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# **Life Course Considerations in Environmental Health: Developmental Neurotoxicity of Domoic Acid at Doses Below Acute Effect Levels in Adult Humans**

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#### **ABSTRACT**

**Background:** Current US federal action levels for domoic acid (DA) in seafood are based on acute toxicity observed in exposed adult humans. Life course considerations have not been incorporated. The potential for developmental neurotoxicity (DNT) at permissible DA levels has previously been noted, but not methodically assessed.

**Methods:** Studies of DNT following DA exposure in experimental and wild animals were identified through a comprehensive search strategy. Evidence from papers meeting inclusion criteria was evaluated for specific outcomes reported for doses at which adverse effects were observed. Exposure levels associated with DNT were compared with those known to cause adult toxicity. The findings are discussed in the context of the well-characterized mechanism of DA neurotoxicity, as well as the toxicokinetics of DA across species and life stages.

**Conclusions:** DNT outcomes were reported with a no observed adverse effect level (NOAEL) 10 times lower than the NOAEL of 0.75 mg DA/kg for acute effects in adults. Apart from reviewing current regulatory action levels, public health outreach messaging to health care professionals and sensitive populations, such as pregnant or breastfeeding women, should be considered as a means of increasing awareness about risk for DNT from consumption of potentially DA-contaminated seafood.

#### **1 | Introduction**

Many researchers have expressed concerns as to whether current safety considerations for the presence of domoic acid (DA) in seafood are sufficient to protect individual consumers throughout the course of their lives (Costa, Giordano, and Faustman [2010;](#page-14-0) Doucette and Tasker [2016](#page-15-0); Grattan [2022;](#page-15-1) Panlilio et al. [2023;](#page-16-0) Petroff et al. [2021](#page-16-1); Shum et al. [2020\)](#page-16-2). The sensitivity of age-based sub-populations can vary due to the developmental stage of target tissues (e.g., brain), as well as with life-stage changes in toxicokinetic (TK) factors such as rates of absorption and elimination (Lanphear [2015](#page-15-2)).

Compared to the adult brain, developing brains are generally more sensitive to the effects of neurotoxicants (Costa, Guizzetti, and Vitalone [2004;](#page-14-1) Lanphear [2015;](#page-15-2) Rodier [1995](#page-16-3)). Prenatal and early postnatal brain development are characterized by rapid neural cell proliferation and migration, maturation of receptor and transmitter systems, increasing numbers of synaptic connections, and production of myelin (Rodier [1995](#page-16-3)). Interference with any of these essential developmental processes can adversely impact eventual mature brain structure and/or function (Costa, Guizzetti, and Vitalone [2004\)](#page-14-1). Simultaneously, an immature blood–brain barrier may not provide the developing brain with the same degree of protection from circulating

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toxins as afforded to an adult brain by a mature barrier (Costa, Guizzetti, and Vitalone [2004](#page-14-1); Rodier [1995\)](#page-16-3).

The neurotoxic effects of DA are consistent among adults of the species evaluated including humans, laboratory animals, and affected wildlife (Goldstein et al. [2008](#page-15-3); Grant et al. [2010](#page-15-4); Iverson and Truelove [1994\)](#page-15-5). Experimental evidence indicates that developing brains are more sensitive than adult brains to DA-induced neurotoxicity (Costa, Giordano, and Faustman [2010](#page-14-0); Petroff et al. [2021\)](#page-16-1).

For this commentary, we have compiled the empirical evidence from studies of DA-induced developmental neurotoxicity (DNT) in experimental and wild animals. Specific impacts observed at the lowest DA doses with an adverse effect were compared across species and developmental stages. While the exact timing of key neurodevelopmental events with respect to the pre- versus post-natal periods may vary among species (Clancy et al. [2008\)](#page-14-2), the sequence of events appears to be highly conserved (Clancy, Darlington, and Finlay [2001\)](#page-14-3). The results are discussed in the context of available background information on mechanisms of DA neurotoxicity and the TK of DA at various life stages. The detailed information presented in this commentary will be valuable for communicating risks of DA exposure throughout life.

# **1.1 | Sources of DA Exposure**

DA is a potent neurotoxin produced by algal diatoms. On the west coast of the US (OEHHA [2022a;](#page-16-4) Petroff et al. [2021\)](#page-16-1) and in many other jurisdictions worldwide (Bates et al. [2018\)](#page-14-4), diatoms of the genus *Pseudo-nitzschia* are the most common source. Shellfish and fish can accumulate DA by feeding in waters contaminated with DA-containing cells of *Pseudo-nitzschia* (Bernstein et al. [2021;](#page-14-5) Lefebvre, Frame, and Kendrick [2012](#page-15-6); Lefebvre et al. [2007\)](#page-15-7). Accumulated DA can then be passed on to higher trophic levels (OEHHA [2022a\)](#page-16-4) including seabirds (Gibble et al. [2021](#page-15-8)), marine mammals (Bowen et al. [2022;](#page-14-6) Goldstein et al. [2008](#page-15-3); Kreuder [2005;](#page-15-9) Moriarty et al. [2021](#page-16-5)), and humans (Fritz et al. [1992\)](#page-15-10).

