- 1 Title: Pleiotropy increases with gene age in six model multicellular eukaryotes
- 2 Authors: Martin, Reese<sup>1,2\*</sup>, and Tate, Ann.T.<sup>1,2\*</sup>
- <sup>1</sup>Department of Biological Sciences, Vanderbilt University, Nashville TN, 37235
- 4 <sup>2</sup>Evolutionary Studies Initiative, Vanderbilt University, Nashville, Tennessee, USA
- 5 \*Email address: reese.a.martin@vanderbilt.edu
- 6 <sup>a</sup>Corresponding email address: <u>a.tate@vanderbilt.edu</u>. ORCID: 0000-0001-6601-0234
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#### 10 Abstract

11 Fundamental traits of genes, including function, length and GC content, all vary with gene age. Pleiotropy, where a single gene affects multiple traits, arises through selection for 12 13 novel traits and is expected to be removed from the genome through subfunctionalization 14 following duplication events. It is unclear, however, how these opposing forces shape the 15 prevalence of pleiotropy through time. We hypothesized that the prevalence of pleiotropy would 16 be lowest in young genes, peak in middle aged genes, and then either decrease to a middling 17 level in ancient genes or stay near the middle-aged peak, depending on the balance between 18 exaptation and subfunctionalization. To address this question, we have calculated gene age and 19 pleiotropic status for several model multicellular eukaryotes, including Homo sapiens, Mus 20 musculus, Danio rerio, Drosophila melanogaster, Caenorhabditis elegans, and Arabidopsis 21 thaliana. Gene age was determined by finding the most distantly related species that shared an 22 ortholog using the Open Tree of Life and the Orthologous Matrix Database (OMAdb). 23 Pleiotropic status was determined using both protein-protein interactions (STRINGdb) and 24 associated biological processes (Gene Ontology). We found that middle-aged and ancient genes 25 tend to be more pleiotropic than young genes, and that this relationship holds across all species 26 evaluated and across both modalities of measuring pleiotropy. We also found absolute 27 differences in the degree of pleiotropy based on gene functional class, but only when looking at 28 biological process count. From these results we propose that there is a fundamental relationship 29 between pleiotropy and gene age and further study of this relationship may shed light on the mechanism behind the functional changes genes undergo as they age. 30

31 Impact statement: Pleiotropy, the phenomenon where a single gene acts on multiple traits, is
32 fundamental to genomic organization and has profound consequences for fitness. This work

- 33 identifies a previously unknown relationship between pleiotropy and gene age, highlighting the
- 34 dynamism of pleiotropy across time. This relationship holds across six distantly related model
- 35 organisms, suggesting that it could be a highly generalizable finding, at least among multicellular
- 36 eukaryotes. Following from this work, future investigation into mechanisms dictating the
- 37 prevalence of pleiotropy at the gene or cellular level could provide fundamental insight into the
- 38 maintenance of pleiotropy despite the potential for constraining rapid adaptation.

### 39 Introduction

40 Pleiotropy is the phenomenon in which a single gene effects multiple traits, and has wide ranging effects on development (Cheverud, 1996), genetic disease (Sivakumaran et al., 2011; 41 42 Ittisoponpisan et al., 2017), signaling robustness (Guillaume & Otto, 2012; Papakostas et al., 2014), the evolution of new traits (Lenski et al., 2003; Armbruster et al., 2009) and organismal 43 44 adaptability (Fraïsse et al., 2019; Hämälä et al., 2020; Kinsler et al., 2020). Pleiotropy is 45 expected to arise in the genome as a result of exaptation, the process through which existing genes and genetic architecture are coopted for use in novel traits. The role of exaptation in 46 47 promoting pleiotropy has been demonstrated theoretically using computational models of 48 complex trait evolution (Lenski et al., 2003) and empirically in the flowering vine genus 49 Dalechampia (Armbruster et al., 2009), where pollinator attractant traits are coopted for plant 50 defense and vice versa. Pleiotropy has also been proposed to arise as a result of co-selection on 51 traits, such as the integration between the growth of the cranial vault and facial masticatory 52 apparatus reported in several primates (Cheverud, 1996). These mechanisms for increasing the 53 number of pleiotropic genes and the number of traits that pleiotropic genes affect suggest that the 54 prevalence of pleiotropy should be positively correlated with gene age. Supporting this idea in 55 humans, older genes have elevated protein-protein interactions (PPI), a commonly used metric to determine the pleiotropic status of a gene (Yin et al., 2016). 56 The prevalence of pleiotropy does not strictly increase over time, as subfunctionalization 57 58 following gene duplication events decreases the frequency of pleiotropy in the genome (Lynch &

