



Apelin – A New Kid on the Block in Periodontology

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Summary: Periodontitis is associated with numerous systemic diseases, and it has been shown that these associations are partly causal in nature. It is assumed that such interactions between periodontal and systemic diseases are also mediated via adipokines. Apelin, an adipokine about which there is little research in the dental field, is also produced together with its receptor in periodontal cells. The aim of this review was to summarize the currently available literature on the apelin-APJ system to better understand the pathomechanistic relationship between periodontitis and obesity and to determine the potential clinical relevance of apelin for diagnostics and therapy. In vitro studies suggest that apelin can enhance bacterial-induced synthesis of proinflammatory and proteolytic molecules, indicating a significant etiopathogenic role of this adipokine. Since serum levels of apelin are elevated in diabetes and/or obesity, it is possible that such systemic diseases promote the development and progression of periodontitis via apelin. On the other hand, it is also conceivable that apelin from the periodontium influences such systemic diseases. Further research is needed to better understand the role of apelin in the periodontium and the entire oral cavity, but also in the interactions between periodontal and systemic diseases. In particular, clinical intervention studies are needed to further decipher the etiopathogenic role of apelin in periodontitis.

Keywords: apelin, adipocytokine, adipokine, periodontitis, periodontium

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PERIODONTITIS AND ITS LINK TO SYSTEMIC DISEASES

Periodontitis is a chronic inflammatory disease of the periodontium characterized by irreversible bone and attachment loss.⁶² If this disease remains untreated, it can lead to tooth loss, reduced quality of life and negative effects on the entire

organism. Periodontitis is caused by microbial dysbiosis associated with an inadequate immune and inflammatory response.¹⁰ Microorganisms on the root surface can cause periodontitis directly, but more importantly indirectly by triggering an immunoinflammatory response in the periodontal tissues to reduce the microbial attack. If the inflammatory processes persist for too long and/or are too intense and/or misdirected,

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the disease becomes chronic and the periodontal soft and hard tissues are destroyed. There is clear evidence that a number of risk factors such as smoking, genetic and epigenetic predisposition, psychological stress and occlusal overload or misload can contribute to the development and progression of periodontitis.^{38,53,63} Various systemic diseases or conditions such as diabetes mellitus, obesity, and metabolic syndrome have also been shown to increase the risk.^{5,36,52,54,66} On the other hand, periodontitis may also promote the development and progression of these and other systemic diseases and conditions, meaning that there are not only significant associations between periodontitis and numerous systemic diseases, but also causal relationship at least in part, and this causality is even bidirectional. The aim of this review was to summarize the currently available literature on the apelin-APJ system to better understand the pathomechanistic relationship between periodontitis and obesity and to determine the potential clinical relevance of apelin for diagnostics and therapy.

ADIPOKINES

Several possible pathomechanisms have been described, and adipokines may play an important role in the link between periodontitis and metabolic diseases.^{18,36,88} A variety of immunoinflammatory but also structural cells of the periodontium, e.g. periodontal ligament (PDL) cells, osteoblasts, gingival fibroblasts and epithelial cells, as well as a large number of inflammatory mediators such as cytokines are involved in the etiopathogenesis of periodontitis. A special group of cytokines are adipokines (adipocytokines). It was originally assumed that these cytokines were only produced by adipocytes, hence named after these cells. However, it later became clear that such adipokines are also produced by other cells and tissues, where they exert physiological and pathophysiological significance.^{18,25} Thus, adipokines represent a pathomechanistic link between obesity and systemic diseases because adipose tissue is not only a pure energy store but also an active metabolic organ, whose secretion of adipokines is dysregulated when excessive.⁶¹ These adipokines can regulate glucose, fat, and bone metabolism as well as angiogenesis, play an important role in the development of cancer, influence the sense of hunger and/or thirst, but can also exhibit pro- or anti-inflammatory effects.^{15,23,43}

Presence and Role of Adipokines in Periodontium

Interestingly, the presence of such adipokines has also been demonstrated in the periodontium, suggesting that such adipokines may also play an important role in the development and progression of periodontitis.^{16–18,37,55,56,59,77} In recent years, the role of adipokines such as visfatin,^{17,55} resistin,^{2,56} leptin,^{59,69} and adiponectin^{58,89} in the periodontium has been increasingly studied. In PDL cells, for example, visfatin (nicotinamide phosphoribosyltransferase, pre-B-cell colony-promoting factor) has been shown to increase the expression of matrix metalloproteinase (MMP) 1 and chemokine C-C motif ligand (CCL) 2.⁵⁷ Leptin has been shown to enhance tumor necrosis factor (TNF)- α gene expression.⁶⁷ Resistin is another adipokine

with proinflammatory properties, as it induces the expression of proinflammatory cytokines in the periodontium.^{19,56} In contrast, adiponectin was able to counteract on the stimulated increase in the expression of proinflammatory mediators such as interleukin (IL)-1 β , IL-6, IL-8, MMP1 and -3 in PDL cells, suggesting an anti-inflammatory effect.^{41,58} Overall, there is fundamental evidence that resident cells of the periodontium and immunoinflammatory cells can produce adipokines and several of these adipokines exert proinflammatory and proteolytic effects on periodontal tissues, thereby contributing to periodontal inflammation and destruction seen in periodontitis.

