

Colorimetric Determination of Dissolved Oxygen: Assessment of Methodological Influences on Iodine Spectra, Isosbestic Point, Precision and Accuracy

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and iodine concentrations on oxygen concentration determinations are ambiguous. We found that, while iodine concentration and absorbance wavelength impart minor influences, temperature changes during sample analysis alter the fractionation of two major iodine species $(I_2 \text{ and } I_3^-)$ and affect their spectra, with the latter effect imparting the greatest influence on oxygen concentrations. An empirical molar extinction coefficient was defined for the mixture (consisting of 96% I_3^- and 4% I_2), yielding values of 2282, 1115,

and 792 M⁻¹ cm⁻¹ at 25 °C at the most commonly used wavelengths of 430, 456, and 466 nm, respectively. Altering the temperature led to variances in absorbance by 0.33%, 0.42%, and 0.51% °C[−]¹ at 430, 456, and 466 nm, respectively. Therefore, for absorbances measured at 456 nm, a room temperature correction can be applied to the empirical molar coefficient by 1115 M⁻¹ cm⁻¹ × [1 + (room temperature − 25 °C) × 0.0042]. Reducing room temperature differences between samples and calibration solutions became the most important factor in maintaining the precision and accuracy of the measurement. With these precautions, a precision of ∼0.2% (coefficient of variation) over the normal surface water oxygen concentration of around 250 *μ*M can be readily achieved at all tested wavelengths. This method offers a linear oxygen concentration range of 0−650 *μ*M.

1. INTRODUCTION

The precise quantification of dissolved oxygen in freshwater and seawater is crucial to identifying subtle environmental differences in both temporal and spatial trends. Several methods are available to measure dissolved oxygen, including the Winkler titration, $\frac{1}{2}$ the colorimetric detection of iodine by spectrophotometer, $^{3-5}$ $^{3-5}$ $^{3-5}$ $^{3-5}$ $^{3-5}$ and various oxygen sensors. 6,7 6,7 6,7 Although oxygen sensors are becoming increasingly common in laboratory and field studies,^{[6](#page-9-0)} data quality by this method still heavily relies on precise, accurate and reliable wet-chemical calibration. The classic Winkler titration is the first choice for this purpose. This method employs three analytical steps: oxygen fixation with a manganese hydroxide oxide precipitate, liberation of iodine by adding acid in the presence of excess iodide, and titration with thiosulfate. Since the titration process requires several hands-on steps, which can be time-consuming, several colorimetric methods^{[4,5,8](#page-9-0)} have been proposed as alternatives to the titration procedure, especially for clean waters like open oceans and nonturbid freshwaters. The advantages of using colorimetric methods are not only due to their simplicity and high precision but also to the effectiveness

of eliminating iodine vaporization loss during the measurement.

Although the colorimetric approach has been used as a routine tool for seagoing expeditions over the last three decades, several analytical concerns still need clarification.

(a) *Regarding reagent strengths*. Since the colorimetric method has been established for measuring a mixture, it is essentially empirical. It is assumed that the three species (i.e., I_2 , I_3^- , and I^-) have reached equilibrium in the final solution. However, in reality, the oxygen within the sample and the added $KIO₃$ both consume I^- , thus the ratio of iodine species can be altered from one

Received: October 5, 2024 Revised: October 28, 2024 Accepted: October 31, 2024 Published: November 11, 2024

sample to another. The fractionation of iodine species must be calculated to identify these influences.

- (b) *Regarding the wavelength chosen for detection*. A wavelength of 456 nm was first proposed by Pai et al. $⁴$ and</sup> then 430 nm by Roland et al.^{[5](#page-9-0)} However, Labasque et al.^{[8](#page-9-0)} suggested that the measurement should be carried out at an "isosbestic" wavelength of 466 nm where both I_2 and I3 [−] species share the same molar extinction coefficient. According to this concept, the determination will not be affected if the iodine fraction is altered. As shown in Labasque et al. 8 and as will be shown below, there was a significant temperature effect on the isosbestic point (the absorbance may change approximately 0.5% per 1 °C at 466 nm). The reason for this effect remains to be identified.
- (c) *Regarding how to minimize the temperature effect*. Ocean water samples (i.e., as a depth profile) can be collected from in situ temperatures ranging between 2 and 35 °C. Additionally, adding sulfuric acid during the last step will suddenly release heat before detection. Analytical criteria need to be established to ensure all measurements (as well as the standardization) are made under the same conditions.

These concerns will be carefully examined in this study.

1.1. Equations Involved in the Colorimetric Winkler Reactions. While basic equations for the Winkler method are available in numerous literature sources, $4,5,8$ $4,5,8$ $4,5,8$ $4,5,8$ we present more detailed equations related to colorimetry, specifically, to focus on how the fractionation of iodine alters absorbance with increases in oxygen or iodate levels.