59 Wagner, 2008; Schmid & Sánchez-Villagra, 2010; Guillaume & Otto, 2012).

60 Subfunctionalization in duplicated genes is predicted to arise through a process of

61 complimentary degenerate mutations in the duplicated genes, leading to loss or degradation of

62 multiple functions in the gene duplicates (Force *et al.*, 1999) and promoting the maintenance of 63 duplicated genes in the genome (Lynch & Force, 2000). In the case where the parent gene's functions are completely segregated between the copies with no overlap in function, the overall 64 65 prevalence of pleiotropy in the genome should be reduced as two genes are now performing the 66 work of a single gene. Aside from subfunctionalization removing pleiotropic genes from the 67 genome, a gene's future adaptability is limited by the acquisition of novel traits (Fraïsse *et al.*, 2019), placing a soft limit on the number of processes a gene can be involved in. These forces, 68 acting in opposition to the constant growth in pleiotropy from exaptation and co-selection, could 69 70 produce a wide range of distributions of pleiotropy across the gene age spectrum. 71 While we have insight into some relationships between the age of a gene and its 72 fundamental characteristics – old genes tend to be more essential (Chen et al., 2012), for 73 example, and have increased GC content and length (Yin *et al.*, 2016) – we do not have a clear 74 picture of how these different forces shake out to affect relationships between gene age and 75 pleiotropy, in terms of both mean and variance among genes and species. For example, it is 76 unclear if changes in the prevalence of pleiotropy over time are consistent among different 77 metrics of estimating pleiotropy, such as PPI and biological process count. It is also unclear if the 78 relationship between gene age and pleiotropy depends on the focal organism or if it is 79 generalizable to large swathes of the tree of life. While previous studies have suggested that 80 pleiotropic genes are more essential than their non-pleiotropic counterparts (Ittisoponpisan *et al.*, 81 2017), this could be related to either the essentiality of old genes or the tendency of pleiotropic proteins to be hubs in gene signaling networks (De Bruyne et al., 2014; Sadhukhan et al., 2021). 82 83 Furthermore, the exact dynamics of how the prevalence of pleiotropy changes with a gene's age

84 are unknown; while previous work demonstrated significant differences in PPI between old and 85 young genes, little is known about changes on a more continuous scale (Yin et al., 2016). 86 We hypothesized that young genes would have the lowest prevalence of pleiotropy, 87 accounting for the limited time they would have had to become involved in other traits. This 88 expectation also aligns with previous work showing that young genes are less essential than old 89 genes (Chen et al., 2012) and that pleiotropic genes tend to be highly essential. We hypothesized 90 that middle-aged genes would be more pleiotropic than young genes, reflecting the increased 91 time for genes to be recruited into other traits via exaptation. For the oldest genes we expected 92 either that the prevalence of pleiotropy would continue to climb or that it would reach an 93 equilibrium state, meaning the mean prevalence of pleiotropy in all age bins past a certain age 94 would be equal, reflecting the net outcomes of forces adding and removing pleiotropy from the 95 genome. 96 To address our hypothesis, we calculated the age and pleiotropic status of protein-coding 97 genes from Homo sapiens, Mus musculus, Danio rerio, Drosophila melanogaster,