Apelin and its Receptor

Apelin was first isolated and described by Tatemoto et al. in 1998,⁷³ while a G protein-coupled receptor was discovered in humans with a gene locus on chromosome 11 locus 11q12,⁶⁰ which turned out to be the endogenous apelin receptor (APJ), was already described in 1933. The gene Xq25-q26.3 encodes the protein biosynthesis for preproapelin, a polypeptide of 77 amino acids which is converted into the 55 residue proapelin by removing the N-terminal signal peptide.⁴⁴ At least four primary bioactive subforms can be formed from proapelin by single- or multi-stage proteolytic cleavages that are still not fully understood, with the C-terminus being retained: Apelin-36, -17, -13, and the N-terminal pyroglutamate-modified Pyr(1)-apelin-13.⁶⁴ The proprotein convertase subtilisin/kexin type 3 (PCSK3) / Furin has been shown to directly convert proapelin into apelin-13.⁷⁰ The receptor-binding C-terminus of these peptides is highly conserved across species in humans as well as in rats, mice, and cows. Rapid further proteolytic degradation determines duration and biological effectiveness of their action, with Pyr(1)-apelin-13, which is detected in human plasma, being the most bioactive subform.⁶⁴ C-terminal cleavage of apelin-13 and Pyr(1)-apelin-13 by angiotensin converting enzyme 2 (ACE2) and at a lower rate by prolyl carboxypeptidase (PRCP) results in an still biologically active apelin-12, while proteolysis by the metalloproteinase neprilysin (NEP) results in apelin-7 and -8, which are biologically inactive.

ROLE OF APELIN IN HEALTH AND DISEASE

Apelin was first isolated in tissues of the central nervous system, and the molecule was thought to play an essential role in central signal transduction.⁴⁴ Over time, the apelin-APJ complex was discovered in virtually all organs, notably the cerebellum, vascular endothelium, heart, lungs, kidneys, liver, adrenal glands and adipose tissue,^{32,39} indicating that apelin has a broad spectrum of physiological effects, including regulation of metabolism, inflammation and apoptosis.^{65,78} Studies have shown differential expression of apelin in different tissues: low expression in kidney, liver and pancreatic tissue, medium expression in skeletal muscle and high expression in chondrocytes, endothelial cells, skin, brain, spleen, thymus, lung, and also in adipose tissue.²² The fact that apelin is secreted by adipocytes and thus falls under the definition of an adipokine was first confirmed by a research group in 2005.¹³ With regard to periodontal diseases, the demonstrated involvement of the

apelin-APJ axis, particularly apelin-13, in the regulation of bone metabolism, the influence on apoptosis, proliferation and differentiation of osteoblasts in animal and human models, and the response of macrophages may be of potential relevance.^{26,72,81} In the heart, apelin is predominantly expressed constitutively in endocardial and vascular endothelial cells, while the APJ receptor is also expressed in vascular smooth muscle cells and cardiomyocytes.⁴⁰ In isolated rat hearts, apelin induced a potent dose-dependent, saturable positive inotropic effect,⁷¹ and at least apelin-36, apelin-13, and Pyr(1)-apelin-13 lower blood pressure through peripheral vasodilation via nitric oxide (NO) secretion.^{35,44} It has been observed that apelin-13 promotes angiogenesis in animal models following myocardial infarction through increased proliferation, migration, and induction of vascular endothelial growth factor (VEGF) in endothelial cells.⁸⁷ The apelin-APJ system further exerts anti-inflammatory and anti-fibrotic actions in part by counteracting the actions of the renin-angiotensin system and involved in the molecular actions of statins.¹⁴ Loss of apelin after myocardial infarction is related to a proinflammatory response (TNF- α , interleukin 1 β), increased macrophage infiltration and infarct size, adverse remodeling, aggravated systolic dysfunction, and heart failure.^{74,76,78} Deficiency or overabundance of apelin-APJ signaling, particularly by apelin-13, is not only involved in inflammatory cardiovascular dysfunctions but also inflammatory processes in other organs.⁷⁸ In lung tissue, apelin-13 stimulates alveolar formation and reduces inflammatory parameters.⁷⁵

Effects of the apelin-APJ system on energy metabolism extend to both glucose and lipid metabolism including the modulation of insulin secretion^{12,33} and therefore plays a key role in metabolic diseases such as type 2 diabetes mellitus (T2DM) and obesity (Fig 1).⁴⁶

In regard of glucose metabolism, decreased glycemia observed in wild type and insulin-resistant, obese mice injected with apelin-13 was mainly due to an increase in glucose utilization additive to insulin in white adipose tissue and skeletal muscles dependent on the energy sensor adenosine monophosphate dependent kinase (AMPK), Akt, and endothelial nitric oxide synthase (eNOS).²⁰ Insulin-stimulated glucose uptake was also increased in insulin-resistant 3T3-L1 adipocytes by Pyr(1)-apelin-13⁹⁰ and apelin-13 stimulated AMPK-dependent glucose uptake also in human adipose tissue *ex vivo*.⁷ Similar findings have been demonstrated in other tissues: In the intestinal tract²¹ and cardiac muscle cells,⁸² Pyr(1)-apelin-13 induced glucose uptake via GLUT2 and GLUT4, respectively. It has been shown that serum and plasma levels of apelin are increased in patients with T2DM^{28,34,50} and the adipokine has an impact on blood glucose regulation via positive modulation of glucose transport in different cell types^{86,90} and enhancement of insulin sensitivity.^{1,4} Based on these observations and the properties of apelin, it has been speculated to make a physiological compensation for insulin deficiency.