(i) Oxygen is fixed by adding Mn^{2+} and NaOH in a flaredmouth bottle, usually referred to as a biological oxygen demand (BOD) bottle, to form precipitates

$$
O_2 + 2Mn^{2+} + 4OH^- \Leftrightarrow 2MnO(OH)_2 \tag{1}
$$

(ii) Iodine is released by dissolving the precipitates with acid in the presence of iodide

> $MnO(OH)_{2} + 4H^{+} + 2I^{-} \Leftrightarrow Mn^{2+} + I_{2} + 3H_{2}O$ (2)

(iii) Iodine further reacts with excess iodide to form triiodide

$$
I_2 + I^- \Leftrightarrow I_3^- \tag{3}
$$

(iv) The final fractions of each iodine species are dependent upon the concentration of iodide present and the stability constant K for I_3^-

$$
K = \frac{[I_3^-]}{[I_2][1^-]} \quad K = 725 \text{ at } 25 \text{ °C}
$$
 (4)

According to Burger and Liebhafsky^{[9](#page-9-0)} the K value at various temperatures *T* (°C) can be estimated by an empirical equation

$$
K = 1840/(T + 273.15) + 0.415
$$
 (5)

(v) The molar extinction coefficient ε_2 for I_2 can be obtained by preparing a known concentration of pure molecular iodine solution and measuring its absorbance

$$
\varepsilon_2 = \text{Abs}/b/[\text{I}_2] \tag{6}
$$

where b is the cuvette light path (usually 1 cm).

(vi) The molar extinction coefficient ε_3 for I_3^- can be calculated by measuring the absorbance of a mixed solution of I_2 and NaI and solving for the triiodide fraction. Assuming the final concentration of triiodide is *x*, *A* and *B* are the initial concentrations of I_2 and I^- , the equilibrium can be expressed as

$$
K = \frac{x}{(A - x)(B - 2A - x)}
$$
(7)

where $B > 2A$. The final concentration of triiodide can be solved as

$$
[I_3^-] = x = [(B - A + (1/K))
$$

$$
- \sqrt{(B - A + (1/K))^2 - 4(AB - 2A^2)}]/2
$$
(8)

And the final concentration of I_2 is

$$
[I_2] = B - x \tag{9}
$$

The absorbance measured refers to a combination of two major iodine species

$$
Abs = \varepsilon_2 b[I_2] + \varepsilon_3 b[I_3^-]
$$
\n(10)

Thus, the value of ε_3 can be estimated following the equation below when ε_2 is obtained previously

$$
s_3 = \frac{\text{Abs} - \varepsilon_2 b[I_2]}{b[I_3^-]}
$$
 (11)

(vii) In a mixed $I_2 + I_3$ ⁻ solution, a molar ratio *r* can be defined as

$$
r = \frac{\begin{bmatrix} I_3 \end{bmatrix}}{\begin{bmatrix} I_2 \end{bmatrix}}\tag{12}
$$

The sum of I_2 and I_3 ⁻ gives total iodine $\sum I_2$

$$
\left[\sum I_2\right] = \left[I_2\right] + \left[I_3\right] \tag{13}
$$

If *r*, *b*, ε_2 , and ε_3 are all fixed values, then eq 10 can be simplified as

$$
Abs = \varepsilon_{\rm T} \times b \times \left[\sum I_2 \right] \tag{14}
$$

where ε_{T} can be regarded as the "empirical molar extinction coefficient" for total iodine (i.e., the sum of I_2) and I_3^-) at a given I^- concentration and a given temperature.

(viii) The calibration is usually made by adding a known amount of iodate to an iodide solution, which generates the brownish iodine color

$$
IO_3^- + 5I^- + 6H^+ \Leftrightarrow 3I_2 + 3H_2O \tag{15}
$$

In this reaction, each mole of iodate added will produce 3 mol of molecular iodine while consuming 5 mol of iodide, followed by a fraction of molecular iodine consuming another mole of iodide to form triiodide. Assuming the final concentration of triiodide is *x*, *C* and *B* are the initial concentrations of iodine and initial iodide (where *B* > 5*C*), the equilibrium can be expressed as

Figure 1. Layout of the water-circulation type thermostat spectrophotometer for the colorimetric Winkler method. (a) A thermo jacket outside the spectrophotometer holds a 60 mL BOD bottle, while a thermo cell holder inside the spectrophotometer contains a Hellma 1 cm dome-type flow cuvette. (b) The ramping rates for heating and cooling are roughly equal at 0.5 °C min[−]¹ . The efficiency of the water flow-circulating thermostat was demonstrated by filling the cuvette with an iodine solution and observing the variation of absorbance due to a change in the temperature setting. The absorbance indicates that when it becomes flat, the temperature has reached a steady state. The ramping rates for heating and cooling were similar at 0.5 °C min^{−1}. If the temperature of the sample is different from the desired temperature, an appropriate waiting time is required before the scanning or detection operation.

$$
K = \frac{x}{(C - x)(B - 5C - x)}
$$
(16)

Accordingly, the final concentration of triiodide will be

$$
\begin{aligned} \left[I_3^-\right] &= \left[\left(B - 4C + (1/K)\right) \right] \\ &- \sqrt{\left(B - 4C + (1/K)\right)^2 - 4(BC - 1SC^2)} \right] / 2 \end{aligned} \tag{17}
$$

The final concentration of molecular iodine is

$$
[I_2] = C - x \tag{18}
$$

Measuring the absorbance of this solution and calculating by eq [14](#page-1-0) an ε_T value can be obtained in the spiking experiment

$$
\varepsilon_{\rm T} = \text{Abs}/b / \Big[\sum I_2 \Big]
$$
 (19)

(ix) There are two types of reagent blanks involved in the colorimetric method, namely RB_1 and RB_2 , representing the iodine impurities in the R2 reagent $(NaI + NaOH)$ and oxygen contents in both R1 (Mn^{2+}) and R2 reagents. The contributions of $RB₁$ to the absorbance can be obtained experimentally by carrying out the same procedure for oxygen but adding reagents in reverse order (i.e., add R3 and R2 first, then R1). In our experience, the absorbance of $RB₁$ is very small, mostly 0.000 and rarely exceeds 0.001.