98 Caenorhabditis elegans, and Arabidopsis thaliana. We selected these organisms in an attempt to 99 cover a wide range of multicellular eukaryotic life, but were limited to model organisms with 100 well-annotated genomes and protein resources. These organisms, while limited in scope 101 compared to the true diversity of the tree of life, allow us to identify potentially generalizable 102 and species-specific trends in the prevalence of pleiotropy. For a given species, a gene's age was 103 determined by finding the most distantly related common ancestor that shared an ortholog of that 104 gene. Orthologs were collected from the Orthologous Matrix Database (OMAdb) (Altenhoff et 105 al., 2021) and common ancestors were identified on a phylogeny generated from the Open Tree 106 of Life (synthesis 14.8) (Redelings & Holder, 2017) that included 1900 species from the

107	OMAdb. To evaluate the prevalence of pleiotropy in genes, we used Gene Ontology (Gene
108	Ontology Consortium, 2021) Biological process (BP) labels for each gene, as well as protein-
109	protein interactions (PPI) from the String DB (Szklarczyk et al., 2023). We elected to use both
110	Gene Ontology Biological processes and PPI as these are complimentary measures of pleiotropy
111	(He & Zhang, 2006). Biological processes assess the kinds of distinct actions a protein may carry
112	out, often with different active sites, while PPI measures total interacting partners and does not
113	capture what a protein is doing with those partners, or which active sites are involved in the
114	interaction (He & Zhang, 2006). Our primary goal with this study is to provide a high-level but
115	generalizable understanding of the evolution of pleiotropy, laying the groundwork for future
116	exploration of this fundamental genomic phenomenon.

#### 117 Methods

Because the datasets for each species were retrieved from databases that used similar terms and formatting, we were able to apply the same pipelines for the analysis of each species individually. The one exception is *A. thaliana*, which lacked gene duplication data in the Ensembl database and was therefore excluded from singleton vs duplicate analysis. With that in mind, the following sections reference the *H. sapiens* dataset and its analysis but apply to all species unless otherwise noted.

- 124 For each species we retrieved the following number of genes: *Homo sapiens* (n = 19,467), *Mus*
- 125 musculus (n = 21,128), Danio rerio (n = 27,897), Drosophila melanogaster (n = 13,659),
- 126 *Caenorhabditis elegans* (n = 16,050), and *Arabidopsis thaliana* (n = 25,125).
- 127 Gene age determination

The entire set of ortholog groups was downloaded from the OMA database (Altenhoff *et al.*, 2021), and the full database was trimmed to only include those ortholog groups with an entry

130 from the focal species (e.g. *H. sapiens*). From these ortholog groups, a list was generated 131 including each unique species that were present in a *H. sapiens* ortholog group. A phylogenetic 132 tree including only these unique species was trimmed from the Open Tree of Life. The Open 133 Tree of Life is a purely relational tree and does not include branch length data, so all branch 134 lengths were set to 1 during the creation of the OMA-specific phylogeny. The distance from the 135 root of the tree to the most recent common ancestor of each species and *H. sapiens* was 136 calculated and the age of each ortholog was set to the age of the oldest common ancestor 137 between *H. sapiens* and the species in which the ortholog was found. For example, if an ortholog 138 was present in several species, with the oldest one being A. thaliana, then the age of the ortholog 139 would be the distance of the last common ancestor between *H. sapiens* and *A. thaliana* to the 140 root of the tree. This means that orthologs with a lower distance from the base of the tree are 141 older than those with a greater distance. If an ortholog could not have an age assigned to it for 142 any reason, it was excluded from the analysis. This process removed 12 A. thaliana genes, 7 D. 143 rerio genes, and no genes for C. elegans, D. melanogaster, or M. musculus. 455 genes were 144 removed in H. sapiens; while this is noticeably more than the other species, it still left 98% of the 145 initial human gene set intact.

