [2006;](#page-11-4) Kinoshita et al. [2009\)](#page-11-5). Medaka can be reared in the laboratory, which facilitates the genetic analysis of various phenotypes (e.g., Myosho et al. [2018;](#page-12-8) Ansai et al. [2021;](#page-10-6) Montenegro et al. [2022;](#page-12-9) Flury et al. [2022](#page-10-7); Hilgers et al. [2022\)](#page-11-6). In addition, medaka exhibit intraspecific variation in egg number (Leaf et al. [2011\)](#page-11-7) and interspecific variation in spawning interval (Montenegro et al. [2022;](#page-12-9) Hilgers et al. [2022](#page-11-6)). In particular, the two species found in the Japanese archipelago (*Oryzias latipes* and *O. sakaizumii*) have been observed to spawn up to 50 eggs upon reaching sexual maturity. These two species are suitable for quantitative analysis of size-fecundity relationships due to their similar adult size ranges, approximately 20–35mm in standard length (SL; e.g., Awaji and Hanyu [1987;](#page-10-8) Kawajiri et al. [2014\)](#page-11-8). A major empirical challenge in quantifying the size-fecundity relationship in fishes is that fecundity generally increases exponentially with size (White et al. [2022\)](#page-13-0), complicating the extrapolation and comparison of estimates between species with different size ranges. Thus, the similar adult size range of the two species makes them well-suited to genetic analyses of reproductive traits. Although the two species are genetically distinct, with an estimated 2.45% *p*-distance in genome-wide nuclear single nucleotide variants between reference sequences (Ichikawa et al. [2017\)](#page-11-9), they can be hybridized easily in captivity (Sakaizumi, Yasushi, and Satoshi [1992\)](#page-12-10). Consequently, quantitative trait locus analysis in $F₂$ individuals can be performed by using interspecific crosses (e.g., Tsuboko et al. [2014](#page-13-4); Kawajiri et al. [2014,](#page-11-8) [2015](#page-11-10); Yassumoto et al. [2020](#page-13-5)), which enables the analysis of the genetic basis of size-fecundity relationships in the two species.

We hypothesize that adaptation to environmental conditions drives the evolution of the size-fecundity relationships in the two species. *Oryzias latipes* and *O. sakaizumii* exhibit a parapatric distribution, in which *O. sakaizumii* tends to be distributed at higher latitudes than *O. latipes* in Japan (Figure [1\)](#page-2-0). This distribution suggests that *O. sakaizumii* has evolved life-history traits to cope with the harsher, more seasonal environments of higher latitudes. Common environment experiments have shown differences in growth, feeding activity, and maturation exist between these two medaka species (Yamahira and Takeshi [2008;](#page-13-6) Suzuki, Miyake, and Yamahira [2010;](#page-12-11) Fujimoto, Miyake, and Yamahira [2015](#page-10-9); Shinomiya et al. [2023\)](#page-12-12). Evidence obtained from seasonal fluctuations in gonad weight in wild populations (Awaji and Hanyu [1987](#page-10-8); Fujimoto et al. [2021](#page-10-10)) combined with evidence of genetic variation in photo-thermal responses to ovarian maturation (Shinomiya et al. [2023](#page-12-12)) suggests that constraints on physiologically suitable reproductive periods at elevated latitudes may impose severe selection pressures on life-history traits in *O. sakaizumii* relative to *O. latipes*. Consequently, natural selection may favor relatively high fecundity in *O. sakaizumii* compared to *O. latipes* in order to optimize lifetime reproductive success within shorter reproductive seasons (e.g., Pincheira-Donoso and Hunt [2017;](#page-12-13) Tarr et al. [2019](#page-12-14)).

However, no ecological studies have quantitatively compared the size-fecundity relationships and spawning season duration, that is, the duration of female reproduction, between wild populations of *O. latipes* and *O. sakaizumii*. In this study, we examined these demographic characters to determine whether *O. sakaizumii* demonstrates a propensity to produce more eggs than *O. latipes* in their natural habitats. Information on the reproductive seasonality and fecundity of wild populations will provide insights into the adaptive evolution of life history and the selection pressures acting on these wild populations.

In addition, to clarify the contribution of genetic variation to fecundity in these species, common-environment experiments were conducted using laboratory-reared offspring. We also conducted QTL analysis using interspecific $F₂$ hybrids to identify which chromosomal region is associated with interspecific variation in size-fecundity relationships. These findings provide a promising foundation for elucidating the genetic and physiological mechanisms underlying the adaptive evolution of the sizefecundity relationship.

2 | Materials and Methods

2.1 | Fish and Field Collection

Medaka, or the *Oryzias latipes* species complex, are small freshwater fish distributed in rice fields and low-lying wetlands throughout Japan, Korea, and China (Iwamatsu [2006;](#page-11-4) Figure [1a\)](#page-2-0). Molecular phylogenetic analyses have shown that the Japanese populations of the *O. latipes* species complex consist of two genetically distinct groups (Katsumura et al. [2019;](#page-11-11) Takehana et al. [2003,](#page-12-15) [2004\)](#page-12-16); the 'Northern Japan group' or *O. sakaizumii*, which is distributed along the coast of the Sea

FIGURE 1 | (a) Distribution range of *Oryzias latipes* (solid line) and *O. sakaizumii* (dotted line) in Japan (Takehana et al. [2003](#page-12-15); Katsumura et al. [2019\)](#page-11-11). Solid and open circles on the map represent the geographic locations of *O. latipes* and *O. sakaizumii* collection sites, respectively. Photographs of (b) *O. sakaizumii* female, (c) *O. sakaizumii* male, (d) *O. latipes* female, and (e) *O. latipes* male. (f, g) Egg spawning behavior during the mating sequence of *O. sakaizumii*. All photographs were taken by Shingo Fujimoto.

of Japan (Asai, Senou, and Hosoya [2011;](#page-10-11) Figure [1b,c](#page-2-0)), and the 'Southern Japan group' or *O. latipes*, which is distributed along the coasts of the Pacific and the East China Sea (Figure [1d,e\)](#page-2-0). The original map was generated using Generic Mapping Tools (Wessel et al. [2013\)](#page-13-7).

In the laboratory, medaka reach sexual maturity at about 3months of age (e.g., Kawajiri et al. [2014\)](#page-11-8). The median life span of *O. latipes* under laboratory conditions is approximately 22months (Ding et al. [2010](#page-10-12)). Both species have an ecological longevity of less than 2years (e.g., Awaji and Hanyu [1987;](#page-10-8) Egami, Terao, and Iwao [1988;](#page-10-13) Fujimoto et al. [2021\)](#page-10-10). Ovarian maturation is triggered by warmer temperatures and longer photoperiod (Koger et al. [1999;](#page-11-12) Shinomiya et al. [2023](#page-12-12)). Consequently, in the wild, the reproductive season is limited to the warm season, with gonad weight increasing from April to August (Awaji and Hanyu [1987](#page-10-8); Fujimoto et al. [2021\)](#page-10-10). During the reproductive season, individuals produce 10–30 eggs almost daily. Mating typically occurs between one male and one female in the water column (Figure [1f\)](#page-2-0). After the female releases the eggs, the male fertilizes the eggs externally while they are attached to the female's genital pore (Figure [1g\)](#page-2-0).

In this study, we collected individuals from two wild medaka populations: *O. sakaizumii* were collected from Aomori (40°50′N, 140°49′E) and *O. latipes* from Okinawa (26°49′N, 128°16′E) (Figure [1a](#page-2-0)). Aomori and Okinawa are located near the northern and southern distribution limits of *O. sakaizumii* and *O. latipes*, respectively (Iwamatsu [2006\)](#page-11-4). Fish from each population were transferred to our laboratory, where they were housed in acrylic tanks for use as stock populations and for QTL analysis.

Throughout the laboratory rearing experiment described below, the water temperature of the tanks was maintained at 26°C (±0.1°C) and the photoperiod was maintained at 14 L:10D. Fish were fed daily on a diet consisting of newly hatched (24-h-old) nauplii of *Artemia franciscana* (Argent Chemical Laboratories, Redmond, WA, USA) and dry feed (Fancy Guppy, Hikari Tropical, Kyorin Corporation, Tokyo, Japan).

2.2 | Spawning Season Assessment in the Wild

To examine spawning season and size-fecundity relationships in the wild, we collected wild individuals from Aomori city (40°49′N, 140°48′E), and Kunigami village, Okinawa Island (26°49′N, 128°16′E) (Figure [1a\)](#page-2-0). The collections in Aomori were conducted at 2-week intervals between April and August in 2013. Similarly, collection in Okinawa were mainly conducted between March and October 2012. To confirm the non-spawning season in Okinawa, additional collection was performed in December 2012 and February 2013. Moreover, to assess the possible role of two large typhoons that passed over Okinawa Island in 2012 on the termination of medaka spawning behavior, we conducted additional collections in Okinawa between August and October in 2013. We collected 62–1472 individuals per collection effort (Table [S1](#page-13-8)). The collected fish were sexed based on sexually dimorphic characters (e.g., Kawajiri et al. [2014](#page-11-8)), and the adult sex ratio (adult males/adult males and females) in each collection was determined (Aomori: 0.44 ± 0.06 , Okinawa: 0.51 ± 0.05 , mean \pm SD, Table [S1\)](#page-13-8).

Following collection, we randomly selected 20 adult females (i.e., >20mm SL to the nearest 1mm using a ruler) and 20 adult males and transported them to a laboratory to evaluate seasonal changes in the proportion of sexually mature males and females. The following trials were repeated immediately after each field collection (Figure [S1](#page-13-8)). To evaluate the reproductive ability of

wild males and females, mating trials were conducted using either a single focal female or male paired with stock males and females, respectively. The stock individuals used in the mating trials consisted of wild males collected in Okinawa (verified in a preliminary survey) and laboratory-reared offspring of females from Aomori (Figure [S1](#page-13-8)). Using fertile stock individuals, we were able to independently assess the reproductive ability of wild individuals in both sexes.

We estimated the ratios of sexually mature individuals using a generalized additive model (GAM, Wood [2006\)](#page-13-9). The GAM was estimated using the "gam" function in the "mgcv" package in the R program (Wood [2011](#page-13-10)). The outcome of the mating experiment for individual males (fertilization success/failure) and females (no clutch, one clutch, or two clutches) was treated as the response variable with a binomial distribution. We assumed a seasonal trend and fitted a smoothed curve between the collection dates, considering sex, population, and their interactions as explanatory variables. The significance of each term in the GAM was evaluated using a likelihood-ratio test, with each term systematically dropped to assess its relative contribution (Tables [S2](#page-13-8) and [S3\)](#page-13-8). Upon confirming the significance of each term, we selected the best-fitting model (Model 5, Table [S3](#page-13-8)) to represent the seasonal variation in the proportion of mature individuals.