# **1.2 | Acute Effects of DA (1987 Amnesic Shellfish Poisoning Outbreak)**

The acute effects of DA poisoning in adults came to wide attention following a 1987 outbreak involving over 100 people who consumed cultured mussels sourced from eastern Prince Edward Island, Canada (Perl, Bedard, Kosatsky, Hockin, Todd, Remis, et al. [1990\)](#page-16-6). Symptoms ranged from gastrointestinal (GI) distress, to disorientation, seizure, coma, and death (Perl, Bedard, Kosatsky, Hockin, Todd, Remis, et al. [1990\)](#page-16-6). DA poisoning is also referred to as "Amnesic Shellfish Poisoning" (ASP) because temporary or permanent loss of short-term memory is a characteristic symptom at higher exposure levels (Grant et al. [2010](#page-15-4); Perl, Teitelbaum, et al. [1990;](#page-16-7) Teitelbaum et al. [1990](#page-17-0)).

Ingested doses of DA were estimated for 10 individuals from the 1987 outbreak based on DA concentrations in meal remnants and estimated portions consumed (Perl, Bedard, Kosatsky, Hockin, Todd, Remis, et al. [1990](#page-16-6)). A single individual estimated to have consumed 20mg of DA had no adverse GI or neurological effects (EFSA [2009\)](#page-15-11). With doses normalized to 60kg body weight (bw), the lowest observed adverse effect level (LOAEL) was determined to be 1.0mg DA/kg bw, which was associated with mild gastrointestinal effects (Iverson and Truelove [1994\)](#page-15-5). Severe illness requiring hospitalization was associated with an estimated dose of 4mg DA/kg bw (EFSA [2009](#page-15-11); Perl, Bedard, Kosatsky, Hockin, Todd, Remis, et al. [1990\)](#page-16-6).

# **1.3 | Basic Mechanism of Action (Kainic Acid Analog)**

DA is a high-affinity structural analog of kainic acid (Figure [1\)](#page-2-0), and hence a selective agonist of kainate receptors (a subtype of ionotropic glutamate receptors) (Larm, Beart, and Cheung [1997\)](#page-15-12). At sufficient concentrations, agonists of glutamate receptors can over-stimulate neurons to death in a process known as excitotoxicity. Excitotoxicity contributes to impairment of learning and memory as well as to frank neurodegeneration in the hippocampus and neocortex (AOP-48 [2023;](#page-14-7) Larm, Beart, and Cheung [1997\)](#page-15-12). DA has a higher affinity for kainate receptors than kainic acid itself, resulting in 3–20 fold greater potency for downstream effects (Larm, Beart, and Cheung [1997;](#page-15-12) Costa, Giordano, and Faustman [2010](#page-14-0)).

The Adverse Outcome Pathway (AOP) for ionotropic glutamate receptors in adult brain (Figure [2\)](#page-2-1) illustrates the sequence of key events from receptor binding through apical expression as deficits in learning and memory (AOP-48 [2023](#page-14-7)). The AOP is intended to apply to all types of glutamate ionotropic receptors, including kainate receptors, involved in basal excitatory synaptic transmission and synaptic plasticity, which under physiological conditions are critical for normal learning and memory. DA's action as an agonist for glutamate receptors is sufficiently well-understood and specific to support its use as a positive control for certain in vitro assays used in large-scale screening of compounds for potential DNT (USEPA [2020](#page-17-1)). While not specifically validated for developing brains, current evidence tends to support the pathway's applicability (AOP-48 [2023\)](#page-14-7).

Data from humans, mice, and rats used to construct the AOP were characterized as strong evidence (AOP-48 [2023](#page-14-7)). Binding of DA to ionotropic glutamate receptors and the resulting adverse neurological effects have been documented broadly in additional vertebrate species including non-human primates, marine mammals, birds, and finfish (Lefebvre, Frame, and Kendrick [2012](#page-15-6); Lefebvre et al. [2007;](#page-15-7) Lefebvre [2001\)](#page-15-13). While fish treated with DA by injection are susceptible to DA-caused excitotoxicity, they may be less sensitive to oral doses of DA when compared to other species (Anderson et al. [2021;](#page-14-8) Lefebvre, Frame, and Kendrick [2012;](#page-15-6) Lefebvre et al. [2007](#page-15-7); Lefebvre [2001\)](#page-15-13). Fish die-offs have not been observed to result from *Pseudo-nitzschia* blooms, even when DA levels in fish digestive tract tissues were high (e.g., >50mg DA/kg viscera) (Bernstein et al. [2021;](#page-14-5) Lefebvre, Frame, and Kendrick [2012](#page-15-6); Lefebvre et al. [2007\)](#page-15-7).