Only a few studies describe the effects of apelin on lipid metabolism. While there are conflicting results regarding the influence on lipolysis in mice and humans, apelin stimulates utilization of lipids.¹² Apelin-13 was shown to AMPK-dependent increase fatty acid oxidation, oxidative capacity, and mitochondrial biogenesis in skeletal muscle of high-fat diet induced

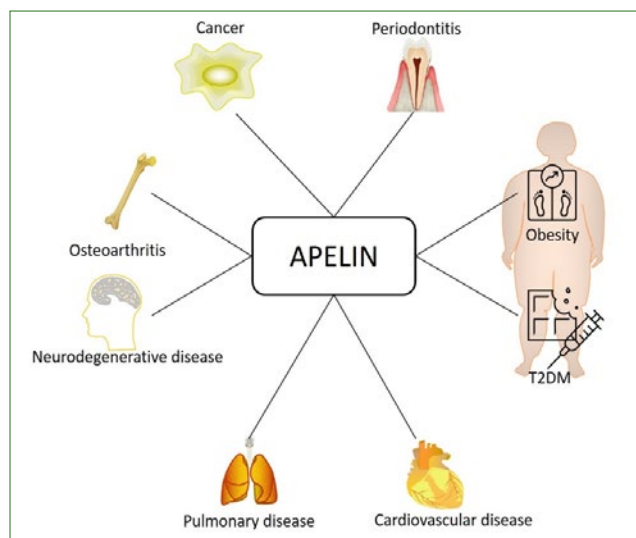


Fig 1 Association of apelin with different systemic diseases.

obese and insulin-resistant mice.⁸ In addition, apelin-over-expressing transgenic mice showed resistance to obesity related to increased lymphatic and blood vessel integrity preventing vascular permeability related fatty acid uptake in adipose tissue and increased blood vessel formation in skeletal muscle.⁸³ Accordingly, apelin knockout mice showed excessive weight gain. Elevated apelin concentrations in human serum have also been found in obesity,^{9,42} but normalized with dietary changes and calorie reduction.¹²

It therefore seems likely that the metabolic effects induced by elevated plasma apelin concentrations in metabolic diseases such as obesity and T2DM are, at least initially, part of a compensatory mechanism to maintain functions such as insulin sensitivity.

In addition to its physiological effects, apelin is also involved in pathological processes such as osteoarthritis and cancer.^{79,84} Data on the role of the apelin/APJ system in cancer vary widely depending on apelin subform and tissue type.^{6,80} However, the adipokine clearly has a modulatory influence on various cancers such as prostate,⁴⁸ gastric,²⁴ ovarian,³¹ lung,¹¹ and oral squamous cell carcinomas.²⁹ Pathomechanistic explanations could be apelin-induced increased cell migration in various tumor tissues,^{24,29,49} increased expression of pro-angiogenic factors in tumor cells and/or apelin-stimulated proliferation of endothelial cells in tumor tissues.³

PRESENCE AND POTENTIAL ROLE OF APELIN IN PERIODONTIUM

The three-way molecular pathological interplay in the chronic multimorbidity of periodontitis, diabetes and obesity, which is

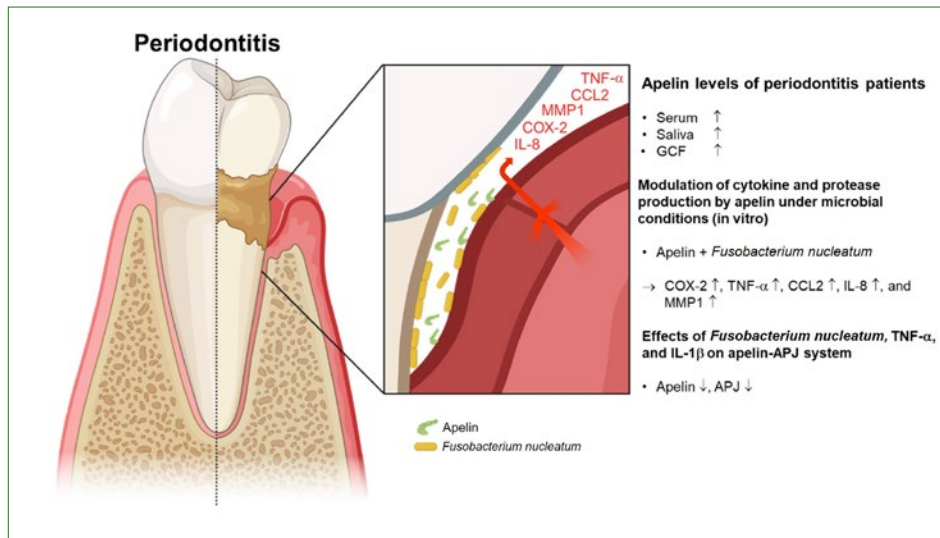


Fig 2 Summary of studies on the role of apelin in periodontal tissues and periodontitis.

maintained by common biological mechanisms such as inflammation and insulin resistance, is of great importance to dentists.⁵² The energy metabolism modulation by proinflammatory and antiinflammatory effects of adipokines such as leptin, adiponectin, resistin, visfatin, and vaspin are also part of this molecular pathological interplay.^{27,47,68} Based on the positive or negative correlations observed between molecular and clinical status, which are also shown in the positive correlation of the severity of diabetes and periodontitis, increased or decreased adipokine levels in body fluids could both be an indication of the severity of these metabolic diseases and chronic periodontitis and also mutually promote the development and progression of such multimorbid status. Poorly controlled diabetics have a threefold increased risk of periodontitis, and severe periodontitis can further worsen the derailed metabolic situation. Moreover, successful periodontal therapy reduces HbA1c levels in type 2 diabetes mellitus (T2DM) by an average of 0.4 %, demonstrating that the relationship between periodontitis and diabetes is causal in nature.⁶⁶ The increased serum/plasma concentrations of the adipokine apelin in diabetes and obesity⁴⁶ are therefore also of interest. Recently, Hirani et al³⁰ investigated the serum levels of apelin in periodontally and systemically healthy individuals and in periodontitis patients with and without T2DM. The study showed that apelin concentrations were significantly higher in the periodontitis group than in the healthy control group. Furthermore, apelin concentrations were highest in patients suffering from both periodontitis and T2DM suggesting that apelin plays a role in inflammation and glucose regulation. Sarhat et al. examined the apelin concentration in the saliva of periodontally diseased diabetics as well as periodontally and systemically healthy individuals.⁶⁸ They also found the highest apelin concentrations in patients with periodontitis and T2DM. The increased systemic apelin