During the mating trial, the experimental fish were housed within a polypropylene container (160×120×80mm) immersed into larger polypropylene tanks (900×620×210mm, water depth 60mm). The container had two holes (30mm diameter) on each side and was covered with a dip-net (150×125×70mm, 32-μm mesh), so that each pair spawned inside the dip-net. Water in the tanks was circulated with a submersible pump and maintained at 26° C \pm 1°C using immersion heaters. Each pair was allowed to mate freely, and the following morning (ca. 10:00AM–12:00PM), we checked for the presence of fertilized eggs, and fish were paired in this way for two consecutive days. After the mating trials, each wild fish was photographed and SL was measured based on these photographs (Optio x90, RICOH, Tokyo, Japan). The wild fish were then released at the collection site. Considering the habitat area and the population density, some of the individuals may have been recaptured and measured.

For the statistical analysis of size-fecundity relationships in wild individuals, we used data from 71 spawned females from Aomori and 188 spawned females from Okinawa. We constructed a linear model to compare the size-fecundity relationship between the two populations. Egg number was treated as the response variable, while female SL, population, and their interaction were treated as the explanatory variables. The significance of the explanatory variables was estimated using an *F* test. All statistical analyses were performed in R ver. 4.1.1 (R Core Team [2021\)](#page-12-17). The linear model analysis was conducted using the "lm" function implemented in the MASS package. Type 3 ANOVA was conducted employing the "Anova" function in the "car" package (Fox and Weisberg [2019](#page-10-14)).

2.3 | Establishment of F₂ Hybrids for QTL Analysis

We reared F_2 hybrid individuals from previous studies (Fujimoto et al. [2014;](#page-10-15) Kawajiri et al. [2014,](#page-11-8) [2015\)](#page-11-10), which we used

for QTL mapping in this study. In May 2011, we collected 34 adults from Aomori and 50 from Okinawa, with a sex ratio of approximately 1:1. We then established two $F₂$ progeny lines: $F₂$ progeny from $F₁$ hybrids between an Aomori female and an Okinawa male (AFOM), and F_2 progeny from F_1 hybrids between an Okinawa female and an Aomori male (OFAM) (Figure [S1\)](#page-13-8). One male from one population and one female from the other population were randomly selected from the wild fish stock kept in the laboratory. The pair was maintained in a small polypropylene container $(15\times11\times8$ cm) placed within an acrylic tank ($75 \times 60 \times 45$ cm). Fertilized eggs were obtained from the mated pair and the resulting F_1 individuals were transferred to small acrylic containers (25×20×23 cm) immersed in the larger acrylic tanks. These F_1 individuals were then reared in the same containers. Following maturation, the F₁ individuals were allowed to mate freely, and newly fertilized eggs of F_2 individuals were collected.

2.4 | Phenotypic Measurements

We performed mating experiments to measure the egg numbers and mating behaviors in F_2 individuals for QTL analysis. The sex of each $F₂$ individual was determined at 124 days postfertilization, after which each F_2 individual was assigned to the mating trials described below. In total, we analyzed 92 OFAM females, 78 OFAM males, 87 AFOM females, and 101 AFOM males. $F₂$ females were mated with wild stock males from Okinawa and F_2 males were mated with laboratory-reared offspring females from Aomori. These stock individuals were the same as those used in the mating trials with wild populations (Figure [S1\)](#page-13-8).

One day before each mating trial, we randomly selected one mating partner from the stock fish in the laboratory. The F_2 individual and a mating partner were placed in a small acrylic tank $(14 \times 7.5 \times 15 \text{ cm}, 12 \text{-cm}$ water depth), which was immersed in a temperature-controlled (26 $^{\circ}$ C) acrylic tank (75 \times 60 \times 45 cm). The female and male were separated by an opaque partition. The following morning at approximately 9:30–10:00AM, the trial was initiated by removing the partition, and the male and female were allowed to mate freely in the tank. We recorded their behavior for 1h using a digital video camera (HDR-CX180, Sony Corporation, Tokyo, Japan). After the trial was completed, the number of eggs was counted for the $F₂$ females at approximately 17:00–18:00PM in the evening. The mating trial was repeated twice for each $F₂$ female and male for 2 days in succession. In the event that the F_2 female spawned clutches on both trial days, the average number of eggs was calculated. As a result, we obtained egg number data for 65 OFAM females and 60 AFOM females.

In medaka fishes, factors such as age, nutrient condition, and ambient temperature have a marked effect on egg production (e.g., Leaf et al. [2011](#page-11-7); Hemmer-Brepson et al. [2014\)](#page-11-13). To exclude these environmental and demographic factors between wild populations, a common environment experiment was conducted, and the egg numbers of laboratory-reared offspring females were measured in 2021. The number of eggs produced by 17 Okinawa females and 21 Aomori females was measured in a manner similar to that described for F_2 individuals.

We also analyzed the mating behaviors of the $F₂$ females using the video recordings. In addition to the number of courtship behaviors observed until spawning, the time interval between the time of first courtship and successful egg spawning was recorded as spawning latency, as this is an indicator of female mate preference in medaka (e.g., Fujimoto et al. [2014;](#page-10-15) Ogino et al. [2023](#page-12-18)). Wrapping rejections (i.e., when a female disengages from male wrapping without releasing eggs) were also counted and used as an index of female mate preference. In the mating experiment of the F₂ females, we used wild Okinawa males as mating partners.

The mating behaviors of $F₂$ males were also examined. To evaluate the sexual motivation of $F₂$ males, courtship latency, that is, the time interval between the removal of the separator and the initiation of courtship, frequency of approaches, and frequency of quick circles, that is, a male courtship behavior in which males approach females from below and then swim rapidly in a circle around the female (Ono and Uematsu [1957\)](#page-12-19), were measured. For F_2 male phenotypic measurements, we used laboratory-reared Aomori females as mating partners. $F₂$ males of pairs that did not spawn within 1h were not used for subsequent QTL analysis, because male sexual motivation may have been affected by the reproductive condition of the mating partner (e.g., experimental females may not have produced mature eggs). We obtained courtship behaviors for 52 OFAM males and 80 AFOM males. These behavioral data of $F₂$ males were originally published in Fujimoto et al. [\(2014\)](#page-10-15).

2.5 | Genotyping and QTL Mapping

For genotyping the F_2 progeny, we used a custom VeraCode GoldenGate Genotyping Assay with 384 SNP panels (Illumina, San Diego, CA, USA) for Japanese medaka (Kawajiri et al. [2014\)](#page-11-8). The linkage map of the OFAM family and the AFOM family was constructed in a previous study (Kawajiri et al. [2014,](#page-11-8) [2015](#page-11-10)). To assess the relationship between QTL for the number of eggs laid and observations of mating behavior, we employed multiple QTL mapping with the R/qtl package (Broman and Sen [2009;](#page-10-16) Arends et al. [2010](#page-10-17)). The false discovery rate (FDR) of logarithm of the odds (LOD) scores among peaks in multiple chromosomes were determined with 1000 bootstrap permutations (significant: $FDR < 0.05$, suggestive: $0.05 < FDR < 0.10$). R/qtl was also used to compute 95% Bayesian credible intervals (95% BI) for QTL peak positions. QTL analyses for each phenotype were performed separately, with datasets treated by family and sex (Table [S4\)](#page-13-8).

In addition, the proportion of variance explained (PVE) for each phenotype was calculated to estimate the genetic variance (Table [S4\)](#page-13-8). PVE was calculated in three ways: (a) an estimate based on the covariance matrix between individuals using all genome-wide genetic markers, (b) an estimate using only markers on the chromosome where the QTL peak was detected, and (c) an estimate using the genetic marker closest to the QTL peak. Genome-wide PVE was calculated using the "mmer" function implemented in the "sommer" package (Covarrubias-Pazaran [2016](#page-10-18)). To minimize the risk of false positives in the QTL analysis, phenotypes with low genetic variance (genomewide PVE<1%) were excluded from the results, even if QTL peaks were detected (Table [S4](#page-13-8)).

We investigated whether the QTL for egg number affected the allometric relationship between body size and egg number. The nearest genetic marker to the peak LOD score was selected for QTL mapping of egg number. Subsequently, a linear model was employed, followed by an ANOVA to assess the significance of the genetic marker, body size, and their interaction. A type 3 ANOVA was executed using the "Anova" function in the "car" package to mitigate compounding effects arising from the order of the terms (Fox and Weisberg [2019\)](#page-10-14).

2.6 | Gene Ontology Analysis

To explore the functional characteristics of genes located within the identified QTLs, we conducted gene ontology (GO) analysis. Gene names and GO terms annotations were downloaded from the Ensembl release 109 [\(https://feb2023.archive.ensembl.](https://feb2023.archive.ensembl.org/) [org/\)](https://feb2023.archive.ensembl.org/) for the *O. latipes* HdrR genome database (ASM223467v1, Ichikawa et al. [2017](#page-11-9)). Given that the reference genome of the HdrR strain in *O. latipes* (ASM223467v1) was better annotated than the HNI strain in *O. sakaizumii* (ASM223471v1), we chose to use the HdrR genome for our analysis. For example, the HNI strain lacks an extensive ortholog database, and gene symbols were undetermined in for most Ensembl gene IDs, whereas this information is available for the HdrR strain of *O. latipes*. We performed GO enrichment analysis using gene annotation information from the HdrR genome and ShinyGO ver. 0.80 (Ge, Jung, and Yao [2020\)](#page-11-14). Enrichment analysis was performed separately for Ensembl gene IDs located within each QTL. Statistical significance of the enrichment was determined as $FDR < 10^{-5}$. We examined the top 10 GO terms ranked by the FDR of fold enrichment. In addition, we also identified genes associated with GO terms such as "reproduction", "oogenesis", or "behavior" using a custom script and the "biomaRt" package (Durinck et al. [2005\)](#page-10-19).