 $RB₂$ refers to the oxygen content within the dense R1 $(3 M MnCl₂ with a density of 1.30 g mL⁻¹) and R2 (4$ M NaI and 8 M NaOH with a density of 1.68 ${\rm g\;mV}^{-1})$ reagents. It is difficult to measure directly but can be evaluated according to Murray et al. 10 10 10 who measured the oxygen concentrations in these two dense reagents. By calculating the amount of reagent used in a sample

$$
[O_2]_{RB2} = ([O_2]_{R1}V_1 + [O_2]_{R2}V_2)/(V_b + V_3)
$$
 (20)

where V_1 , V_2 , and V_3 are added volumes for R1, R2, and R3. The excess $[O_2]$ for this reason is approximately 0.5 *μ*M, which can be subtracted directly from each measured result.

(x) Converting measured absorbance to oxygen concentration: During the oxygen fixation step, for a bottle with an inner volume of $V_{\rm b}$, the actual volume of sample is $V_{\rm b}$ $-V_1 - V_2$ (e.g., 60 – 0.5 – 0.5 = 59 mL), and the final volume after adding R3 is $V_b + V_3$ (e.g., 60 + 0.5 = 60.5) mL). Therefore, the resultant total iodine will be

$$
\left[\sum I_2\right] = 2 \times [O_2]_{\text{sample}} \times (V_b - V_1 - V_2)
$$

$$
/(V_b + B_3)
$$
(21)

By combining this equation with eq [13](#page-1-0) and considering both reagent blanks, $[O_2]_{\text{sample}} = (Abs - RB1)/2 \times (V_b)$ $(V_3)/(\tilde{V}_b - V_1 - V_2) - [\dot{O}_2]_{R B 2}$ or

$$
[O_2]_{\text{sample}}(\mu M) = \frac{(Abs - RBI)}{2 \times b \times \epsilon_{\text{T}}} \times \frac{(V_b + V_3)}{(V_b - V_1 - V_2)}
$$

× 10⁶ - 0.5 (22)

The dilution term of $(V_b + V_3)/(V_b - V_1 - V_2)$ is nearly constant, even though the bottle volume V_b may vary significantly. For example, in the Shibala procedure, 4 the value is $60.5/59 = 1.0254$; in the Labasque^{[8](#page-9-0)} procedure, it is 146.9/144.2 = 1.019. A bottle volume error of \pm 5% will only give a final variation of ± 0.1 %. Thus, for

Figure 2. Illustration of the colorimetric Winkler method for oxygen measurement and calibration (following procedure outlined by Pai et al.^{[4](#page-9-0)}). Reagents used are R1 (containing 3 M Mn²⁺), R2 (containing 4 M NaI and 8 M NaOH) and R3 (10 N sulfuric acid). Potassium iodate (KIO₃) is used as a standard. (a) Oxygen in a sample is fixed by adding R1 and R2 to form a precipitate. Upon measurement, the precipitate is dissolved by adding R3, releasing iodine and giving a brown color. The absorbance of the mixture is proportional to the $[O_2]$ in the original sample. (b) Distilled water (DW) is added with the same amounts of reagents but in reverse order. No precipitation is formed in the bottle, to which a known amount of $KIO₃$ is added, which generates an iodine color. An empirical molar extinction coefficient for this mixture can be obtained for calibration purposes.

example, if ε_T is measured to be 1115 M⁻¹ cm⁻¹ at 25 $\rm{^{\circ}C}$ using the Shibala procedure, $\rm{^{\circ}}$ [eq](#page-2-0) 22 can be simplified as

$$
[O_2](\mu M) = Abs(corr) \times 460 - 0.5
$$
 (23)

2. EXPERIMENTAL SECTION

2.1. Thermostat Spectrophotometer. Since the present study focuses on the temperature effect on the iodine color, most experiments were carried out under very precise temperature control. To do this, a double-beam spectrophotometer (Shimadzu 1800A, Japan) was equipped with a watercirculation type temperature-controlled flow system, including a sample-loading jacket and a cuvette holder ([Figure](#page-2-0) 1a). The temperature of the water tank was controlled by a thermostat (Firstek, Taiwan) with a 1 KW heater and a 700 W cooler. A Hellma 1 cm dome-type flow cuvette (Hellma GmbH & Co, Germany) was used, which was surrounded by a thermo holder. The sample, contained within a 60 mL BOD bottle (Wheaton) was first placed in the thermo jacket and held for several minutes before being siphoned up to fill the cuvette volume. Then the sample was trapped in the cuvette to wait for another several minutes to reach a steady state before proceeding to either spectral scanning or to measure at a fixed wavelength. The ramping rate of the thermostat system was checked by an iodine color solution and found to be about 0.5 °C min[−]¹ for both the heating and cooling processes ([Figure](#page-2-0) 1b).