concentrations in diabetes and obesity may be explained by its compensation for the reduced insulin sensitivity in T2DM. However, it is possible that resistance/tolerance to apelin develops over a longer period of time, as is also known for leptin in metabolic diseases.⁵¹ The consequence would then be an increased apelin level, as is the case for insulin. Such sustained elevation of apelin levels may negatively affect the periodontium. Our research group has recently demonstrated for the first time that apelin concentrations are not only altered systemically in serum and saliva, but also locally in gingival crevicular fluid (GCF) samples.⁸⁵ We compared the GCF apelin levels of individuals with clinically healthy periodontal tissues with those of patients suffering from gingivitis or periodontitis, all of whom were systemically healthy. We found that periodontitis patients had the highest GCF apelin levels, followed by gingivitis patients and healthy controls. This suggests that apelin may have a potential as a diagnostic biomarker in periodontology. Systemically elevated apelin concentrations have also been found in association with other systemic diseases.⁶ Future studies are warranted to clarify whether increased systemic apelin concentrations in these systemic diseases are also reflected in GCF, gingival tissue, and saliva samples.

To date, very little is known about the effects of apelin subforms on periodontal cells. We have recently addressed this issue in numerous in vitro experiments.¹⁶ As expected, the periodontopathogen *Fusobacterium nucleatum* (*F. nucleatum*) caused an upregulation of molecules associated with periodontal inflammation and destruction in our experiments. Interestingly, adding proteolytically unprocessed preproapelin increased *F. nucleatum*-induced expressions of CCL2 and MMP1 at 24 h and of COX2, CCL2, IL-8, TNF-α and MMP1 at 48 h. In addition, preproapelin was able to significantly increase the inhibition of RUNX2 induced by *F. nucleatum* at 24 h. Preproapelin

alone had no significant effect on the expression of the above molecules indicating *F. nucleatum* dependent processing of preproapelin. These in-vitro results suggest that certain proteolytically processed apelin subforms may contribute to periodontal inflammation and tissue destruction by periodontopathogens. Elevated concentrations of certain apelin subforms in systemic diseases could therefore represent a pathomechanistic link for the association between periodontitis and certain systemic diseases. Moreover, our in vitro experiments showed that apelin and its receptor are constitutively expressed in PDL cells and that their spontaneous expression is also regulated by *F. nucleatum*. In a study by Lee et al,⁴⁵ incubation of PDL cells and gingival fibroblasts with the inflammatory mediator TNF- α led to a downregulation of apelin. Our findings together with those of Lee et al⁴⁵ suggest that the local apelin-APJ system in the periodontium is downregulated, at least initially, during periodontal microbial dysbiosis and inflammation (Fig 2). As dysregulation of the apelin-APJ system tends to have proinflammatory effects, the initial downregulation of apelin and its receptor may represent a transient, protective host compensatory mechanism to limit inflammation and associated tissue destruction. Nevertheless, the experiments also showed that this potentially protective inhibition of apelin and its receptor was no longer present after a certain period of time, suggesting that the apelin-APJ system may play a crucial role in the pathogenesis of periodontitis during prolonged periodontal inflammation. However, the initial downregulation could also be the result of either dysregulation (deficiency/overabundance) of the apelin-APJ system or the distribution of apelin forms due to altered processing or maybe both.

Further studies are required to clarify whether periodontal therapy alters apelin levels and its proteolytic processing in GCF, saliva, gingiva and serum. In that case, apelin subforms could serve as biomarkers for diagnostic purposes, as has been suggested for T2DM. In the early stages, periodontal diseases usually do not show severe subjective complaints or clinical symptoms, which often delays the patient's visit to a specialist. However, an early diagnosis would be extremely important, as treatment at this point is efficient to avoid consequences of the disease. Probing pocket depth, clinical attachment loss, bleeding on probing, gingival index and plaque index are the most important clinical periodontal parameters and, together with radiographic assessment of alveolar bone loss, form the current basis of clinical periodontal diagnosis. Given the episodic nature of periodontal disease, these diagnostic tools are inadequate. Furthermore, clinical signs of tissue destruction provide information about the past, but only very limited information about the current status of periodontal disease activity. One of the challenges in periodontal diagnostics is to find a reliable clinical and/or biochemical biomarker that provides a valid indication of ongoing and/or future tissue destruction. Such a biomarker could enable dental care providers to offer individually tailored treatment plans and adjust the frequency of recalls during supportive periodontal therapy. Modulating the apelin-APJ system could lead to new immunomodulatory strategies in the treatment of periodontitis.

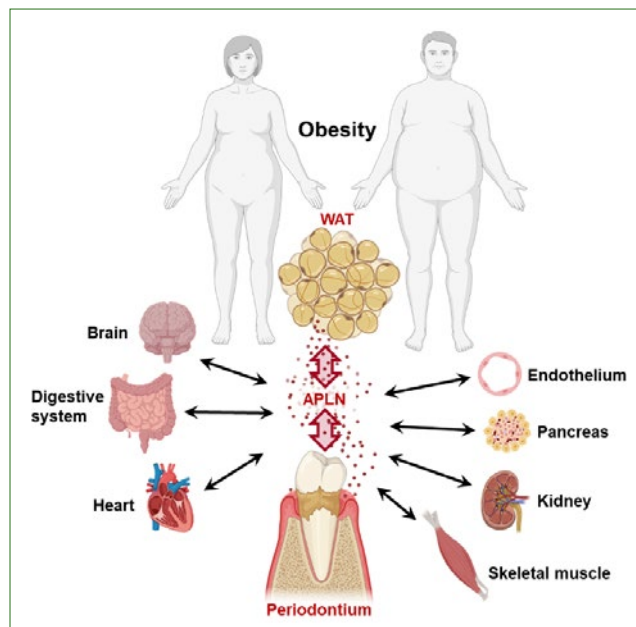


Fig 3 Schematic of apelin-based multidirectional organ interactions between periodontitis and obesity. WAT: white adipose tissue; APLN: apelin.