The original genetic markers used for QTL analysis (Kawajiri et al. [2014,](#page-11-8) [2015\)](#page-11-10) were based on an earlier version of the reference genome of the HdrR strain in *O. latipes* (ASM31367v1, Kasahara et al. [2007\)](#page-11-15). Subsequently, the genomic positions of these genetic markers were lifted from the older reference (ASM31367v1) to the newer version (ASM223467v1) using the LiftOver software package (Hinrichs et al. [2006\)](#page-11-16); we performed this liftover using "flo" ([https://github.com/wurmlab/flo/,](https://github.com/wurmlab/flo/) Pracana et al. [2017\)](#page-12-20), a pipeline that employs the UCSC tools (Kent [2002\)](#page-11-17).

3 | Results

3.1 | Interspecific Variation in Fecundity

Compared to the wild Okinawa population, Aomori females tended to produce more eggs in each clutch (Figure [2a](#page-5-0)). The results of the linear model analysis showed that the SL of females was positively correlated with the average number of eggs per clutch (SL; Estimate \pm SE = 1.13 \pm 0.15, *df* = 1, *F* = 59.58, *p* < 0.0001), and that the Aomori population tended to produce more eggs than the Okinawa population (population; Estimate \pm SE = 11.1 \pm 3.87, df = 1, F = 8.28, p = 0.004). Similarly, the interaction between SL and population was also

FIGURE 2 | (a) Size-fecundity relationship of *Oryzias latipes* (Okinawa), and *O. sakaizumii* (Aomori) females in the wild. (b) Average egg number in laboratory-reared offspring females from Okinawa and in Aomori. (c) Seasonal change in the proportion of sexually mature individuals during 2-day observation period in Aomori and (d) in Okinawa (circles: females, triangles: males). Solid and dashed lines (estimated using a generalized additive model) represent males and females, respectively.

significant (population \times SL; Estimate \pm SE = -0.51 \pm 0.15, $df=1$, $F=12.14$, $p=0.0006$), suggesting that the increase in the number of eggs produced by females from Aomori increased at a higher rate with body size compared to females from Okinawa (Figure [2a\)](#page-5-0).

A significantly larger proportion of females from Aomori spawned two clutches compared to females from Okinawa (Fisher's exact test, $p < 0.001$, Figure [S2\)](#page-13-8); approximately 40% of females from Aomori produced two clutches during the two consecutive days of the experiment, but only 10% of Okinawa females produced two clutches (Figure [S2](#page-13-8)). The results showed that the Aomori population has a higher fecundity than the Okinawa population in the wild.

A difference in the average number of eggs was also observed in lab-reared females (Figure [2b](#page-5-0)), indicating that differences in average egg number observed between wild populations persisted when environmental and demographic factors were excluded. When SL was considered in the linear model, the variance in egg number was observed between Aomori and Okinawa (Estimate ± SE=42.14±15.48, *df*=1, *F*=7.41, *p*=0.0102). In this experiment, laboratory reared offspring females did not show a significant difference in the mean SL between females from Aomori and Okinawa (*t*=−1.56, *df*=32.20, *p*=0.1286). The mean SL was 25.0 ± 1.2 mm in the Aomori population ($N=17$) and 24.4 ± 1.4 mm in the Okinawa population (mean \pm SD, N =21). The trend in female SL was consistently observed in wild populations assessed for reproductive ability $(t=-0.46, df=237.3, p=0.65)$. The mean SL was 25.2 ± 3.0 mm in the Aomori population ($N=157$) and 24.4 ± 1.4 mm was in the Okinawa population ($N=437$).

3.2 | Spawning Season in the Wild

The spawning season of the Aomori population was shorter than the Okinawa population (Figure [2c,d;](#page-5-0) Table [S2\)](#page-13-8). Wild fish from Aomori spawned from May to July, whereas those in Okinawa spawned from March to October (Figure [2c,d;](#page-5-0) Table [S2](#page-13-8)). A higher proportion of females matured during the peak of the spawning season in the Aomori population (June, 80%) compared to the Okinawa population (May–September, 50%–60%) (Figure $2c,d$). The seasonal trend in the proportion of mature individuals differed significantly, not only be-tween populations (Table [S2,](#page-13-8) Model 2 vs. Model 3: $df = 5.3$, $deviance = 141.0, p < 0.0001$, but also between sexes and their interactions (Model 3 vs. Model 4: *df* = 9.4, *deviance* = 92.7, *p* < 0.0001; Model 4 vs. Model 5: *df* = 1.2, *deviance* = 283.0, *p* < 0.0001). Almost all of the males from both populations were sexually mature at the peak of the female spawning season (Figure [2c,d](#page-5-0)).

3.3 | QTLs Associated With Phenotype

Six QTLs showed associations with the observed phenotypes (Table [1](#page-6-0); Table [S4](#page-13-8)) and three QTLs showed statistically significant (Table [1;](#page-6-0) Figure [3a–d\)](#page-7-0). No significant and suggestive

a0.05<FDR<0.10. *FDR<0.05. QTLs were shared between the AFOM and OFAM strains (Figure [3a–d](#page-7-0); Table [1](#page-6-0); Table [S4](#page-13-8); Figure [S3](#page-13-8)). The QTLs associated with egg number were located on chromosome 23 in the OFAM strain (Table [1](#page-6-0); Figure [3b\)](#page-7-0). Analysis revealed that the average egg number was higher in alleles of the Aomori genotype compared to those of Okinawa at the nearest genetic marker of the peak LOD score (Figure 3e, f).

Furthermore, we also assessed the influence of QTLs on the relationship between size and fecundity using a linear model ($F_{5,59} = 8.11$, adj $R² = 0.36$, $p = 7.07e⁻⁰⁶$). The results showed that egg number increased in individuals with larger body size possessing the Aomori homozygous genotype $(N=10, 10)$ slope $=$ 4.92, intercept $=$ -106.73) and heterozygous genotype $(N=42, \text{ slope}=1.55, \text{ intercept}=-25.25)$, whereas the slope of egg number was significantly smaller in individuals with the Okinawa homozygous genotype $(N=13, \text{ slope} = -0.10,$ intercept = 13.75) (Figure $3f$). In the linear model, the main effect of SL was statistically significant $(df=1, F_{1,59} = 8.83,$ *p* = 0.004) and the interaction between SL and genotype was also significant ($df = 2$, $F = 3.20$, $p = 0.048$), whereas the main effect of genotype was not statistically significant $(df=2, F_{2,59} = 2.41$, p = 0.099). To confirm that the detected egg number QTL had a statistically significant effect on the size-fecundity relation ship, we also examined the distribution of slope differences in the size-fecundity relationship using all genetic markers. Two genetic markers (OL_23_19103934, OL_23_20828757) located within the egg number QTL were among the top 5% (13/273 markers) in the distribution, supporting the finding that the egg number QTL on chromosome 23 affected the sizefecundity relationship.

Suggestive QTLs related to the female behaviors, specifi cally spawning latency and wrapping rejection, were identi - fied on chromosomes [1](#page-6-0)3, 18, and 24 (Table 1, Table [S4\)](#page-13-8). For both QTL 13 and 18, individuals with Aomori alleles tended to take longer to spawn than those with the Okinawa alleles (Figure [S4e,f](#page-13-8)). Similarly, QTL 24, individuals with Aomori al leles exhibited a higher frequency of rejection behavior com pared to those with Okinawa alleles (Figure [S4c\)](#page-13-8). In AFOM males, a significant QTL associated with SL was mapped to chromosome 12 (Table [1,](#page-6-0) Figure [3a](#page-7-0)). The frequency of approaching, which are indicator of male sexual motivation, demonstrated an association with chromosome 20 (Table [1,](#page-6-0) Figure [3d](#page-7-0)). Individuals with Aomori alleles exhibited a higher frequency of courtship behavior compared to those with Okinawa alleles (Figure [S4b\)](#page-13-8).

We performed GO enrichment analysis for each of the six QTLs (Table [1](#page-6-0); Table [S5](#page-13-8)). The significant enrichment was observed QTL of wrapping rejection in chromosome 24, including the term "Chemokine binding" (Table [S5](#page-13-8), Fold Enrichment = 52.97 , FDR < $1e^{-5}$). No significant enrichment was observed for the other QTLs (Table [S5](#page-13-8)). The GO terms with the lowest enrichment FDR for each QTL are listed below: for the QTL associated with SL, "response to heat" (Fold Enrichment = 15.64, FDR = 0.0023); spawning latency in chromosome 13, "G protein-coupled receptor activity" (Fold Enrichment = 4.68, FDR = 0.0019); spawning latency in chromosome 18, "microtubule associated complex" (Fold Enrichment = 9.94 , $FDR = 0.1784$; courtship frequency,

FIGURE 3 | Quantitative trait locus analysis of the F_2 hybrids between Okinawa and Aomori. (a, c) Logarithm of the odds (LOD) score for the standard length, (b) average egg number, (d) frequency of approaching. Lines show significance levels of false discovery rate (FDR) by genomewide permutation tests with multiple peaks (solid line: FDR <0.05, dotted line: 0.05 <FDR <0.10). (e) Average egg number for genotypes of single nucleotide polymorphism (SNP) marker OL_C23_19103934, which had the highest LOD score among the markers. (f) Size-fecundity relationship for genotypes of the SNP marker OL_C23_19103934. Strain refers to F_2 progeny derived from F_1 hybrids between an Aomori female and an Okinawa male (AFOM, red lines), or F_2 progeny derived from F_1 hybrids between an Okinawa female and an Aomori male (OFAM, blue lines).

"regulation of B cell proliferation" (Fold Enrichment = 271.26, $FDR = 0.0025$; and egg number, "glycoprotein metabolic process" (Fold Enrichment = 9.44 , FDR = 0.0475).

In addition, to identify the candidate genes within the three significant QTL regions falling within the 95% Bayesian credible intervals (Table [1](#page-6-0)), we conducted a search for genes associated with "reproduction", "oogenesis", and "behavior" utilizing the GO database (ASM223467v1, Ensembl 109, [https://feb2023.](https://feb2023.archive.ensembl.org) [archive.ensembl.org](https://feb2023.archive.ensembl.org)). QTLs located on chromosomes 20 and 23 were found to harbor three genes related to "reproduction", namely THAP Domain Containing 6 (*THAP6*), and insulinlike growth factor 1 gene (*igf1*) (Table [S6\)](#page-13-8). Notably, the QTL region on chromosome 23 contained *igf1*, as well as the leptin gene (*lep-b*), a gene potentially related to reproduction; however, this information was not available in the GO database for ASM223467v1. Similarly, one gene related to "behavior", Pappalysin 1 (*pappaa*), were identified on chromosomes 12 (Table [S6\)](#page-13-8).