#### 146 Age binning

The number of orthologs assigned to any specific age is widely variable, with older age groups, such as those genes present in the last common ancestor of all eukaryotes, comprising thousands of genes while more modern age groups, like the genes found only in primates, could have tens of genes or no genes at all. To overcome this uneven sampling across ages, orthologs were grouped into age bins with at least 1000 orthologs. This binning process started in the oldest ages and progressed by collapsing ages into a single bin until the total ortholog count

153 exceeded 1000, then a new bin would be created and the process would repeat. Due to

154 differences in total gene counts and the specific distributions of genes, each species has a slightly

155 different count of total age bins.

#### 156 Determining the prevalence of pleiotropy

157 The prevalence of pleiotropy was determined on a per-gene basis using two methods: 158 Gene Ontology unique biological processes (BP) and STRING database protein-protein 159 interactions (PPI). These measures were selected because they provide complimentary insight 160 into the pleiotropic characteristics of a gene. A gene's BP count relates to a purely functional 161 view of pleiotropy, where each unique function the gene carries out is counted as a BP and 162 having a greater BP count indicates that the gene affects more traits (He & Zhang, 2006; 163 Papakostas et al., 2014; Williams et al., 2023). A gene's PPI count instead is agnostic to 164 annotated functional traits and focuses instead on the number of other proteins it interacts with, 165 which provides a different kind of proxy for the degree of pleiotropy (He & Zhang, 2006; 166 Papakostas et al., 2014; Williams et al., 2023). BP counts were calculated by retrieving Gene 167 Ontology data associated with that gene from the OMA database using the omadb python API 168 (Altenhoff et al., 2021). All GO evidence types were used to determine total biological process 169 involvement, but multiple entries of the same process were ignored. PPI counts were determined 170 for each protein by counting the number of interactions in the STRING database that surpassed 171 the .66 confidence score threshold. Confidence scores are provided for each interaction in 172 STRING, generated using the available evidence for a protein-protein interaction, and the chosen 173 threshold indicated high confidence that the given interaction exists (Szklarczyk et al., 2023). 174 For both BP counts and PPI, larger numbers indicated more pleiotropy. At no point did we

deploy a hard cutoff to determine if a gene was pleiotropic or non-pleiotropic, instead opting toderive distributions of these pleiotropic measures across ages.

#### 177 Determination of broad function

178 Using the Gene Ontology hierarchy of terms, we determined the broad category that each 179 specific GO term was associated with, for example pyrimidine nucleobase biosynthetic process 180 is a specific (leaf) term that falls under the *metabolic process* GO term. The broad categories into 181 which we grouped genes were the child terms of the Biological Process term (GO:0008150), 182 which is the root of the biological process ontology. To each protein we then assigned 183 association to a broad functional category if one of its functions fell in that category. Ultimately, 184 16 functional categories were present in each species, but for simplicity we elected to analyze 185 only the metabolic, cellular, and developmental processes as these were among the most 186 abundant in each species. We plotted immune system processes for the *M. musculus* and *H.* 187 sapiens datasets as well, because the immune system is thought to be highly pleiotropic 188 (Sivakumaran *et al.*, 2011) and provided a basis of comparison to other selected processes. We 189 plotted immune system processes only for M. musculus and H. sapiens because other species had 190 immune system process gene counts that were too low for effective comparison.

#### 191 Duplicated gene analysis

Using the ensembl biomart (Harrison *et al.*, 2024), we identified the total set of
duplicated genes for *H. sapiens*, *M. musculus*, *D. rerio*, *D. melanogaster*, and *C. elegans*. *A. thaliana* was excluded from this analysis because it was not available in the biomart dataset.
Each gene was then assigned a value based on the number of paralogs it had in the dataset, and
any gene with at least one paralog was considered to be duplicated in further analysis.