SUMMARY

Periodontitis is associated with numerous systemic diseases, and it has been shown that these associations are partly causal in nature. It is assumed that such interactions between periodontal and systemic diseases are also mediated via adipokines. Apelin is an adipokine that has been little studied in relation to periodontitis to date. This adipokine is produced together with its receptor in periodontal cells. In vitro studies suggest that apelin can enhance bacterial-induced synthesis of proinflammatory and proteolytic molecules, indicating a significant etiopathogenetic role of this adipokine. Since the serum levels of apelin are elevated in diabetes and/or obesity, it is possible that these systemic diseases promote the development and progression of periodontitis via apelin. On the other hand, it is also conceivable that apelin from the periodontium influences such systemic diseases (Fig 3). Further research is needed to better understand the role of apelin subforms in the periodontium and the entire oral cavity, but also in the interactions between periodontal and systemic diseases. In particular, clinical intervention studies are needed to further decipher the etiopathogenetic role of apelin subforms in periodontitis. Further basic research is also needed on the regulatory effects of apelin subforms on periodontal wound healing and soft and hard tissue metabolism. A better understanding of the apelin-APJ system could pave the way for new diagnostic and therapeutic strategies in the treatment of periodontitis.

REFERENCES

- Akcilar R, Turgut S, Caner V, Akcilar A, Ayada C, Elmas L, et al. The effects of apelin treatment on a rat model of type 2 diabetes. *Adv Med Sci* 2015;60(1):94–100.
- Akram Z, Rahim ZHA, Taiyeb-Ali TB, Shahdan MSA, Baharuddin NA, Vaithilingam RD et al. Resistin as potential biomarker for chronic periodontitis: A systematic review and meta-analysis. *Arch Oral Biol* 2017;73:311–320.
- Al-Abd AM, Alamoudi AJ, Abdel-Naim AB, Neamatallah TA, Ashour OM. Anti-angiogenic agents for the treatment of solid tumors: Potential pathways, therapy and current strategies - A review. *J Adv Res* 2017;8(6):591–605.
- Alipour FG, Ashoori MR, Pilehvar-Soltanahmadi Y, Zarghami N. An overview on biological functions and emerging therapeutic roles of apelin in diabetes mellitus. *Diabetes Metabol Syndr* 2017;11(suppl 2):S919-S923.
- Amar S, Leeman S. Periodontal innate immune mechanisms relevant to obesity. *Molec Oral Microbiol* 2013;28(5):331–341.
- Antushevich H, Wojcik M. Apelin in disease. *Clinica Chimica Acta Int J Clin Chem* 2018;483:241-248.
- Attané C, Daviaud D, Dray C, Dusaulcy R, Masseboeuf M, Prévot D et al. Apelin stimulates glucose uptake but not lipolysis in human adipose tissue ex vivo. *J Molec Endocrinol* 2011;46(1):21–28.
- Attané C, Foussal C, Le Gonidec S, Benani A, Daviaud D, Wanecq E et al. Apelin treatment increases complete Fatty Acid oxidation, mitochondrial oxidative capacity, and biogenesis in muscle of insulin-resistant mice. *Diabetes* 2012;61(2):310–320.
- Ba H-J, Chen H-S, Su Z, Du M-L, Chen Q-L, Li Y-H et al. Associations between serum apelin-12 levels and obesity-related markers in Chinese children. *PLoS One* 2014;9(1):e86577.
- Belibasakis GN, Belström D, Eick S, Gursoy UK, Johansson A, Könönen E. Periodontal microbiology and microbial etiology of periodontal diseases: Historical concepts and contemporary perspectives. *Periodontol* 2000 2023; doi: 10.1111/prd.12473 [Epub ahead of print].
- Berta J, Kenessey I, Dobos J, Tovari J, Klepetko W, Jan Ankersmit H et al. Apelin expression in human non-small cell lung cancer: role in angiogenesis and prognosis. *J Thorac Oncol* 2010;5(8):1120–1129.
- Bertrand C, Valet P, Castan-Laurell I. Apelin and energy metabolism. *Front Physiol* 2015;6:115.
- Boucher J, Masri B, Daviaud D, Gesta S, Guigné C, Mazzucotelli A et al. Apelin, a newly identified adipokine up-regulated by insulin and obesity. *Endocrinol* 2005;146(4):1764–1771.
- Chapman FA, Maguire JJ, Newby DE, Davenport AP, Dhaun N. Targeting the apelin system for the treatment of cardiovascular diseases. *Cardiovasc Res* 2023;119(17):2683–2696.
- Clemente-Suárez VJ, Redondo-Flórez L, Beltrán-Velasco AI, Martín-Rodríguez A, Martínez-Guardado I, Navarro-Jiménez E et al. The role of adipokines in health and disease. *Biomed* 2023;11(5):1290.
- Cores Ziskoven P, Nogueira AVB, Gutierrez LS, Weusmann J, Eick S, Buduneli N et al. Apelin enhances the effects of *Fusobacterium nucleatum* on periodontal ligament cells in vitro. *Int J Molec Sci* 2023;24(5):4733.
- Damanaki A, Nokhbehsaim M, Eick S, Götz W, Winter J, Wahl G et al. Regulation of NAMPT in human gingival fibroblasts and biopsies. *Mediators Inflamm* 2014;2014:912821.
- Deschner J, Eick S, Damanaki A, Nokhbehsaim M. The role of adipokines in periodontal infection and healing. *Molec Oral Microbiol* 2014;29(6):258–269.
- Devanoorkar A, Kathariya R, Guttiganur N, Gopalakrishnan D, Bagchi P. Resistin: a potential biomarker for periodontitis influenced diabetes mellitus and diabetes induced periodontitis. *Dis Mark* 2014;2014:930206.
- Dray C, Knauf C, Daviaud D, Waget A, Boucher J, Buléon M, et al. Apelin stimulates glucose utilization in normal and obese insulin-resistant mice. *Cell Metabol* 2008;8(5):437–445.
- Dray C, Sakar Y, Vinel C, Daviaud D, Masri B, Garrigues L, et al. The intestinal glucose-apelin cycle controls carbohydrate absorption in mice. *Gastroenterol* 2013;144(4):771–780.
- Falco M de, Luca L de, Onori N, Cavallotti I, Artigiano F, Esposito V, et al. Apelin expression in normal human tissues. *In Vivo (Athens, Greece)* 2002; 16(5):333–336.