2.2. Procedure for Routine Measurement Using an Ordinary Spectrophotometer. The routine procedure for measuring oxygen can be described graphically, as shown in Figure 2a. It is based on the protocol suggested by Pai et al.^{[4](#page-9-0)} (also known as the Shibala procedure). For this, the water

sample is filled into a BOD bottle (volume $V_b = 60$ mL), to which are added 0.5 mL of reagent R1 (containing 3 M $MnCl₂$), 0.5 mL of reagent R2 (containing 8 M NaOH and 4 M NaI), the bottle is then stoppered, well mixed and allowed to stand for few hours until all precipitates are settled down to the bottom. Upon measurement, the bottle is opened and placed on a stir plate, and an aliquot of 0.5 mL of R3 (10 N sulfuric acid) is added while stirring. The brownish-colored solution is promptly sipped and transferred to fill a flow cuvette installed in a spectrophotometer, and the absorbance is recorded. The brownish color results from a mixture of molecular iodine (I_2) and triiodide (I_3^-) . Since their molar ratio is fixed, the absorbance is directly proportional to the oxygen concentration of the sample. DDW refers to double distilled water.

The procedure for the standardization is illustrated in Figure 2b. A BOD bottle filled with distilled water is treated with the same amounts of reagents but in reverse order. As the acidic R3 is added first, no precipitation will form; thus, the absorbance of this solution refers to the reagent blank, which has the same matrix as the oxygen measurement. The addition of known amounts of $KIO₃$ to this solution produces an iodine color, and the absorbance obtained can be used to construct a calibration curve for the quantification of oxygen.

2.3. Definition of "Temperature". In this study, two types of spectrophotometers were used. A thermostat spectrophotometer was employed to scan the spectra of pure I_2 and I_3 ⁻ solutions to identify the isosbestic point, while an ordinary spectrophotometer was used for routine measurements. To avoid any potential misunderstandings or confusion for readers, we here define the term "temperature" more clearly and precisely as follows:

 T_b : Temperature of the circulating water in the thermostat, which might slightly differ from the actual temperature of the

Table 1. Theoretical Calcultion of Iodine Fractionation Affected by Increasing Temperature Using Various Protocols*^a*

a Calculations were based on an assumed oxygen concentration of 250 *μ*M.

Table 2. Theoretical Calculation of Iodine Fractionation Affected by Increasing Oxygen Concentration Using Various Protocols*^a*

	vol. of bottles (mL)	vol. of R2 (mL)	$[I^-]$ in R2 (M)	final volume (mL)	assumed $[O_2]$ (μM)	fractionation		
protocol						$(I_3^-)\%$	(I_2) %	change of $(I_3^-)\%$ per 100 μ M O ₂
Shibala	60	0.5	$\overline{4}$	60.5	100	95.93%	4.07%	
					200	95.86%	4.14%	$-0.07%$
					300	95.78%	4.22%	
Labasque	146	0.9	$\overline{4}$	146.9	100	94.55%	5.45%	
					200	94.42%	5.58%	$-0.14%$
					300	94.29%	5.71%	
APHA	300	\perp	0.87	301	100	63.20%	36.80%	
					200	57.53%	42.47%	$-11.14%$
					300	50.38%	49.62%	
Grasshoff	50	0.5	3.61	50.5	100	96.23%	3.77%	
					200	96.17%	3.83%	$-0.06%$
					300	96.10%	3.90%	
	$a \cap 1$ \cdots 1 \cd							

a Calculations were made at a temperature of 25 °C.

flow cuvette if it is significantly higher or lower than room temperature.

*T*r: Room temperature in the laboratory where routine measurements are conducted.

*T*w: Temperature of the sample liquid after it has been left in the laboratory for at least 4 h to equilibrate with the surrounding environment. It is obtained by reading a thermometer placed in a bottle filled with water next to the spectrophotometer.

T_c: Temperature of the sample immediately after acidification with R3 sulfuric acid, which releases heat and temporarily raises the temperature by roughly 0.8 °C.

*T*a: Actual temperature of the sample liquid trapped in the cuvette upon detection, which cannot be measured directly.

Since only T_w can be reliably recorded, in subsequent routine practice with an ordinary spectrophotometer, the term "temperature" should refer to T_w .

3. RESULTS AND DISCUSSION

3.1. Comparison of Existing Protocols. The two most adopted colorimetric protocols $4,8$ $4,8$ $4,8$ and two titration meth $\mathrm{ods}^{11,12}$ $\mathrm{ods}^{11,12}$ $\mathrm{ods}^{11,12}$ were compared based on bottle volumes, reagent strengths, and possible conditions that may cause variations in the final iodine fractionation. For simplicity, these four methods are referred to as Shibala, Labasque, APHA and Grasshoff in text and tables.

fractionation

According to eq [4](#page-1-0), the iodine fraction is heavily related to two factors, i.e., the I⁻ concentration and the formation constant K; the latter is a function of temperature. Theoretical calculations on alterations to the fractionation of iodine due to temperature are made and shown in Table 1. The original data provided by Burger and Liebhafsky⁹ showed a clear linear relationship between ln *K* and the reciprocal of temperature, leading to an empirical equation shown in eq [5](#page-1-0). In this study, the *K* values were calculated to be 806, 725, 655, and 593 at temperatures of 20, 25, 30, and 35 °C, respectively. Four protocols were compared using those *K* values, assuming the oxygen concentration was 250 *μ*M. For example, for the Shibala protocol an aliquot of 0.5 mL of R2 (containing 4 M I⁻) was added to a 60 mL bottle, resulting in an initial I⁻ concentration of 0.033 M in the final 60.5 mL solution. At 25 $^{\circ}$ C with a *K* value of 725, the final I₃^{$-$} fraction is 95.82%, which