197

#### 198 Results

#### 199 The prevalence of pleiotropy increases with gene age across eukaryotes

200 Generally speaking, genes in middle-aged and older time bins have more pleiotropy, as 201 measured by biological process count, than genes in younger time bins (Fig. 1). This finding is 202 also recapitulated in the protein-protein interaction (PPI) results (Fig. S1). In older age bins the 203 distribution of biological processes tends to be unimodally distributed around the median value, 204 but younger genes develop skewed or bimodal distribution of biological process count, with a 205 prominent peak near the mean and a second peak around the single-process baseline. These 206 secondary peaks are not observed in the PPI plots, where even the age bins with the lowest mean 207 PPI are still well above the floor of 1 PPI. The statistical relationship between gene age and BP 208 count was tested using a one-way ANOVA, and for each species there is a significant 209 relationship between gene age and BP count (Table 1) as well as between gene age and PPI 210 (Table S1).

211 While, broadly speaking, younger gene age corresponds to less pleiotropy than middle 212 and older age, the qualitative dynamics of this pattern are relatively species-specific. In H. 213 sapiens and *M. musculus*, the mean BP rapidly climbs as genes age and then levels out across the 214 remaining ages. In D. rerio there is more variability in BP accumulation as age increases, with 215 some age bins having lower mean BP counts than the next youngest age bin. D. melanogaster 216 and A. thaliana exhibit a stepwise increase in BP count rather than a smooth accumulation of 217 biological processes. C. elegans diverges the most from the other organisms, with its youngest 218 age bins having approximately equal BP counts, and only the very old genes increasing in mean 219 BP count. These patterns only manifest in the BP plots; when looking at PPI there is a 220 universally smooth trend of constant accumulation of interactions as orthologs age.

### 221 The prevalence of pleiotropy differs across age and functional groups

222 The prevalence of pleiotropy is distinct among groups of functions (Figs. 2, S2). 223 Metabolic processes almost always have the lowest prevalence of pleiotropy, and in H. sapiens 224 and *M. musculus*, the highest prevalence of pleiotropy occurs in the immune gene set in the 225 oldest age group. For D. rerio, D. melanogaster, C. elegans, and A. thaliana (for which immune 226 processes were not included) the developmental genes in the oldest age are the most pleiotropic. 227 Interestingly these differences are not consistent between the PPI and BP plots, likely due to the 228 kind of pleiotropy they represent (Fig. S2). Affirming our genome-wide findings, no young 229 functional group has a higher prevalence of pleiotropy than the corresponding old functional 230 group in the same organism. For each species, a two-way ANOVA was conducted to evaluate 231 the relationship between age category, trait, and BP. The relationships between both age group 232 and BP as well as trait and BP were significant for all species (Table 2). The relationship 233 between age group and PPI was significant for all species, and the relationship between trait and 234 PPI was significant in all species except for *D. rerio* (Table S2). 235 The number of genes in each functional group differs significantly, so we calculated a 236 bootstrapped mean for each functional group based on the group with the smallest number of 237 orthologs per species. We then determined 95% confidence intervals around these bootstrapped 238 means and found that these means were distinct based on their disjoint confidence intervals

(Figs. S4, S5), suggesting that the observed differences in distributions were likely not due tosample size.

#### 241 Gene duplication events do not decrease the prevalence of pleiotropy

For each species, a two-way ANOVA was conducted to evaluate the extent to which geneage and duplication status could explain biological process count. The relationships between age

244 and BP as well as duplication status and BP were significant for all species (Table 3). The 245 relationship between gene age and PPI was significant for all species, and the relationship 246 between duplication status and PPI was significant in all species except D. melanogaster (Table 247 S3). Genes without paralogs generally have fewer biological processes associated with them than 248 genes with paralogs. Several individual age bins across the species show duplicated genes with a 249 prevalence of pleiotropy that was less than or equal to the non-duplicated genes, but the overall 250 trend supports increased pleiotropy in genes with paralogs (Fig. 3). This trend is also seen when 251 PPI is the measure of pleiotropy (Fig. S3). Furthermore, the exact trends between species vary, 252 with some species showing consistent differences between duplicated and singleton genes 253 throughout the entire age range (H. sapiens, D. rerio, D. melanogaster) while others have 254 inconsistent patterns across gene age (M. musculus). The C. elegans plots are challenging to 255 interpret due to the extremely limited duplicated gene counts.