- Fasshauer M, Blüher M. Adipokines in health and disease. *Trends Pharmacol Sci* 2015;36(7):461–470.
- Feng M, Yao G, Yu H, Qing Y, Wang K. Tumor apelin, not serum apelin, is associated with the clinical features and prognosis of gastric cancer. *BMC Cancer* 2016;16(1):794.
- Fève B, Bastard C, Fellahi S, Bastard J-P, Capeau J. New adipokines. *Annal Endocrinol* 2016;77(1):49–56.
- Gong G, Wan W, Liu X, Yin J. Apelin-13, a regulator of autophagy, apoptosis and inflammation in multifaceted bone protection. *International Immunopharmacology* 2023;117:109991.
- Guo Z, Peng Y, Hu Q, Liu N, Liu Q. The relationship between leptin and periodontitis: a literature review. *PeerJ* 2023;11:e16633.
- Habchi M, Duvallard L, Cottet V, Brindisi M-C, Bouillet B, Beacco M, et al. Circulating apelin is increased in patients with type 1 or type 2 diabetes and is associated with better glycaemic control. *Clin Endocrinol* 2014;81(5):696–701.
- Heo K, Kim YH, Sung HJ, Li HY, Yoo CW, Kim JY, et al. Hypoxia-induced up-regulation of apelin is associated with a poor prognosis in oral squamous cell carcinoma patients. *Oral Oncol* 2012;48(6):500–506.
- Hirani T, Kumar S, Patel V, Hirani S, Mohammed I, Shishoo D. Expression of apelin among the individuals of chronic periodontitis, with and without type ii diabetes mellitus: A study using enzyme-linked immunosorbent assay. *Adv Hum Biol* 2020;10(3):182.
- Hoffmann M, Fiedor E, Ptak A. Bisphenol A and its derivatives tetrabromobisphenol A and tetrachlorobisphenol A induce apelin expression and secretion in ovarian cancer cells through a peroxisome proliferator-activated receptor gamma-dependent mechanism. *Toxicol Lett* 2017;269:15–22.
- Hosoya M, Kawamata Y, Fukusumi S, Fujii R, Habata Y, Hinuma S et al. Molecular and functional characteristics of APJ. Tissue distribution of mRNA and interaction with the endogenous ligand apelin. *J Biol Chem* 2000;275(28):21061–21067.
- Hu G, Wang Z, Zhang R, Sun W, Chen X. The role of apelin/apelin receptor in energy metabolism and water homeostasis: a comprehensive narrative review. *Front Physiol* 2021;12:632886.
- Hu H, He L, Li L, Chen L. Apelin/APJ system as a therapeutic target in diabetes and its complications. *Molec Genet Metabol* 2016;119(1-2):20–27.
- Japp AG, Cruden NL, Amer DAB, Li VKY, Goudie EB, Johnston NR et al. Vascular effects of apelin in vivo in man. *J Amer Coll Cardiol* 2008;52(11):908–913.
- Jepsen S, Suvan J, Deschner J. The association of periodontal diseases with metabolic syndrome and obesity. *Periodontol* 2000 2020;83(1):125–153.
- Johnson RB, Serio FG. Leptin within healthy and diseased human gingiva. *J Periodontol* 2001;72(9):1254–1257.
- Kinane DF, Lappin DF, Culshaw S. The role of acquired host immunity in periodontal diseases. *Periodontol* 2000 2024; doi: 10.1111/prd.12562 (Epub ahead of print).
- Kleinz MJ, Davenport AP. Immunocytochemical localization of the endogenous vasoactive peptide apelin to human vascular and endocardial endothelial cells. *Regulat Peptid* 2004;118(3):119–125.
- Kleinz MJ, Skepper JN, Davenport AP. Immunocytochemical localisation of the apelin receptor, APJ, to human cardiomyocytes, vascular smooth muscle and endothelial cells. *Regulat Peptid* 2005;126(3):233–240.
- Kraus D, Winter J, Jepsen S, Jäger A, Meyer R, Deschner J. Interactions of adiponectin and lipopolysaccharide from *Porphyromonas gingivalis* on human oral epithelial cells. *PLoS One* 2012;7(2):e30716.
- Krist J, Wieder K, Klötting N, Oberbach A, Kralisch S, Wiesner T, et al. Effects of weight loss and exercise on apelin serum concentrations and adipose tissue expression in human obesity. *Obes Facts* 2013;6(1):57–69.
- Krysiak R, Handzlik-Orlik G, Okopien B. The role of adipokines in connective tissue diseases. *Eur J Nutr* 2012;51(5):513–528.
- Lee DK, Cheng R, Nguyen T, Fan T, Kariyawasam AP, Liu Y, et al. Characterization of apelin, the ligand for the APJ receptor. *J Neurochem* 2000;74(1):34–41.
- Lee GS, Song WH, Kim SJ, Kim YG, Ryu JH. Apelin-APJ axis inhibits TNF-alpha-mediated expression of genes involved in the inflammatory response in periodontal ligament cells. *Int J Oral Biol* 2019;44(4):182–190.
- Li C, Cheng H, Adhikari BK, Wang S, Yang N, Liu W, et al. The role of apelin-APJ system in diabetes and obesity. *Front Endocrinol* 2022;13:820002.
- Li S, Li H, Kong H, Wu SY, Cheng CK, Xu J. Endogenous and microbial biomarkers for periodontitis and type 2 diabetes mellitus. *Front Endocrinol* 2023;14:1292596.
- Lin T-H, Chang SL-Y, Khanh PM, Trang NTN, Liu S-C, Tsai H-C et al. Apelin promotes prostate cancer metastasis by downregulating TIMP2 via increases in miR-106a-5p expression. *Cells* 2022;11(20):3285.
- Lv D, Li L, Lu Q, Li Y, Xie F, Li H. et al. PAKI-cofilin phosphorylation mediates human lung adenocarcinoma cells migration induced by apelin-13. *Clin Experim Pharmacol Physiol* 2016;43(5):569–579.
- Ma W-Y, Yu T-Y, Wei J-N, Hung C-S, Lin M-S, Liao Y-J, et al. Plasma apelin: A novel biomarker for predicting diabetes. *Clinica Chimica Acta Int J Clin Chem* 2014;435:18–23.
- Mantzoros CS, Magkos F, Brinkoetter M, Sienkiewicz E, Dardeno TA, Kim S-Y, et al. Leptin in human physiology and pathophysiology. *Amer J Physiol Endocrinol Metabol* 2011;301(4):E567-84.
- Marruganti C, Suvan JE, D'Aiuto F. Periodontitis and metabolic diseases (diabetes and obesity): Tackling multimorbidity. *Periodontol* 2000 2023; doi: 10.1111/prd.12536 (Epub ahead of print).
- Miron RJ, Estrin NE, Sculean A, Zhang Y. Understanding exosomes: Part 3-therapeutic + diagnostic potential in dentistry. *Periodontol* 2000 2024;94(1):415–482.