4 | Discussion

4.1 | Fecundity Increases at Higher Latitudes

Species distributed at high latitudes tend to have higher fecundity, even if they are of the same size (Pincheira-Donoso and Hunt [2017](#page-12-13); Sparks et al. 2022, Álvarez-Noriega et al. [2023;](#page-10-3) Figure [2\)](#page-5-0). This tendency is not limited to fishes (Winemiller and Rose [1992;](#page-13-11) Vila-Gispert, Moreno-Amich, and Garcia-Berthou [2002](#page-13-12); Álvarez-Noriega et al. [2023](#page-10-3)), and is found in a variety of taxa (Pincheira-Donoso and Hunt [2017](#page-12-13), Tarr et al. [2019,](#page-12-14) Sparks et al. [2022\)](#page-12-21). In addition, numerous high-latitude fish species have been reported to increase the number of eggs that they produce in a manner that is disproportionate to body size (Álvarez-Noriega et al. [2023](#page-10-3)). Geographic variation in fecundity is considered to reflect an adaptive advantage to higher latitude environments, for example, higher mortality and/or larger temporal change of food availability in more seasonal environments (i.e., higher latitudes) (Pincheira-Donoso and Hunt [2017;](#page-12-13) Tarr et al. [2019](#page-12-14); Álvarez-Noriega et al. [2023](#page-10-3)). However, unlike studies on fruit flies, numerous studies on wild animals encounter challenges in distinguishing between the relative contributions of genetic and environmental variations (Bunger et al. [2005;](#page-10-2) Green and Extavour [2014\)](#page-11-0). Our study; however, demonstrates the existence of interspecific variation in the size-fecundity relationship and suggests a potential genetic basis in medaka (Figures [2](#page-5-0) and [3](#page-7-0)). Specifically, *O. sakaizumii* females from Aomori exhibit genetically higher egg production compared to their congener, *O. latipes* from Okinawa (Figures [2b](#page-5-0) and [3](#page-7-0)). This system provides a valuable foundation for clarifying the genetic and physiological mechanisms underlying the adaptive evolution of life-history traits.

The Aomori population exhibits a markedly shorter spawning season compared to the Okinawa population in their natural habitat (Figure $2c,d$). At higher latitudes, where the period suitable for reproduction tends to be short, natural selection favors life-history strategies that maximize the number of offspring per reproductive event (Conover and Schultz [1995;](#page-10-20) Kokita [2004;](#page-11-18) Pincheira-Donoso and Hunt [2017](#page-12-13); Tarr et al. [2019\)](#page-12-14). *O. sakaizumii* occupies one of the highest latitudinal ranges in the genus *Oryzias* (e.g., Sumarto et al. [2020](#page-12-22)). Consistent with the paradigm of adaptive evolution at higher latitudes, *O. sakaizumii* may have undergone evolutionary changes towards increased fecundity as this species expanded its northernmost range (Takehana et al. [2003;](#page-12-15) Katsumura et al. [2019\)](#page-11-11).

The concepts of thermal plasticity in gonad maturity provides support for the adaptive evolution of reproductive physiology in medaka fishes. Following sexual maturation in females, lower water temperature plastically suppresses gonad maturation and reduces egg production in both *O. latipes* (Hirshfield [1980;](#page-11-19) Koger et al. [1999;](#page-11-12) Hemmer-Brepson et al. [2014](#page-11-13)) and *O. sakaizumii* (Shinomiya et al. [2023](#page-12-12)), as demonstrated in temperaturecontrolled experiments. In addition, habitat temperatures are typically lower at high latitudes, for example, the average water temperature in Aomori during the spawning season is 15.7°C±4.3°C (from May 1, 2013 to July 26, 2013), and that of Okinawa is 22.3° C \pm 3.4°C (from March 8, 2012 to October 15, 2012) (see also Yamahira et al. [2007](#page-13-13); Shinomiya et al. [2023\)](#page-12-12). Consequently, the observation that Aomori females produce more eggs than Okinawa females in their natural environ-ment (Figure [2a](#page-5-0)), contrary to the pattern anticipated from thermal plasticity, suggests adaptive evolution in fecundity. Such counter-gradient variation in fecundity traits has been reported in latitudinal populations of several fishes and birds (e.g., Conover and Schultz [1995](#page-10-20), Kokita [2004](#page-11-18); Pincheira-Donoso and Hunt [2017](#page-12-13); Sparks et al. [2022](#page-12-21)).

4.2 | Genetic Basis of the Size-Fecundity Relationship

From an individual perspective, ontogenetic changes or plasticity in reproductive investment represent critical factors in enhancing an individual's lifetime reproductive success (Kooijman [2010;](#page-11-2) Simmons, Lupold, and Fitzpatrick [2017;](#page-12-3) White et al. [2022](#page-13-0)). The regulation of egg production through nutrient metabolism may constitute one of the physiological mechanisms underlying ontogenetic changes in reproductive investment (Copeland et al. [2011;](#page-10-21) Dantzer and Swanson [2012](#page-10-22)). For example, in *Drosophila* species, *igf1* expression plastically alters ovariole number in response to food availability (Green and Extavour [2014\)](#page-11-0). Furthermore, a GWAS study conducted in chickens showed that *igf1* exhibits pleiotropic effects on both body weight and egg production (Wolc et al. [2014](#page-13-2)). Interestingly, a transgenic medaka strain that overexpress growth hormone and *igf1* shows faster somatic growth, but lower fecundity compared to the wild type (Komine et al. [2016\)](#page-11-20). In our study, the QTL for egg number on chromosome 23 includes both *igf1* and *lep-b* (Table [1;](#page-6-0) Table [S4\)](#page-13-8). Both IGF signaling genes and leptin are associated with somatic growth and gonad development in vertebrates (Copeland et al. [2011](#page-10-21); Dantzer and Swanson [2012;](#page-10-22) Chisada et al. [2014;](#page-10-23) Ndandala et al. [2022\)](#page-12-23), suggesting that these genes may influence interspecific variations in the size-fecundity relationship.

In this study, our QTL analysis focused on evaluating egg numbers in mating trials. If spawning fails to occur due to factors such as sexual motivation or mate preference in either of the sexes (e.g., Yokoi et al. [2020;](#page-13-14) Daimon et al. [2022;](#page-10-24) Kondo et al. [2023](#page-11-21)), then the detection of QTLs for egg number may potentially be biased by sexual motivation. However, the QTLs for egg number do not overlap with the QTL peaks associated with female spawning latency or male courtship frequency (Table [1;](#page-6-0) Table [S4;](#page-13-8) Figure [3\)](#page-7-0). These findings suggest that the observed QTLs for egg numbers are unlikely statistical confounders related to the sexual motivation of $F₂$ individuals.

In addition, the peaks of the six QTLs were not consistent be-tween the AFOM and OFAM families (Table [S4;](#page-13-8) Figure [S3\)](#page-13-8), which may be due to genetic variation within the population. Since wild individuals were used as grandparents in the QTL analysis, individual differences in genetic background may exist between the two families. All four of the QTLs of associated with mating behaviors tended to show effects opposite to those expected from phenotypic differences in wild populations reported in previous studies (Figure [S4,](#page-13-8) e.g., Kagawa [2014,](#page-11-22) Fujimoto, Miyake, and Yamahira [2015\)](#page-10-9), further suggesting genetic variation within wild populations. Moreover, genetic studies in medaka reported that sex-specific alleles, possibly related to sex determination, are distributed across multiple chromosomes (Shinomiya et al. [2010;](#page-12-24) Kitano et al. [2023;](#page-11-23) Shinya, Kimura, and Naruse [2023](#page-12-25)), indicating that males and females of the same population might carry different alleles at these loci. Given the genetic variation within wild populations, further validation is necessary to more comprehensively determine whether the QTLs detected in this study affect phenotypes in a sex- and species-specific manner. This will require enhanced research methods, such as breeding designs with replicates or genome-wide association studies using improved genetic markers in the wild populations.

In this study, the PVE for egg number calculated with genomewide markers was 16.2% and 28.2% for OFAM and AFOM, respectively (Table [S4\)](#page-13-8). The PVE values were similar to the heritability of 0.24 obtained in a genome-wide association study of salmonids using 57,000 SNPs (D'Ambrosio et al. [2020](#page-10-5)). QTL and GWAS studies conducted in domesticated animals have revealed that size and fecundity are often regulated by distinct chromosomal regions (Bunger et al. [2005;](#page-10-2) Minvielle et al. [2005;](#page-11-3) Wright et al. [2010](#page-13-1); Wolc et al. [2014](#page-13-2); D'Ambrosio et al. [2020\)](#page-10-5). Similarly, in medaka, numerous QTLs associated with body size do not overlap with QTLs for egg number. For instance, the QTL for body weight is located on chromosome 4 (Yassumoto et al. [2020\)](#page-13-5), and the QTLs for SL are located on chromosomes 12 and 24 (Table [1](#page-6-0); Kawajiri et al. [2014](#page-11-8)). Such a genetic architecture, characterized by multiple genetically independent chromosomal regions, may facilitate the adaptive evolution of the size-fecundity relationship.

4.3 | Relationship Between Fecundity Evolution and Proportion of Spawning Females

We observed differences in the proportion of spawning fe-males among wild populations (Figure [2c,d;](#page-5-0) Figure [S2](#page-13-8)). This difference in spawning among females in the wild is primarily attributed to the higher frequency of spawning in the Aomori females compared to the Okinawa females (Figure [S2](#page-13-8)), resulting in a less male-biased operational sex ratio (OSR) in the Aomori population (i.e., the number of spawned females to the total number of adult males and females). Since the numbers of adult males and females being nearly equal in both populations (Table $S1$), the proportion of spawning females significantly influences the bias in OSR. On average, the Aomori population tends to exhibit a less male-biased OSR $(0.65 \pm 0.13;$ Figure [2c](#page-5-0)) compared to the Okinawa population (0.75 ± 0.12) ; Figure [2d\)](#page-5-0) throughout the spawning season. These results suggest that fecundity evolution affects OSR as a by-product of life-history adaptation.