Ionotropic glutamate receptors, including kainate receptors, have also been identified in tissues of the GI tract (Baj et al. [2019](#page-14-9)). These receptors are part of the "microbiota–gut– brain axis," and modulation of their activity may influence brain as well as gut function. While vomiting and other GI effects have



**Kainic Acid** 

**FIGURE 1** | Chemical structures of domoic acid, and analogues glutamic acid and kainic acid.

<span id="page-2-0"></span>

<span id="page-2-1"></span>**FIGURE 2** | Schematic diagram of the adverse outcome pathway (AOP) for domoic acid and similar compounds. Modified from the AOP "Binding of agonists to ionotropic glutamate receptors in adult brain leading to excitotoxicity that mediates neuronal cell death, contributing to learning and memory impairment" (AOP 48, 2023) available online at:<https://aopwiki.org/aops/48>.

been reported from DA exposure in humans and non-human primates, the specific involvement of ionotropic glutamate receptors in these symptoms has not been definitively established.

#### **1.4 | Basic Toxicokinetics (TK) of DA**

DA is poorly absorbed from the GI tract, and is rapidly eliminated (e.g., half-life <110min reported for non-human primates) with approximately 75% metabolically unchanged in urine and feces (Suzuki and Hierlihy [1993](#page-17-2); Truelove and Iverson [1994;](#page-17-3) Truelove et al. [1997](#page-17-4); Jing et al. [2018](#page-15-14); Costa, Giordano, and Faustman [2010\)](#page-14-0). Distribution and elimination of DA has been studied in pregnant and non-pregnant animals (Jing et al. [2018;](#page-15-14) Maucher Fuquay et al. [2012a;](#page-16-8) Shum et al. [2020\)](#page-16-2), as well as in fetuses and neonates (Maucher Fuquay et al. [2012b;](#page-16-9) Maucher and Ramsdell [2005\)](#page-16-10).

The results from TK studies of acutely dosed DA differ between the injection and oral routes of exposure. In adult female cynomolgus monkeys (*Macaca fascicularis*), a single intravenous (IV) dose of  $5\mu$ g DA/kg bw (0.005 mg/kg) had a mean elimination half-life of 1.2h (Jing et al. [2018](#page-15-14)). Acute oral doses of 0.075 and 0.15mg DA/kg bw produced a mean terminal half-life of 11.3h with the difference between routes indicating "flip-flop kinetics," or a rate of oral absorption that is slower than the rate of elimination and hence rate-limiting (Jing et al. [2018\)](#page-15-14).

The TK of chronic oral dosing with DA at 0.075 and 0.15mg/ kg bw was studied in adult female cynomolgus monkeys before, during, and after pregnancy (Shum et al. [2020](#page-16-2)). At the time of delivery, DA was detected in infant plasma and amniotic fluid. TK modeling of maternal-fetal DA disposition suggested that placental transport introduces DA to the fetus via cord blood. The fetus subsequently eliminates DA into the amniotic fluid,

which is then recirculated by fetal swallowing (Figure [3\)](#page-3-0). DA levels were 4.5–7.5X higher in amniotic fluid than in fetal plasma, suggesting that DA accumulates in amniotic fluid.

Distribution of DA in maternal and fetal tissues following gestational dosing is broadly consistent in experimental rats and monkeys (Maucher Fuquay et al. [2012a,](#page-16-8) [2012b;](#page-16-9) Shum et al. [2020\)](#page-16-2), as well as in samples collected from fetal tissues and fluids of stranded sea lions (Brodie et al. [2006;](#page-14-10) Goldstein et al. [2009;](#page-15-15) Lefebvre et al. [2018\)](#page-15-16). Following treatment of pregnant rats on gestation day (GD) 20 with 1.0mg DA/kg bw IV, DA levels in maternal brain and cerebrospinal fluid (CSF) peaked at 15min following treatment and were undetectable by 12–24h (Maucher Fuquay et al. [2012a\)](#page-16-8). Fetal plasma DA levels peaked at 60min post-dosing (Maucher Fuquay et al. [2012b\)](#page-16-9). Post-treatment DA levels in amniotic fluid and fetal brain did not show evidence of elimination over 24h. Retention of DA in the amniotic fluid of these experimental animals (Maucher Fuquay et al. [2012b\)](#page-16-9) reinforces the suggestion of continuous fetal re-exposure, in turn increasing the relative risk for fetal harm from a given maternal dose of DA.