Figure 3. (a) Direct wavelength scan (400–700 nm) of two iodine solutions at various temperatures ($T_b = 20-35$ °C). The pure I₂ solution containing 0.0563 g I₂ flakes in 1 L gives an [I₂] of 221 μ M. The NaI+I₂ solution is a mixture of 0.0527 g I₂ flakes and 34.54 g NaI in 1 L. (b) Molar extinction coefficients (400–480 nm) for I₂ and I₃ [−] at different temperatures: (blue) 20 °C (green) 25 °C (yellow) 30 °C (red) 35 °C. (c) Molar extinction coefficients for I_2 (no iodide) and I_3^- (with different iodide concentrations). (d) The molar extinction coefficients for I_3^- at 430, 456, and 466 nm show a linear correlation with the concentration of iodide. The intercept of the regression indicates a hypothetical value.

will be reduced to 94.94% if the temperature is raised to 35 °C, as calculated according to eqs [8](#page-1-0) and [9](#page-1-0). The reducing trend is estimated to be −0.09% per 1 °C. The Labasque protocol uses the same R2 but adds 0.9 mL to a 146 mL bottle to give an initial I [−] concentration of 0.0245 M in the final solution. At 25 $^{\circ}$ C the final I₃⁻ fraction is 94.36% and will reduce to 93.49% at 35 °C. The decreasing trend is ca. −0.12% per 1 °C.

For the APHA method, in which an aliquot of 1 mL of 0.87 M I[−] is added to a 300 mL bottle, the starting I[−] concentration is 0.0029 M. It does not provide enough $\left[\text{I}^-\right]$ to give a high $\text{I}_3^$ fraction value. At 25 °C the I_3^- fraction is 54.17% and drops to 49.5% at 35 °C with a decreasing trend of −0.87% per 1 °C. The increase of I_2 fraction may induce vaporization loss, $13,14$ therefore it is not recommended for the colorimetric approach. In contrast, the Grasshoff protocol provides a more reasonable high I⁻ concentration (0.0357 M) and results in high $I_3^$ fraction of 96.13% at 25 °C, similar to that of the two colorimetric protocols.

In general, raising the temperature will lead to a slight decrease in I_3^- and an increase in I_2 . However, by using an I⁻ concentration of >0.03 M, these variations can be effectively limited to a negligible level within the normal oxygen concentration range.

3.2. Change of Iodine Fractionation Due to Oxygen Consumption. When oxygen is fixed and converted to $I₂$, it consumes I^- as indicated by eqs [2](#page-1-0) and [7,](#page-1-0) thus causing a slight

decrease in I^- concentration and changing the final I_2 fractionation. A theoretical calculation is also made for the four protocols. The results are summarized in [Table](#page-4-0) 2.

For the Shibala procedure, the starting I⁻ concentration is 0.033 M within the bottle. Assuming the oxygen concentration of a sample is 100 μ M, the final concentrations for I⁻, I₂ and I3 [−] will be 32855, 7.95, and 187.1 *μ*M, respectively, with a $\left[1_{3}^{-}\right]/\left[1_{2}\right]$ ratio of 23.55. This means that 95.93% of total iodine will be I_3^- and 4.07% will be I_2 in the final solution. Increasing the oxygen concentration from 100 to 300 μ M, $I_3^$ only drops slightly to 95.78%. The decreasing trend is ca. −0.07% per 100 *μ*M O₂.

For the Labasque procedure, at a dissolved oxygen concentration of 100 μ M, the final concentrations of I^- , I₂ and I_3 ⁻ will be 24299, 10.7, and 185.6 μ M, with a molar ratio of 17.35 and a final I_3^- of 94.55%. Increasing the oxygen concentration from 100 to 300 *μ*M decreases the fraction slightly to 94.29%, and the decreasing trend is also very small, at −0.14% per 100 *μ*M O₂.

Calculations of the two titration protocols are also given in [Table](#page-4-0) 1. It is clear that the APHA procedure uses a comparatively larger bottle but weaker reagent strength for I⁻, giving a very low $[I_3^-]/[I_2]$ ratio of 1.72 at an $[O_2]$ of 100 μ M. The ratio drops further as $[O_2]$ increases, which indicates that the I₂ fraction turns gradually to nearly 50% at an $[O_2]$ of 300 *μ*M. The Grasshoff procedure uses a smaller bottle and a

stronger I^- strength, giving a much higher $[I_3^-]/[I_2]$ ratio of 25.5 at an $[O_2]$ of 100 μ M, this is comparable to the high ratios provided by the Shibala and Labasque procedures.

3.3. The Iodine Spectra. Although the absorbance measured by the colorimetric method refers to a mixture of I_2 and I_3^- , it is possible to obtain their contribution individually by observing the spectra of a pure I_2 standard solution and a nearly pure I_3^- solution (following eqs [6](#page-1-0)–[19\)](#page-2-0). Two solutions were prepared for this purpose and they were scanned between 400 and 700 nm at 20, 25, 30, and 35 °C. The first solution contained 0.0563 g I_2 flakes (purity 99.8%) in 1 L of water, giving an $[I_2]$ of 224 μ M. The second solution contained 0.0527 g of I_2 flakes and 35.54 g of NaI in 1 L giving a final [I⁻] of 0.237 M, a total [I₂] of 207 μ M, and a I₃⁻ fraction of >99%.