54. Nibali L, Tatarakis N, Needleman I, Tu Y-K, D'Aiuto F, Rizzo M, et al. Clinical review: association between metabolic syndrome and periodontitis: a systematic review and meta-analysis. *J Clin Endocrinol Metab* 2013;98(3):913–920.
55. Nogueira AVB, Nokhbehshaim M, Eick S, Bourauel C, Jäger A, Jepsen S et al. Regulation of visfatin by microbial and biomechanical signals in PDL cells. *Clin Oral Invest* 2014;18(1):171–178.
56. Nogueira AVB, Nokhbehshaim M, Tekin S, Molon RS de, Spolidorio LC, Memmert S, et al. Resistin is increased in periodontal cells and tissues: in vitro and in vivo studies. *Mediators Inflamm* 2020.
57. Nokhbehshaim M, Eick S, Nogueira AVB, Hoffmann P, Herms S, Fröhlich H et al. Stimulation of MMP-1 and CCL2 by NAMPT in PDL cells. *Mediators Inflamm* 2013;2013:437123. doi: 10.1155/2013/437123.
58. Nokhbehshaim M, Keser S, Nogueira AVB, Cirelli JA, Jepsen S, Jäger A et al. Beneficial effects of adiponectin on periodontal ligament cells under normal and regenerative conditions. *J Diabetes Res* 2014;2014:796565. doi: 10.1155/2014/796565.
59. Nokhbehshaim M, Keser S, Nogueira AVB, Jäger A, Jepsen S, Cirelli JA, et al. Leptin effects on the regenerative capacity of human periodontal cells. *Int J Endocrinol* 2014;2014:180304. doi: 10.1155/2014/180304.
60. O'Dowd BF, Heiber M, Chan A, Heng HH, Tsui LC, Kennedy JL, et al. A human gene that shows identity with the gene encoding the angiotensin receptor is located on chromosome 11. *Gene* 1993;136(1-2):355–360.
61. Ouchi N, Parker JL, Lugus JJ, Walsh K. Adipokines in inflammation and metabolic disease. *Nat Rev Immunol* 2011;11(2):85–97.
62. Papapanou PN, Sanz M, Buduneli N, Dietrich T, Feres M, Fine DH et al. Periodontitis: Consensus report of workgroup 2 of the 2017 World Workshop on the Classification of Periodontal and Peri-Implant Diseases and Conditions. *J Periodontol* 2018;89(suppl 1):173–182.
63. Passanezi E, Sant'Ana ACP. Role of occlusion in periodontal disease. *Periodontology* 2000 2019;79(1):129–150.
64. Pisarenko OI, Studneva IM. Apelin C-terminal fragments: biological properties and therapeutic potential. *Biochem Biokhimiia* 2023;88(11):1874–1889.
65. Pitkin SL, Maguire JJ, Bonner TI, Davenport AP. International Union of Basic and Clinical Pharmacology. LXXIV. Apelin receptor nomenclature, distribution, pharmacology, and function. *Pharmacol Rev* 2010;62(3):331–342.
66. Preshaw PM, Alba AL, Herrera D, Jepsen S, Konstantinidis A, Makrilakis K, et al. Periodontitis and diabetes: a two-way relationship. *Diabetologia* 2012; 55(1):21–31.
67. Procaccini C, Jirillo E, Matarese G. Leptin as an immunomodulator. *Molec Asp Med* 2012;33(1):35–45.
68. Sarhat ER, Rmaid ZJ, Jabir TH. Changes of salivary interleukine-17, apelin, omentin and vaspin levels in normal subjects and diabetic patients with chronic periodontitis. *ATMPH* 2020;23(01):135–141.
69. Shimada Y, Komatsu Y, Ikezawa-Suzuki I, Tai H, Sugita N, Yoshie H. The effect of periodontal treatment on serum leptin, interleukin-6, and C-reactive protein. *J Periodontol* 2010;81(8):1118–1123.
70. Shin K, Pandey A, Liu X-Q, Anini Y, Rainey JK. Preferential apelin-13 production by the proprotein convertase PCSK3 is implicated in obesity. *FEBS open bio* 2013;3:328–333.
71. Szokodi I, Tavi P, Földes G, Voutilainen-Myllylä S, Ilves M, Tokola H, et al. Apelin, the novel endogenous ligand of the orphan receptor APJ, regulates cardiac contractility. *Circul Res* 2002;91(5):434–440.