256

#### 257 Discussion

258 In this study, we have identified a potentially generalizable relationship between gene age 259 and the prevalence of pleiotropy. Our results suggest that young genes accumulate functions as 260 they age, eventually trending toward a plateau that could be indicative of a carrying capacity of 261 function rather than a balance between gain and loss of function rates. We have also shown that 262 genes belonging to metabolic, developmental, cellular, and immune functional groups differ in 263 their prevalence of pleiotropy, and these differences are preserved within age groups. These 264 results lay the groundwork to better understand the evolution and maintenance of pleiotropy in 265 multicellular eukaryotes.

266 While previous studies suggest that both PPI and BP act as complementary proxies of 267 pleiotropy (Williams et al., 2023), there are differences between the two measures that may 268 explain the qualitative differences observed. Namely, PPI measures the number of interacting 269 partners a protein has and is agnostic to the action of the focal protein on those partners. BP 270 describes distinct actions a protein carries out, but any one of those processes could have many 271 protein-protein interactions associated with it. In the most extreme cases, as with the human 272 CAPN13 protein, a protein can be associated with a single BP (proteolysis) and have more than 273 750 annotated protein-protein interactions (Sorimachi et al., 2011). The definition of pleiotropy 274 based on BP count is closely related to the concept of moonlighting, where a protein carries out 275 distinct actions in different contexts (Matos et al., 2022). We could therefore interpret our results 276 as indicating that gaining more interactions is easier than developing novel functionality, which 277 matches expectations of protein evolution (i.e. micro- and macrotransitions) (Jayaraman et al., 278 2022). This explanation concurs with the observed results, where increases in PPI are largely 279 consistent between ages (Fig. S2) while increases in BP are more abrupt and taper off 280 significantly with age (Fig. 2).

281 Previous work has implicated immune systems as being highly pleiotropic (Sivakumaran 282 et al., 2011; Williams et al., 2023), and our work has expanded on these findings by directly 283 comparing the abundance of pleiotropy in genes across a set of traits, enabling us to determine 284 more systemic variations in the prevalence of pleiotropy. We have shown that metabolic genes 285 tend to have low levels of pleiotropy, as measured by PPI and BP, compared to cellular process 286 and developmental genes. These findings are largely stable across the species we investigated 287 (Figs. 2, S2). It is unclear if these relationships are due to differences in the ability of each trait to 288 tolerate pleiotropy or if some traits are simply more liable to become pleiotropic through

289 evolutionary processes. We do not expect that the differences observed are strictly due to the 290 importance (in a fitness sense) of the traits in question as development, metabolism, and cellular processes are all fundamental to the survival and reproduction of multicellular organisms. The 291 292 difference could instead be attributed to the nature of the proteins that are associated with each 293 process. For example, signaling proteins tend to be more pleiotropic than other protein classes 294 (Williams et al., 2023), potentially indicating that the abundance of signaling proteins associated 295 with immunity and development are partially responsible for inflating the prevalence of 296 pleiotropy in genes associated with these traits.