The absorption spectra of the two solutions are shown in [Figure](#page-5-0) 3a. There are two major spectra for the pure I_2 and the I_2 +NaI mixture, each consisting of 4 lines revealing the spectra at 4 different temperatures. The two bunches of spectra have two crosses, matching the results reported by $Morel¹⁵$ and Labasque et al., 8 who described that the two species have identical absorptivities at 466 and 580 nm, respectively.

The influence of temperature on the absorption spectra was statistically significant, but the trends were opposite to that of the fractionation of iodine species. In [Figure](#page-5-0) 3b, the absorbance scale was converted to molar extinction coefficient scale. For the pure I_2 solution, the highest absorption occurred at 460 nm. The molar extinction coefficient (ε_2) measured at 20 °C was higher than that at 35 °C, with values of 729 and $708 \, \text{M}^{-1} \text{cm}^{-1}$, respectively (Table 3). The decreasing trend

Table 3. Effect of Temperature on the Molar Extinction **Coefficients**

was -0.15% °C⁻¹. For the predominantly I_3^- solution, the spectrum appeared to shift up or to the right as temperature increased. At 456 nm, the molar extinction coefficient (ε_3) was 1145 M[−]¹ cm[−]¹ at 20 °C and 1222 M[−]¹ cm[−]¹ at 35 °C (Table 3), with an increasing trend of 0.44% $^{\circ} \mathrm{C}^{-1}.$

3.4. The Effect of Iodide Concentration. In this experiment, six solutions were prepared by transferring

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different amounts of NaI (i.e., 0.571, 1.103, 2.027, 3.025, 6.116, and 31.711 g) (mol. wt. = 149.89) with distilled water into a 100 mL flask. To each flask, 1 mL of 4972 μM KIO₃ solution and 0.1 mL of 10 N sulfuric acid were added, and the final volume was adjusted to the mark. These solutions contained starting concentrations of 0.038, 0.074, 0.135, 0.202, 0.408, and 2.116 M of I [−]. These solutions were loaded into the flow cuvette at a fixed temperature of 25 \degree C for scanning between 430 and 480 nm. The results are shown in [Figure](#page-5-0) 3c and [Table](#page-7-0) 4.

Under extremely high concentrations of I^- (e.g., >0.2 M), the I_3^- fraction exceeds 99% ([Table](#page-7-0) 4), but the absorbance still increases with I⁻ concentration. This phenomenon could be attributed a further complexation of I_3 ⁻ to (I_5^-) .^{[16](#page-10-0)} However, in practical applications, I⁻ concentrations typically do not exceed 0.04 M, rendering the formation of I_5^- negligible.

3.5. The Isosbestic Point. Labasque et al.⁸ suggested that the colorimetric measurements for $[O_2]$ should be made at an isosbestic point of 466 nm, at which the molar extinction coefficients for both I_2 and I_3^- species are identical. However, the isosbestic point was found to be around 470 nm in the current study. Experimental results have demonstrated that the isosbestic point shifts toward higher wavelengths and lower intensities by increasing temperature or [I[−]].

For instance, at constant $[I^-]$, the isosbestic point (λ_{iso}) was found to be 468.7 nm at 20 °C, but drifted to 472.1 nm as the temperature increased to 35 °C [\(Figure](#page-5-0) 3b and [Table](#page-7-0) 5). The corresponding molar extinction coefficient (*ε*iso) dropped from 716.5 M^{-1} cm⁻¹ at 20 °C and to 687.6 M^{-1} cm⁻¹ at 35 °C.

Iodide concentrations can also influence the isosbestic point. Under a fixed temperature of 25 °C, the isosbestic point drifted in a similar way as described above. At an [I[−]] of 0.038 M, the isosbestic wavelength was 468.6 nm and drifted to a higher wavelength as concentrations increased ([Figure](#page-5-0) 3c and [Table](#page-7-0) [5](#page-7-0)). At an extremely high [I[−]] of 2.116 M, the isosbestic wavelength $(\lambda_{\rm iso})$ was found to be 479.8 nm. The isosbestic molar extinction coefficient $(\varepsilon_{\text{iso}})$ dropped from 710.2 to 657.4 M^{-1} cm⁻¹.

3.6. Effect of Ionic Strength. For samples with high ionic strength, such as seawater, there is a concern that the activity coefficients of different species might change, potentially altering the formation constant *K*. However, previous studies have shown that the iodine spectra are not affected by salinity between wavelengths of [4](#page-9-0)30 and 470 nm.⁴ This may be explained according to [eq](#page-1-0) 4, given that both I^- and I_3^- are monovalent ions and are present in the numerator and denominator, their activity coefficients tend to cancel each other out. This leaves only a small variation due to the activity of the nonvalent I_2 species. In an I_3^- -dominating mixed solution, such variation is very small and can be reasonably ignored.

3.7. Oxygen Measurement and Calibration of ϵ_{T} **by Spiking with KIO₃.** The calibration of ε_{T} was made by spiking various amounts of $KIO₃$ standard solution to a distilled water sample and adding all the reagents in reverse order. The bottle was placed on a stir plate, and known amounts of $KIO₃$ were added while stirring (e.g., 1.00–6.50 mL of 5000 μM KIO₃). The absorbances recorded (at 456 nm in this case) were used to calculate the empirical molar extinction coefficient ε_T following eqs [19](#page-2-0) and [20](#page-2-0). Results are shown in [Table](#page-8-0) 6. Basically, the empirical molar extinction coefficients were very consistent in the lower concentration range (0−1000 *μ*M $\sum I_2$), to be 1115 ± 3 M⁻¹ cm⁻¹ at $T_w = 25$ °C. The upper

 a Each NaI solution was added with 1 mL of 0.4972 mM KIO₃. ^{*b*}Experiments were carried out at 25 °C.