Few reports have investigated whether OSR varies with environmental gradients, such as latitude and/or altitude (Hettyey et al. [2005](#page-11-24); Monteiro and Lyons [2012](#page-12-26); Petry et al. [2016](#page-12-27); Machado et al. [2016](#page-11-25); Monteiro et al. [2017](#page-11-26); Garcia-Roa et al. [2020\)](#page-11-27). In a case study on pipefish, sex-specific thermal plasticity in reproduction has been proposed as a physiological mechanism driving variations in OSR among latitudinal populations (Monteiro and Lyons [2012](#page-12-26); Monteiro et al. [2017](#page-11-26)). Although our findings initially appear to align with those of the pipefish study, thermal plasticity in medaka does not account for the observed spawning intervals at higher latitudes with cooler climates (Hirshfield [1980](#page-11-19); Koger et al. [1999](#page-11-12); Hemmer-Brepson et al. [2014](#page-11-13)). Consequently, the disparities in OSR among medaka populations should be attributed to the evolutionary changes in female reproductive traits, such as spawning in-terval (Figure [S1\)](#page-13-8) (e.g., Montenegro et al. [2022](#page-12-9)) and/or the synchronous reproduction of *O. sakaizumii* females in specific seasons (Figure [2c,d](#page-5-0)) (Shinomiya et al. [2023](#page-12-12)). Evolutionary shifts in female reproductive traits along environmental gradients have been extensively reported in several animal species (Pincheira-Donoso and Hunt [2017;](#page-12-13) Álvarez-Noriega et al. [2023\)](#page-10-3). Given that females tend to be more sensitive to

environmental cues than males in terms of reproductive decisions (Morbey and Ydenberg [2001;](#page-12-28) Nakazawa and Hsu [2023](#page-12-29)), adaptations in female reproductive life-history may significantly influence OSR along environmental gradients in other animal taxa as well.

OSR typically affects the variance that exists in reproductive success, thereby influencing opportunities for sexual selection through individual sexual interactions (Shuster and Wade [2003;](#page-12-30) Klug et al. [2010](#page-11-28); Janicke and Morrow [2018](#page-11-29)), including mating competition and mate choice (e.g., Grant, Bryant, and Soos [1995;](#page-11-30) Grant, Gaboury, and Levit [2000;](#page-11-31) Forsgren et al. [2004;](#page-10-25) Sogabe and Yanagisawa [2007](#page-12-31)). In some animal species, including medaka (Fujimoto, Miyake, and Yamahira [2015;](#page-10-9) Sumarto et al. [2021\)](#page-12-32); sexual selection pressure is hypothesized to be weaker at higher latitudes, as indicated by latitudinal variations in sexual dimorphism (Fujimoto, Miyake, and Yamahira [2015](#page-10-9); Tarr et al. [2019;](#page-12-14) Sumarto et al. [2020](#page-12-22); Dudaniec et al. [2022;](#page-10-26) Murray et al. [2021](#page-12-33)) and mating behavior (Fujimoto, Miyake, and Yamahira [2015;](#page-10-9) Sumarto et al. [2021;](#page-12-32) Matsumura et al. [2023](#page-11-32)). Our findings underscore that the reduced male bias in OSR observed at higher latitudes serves as one of the ecological mechanisms contributing to weaker sexual selection at higher latitudes in ricefish.

5 | Conclusion

We demonstrated interspecific variation in the size-fecundity relationship in two medaka species. Since *O. sakaizumii* is distributed at higher latitudes and has a shorter spawning season than *O. latipes* in the wild, we propose that adaptation to high latitudes is responsible for the relatively high fecundity observed in *O. sakaizumii*. QTL analysis using interspecific F₂ hybrids showed that chromosome 23 is linked to the size-fecundity relationship, supporting the genetic basis and fecundity evolution in high-latitude females. Interestingly, fecundity evolution also results in a less male-biased OSR as a by-product of life-history adaptation in seasonal reproduction. These findings suggest that life-history evolution along latitudinal gradients significantly affects the reproductive characteristics of wild medaka populations.

Author Contributions

S.F., M.Y., and K.Y. conceived and designed the study. S.F., S.T., and K.Y. performed field investigations. S.F., T.M., D.M.F., I.M., B.K.A.S., and K.Y. performed laboratory analyses. S.F., T.M., S.A., and J.K. conducted the analyses. S.F. and K.Y. wrote the first draft. S.F. wrote the submitted manuscript. All authors read and approved the final manuscript.

Acknowledgements

We would like to thank M. Kawajiri, Y. Shiroma, Y. Yoshida, and T. Nojima for their assistance with the mating experiment. Fish collection in Aomori was partially supported by A. Ooyagi. Data analysis was improved by discussion and feedback from A. Sogabe, K. Mochida, and M. Kawajiri. The authors thank FORTE Science Communications (<https://www.forte-science.co.jp/>) for English language editing. This study was supported by the Spatiotemporal Genomics Project promoted by the University of the Ryukyus, with JSPS KAKENHI grants to S.F.

(KAKENHI 13J04092, 19K16232, 24K09610) and Y.K. (KAKENHI 20570019, 23570031), as well as the National Institute of Genetics Cooperative Research Program (2013-A35) to Y.K.

Ethics Statement

All experimental methods related to the handling of live fish were carried out in accordance with the Regulations for Animal Experiments at the University of the Ryukyus. Experiments were approved by the Animal Care Ethics Committee of University of the Ryukyus (Approval No. A2021012). The collection and rearing experiments involving wild *Oryzias latipes* in Okinawa Prefecture was conducted in accordance with the Protection Ordinance of Precious wild animals and plants in Okinawa Prefecture (No. 92, 2021/4/26).

Conflicts of Interest

The authors declare no conflicts of interest.

Data Availability Statement

Analyzed data and custom R scripts have been deposited in Dryad. <https://doi.org/10.5061/dryad.mpg4f4r7d>.

References

Álvarez-Noriega, M., C. R. White, J. Kozłowski, T. Day, and D. J. Marshall. 2023. "Life History Optimisation Drives Latitudinal Gradients and Responses to Global Change in Marine Fishes." *PLoS Biology* 21, no. 5: 1–19.<https://doi.org/10.1371/journal.pbio.3002114>.

Ansai, S., K. Mochida, S. Fujimoto, et al. 2021. "Genome Editing Reveals Fitness Effects of a Gene for Sexual Dichromatism in Sulawesian Fishes." *Nature Communications* 12, no. 1: 1–13. [https://doi.org/10.](https://doi.org/10.1038/s41467-021-21697-0) [1038/s41467-021-21697-0.](https://doi.org/10.1038/s41467-021-21697-0)

Arends, D., P. Prins, R. C. Jansen, and K. W. Broman. 2010. "R/Qtl: High-Throughput Multiple QTL Mapping." *Bioinformatics* 26: 2990– 2992. <https://doi.org/10.1093/bioinformatics/btq565>.

Asai, T., H. Senou, and K. Hosoya. 2011. "A New Ricefish From Northern Japan (Teleostei: Adrianichthyidae)." *Ichthyol Explor Freshwaters* 22, no. 4: 289–299.

Awaji, M., and I. Hanyu. 1987. "Annual Reproductive Cycle of the Wild Type Medaka." *Nippon Suisan Gakkaishi* 53, no. 6: 959–965.

Barneche, D. R., D. R. Robertson, C. R. White, and D. J. Marshall. 2018. "Fish Reproductive-Energy Output Increases Disproportionately With Body Size." *Science* 360, no. 6389: 642–645.

Berger, D., R. Walters, and K. Gotthard. 2008. "What Limits Insect Fecundity? Body Size- and Temperature-Dependent Egg Maturation and Oviposition in a Butterfly." *Functional Ecology* 22, no. 3: 523–529. <https://doi.org/10.1111/j.1365-2435.2008.01392.x>.

Broman, K. W., and Ś. Sen. 2009. *A Guide to QTL Mapping With R/Qtl*. New York: Springer.

Bunger, L., R. M. Lewis, M. F. Rothschild, et al. 2005. "Relationships Between Quantitative and Reproductive Fitness Traits in Animals." *Philosophical Transactions of the Royal Society of London. Series B, Biological Sciences* 360, no. 1459: 1489–1502. [https://doi.org/10.1098/](https://doi.org/10.1098/rstb.2005.1679) [rstb.2005.1679.](https://doi.org/10.1098/rstb.2005.1679)

Charnov, E. L., T. F. Turner, and K. O. Winemiller. 2001. "Reproductive Constraints and the Evolution of Life Histories With Indeterminate Growth." *Proceedings of the National Academy of Sciences of the United States of America* 98, no. 16: 9460–9464. [https://doi.org/10.1073/pnas.](https://doi.org/10.1073/pnas.161294498) [161294498.](https://doi.org/10.1073/pnas.161294498)

Chisada, S., T. Kurokawa, K. Murashita, et al. 2014. "Leptin Receptor-Deficient (Knockout) Medaka, *Oryzias latipes*, Show Chronical Up-Regulated Levels of Orexigenic Neuropeptides, Elevated Food Intake and Stage Specific Effects on Growth and Fat Allocation." *General and Comparative Endocrinology* 195, no. 1: 9–20. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.ygcen.2013.10.008) [ygcen.2013.10.008.](https://doi.org/10.1016/j.ygcen.2013.10.008)

Conover, D. O., and E. T. Schultz. 1995. "Phenotypic Similarity and the Evolutionary Significance of Countergradient Variation." *Trends in Ecology & Evolution* 10, no. 6: 248–252.

Copeland, D. L., R. J. Duff, Q. Liu, J. Prokop, and R. L. Londraville. 2011. "Leptin in Teleost Fishes: An Argument for Comparative Study." *Frontiers in Physiology* 2: 11023. [https://doi.org/10.3389/fphys.2011.](https://doi.org/10.3389/fphys.2011.00026) [00026](https://doi.org/10.3389/fphys.2011.00026).

Covarrubias-Pazaran, G. 2016. "Genome-Assisted Prediction of Quantitative Traits Using the R Package Sommer." *PLoS One* 11, no. 6: e0156744.<https://doi.org/10.1371/journal.pone.0156744>.

Daimon, M., T. Katsumura, H. Sakamoto, et al. 2022. "Mating Experiences With the Same Partner Enhanced Mating Activities of naïve Male Medaka Fish." *Scientific Reports* 12, no. 1: 1–8. [https://doi.](https://doi.org/10.1038/s41598-022-23871-w) [org/10.1038/s41598-022-23871-w](https://doi.org/10.1038/s41598-022-23871-w).