297 Despite our expectation that gene duplications would reduce the prevalence of pleiotropy, 298 the vast majority of duplicated genes either maintain, or even increase, their prevalence of 299 pleiotropy compared to age-matched singleton genes. A review of the functional changes 300 associated with duplicated genes has found that complete subfunctionalization following gene 301 duplication is likely to be rare (Janiak et al., 2019; Kuzmin et al., 2022). Several processes may 302 act together to promote the maintenance of pleiotropy in these genes, including partial 303 subfunctionalization where both genes maintain at least some activity for each function, neo-304 functionalization following subfunctionalization, dosage amplification where the excess gene 305 products associated with multiple copies is beneficial, and backup compensation where having a 306 second copy of a critical gene safeguards against loss of function (Kuzmin et al., 2022). When 307 paralogs form complexes, selective pressure can lead to correlated mutations and provide another 308 avenue for paralogs to actively increase pleiotropy, as one member of a complex acquiring a 309 novel function can force others to acquire that same function (Marchant et al., 2019). Critically, 310 our measures of pleiotropy cannot assess how well duplicated genes carry out their shared 311 functions. Thus, a gene that has lost much but not all of its functional capacity following a

312 duplication event (partial subfunctionalization) would still be pleiotropic in our analyses (Janiak 313 et al., 2019). When looking for direct examples of this kind of interaction in our data we find that 314 many duplicates, such as myogenic differentiation 1 (MYOD1) and myogenic factor 5 (MYF5) in 315 mice, share a significant number of biological processes despite these genes having distinct and 316 independent roles in specialization and differentiation (Conerly et al., 2016). Critically, even 317 though MYF5 does not induce robust transcription while MYOD1 does, it is still associated with 318 the biological process 'regulation of DNA-templated transcription'. We are then left to believe 319 that duplication events play a relatively small role in the reduction of pleiotropy in the genome, 320 supporting the hypothesis that the accumulation of pleiotropic functions is instead limited by 321 reduced evolvability as genes acquire novel functions (Fraïsse et al., 2019).

322 This work highlights the dynamism of pleiotropy and reveals a previously poorly 323 understood link between a gene's pleiotropic status and its age. This relationship holds across six 324 distantly related model organisms, suggesting that it could be generalizable among multicellular 325 organisms. Further work could expand these findings to the single celled eukaryotes and other 326 domains of life to examine their broader generality. It is unlikely that the variance in the 327 prevalence of pleiotropy observed between traits is explained by the genes in the oldest age 328 groups being common to multiple species as we observed similar trends amongst the more 329 species-specific young genes. However, it would be interesting to use the pseudo-replication of 330 the same ortholog present in multiple species to study the accumulation of functions. For 331 example, does the same ortholog have the same functions across each species? If not, are there 332 some species where functions are similar while others have diverged? Such analysis could be 333 expanded to conduct maximum likelihood ancestral state reconstructions to determine if the 334 initial functions of a gene bias the other functions it may evolve over time.

335	Our work also suggests that the observation that immune genes are disproportionately
336	pleiotropic in humans (Sivakumaran et al., 2011) may hold more broadly across species and gene
337	age groups. This raises profound questions about the nature of genomic organization and
338	function. For example, is the prevalence of pleiotropy dictated by the importance (in a fitness
339	related manner) of the trait or is it intrinsic to the protein type and cellular localization that are
340	necessary for the trait to function? Future investigation into mechanisms at the gene or cellular
341	level could provide fundamental insight into the maintenance of pleiotropy despite the potential
342	for constraining rapid adaptation (Guillaume & Otto, 2012; Fraïsse et al., 2019; Williams et al.,
343	2023).

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# 349 Author Contributions

R.A. and A.T.T. conceived the project. A.T.T. provided funding. R.A. and A.T.T.
designed the analyses, and R.A. conducted them. R.A. and A.T.T. wrote the manuscript.

# 353 Conflict of Interest Statement

The authors declare no conflicts of interest.

# 355356 Data Accessibility

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The data and code used to generate these results will be made available on Dryad upon manuscript acceptance (as specified by journal policy).