Table 5. Isosbestic Point Affected by Temperature and I⁻ Concentration

$T({}^{\circ}C)$	$[I^{-}](M)$	λ (iso) (nm)	ε (iso) $(M^{-1} cm^{-1})$								
Change Temperature											
20	0.237	468.7	716.5								
25	0.237	469.9	705.9								
30	0.237	470.9	698.1								
35	0.237	472.1	687.6								
Change $[I^{-}]$											
25	0.038	468.6	710.2								
25	0.074	468.9	709.4								
25	0.135	469.2	708.0								
25	0.202	469.6	706.9								
25	0.408	470.5	704.2								
25	2.116	479.8	657.4								

linear threshold of the calibration curve was defined as the concentration at which the slope value is dropped by 1% comparing to that at low concentration range. In the present case, the linear range was 0−1300 μ M ∑I₂, equivalent to an oxygen concentration range of 0−650 *μ*M.

3.8. Temperature Variation in Field Application. During open ocean field work, samples are often from different depths, which may cover a wide temperature range. A test was made to quantify the equilibration time required to minimize the temperature effect in a batch of environmental samples with different temperatures. To do this, aliquots of distilled water were adjusted to 32, 25, 10, and 5 \degree C, filled into the bottles, and then reagents were added. The temperatures inside those bottles were monitored for 4 h. After 1 h, the maximum temperature difference among the four bottles was reduced to 4.1 °C ([Figure](#page-8-0) 4). The gap was further narrowed to 1.2 °C after 2 h, then 0.5 °C after 3 h, and 0.2 °C after 4 h. Upon adding the sulfuric acid, the temperature of each bottle was suddenly raised by 0.8 °C, and then gradually cooled down to the room temperature of nearly 25 $^{\circ}$ C ([Figure](#page-8-0) 4). If the final temperature difference can be limited to less than 0.2 °C, the temperature influence on sensitivity (0.42% °C[−]¹) can be reasonably controlled to less than 0.1%.

Although a thermostat spectrophotometer is always preferable, most laboratories are not equipped with such thermostat devices. It should also be noted that the temperature in the cuvette chamber is typically warmer than outside the instrument. If the laboratory environment is 25 $\mathrm{^{\circ}C}$,

the actual detection temperature may be higher by 1−2 °C. Consequently, the empirical molar extinction coefficient obtained may be raised by ca. 1%. Analysts may not need to know the exact temperature during detection but can ensure all samples (including the calibration samples) undergo the same process. In this way, a highly precise measurement can still be expected without using a thermostat.

3.9. Precision and Accuracy for Routine Analysis. A replicate test was carried out on an air-saturated distilled water sample pickled at 22.0 °C and measured at $T_w = 24.8$ °C, the resultant absorbances ($n = 6$) were 1.097 \pm 0.002, 0.536 \pm 0.001 and 0.381 \pm 0.001 at wavelengths of 430, 456, and 466 nm, respectively, with a coefficient of variation (c.v.) of ∼0.2% at normal oxygen saturation level. It should be noted again that the variation in bottle volume does not contribute more than 0.1% to the variation in oxygen concentration, as described in [eq](#page-2-0) 22.

The accuracy of this colorimetric method depends solely on the spiking with known amounts of $KIO₃$, deriving an empirical molar extinction coefficient *ε*_T. Before analyzing each batch of samples, calibration must be performed carefully under a constant temperature T_w . In our laboratory, after a decade of experience, we have consistently obtained an $\epsilon_{\rm T}$ value of $1115 \pm 5 \text{ M}^{-1} \text{ cm}^{-1}$ at 456 nm when $T_w = 25 \pm 0.5$ $^{\circ}$ C. If the laboratory temperature deviates from 25 $^{\circ}$ C, a modified empirical equation can be used

$$
\epsilon_{\rm T}(T_{\rm w}) = 1115 \times [1 + (T_{\rm w} - 25) \times 0.0042] \tag{24}
$$

This equation is valid under the conditions where 20 °C < T_w $<$ 30 °C and 0.000 $<$ Abs $<$ 1.400.

4. CONCLUSIONS

The colorimetric Winkler method indeed offers advantages in the amount of time required and simplicity over traditional titration methods for determining dissolved oxygen concentration in water samples. Three factors have been identified to have influences on the precision and accuracy. These include:

- (i) *Alteration of the fractionation due to change of temperature on the formation constant K*. In the Shibala procedure,^{[4](#page-9-0)} the magnitudes are -0.09% °C⁻¹ for I₃⁻ and +2.05% $°C^{-1}$ for I₂. In the Labasque^{[8](#page-9-0)} procedure, the magnitudes are -0.12% °C⁻¹ for I₃⁻ and +2.02% °C⁻¹ for I₂.
- (ii) *Consumption of* Γ *by oxygen leads to a decrease in the* $I_3^$ *but an increase in the* I_2 . The change of fractions in I_3 ⁻

Table 6. Standardization of the Colorimetric Protocol by Spiking Known Amounts of Potassium Iodate at $T_w = 25 \text{ °C}^a$

 a^aA stock solution of 5.00 mM KIO₃ was used for spiking.