D'Ambrosio, J., R. Morvezen, S. Brard-Fudulea, et al. 2020. "Genetic Architecture and Genomic Selection of Female Reproduction Traits in Rainbow Trout." *BMC Genomics* 21, no. 1: 1–14. [https://doi.org/10.1186/](https://doi.org/10.1186/s12864-020-06955-7) [s12864-020-06955-7.](https://doi.org/10.1186/s12864-020-06955-7)

Dantzer, B., and E. M. Swanson. 2012. "Mediation of Vertebrate Life Histories via Insulin-Like Growth Factor-1." *Biological Reviews* 87, no. 2: 414–429. <https://doi.org/10.1111/j.1469-185X.2011.00204.x>.

Ding, L., W. W. Kuhne, D. E. Hinton, J. Song, and W. S. Dynan. 2010. "Quantifiable Biomarkers of Normal Aging in the Japanese Medaka Fish (*Oryzias latipes*)." *PLoS One* 5, no. 10: e13287. [https://doi.org/10.](https://doi.org/10.1371/journal.pone.0013287) [1371/journal.pone.0013287](https://doi.org/10.1371/journal.pone.0013287).

Dudaniec, R. Y., A. R. Carey, E. I. Svensson, B. Hansson, C. J. Yong, and L. T. Lancaster. 2022. "Latitudinal Clines in Sexual Selection, Sexual Size Dimorphism and Sex-Specific Genetic Dispersal During a Poleward Range Expansion." *Journal of Animal Ecology* 91: 1104–1118. <https://doi.org/10.1111/1365-2656.13488>.

Durinck, S., Y. Moreau, A. Kasprzyk, et al. 2005. "BioMart and Bioconductor: A Powerful Link Between Biological Databases and Microarray Data Analysis." *Bioinformatics* 21, no. 16: 3439–3440. [https://doi.org/10.1093/bioinformatics/bti525.](https://doi.org/10.1093/bioinformatics/bti525)

Egami, N., O. Terao, and Y. Iwao. 1988. "The Life Span of Wild Populations of the Fish *Oryzias latipes* Under Natural Conditions." *Zoological Science* 5: 1149–1152.

Flury, J. M., L. Hilgers, F. Herder, et al. 2022. "The Genetic Basis of a Novel Reproductive Strategy in Sulawesi Ricefishes: How Modularity and a Low Number of Loci Shape Pelvic Brooding." *Evolution* 76, no. 5: 1033–1051.<https://doi.org/10.1111/evo.14475>.

Forsgren, E., T. Amundsen, A. A. Borg, and J. Bjelvenmark. 2004. "Unusually Dynamic Sex Roles in a Fish." *Nature* 429, no. 6991: 551– 554.<https://doi.org/10.1038/nature02560.1>.

Fox, J., and S. Weisberg. 2019. *An R Companion to Applied Regression*. Third ed. Thousand Oaks CA: Sage.

Fujimoto, S., M. Kawajiri, J. Kitano, and K. Yamahira. 2014. "Female Mate Preference for Longer Fins in Medaka." *Zoological Science* 31, no. 11: 703–708. [https://doi.org/10.2108/zs140102.](https://doi.org/10.2108/zs140102)

Fujimoto, S., T. Miyake, and K. Yamahira. 2015. "Latitudinal Variation in Male Competitiveness and Female Choosiness in a Fish: Are Sexual Selection Pressures Stronger at Lower Latitudes?" *Evolutionary Biology* 42, no. 1: 75–87. [https://doi.org/10.1007/s1169](https://doi.org/10.1007/s11692-014-9300-9) [2-014-9300-9.](https://doi.org/10.1007/s11692-014-9300-9)

Fujimoto, S., S. Takeda, M. Yagi, and K. Yamahira. 2021. "Seasonal Change in Male Reproductive Investment of a Fish." *Environmental Biology of Fishes* 104, no. 2: 107–118. [https://doi.org/10.1007/s10641-](https://doi.org/10.1007/s10641-021-01059-x) [021-01059-x](https://doi.org/10.1007/s10641-021-01059-x).

Garcia-Roa, R., F. Garcia-Gonzalez, D. W. A. Noble, and P. Carazo. 2020. "Temperature as a Modulator of Sexual Selection." *Biological Reviews* 95, no. 6: 1607–1629. <https://doi.org/10.1111/brv.12632>.

Ge, S. X., D. Jung, and R. Yao. 2020. "ShinyGO: A Graphical Gene-Set Enrichment Tool for Animals and Plants." *Bioinformatics* 36, no. 8: 2628–2629. [https://doi.org/10.1093/bioinformatics/btz931.](https://doi.org/10.1093/bioinformatics/btz931)

Grant, J. W. A., M. J. Bryant, and C. E. Soos. 1995. "Operational Sex Ratio, Mediated by Synchrony of Female Arrival, Alters the Variance of Male Mating Success in Japanese Medaka." *Animal Behaviour* 49: 367–375. [https://doi.org/10.1006/anbe.1995.9998.](https://doi.org/10.1006/anbe.1995.9998)

Grant, J. W. A., L. C. Gaboury, and L. H. Levit. 2000. "Competitor-To-Resource Ratio, a General Formulation of Operational Sex Ratio, as a Predictor of Competitive Aggression in Japanese Medaka (Pisces: Oryziidae)." *Behavioral Ecology* 11, no. 6: 670–675. [https://doi.org/10.](https://doi.org/10.1093/beheco/11.6.670) [1093/beheco/11.6.670.](https://doi.org/10.1093/beheco/11.6.670)

Green, D. A., and C. G. Extavour. 2014. "Insulin Signalling Underlies Both Plasticity and Divergence of a Reproductive Trait in Drosophila." *Proceedings of the Royal Society B: Biological Sciences* 281, no. 1779: 20132673. [https://doi.org/10.1098/rspb.2013.2673.](https://doi.org/10.1098/rspb.2013.2673)

Healy, K., T. H. G. Ezard, O. R. Jones, R. Salguero-Gómez, and Y. M. Buckley. 2019. "Animal Life History Is Shaped by the Pace of Life and the Distribution of Age-Specific Mortality and Reproduction." *Nature Ecology & Evolution* 3, no. 8: 1217–1224. [https://doi.org/10.1038/s4155](https://doi.org/10.1038/s41559-019-0938-7) [9-019-0938-7.](https://doi.org/10.1038/s41559-019-0938-7)

Hemmer-Brepson, C., L. Replumaz, C. Romestaing, Y. Voituron, and M. Daufresne. 2014. "Non-Stressful Temperature Effect on Oxidative Balance and Life History Traits in Adult Fish (*Oryzins latipes*)." *Journal of Experimental Biology* 217, no. 2: 274–282. [https://doi.org/10.1242/jeb.](https://doi.org/10.1242/jeb.096172) [096172](https://doi.org/10.1242/jeb.096172).

Hettyey, A., A. Laurila, G. Herczeg, K. I. Jönsson, T. Kovács, and J. Merilä. 2005. "Does Testis Weight Decline Towards the Subarctic? A Case Study on the Common Frog, *Rana temporaria*." *Naturwissenschaften* 92: 188– 192. [https://doi.org/10.1007/s00114-005-0607-3.](https://doi.org/10.1007/s00114-005-0607-3)

Hilgers, L., O. Roth, A. W. Nolte, et al. 2022. "Inflammation and Convergent Placenta Gene Co-Option Contributed to a Novel Reproductive Tissue." *Current Biology* 32, no. 3: 715–724. [https://doi.](https://doi.org/10.1016/j.cub.2021.12.004) [org/10.1016/j.cub.2021.12.004.](https://doi.org/10.1016/j.cub.2021.12.004)

Hinrichs, A. S., D. Karolchik, R. Baertsch, et al. 2006. "The UCSC Genome Browser Database: Update 2006." *Nucleic Acids Research* 34: 590–598. [https://doi.org/10.1093/nar/gkj144.](https://doi.org/10.1093/nar/gkj144)

Hirshfield, M. 1980. "An Experimental Analysis of Reproductive Effort and Cost in the Japanese Medaka, *Oryzias latipes*." *Ecology* 61, no. 2: 282–292.

Ichikawa, K., S. Tomioka, Y. Suzuki, et al. 2017. "Centromere Evolution and CpG Methylation During Vertebrate Speciation." *Nature Communications* 8, no. 1: 1833. [https://doi.org/10.1038/s41467-017-](https://doi.org/10.1038/s41467-017-01982-7) [01982-7](https://doi.org/10.1038/s41467-017-01982-7).

Iwamatsu, T. 2006. *The Integrated Book for the Biology of the Medaka*. Okayama, Tokyo: University Education Press.

Janicke, T., and E. H. Morrow. 2018. "Operational Sex Ratio Predicts the Opportunity and Direction of Sexual Selection Across Animals." *Ecology Letters* 21, no. 3: 384–391.<https://doi.org/10.1111/ele.12907>.

Kagawa, N. 2014. "Comparison of Aggressive Behaviors Between Two Wild Populations of Japanese Medaka,Oryzias latipes and O. sakaizumii." *Zoological Science* 31, no. 3: 116–121. [https://doi.org/10.2108/](https://doi.org/10.2108/zsj.31.116) [zsj.31.116](https://doi.org/10.2108/zsj.31.116).

Kasahara, M., K. Naruse, S. Sasaki, et al. 2007. "The Medaka Draft Genome and Insights Into Vertebrate Genome Evolution." *Nature* 447, no. 7145: 714–719.<https://doi.org/10.1038/nature05846>.

Katsumura, T., S. Oda, H. Mitani, and H. Oota. 2019. "Medaka Population Genome Structure and Demographic History Described via Genotyping-By-Sequencing." *G3: Genes, Genomes, Genetics* 9, no. 1: 217–228. <https://doi.org/10.1534/g3.118.200779>.

Kawajiri, M., S. Fujimoto, K. Yoshida, K. Yamahira, and J. Kitano. 2015. "Genetic Architecture of the Variation in Male-Specific Ossified Processes on the Anal Fins of Japanese Medaka." *G3: Genes, Genomes, Genetics* 5, no. 12: 2875–2884. [https://doi.org/10.1534/g3.](https://doi.org/10.1534/g3.115.021956) [115.021956.](https://doi.org/10.1534/g3.115.021956)

Kawajiri, M., K. Yoshida, S. Fujimoto, et al. 2014. "Ontogenetic Stage-Specific Quantitative Trait Loci Contribute to Divergence in Developmental Trajectories of Sexually Dimorphic Fins Between Medaka Populations." *Molecular Ecology* 23, no. 21: 5258–5275. [https://](https://doi.org/10.1111/mec.12933) doi.org/10.1111/mec.12933.