72. Tang S, Xie H, Yuan L, Luo X, Huang J, Cui R, et al. Apelin stimulates proliferation and suppresses apoptosis of mouse osteoblastic cell line MC3T3-E1 via JNK and PI3-K/Akt signaling pathways. *Peptides* 2007;28(3):708–718.
73. Tatemoto K, Hosoya M, Habata Y, Fujii R, Kakegawa T, Zou MX, et al. Isolation and characterization of a novel endogenous peptide ligand for the human APJ receptor. *Biochem Res Commun* 1998;251(2):471–476.
74. Tatin F, Renaud-Gabardos E, Godet A-C, Hantelys F, Pujol F, Morfisse F et al. Apelin modulates pathological remodeling of lymphatic endothelium after myocardial infarction. *JCI Insight* 2017;2(12).
75. Visser YP de, Walther FJ, Laghmani EH, van der Laarse A, Wagenaar GTM. Apelin attenuates hyperoxic lung and heart injury in neonatal rats. *Amer J Respir Critic Care Med* 2010;182(10):1239–1250.
76. Wang W, McKinnie SMK, Patel VB, Haddad G, Wang Z, Zhabeyev P, et al. Loss of apelin exacerbates myocardial infarction adverse remodeling and ischemia-reperfusion injury: therapeutic potential of synthetic Apelin analogues. *J Amer Heart Assoc* 2013;2(4):e000249.
77. Wang X, Tang Y, Xiao R. Chemerin contributes to inflammatory responses and suppresses osteogenic differentiation in chronic periodontitis. *Oral Dis* 2023; 29(4):1706–1714.
78. Wang X, Zhang L, Li P, Zheng Y, Yang Y, Ji S. Apelin/APJ system in inflammation. *Int Immunopharmacol* 2022;109:108822.
79. Wang Y-H, Kuo S-J, Liu S-C, Wang S-W, Tsai C-H, Fong Y-C et al. Apelin affects the progression of osteoarthritis by regulating VEGF-dependent angiogenesis and miR-150-5p expression in human synovial fibroblasts. *Cells* 2020;9(3):594.
80. Wysocka MB, Pietraszek-Gremplewicz K, Nowak D. The role of apelin in cardiovascular diseases, obesity and cancer. *Front Physiol* 2018;9:557.
81. Xie H, Tang S, Cui R, Huang J, Ren X, Yuan L, et al. Apelin and its receptor are expressed in human osteoblasts. *Regulat Peptides* 2006;134(2-3):118–125.
82. Xu S, Han P, Huang M, Wu JC, Chang C, Tsao PS, et al. In vivo, ex vivo, and in vitro studies on apelin's effect on myocardial glucose uptake. *Peptides* 2012;37(2):320–326.
83. Yamamoto T, Habata Y, Matsumoto Y, Yasuhara Y, Hashimoto T, Hamajyo H, et al. Apelin-transgenic mice exhibit a resistance against diet-induced obesity by increasing vascular mass and mitochondrial biogenesis in skeletal muscle. *Biochim Biophys Acta* 2011;1810(9):853–862.
84. Yang Y, Lv S-Y, Ye W, Zhang L. Apelin/APJ system and cancer. *Clinica Chimica Acta Int J Clin Chem* 2016;457:112–116.
85. Yoldaş O, Nogueira AVB, Kantar PM, Ziskoven PC, Deschner J, Buduneli N. Gingival crevicular fluid levels of apelin correlates with clinical periodontal diagnosis. *Clin Oral Invest* 2023;28(1):50.
86. Yue P, Jin H, Aillaud M, Deng AC, Azuma J, Asagami T, et al. Apelin is necessary for the maintenance of insulin sensitivity. *American journal of physiology. Endocrinol Metabol* 2010;298(1):E59–67.
87. Zhang B-H, Guo C-X, Wang H-X, Lu L-Q, Wang Y-J, Zhang L-K, et al. Cardioprotective effects of adipokine apelin on myocardial infarction. *Heart Vessels* 2014;29(5):679–689.
88. Zhang L, Meng S, Tu Q, Yu L, Tang Y, Dard MM, et al. Adiponectin ameliorates experimental periodontitis in diet-induced obesity mice. *PLoS One* 2014; 9(5):e97824.
89. Zhu J, Guo B, Gan X, Zhang L, He Y, Liu B, et al. Association of circulating leptin and adiponectin with periodontitis: a systematic review and meta-analysis. *BMC Oral Health* 2017;17(1):104.
90. Zhu S, Sun F, Li W, Cao Y, Wang C, Wang Y, et al. Apelin stimulates glucose uptake through the PI3K/Akt pathway and improves insulin resistance in 3T3-L1 adipocytes. *Molec Cell Biochem* 2011;353(1-2):305–313.