Time (min)	

Figure 4. Changes of temperature in sampling operation. Four water samples with initial temperatures of 5, 10, 25, and 32 °C were filled into BOD bottles following standard procedures, $⁴$ and the temper-</sup> atures were monitored for 240 min. The maximum difference was 4.1 $\rm{^{\circ}C}$ at 60 min, but narrowed down to 1.2, 0.5, and 0.2 $\rm{^{\circ}C}$ after 2, 3, and 4 h. After the addition of 0.5 mL of 10 N sulfuric acid at 4 h, the temperature in each bottle was raised by ca. 0.8 °C.

and I₂ for the Shibala procedure are -0.07% and +1.73% per 100 μ M O₂ change, respectively. Meanwhile, for the Labasque procedure, the changes are −0.14% and +2.34% per 100 μ M O₂ change, respectively.

(iii) *Changes in the absorptivities of both iodine species due to temperature effects.* At 456 nm, the molar extinction coefficient increases by approximately +0.44% °C[−]¹ for I_3 ⁻, decreasing by −0.19% ^oC⁻¹ for I_2 . At 466 nm, the trends are comparable, with an increase of +0.53% $^{\circ}C^{-1}$ for I_3^- and a decrease of -0.18% °C⁻¹ for I_2 .

The combined result, as seen in absorbance, indicates an apparent temperature effect of +0.33%, +0.42%, and +0.51% C^{-1} at wavelengths of 430, 456, and 466 nm, respectively. [Figure](#page-9-0) 5 illustrates this effect by comparing absorbances measured at different room temperatures from those measured at 25 °C.

Based on the above findings, temperature control emerges as the most critical factor influencing the sensitivity, precision, and accuracy of the colorimetric method. While the temperature effect may be unavoidable, it can be minimized by ensuring a sufficiently high [I[−]] (>0.03 M) and allowing all samples, as well as standard solutions, to equilibrate with the

Figure 5. Overall temperature effect on the sensitivity at various wavelengths (430, 456, 466, and 476 nm) demonstrated as relative absorbance (w.r.t. $T_w = 25 \text{ °C}$). Slopes are 0.33, 0.42, 0.51, and 0.62% per 1 °C at the corresponding wavelengths, respectively.

laboratory environment for at least 4 h. By implementing these conditions, the impact of temperature fluctuations on the colorimetric method can be effectively mitigated, thus enhancing the reliability of the $[O_2]$ measurements.

Given that variations caused by iodine fractionation are no longer significant, it is reasonable to conclude that detection does not necessarily need to be at the isosbestic wavelength; a wide range from 430 to 476 nm can be selected for detection, providing flexibility in choosing the most suitable wavelength for different purposes.

Considering the naturally occurring $[O_2]$ range of 100–800 *μ*M in most environmental applications, the sensitivity of detecting at 430 nm would be too high $(A430/A456 = 2.05)$, as it could potentially exceed the linearity range in absorbance. Conversely, detecting at 466 nm might lose significant sensitivity $(A466/A456 = 0.71)$ without necessarily improving precision. Therefore, detection made at 456 nm, as suggested by the Shibala procedure, 4 offers a balanced compromise between sensitivity and linearity, as well as accounting for the influence of the temperature effects.

This work provides valuable analytical guidance for achieving very precise measurements of oxygen concentration using colorimetry. However, it is worth noting that the current application is still limited to oligotrophic waters, such as nonturbid freshwaters and open oceans. Corrections can be applied to turbid waters by adopting the procedures suggested by Roland et al.⁵ Further research will be continued to evaluate the potential of expanding the Shibala 4 method to correct for turbidity and a suite of other interferences including nitrite, hydrogen peroxide, and iodate.

■ **AUTHOR INFORMATION**

Corresponding Author

Brandon M. Stephens − *Institute of Oceanography, National Taiwan University, Taipei 106319, Taiwan;* [orcid.org/](https://orcid.org/0000-0001-7351-2750) [0000-0001-7351-2750](https://orcid.org/0000-0001-7351-2750); Email: bstephens@ntu.edu.tw

Authors

- Su-Cheng Pai − *Institute of Oceanography, National Taiwan University, Taipei 106319, Taiwan;* ● [orcid.org/0000-](https://orcid.org/0000-0003-1000-8834) [0003-1000-8834](https://orcid.org/0000-0003-1000-8834)
- Cheng-Ho Li − *Institute of Oceanography, National Taiwan University, Taipei 106319, Taiwan*

Complete contact information is available at: [https://pubs.acs.org/10.1021/acsomega.4c09084](https://pubs.acs.org/doi/10.1021/acsomega.4c09084?ref=pdf)

Funding

This work was supported by Taiwan's National Science and Technology Council (NSTC), awarded to S.-C.P. under project NSC 101-261-M-002-013 and to B.M.S. under project NSTC 113-2611-M-002-009.

Notes

The authors declare no competing financial interest.

■ **ACKNOWLEDGMENTS**

We would like to thank many past students who have contributed to this work. In particular, we would like to thank Kuan-Lun Huang, Gallon Cheng and Yu-Chen Yen. We would also like to thank Professors G.T.F. Wong and A.C.T. Chen for their invaluable suggestions during the early stages of this study. Part of the data presented in this manuscript were taken from the Master's Thesis of C.H. Li at National Taiwan University.

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