Kent, W. J. 2002. "BLAT—The BLAST-Like Alignment Tool." *Genome Research* 12, no. 4: 656–664.<https://doi.org/10.1101/gr.229202>.

Kinoshita, M., K. Murata, K. Naruse, and M. Tanaka. 2009. *Medaka: Biology, Management, and Experimental Protocols*. USA: John Wiley & Sons.

Kitano, J., S. Ansai, S. Fujimoto, et al. 2023. "A Cryptic Sex-Linked Locus Revealed by the Elimination of a Master Sex-Determining Locus in Medaka Fish." *American Naturalist* 202, no. 2: 231–240. [https://doi.](https://doi.org/10.1086/724840) [org/10.1086/724840.](https://doi.org/10.1086/724840)

Klug, H., J. Heuschele, M. D. Jennions, and H. Kokko. 2010. "The Mismeasurement of Sexual Selection." *Journal of Evolutionary Biology* 23, no. 3: 447–462.<https://doi.org/10.1111/j.1420-9101.2009.01921.x>.

Koger, C. S., S. J. Teh, and D. E. Hinton. 1999. "Variations of Light and Temperature Regimes and Resulting Effects on Reproductive Parameters in Medaka (Oryzias latipes)." *Biology of Reproduction* 61, no. 5: 1287–1293. <https://doi.org/10.1095/biolreprod61.5.1287>.

Kokita, T. 2004. "Latitudinal Compensation in Female Reproductive Rate of a Geographically Widespread Reef Fish." *Environmental Biology of Fishes* 71: 213–224. [https://doi.org/10.1007/s10641-003-0304-z.](https://doi.org/10.1007/s10641-003-0304-z)

Komine, R., T. Nishimaki, T. Kimura, et al. 2016. "Transgenic Medaka That Overexpress Growth Hormone Have a Skin Color That Does Not Indicate the Activation or Inhibition of Somatolactin-α Signal." *Gene* 584, no. 1: 38–46. <https://doi.org/10.1016/j.gene.2016.02.046>.

Kondo, Y., M. Kohda, Y. Koya, and S. Awata. 2023. "Sperm Allocation in Relation to Male–Male Aggression and Courtship in an Externally Fertilizing Fish, the Medaka." *Animal Behaviour* 202: 9–19. [https://doi.](https://doi.org/10.1016/j.anbehav.2023.05.011) [org/10.1016/j.anbehav.2023.05.011](https://doi.org/10.1016/j.anbehav.2023.05.011).

Kooijman, S. A. L. M. 2010. *Dynamic Energy Budget Theory for Metabolic Organisation*. Cambridge: Cambridge University Press.

Leaf, R. T., Y. Jiao, B. R. Murphy, J. I. Kramer, K. M. Sorensen, and V. G. Wooten. 2011. "Life-History Characteristics of Japanese Medaka *Oryzias latipes*." *Copeia* 4: 559–565. [https://doi.org/10.1643/CI-09-190.](https://doi.org/10.1643/CI-09-190)

Machado, G., B. A. Buzatto, S. Garcia-Hernandez, and R. Macias-Ordonez. 2016. "Macroecology of Sexual Selection: A Predictive Conceptual Framework for Large-Scale Variation in Reproductive Traits." *American Naturalist* 188, no. S1: 8–27. <https://doi.org/10.1086/687575>.

Matsumura, K., K. Wakabayashi, R. Kawakami, and T. Miyatake. 2023. "Latitudinal Cline in Reproductive Traits in the Red Flour Beetle *Tribolium castaneum*." *Behavioral Ecology and Sociobiology* 77, no. 7: 1–10. [https://doi.org/10.1007/s00265-023-03359-x.](https://doi.org/10.1007/s00265-023-03359-x)

Minvielle, F., B. B. Kayang, M. Inoue-Murayama, et al. 2005. "Microsatellite Mapping of QTL Affecting Growth, Feed Consumption, Egg Production, Tonic Immobility and Body Temperature of Japanese Quail." *BMC Genomics* 6, no. 87: 1–9. [https://doi.org/10.1186/](https://doi.org/10.1186/1471-2164-6-87) [1471-2164-6-87.](https://doi.org/10.1186/1471-2164-6-87)

Monteiro, N., M. Cunha, L. Ferreira, et al. 2017. "Parabolic Variation in Sexual Selection Intensity Across the Range of a Cold-Water Pipefish: Implications for Susceptibility to Climate Change." *Global Change Biology* 23, no. 9: 3600–3609. [https://doi.org/10.1111/gcb.13630.](https://doi.org/10.1111/gcb.13630)

Monteiro, N., and D. Lyons. 2012. "Stronger Sexual Selection in Warmer Waters: The Case of a Sex Role Reversed Pipefish." *PLoS One* 7: e44251. <https://doi.org/10.1371/journal.pone.0044251>.

Montenegro, J., S. Fujimoto, S. Ansai, et al. 2022. "Genetic Basis for the Evolution of Pelvic-Fin Brooding, a New Mode of Reproduction, in a Sulawesian Fish." *Molecular Ecology* 31, no. 14: 3798–3811. [https://doi.](https://doi.org/10.1111/mec.16555) [org/10.1111/mec.16555](https://doi.org/10.1111/mec.16555).

Morbey, Y. E., and R. C. Ydenberg. 2001. "Protandrous Arrival Timing to Breeding Areas: A Review." *Ecology Letters* 4, no. 6: 663–673. [https://](https://doi.org/10.1046/j.1461-0248.2001.00265.x) doi.org/10.1046/j.1461-0248.2001.00265.x.

Murray, C. M., C. D. McMahan, A. R. Litmer, et al. 2021. "Latitudinal Gradients in Sexual Dimorphism: Alternative Hypotheses for Variation in Male Traits." *Ecology and Evolution* 11, no. 23: 17519–17526. [https://](https://doi.org/10.1002/ece3.8386) doi.org/10.1002/ece3.8386.

Myosho, T., H. Takahashi, K. Yoshida, et al. 2018. "Hyperosmotic Tolerance of Adult Fish and Early Embryos Are Determined by Discrete, Single Loci in the Genus *Oryzias*." *Scientific Reports* 8, no. 1: 6897.<https://doi.org/10.1038/s41598-018-24621-7>.

Nakazawa, T., and Y. H. Hsu. 2023. "Chen IC (2023) why Sex Matters in Phenological Research." *Oikos* 8: e09808. [https://doi.org/10.1111/oik.](https://doi.org/10.1111/oik.09808) [09808](https://doi.org/10.1111/oik.09808).

Ndandala, C. B., M. Dai, U. F. Mustapha, et al. 2022. "Current Research and Future Perspectives of GH and IGFs Family Genes in Somatic Growth and Reproduction of Teleost Fish." *Aquaculture Reports* 26: 101289.<https://doi.org/10.1016/j.aqrep.2022.101289>.

Ogino, Y., S. Ansai, E. Watanabe, et al. 2023. "Evolutionary Differentiation of Androgen Receptor Is Responsible for Sexual Character- Istic Development in a Teleost Fish." *Nature Communications* 14, no. 1: 1428. [https://doi.org/10.1038/s41467-023-37026-6.](https://doi.org/10.1038/s41467-023-37026-6)

Ono, Y., and T. Uematsu. 1957. "Mating Ethogram in *Oryzias latipes*." *Journal of Faculty of Science Hokkaido Universtiy* 13: 197–202.

Orgogozo, V., K. W. Broman, and D. L. Stern. 2006. "High-Resolution Quantitative Trait Locus Mapping Reveals Sign Epistasis Controlling Ovariole Number Between Two Drosophila Species." *Genetics* 173, no. 1: 197–205.<https://doi.org/10.1534/genetics.105.054098>.

Petry, W. K., J. D. Soule, A. M. Iler, et al. 2016. "Sex-Specific Responses to Climate Change in Plants Alter Population Sex Ratio and Performance." *Science* 353, no. 6294: 69–71.<https://doi.org/10.1126/science.aaf2588>.

Pianka, E. R. 1960. "On r- and K- Selection." *American Naturalist* 104, no. 940: 592–597.

Pincheira-Donoso, D., and J. Hunt. 2017. "Fecundity Selection Theory: Concepts and Evidence." *Biological Reviews* 92, no. 1: 341–356. [https://](https://doi.org/10.1111/brv.12232) [doi.org/10.1111/brv.12232.](https://doi.org/10.1111/brv.12232)

Pracana, R., A. Priyam, I. Levantis, R. A. Nichols, and Y. Wurm. 2017. "The Fire Ant Social Chromosome Supergene Variant Sb Shows Low Diversity but High Divergence From SB." *Molecular Ecology* 26, no. 11: 2864–2879.<https://doi.org/10.1111/mec.14054>.

R Core Team. 2021. *R: A Language and Environment for Statistical Computing*. Vienna, Austria: R Foundation for Statistical Computing. [https://www.R-project.org/](https://www.r-project.org/).

Sakaizumi, M., S. Yasushi, and H. Satoshi. 1992. "Electrophoretic Studies of Meiotic Segregation in Inter-and Intraspecific Hybrids Among East Asian Species of the Genus Oryzias (Pisces: Oryziatidae)." *Journal of Experimental Zoology* 264, no. 1: 85–92. [https://doi.org/10.](https://doi.org/10.1002/jez.1402640113) [1002/jez.1402640113](https://doi.org/10.1002/jez.1402640113).

Santure, A. W., I. De Cauwer, M. R. Robinson, et al. 2013. "Genomic Dissection of Variation in Clutch Size and Egg Mass in a Wild Great Tit (*Parus major*) Population." *Molecular Ecology* 22, no. 15: 3949–3962. <https://doi.org/10.1111/mec.12376>.

Sauvage, C., M. Vagner, N. Derome, et al. 2012. "Coding Gene Single Nucleotide Polymorphism Mapping and Quantitative Trait Loci Detection for Physiological Reproductive Traits in Brook Charr, Salvelinus Fontinalis." *G3: Genes, Genomes, Genetics* 2, no. 3: 379–392. <https://doi.org/10.1534/g3.111.001867>.