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### 441 List of Supplemental Figures

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   gene age, and the primary trait associated with a gene.
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- 448 Figure S1: Old genes have an elevated prevalence of pleiotropy compared to young genes
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# 454 Tables

Species	Formula	df	Sum. Sq.	Mean Sq.	F	PR(>F)
H. sapiens	BP~Age	.ge 1 7723.2		7723.26	49.55	<2.01e-12
M. Musculus	BP~Age	1	2.46e4	2.46e4	163.89	<2e-16
D. rerio	BP~Age	1	452	452	18.33	<2e-5
D. melanogaster	BP~Age	1	2.3e4	2.3e4	521.29	<2e-16
C. elegans	BP~Age	1	7079.67	7079.67	359.06	<2e-16
A. thaliana	BP~Age	1	1.25e4	1.25e4	148.39	<2e-16

455 Table 1: ANOVA tables for the relationship between the number of biological processes a gene456 is associated with and the age of that gene.

457

458 Table 2: Two-way ANOVA tables for the relationship between biological process count, gene459 age, and the primary trait associated with a gene.

Species	Factor	df	Sum. Sq.	Mean Sq.	F	PR(>F)
II. gamiona	Age	1	338.05	338.05	453.56	<2e-16
n. sapiens	Traits	4	166.01	41.5	55.56	<2e-16
M Mugaulug	Age	1	359.48	359.48	453.88	<2e-16
M. Musculus	Traits	4	174.26	43.56	55.01	<2e-16
Durania	Age	1	76.46	76.46	140.26	<2e-16
D. rerio	Traits	3	192.18	64.06	117.52	<2e-16
D malanogastar	Age	1	503.41	503.41	922.33	<2e-16
D. meianogasier	Traits	3	150.33	$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	91.81	<2e-16
C alagana	Age	1	87.75	87.75	142.13	<2e-16
C. elegans	Traits	3	182.00	60.67	98.26	<2e-16
A thalian a	Age	1	283.68	283.68	549.60	<2e-16
A. indiiana	Traits	3	292.62	97.54	188.63	<2e-16

460

	Species	Factor	df	Sum.	Mean Sq.	F	PR(>F)
				Sq.			
-	H. sapiens	Age	1	8217.78	8217.78	53.16	<2e-16
		Duplication	1	2.21e4	2.21e4	142.98	<3.21e-13
	M. Musculus	Age	1	1.64e4	1.64e4	110.97	<2e-16
		Duplication	1	4.96e4	4.96e4	334.96	<2e-16
	D. rerio	Age	1	509.25	509.25	21.00	<4.62e-6
		Duplication	1	7658.53	7658.53	315.83	<2e-16
	D. melanogaster	Age	1	2.23e4	2.23e4	517.50	<2e-16
-		Duplication	1	597.49	597.49	13.85	1.99e-4
	C. elegans	Age	1	6864.47	6864.47	348.57	<2e-16
		Duplication	1	455.62	455.62	23.14	1.53e-6

462 Table 3: Two-way ANOVA tables for the relationship between biological process count, gene463 age, and gene duplication.

464

#### 466 Figures



Figure 1: Older genes have an elevated amount of pleiotropy as measured by biological process
(BP) count. The y axis shows violin plots built on the log<sub>10</sub>(biological process count) for all
genes in each age bin. The x axis shows age bins for genes, from oldest on the left to the
youngest on the right. Orthologs that did not have a BP count were excluded from these plots.
The number of genes present in each binned age group is denoted above the corresponding
violin.

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Figure 2: The prevalence of pleiotropy is dependent on gene function. Plots show genes in the oldest and youngest time bins (labeled Oldest and Youngest). The y axis shows violin plots built on the log<sub>10</sub>(biological process count) for all genes in a given age bin, separated into 4 functional groups (metabolic, cellular process, developmental, and immune processes). Orthologs that did not have a BP count were excluded from these plots.

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Figure 3: Genes with paralogs are more pleiotropic than genes without paralogs. The y axis
shows violin plots built on the log<sub>10</sub>(biological process count) for all genes in a given age bin.
The x axis shows age bins for genes, from oldest on the left to the youngest on the right. Genes
with paralogs are shown in orange, genes without paralogs are shown in blue. Orthologs that did
not have a BP count were excluded from these plots.