Shine, R. 2005. "Life-History Evolution in Reptiles." *Annual Review of Ecology, Evolution, and Systematics* 36: 23–46. [https://doi.org/10.1146/](https://doi.org/10.1146/annurev.ecolsys.36.102003.152631) [annurev.ecolsys.36.102003.152631.](https://doi.org/10.1146/annurev.ecolsys.36.102003.152631)

Shinomiya, A., D. Adachi, T. Shimmura, et al. 2023. "Variation in Responses to Photoperiods and Temperatures in Japanese Medaka From Different Latitudes." *Zoological Letters* 9, no. 1: 1–14. [https://doi.](https://doi.org/10.1186/s40851-023-00215-8) [org/10.1186/s40851-023-00215-8](https://doi.org/10.1186/s40851-023-00215-8).

Shinomiya, A., H. Otake, S. Hamaguchi, and M. Sakaizumi. 2010. "Inherited XX Sex Reversal Originating From Wild Medaka Populations." *Heredity* 105, no. 5: 443–448. [https://doi.org/10.1038/hdy.2010.51.](https://doi.org/10.1038/hdy.2010.51)

Shinya, M., T. Kimura, and K. Naruse. 2023. "High-Speed System to Generate Congenic Strains in Medaka." *Genes & Genetic Systems* 98, no. 5: 267–275. [https://doi.org/10.1266/ggs.23-00075.](https://doi.org/10.1266/ggs.23-00075)

Shuster, S. M., and M. J. Wade. 2003. *Mating Systems and Strategies*. Princeton: Princeton University Press.

Simmons, L. W., S. Lupold, and J. L. Fitzpatrick. 2017. "Evolutionary Trade-Off Between Secondary Sexual Traits and Ejaculates." *Trends in Ecology & Evolution* 32, no. 12: 964–976. [https://doi.org/10.1016/j.tree.](https://doi.org/10.1016/j.tree.2017.09.011) [2017.09.011.](https://doi.org/10.1016/j.tree.2017.09.011)

Sogabe, A., and Y. Yanagisawa. 2007. "Sex-Role Reversal of a Monogamous Pipefish Without Higher Potential Reproductive Rate in Females." *Proceedings of the Royal Society B: Biological Sciences* 274, no. 1628: 2959–2963. [https://doi.org/10.1098/rspb.2007.1041.](https://doi.org/10.1098/rspb.2007.1041)

Sparks, M. M., J. C. Kraft, K. M. S. Blackstone, G. G. McNickle, and M. R. Christie. 2022. "Large genetic Divergence Underpins Cryptic Local Adaptation Across Ecological and Evolutionary Gradients." *Proceedings of the Royal Society B: Biological Sciences* 289, no. 1984. [https://doi.org/](https://doi.org/10.1098/rspb.2022.1472) [10.1098/rspb.2022.1472.](https://doi.org/10.1098/rspb.2022.1472)

Stearns, S. C. 1992. *The Evolution of Life Histories*. Oxford, UK: Oxford University Press. [https://doi.org/10.1093/oso/9780198577416.001.0001.](https://doi.org/10.1093/oso/9780198577416.001.0001)

Sumarto, B. K. A., H. Kobayashi, R. Kakioka, et al. 2020. "Latitudinal Variation in Sexual Dimorphism in a Freshwater Fish Group." *Biological Journal of the Linnean Society* 131, no. 4: 898–908. [https://doi.org/10.](https://doi.org/10.1093/biolinnean/blaa166) [1093/biolinnean/blaa166](https://doi.org/10.1093/biolinnean/blaa166).

Sumarto, B. K. A., A. B. Nofrianto, D. F. Mokodongan, et al. 2021. "Variation in Mating Behaviors Between a Tropical and a Temperate Species of Medaka Fishes." *Zoological Science* 38, no. 1: 45–50. [https://](https://doi.org/10.2108/zs200114) doi.org/10.2108/zs200114.

Suzuki, Y., T. Miyake, and K. Yamahira. 2010. "An Acquisition Trade-Off With Fast Growth in a Fish, the Medaka *Oryzias latipes*: Why Do Low-Latitude Ectotherms Grow More Slowly?" *Evolutionary Ecology* 24, no. 4: 749–759.<https://doi.org/10.1007/s10682-009-9331-3>.

Swanson, E. M., and B. Dantzer. 2014. "Insulin-Like Growth Factor-1 Is Associated With Life-History Variation Across Mammalia." *Proceedings of the Royal Society B: Biological Sciences* 281, no. 1782: 20132458. [https://doi.org/10.1098/rspb.2013.2458.](https://doi.org/10.1098/rspb.2013.2458)

Takehana, Y., N. Nagai, M. Matsuda, K. Tsuchiya, and M. Sakaizumi. 2003. "Geographic Variation and Diversity of the Cytochrome b Gene in Japanese Wild Populations of Medaka, *Oryzias latipes*." *Zoological Science* 20, no. 10: 1279–1291.

Takehana, Y., S. Uchiyama, M. Matsuda, S. R. Jeon, and M. Sakaizumi. 2004. "Geographic Variation and Diversity of the Cytochrome b Gene in Wild Populations of Medaka (*Oryzias latipes*) From Korea and China." *Zoological Science* 21, no. 4: 483–491.

Tarr, S., S. Meiri, J. J. Hicks, and A. C. Algar. 2019. "A Biogeographic Reversal in Sexual Size Dimorphism Along a Continental Temperature Gradient." *Ecography* 42, no. 4: 706–716. [https://doi.org/10.1111/ecog.](https://doi.org/10.1111/ecog.03593) [03593.](https://doi.org/10.1111/ecog.03593)

Tsuboko, S., T. Kimura, M. Shinya, et al. 2014. "Genetic Control of Startle Behavior in Medaka Fish." *PLoS One* 9, no. 11: e112527. [https://](https://doi.org/10.1371/journal.pone.0112527) [doi.org/10.1371/journal.pone.0112527.](https://doi.org/10.1371/journal.pone.0112527)

Vila-Gispert, A., R. Moreno-Amich, and E. Garcia-Berthou. 2002. "Gradients of Life-History Variation: An Intercontinental Comparison of Fishes." *Reviews in Fish Biology and Fisheries* 12: 417–427.

Wessel, P., W. H. Smith, R. Scharroo, J. Luis, and F. Wobbe. 2013. "Generic Mapping Tools: Improved Version Released." *Eos, Transactions American Geophysical Union* 94, no. 45: 409–410. [https://doi.org/10.](https://doi.org/10.1002/2013EO450001) [1002/2013EO450001.](https://doi.org/10.1002/2013EO450001)

White, C. R., L. A. Alton, C. L. Bywater, E. J. Lombardi, and D. J. Marshall. 2022. "Metabolic Scaling Is the Product of Life-History Optimization." *Science* 377, no. 6608: 834–839. [https://doi.org/10.1126/](https://doi.org/10.1126/science.abm7649) [science.abm7649](https://doi.org/10.1126/science.abm7649).

Winemiller, K., and K. Rose. 1992. "Patterns of Life-History Diversification in North American Fishes: Implications for Population Regulation." *Canadian Journal of Fisheries and Aquatic Sciences* 49, no. 10: 2196–2218.

Wolc, A., J. Arango, T. Jankowski, et al. 2014. "Genome-Wide Association Study for Egg Production and Quality in Layer Chickens." *Journal of Animal Breeding and Genetics* 131, no. 3: 173–182. [https://doi.](https://doi.org/10.1111/jbg.12086) [org/10.1111/jbg.12086](https://doi.org/10.1111/jbg.12086).

Wood, S. N. 2006. *Generalized Additive Models*. New York: An Introduction with R. Chapman & Hall/CRC.

Wood, S. N. 2011. "Fast Stable Restricted Maximum Likelihood and Marginal Likelihood Estimation of Semiparametric Generalized Linear Models." *Journal of the Royal Statistical Society, Series B: Statistical Methodology* 73, no. 1: 3–36. [https://doi.org/10.1111/j.1467-9868.2010.](https://doi.org/10.1111/j.1467-9868.2010.00749.x) [00749.x.](https://doi.org/10.1111/j.1467-9868.2010.00749.x)

Wright, D., C. J. Rubin, A. Martinez Barrio, et al. 2010. "The Genetic Architecture of Domestication in the Chicken: Effects of Pleiotropy and Linkage." *Molecular Ecology* 19, no. 23: 5140–5156. [https://doi.org/10.](https://doi.org/10.1111/j.1365-294X.2010.04882.x) [1111/j.1365-294X.2010.04882.x](https://doi.org/10.1111/j.1365-294X.2010.04882.x).

Yamahira, K., M. Kawajiri, K. Takeshi, and T. Irie. 2007. "Inter- and Intrapopulation Variation in Thermal Reaction Norms for Growth Rate: Evolution of Latitudinal Compensation in Ectotherms With a Genetic Constraint." *Evolution* 61, no. 7: 1577–1589. [https://doi.org/10.1111/j.](https://doi.org/10.1111/j.1558-5646.2007.00130.x) [1558-5646.2007.00130.x.](https://doi.org/10.1111/j.1558-5646.2007.00130.x)

Yamahira, K., and K. Takeshi. 2008. "Variation in Juvenile Growth Rates Among and Within Latitudinal Populations of the Medaka." *Population Ecology* 50, no. 1: 3–8. [https://doi.org/10.1007/s10144-007-0055-3.](https://doi.org/10.1007/s10144-007-0055-3)

Yassumoto, T. I., M. Nakatsukasa, A. J. Nagano, M. Yasugi, T. Yoshimura, and A. Shinomiya. 2020. "Genetic Analysis of Body Weight in Wild Populations of Medaka Fish From Different Latitudes." *PLoS One* 15, no. 6: e0234803. [https://doi.org/10.1371/journal.pone.0234803.](https://doi.org/10.1371/journal.pone.0234803)

Yokoi, S., K. Naruse, Y. Kamei, et al. 2020. "Sexually Dimorphic Role of Oxytocin in Medaka Mate Choice." *Proceedings of the National Academy of Sciences of the United States of America* 117, no. 9: 4802–4808. [https://](https://doi.org/10.1073/pnas.1921446117) [doi.org/10.1073/pnas.1921446117.](https://doi.org/10.1073/pnas.1921446117)

Zhao, X., C. Nie, J. Zhang, et al. 2021. "Identification of Candidate Genomic Regions for Thermogelled Egg Yolk Traits Based on a Genome-Wide Association Study." *BMC Genomics* 22, no. 1: 610. [https://doi.org/](https://doi.org/10.1016/j.psj.2022.102402) [10.1016/j.psj.2022.102402.](https://doi.org/10.1016/j.psj.2022.102402)

Supporting Information

Additional supporting information can be found online in the Supporting Information section.