Experiments in lactating rats demonstrated the presence of DA in the milk of treated dams and in the plasma of nursing pups given supplemental milk with added DA (Maucher and Ramsdell [2005](#page-16-10)). While at one-hour post-dosing with 1.0mg/ kg bw DA by intraperitoneal injection (IP) DA levels were 16 times lower in milk than in maternal plasma; by 8h post-dosing, remaining DA levels were four times higher in milk than in maternal plasma. Although neonates given milk from these DAexposed females that did not attain measurable plasma levels of DA, neonates did absorb sufficient DA from consuming milk "spiked" with 1.0mg DA/kg bw to result in measurable plasma DA levels.

# **1.5 | Current Basis for US Regulation of DA in Seafood**

Health Canada was the first entity to establish and implement an official regulatory limit of  $20 \mu$ g DA/g (20 ppm) for DA in edible tissue of bivalve shellfish in 1989. Prior to that, Canadian health and regulatory entities had instituted an interim safety threshold in 1988 that restricted shellfish harvest when DA levels exceeded 20 μg DA/g (Todd [1990;](#page-17-5) Wekell, Hurst, and Lefebvre [2004\)](#page-17-6). Health Canada organized a symposium on DA toxicity in Ottawa (April 1989), where scientists, health officials, and regulators met to discuss various facets of the 1987 DA poisoning event, define the acute DA toxicity clinical syndrome now known as ASP, and establish the scientific basis for a safe consumption threshold for DA in shellfish of 20 ppm. Health Canada applied a LOAEL approach to calculate the threshold where 50 mg DA was divided by an approximate 0.200 kg (wet weight) shellfish consumption rate, or 250 ppm, to which a safety factor of 12 was applied (Perl, Bedard, Kosatsky, Hockin, Todd, Remis, et al. [1990;](#page-16-11) Perl, Bedard, Kosatsky, Hockin, Todd, McNutt, et al. [1990;](#page-16-11) Perl, Teitelbaum, et al. [1990](#page-16-7); Todd [1990](#page-17-5); Wekell, Hurst, and Lefebvre [2004](#page-17-6)).

US Food and Drug Administration (FDA) critically reviewed the Health Canada regulatory limit at the FDA Pacific Region 1992 Domoic Acid Workshop in San Pedro, California, which was modeled after the 1989 Canadian symposium (DHHS [1993;](#page-15-17) USFDA [2021](#page-17-7); Wekell, Hurst, and Lefebvre [2004\)](#page-17-6). US FDA adopted the LOAEL approach using 60mg DA as a LOAEL, divided by 0.250kg (wet weight) shellfish consumption rate, or 240ppm, and applied an uncertainty factor (UF) of 10. Twenty ppm was subsequently adopted and implemented as the FDA action level for DA in seafood (DHHS [1993\)](#page-15-17). US FDA later



<span id="page-3-0"></span>**FIGURE 3** | Schematic diagram of maternal-fetal distribution of domoic acid (DA). DA in the maternal circulation is transported across the placenta and into the fetus via the umbilical vein. The fetal circulatory system transports DA to the immature fetal organs, including brain. The fetus eliminates DA into the amniotic fluid, which is swallowed by the fetus, thus recirculating available DA.

evaluated additional DA data in Dungeness Crab and adopted a second action level in 1993 permitting 30 ppm DA in Dungeness Crab viscera (DHHS [1993;](#page-15-17) USFDA [2021](#page-17-7)).

Subsequent to the adoption of federal action limits, the Washington State Department of Health (DOH) conducted a literature review to determine a tolerable daily intake (TDI) for DA (Marien [1996\)](#page-16-12). Overall, Washington DOH determined that the human and nonhuman primate toxicity data supported a no observed adverse effect level (NOAEL) of 0.75mg DA/ kg bw for acute exposure of adult individuals (Marien [1996](#page-16-12)). Due to the similarities between humans and test monkeys in response to doses of DA (Iverson and Truelove [1994\)](#page-15-5), no between-species uncertainty factor (UF) was applied. Washington State health officials established a TDI of 0.075mg DA/kg bw that incorporated a 10-fold UF to account for variation in sensitivity within species (Marien [1996](#page-16-12)). Washington DOH concluded that the TDI was consistent with the US FDA action levels of 30 and 20ppm for crab viscera and clams, respectively, and that the federal limits were sufficiently protective of public health from acute DA toxicity.

# **1.6 | California State Agency Activities Regarding DA**

The Office of Environmental and Health Hazard Assessment (OEHHA) is the lead California agency for evaluating health hazards posed by environmental contaminants and partners with other State agencies to ensure the safety of California fisheries from toxic substances, including DA (OEHHA [2019a\)](#page-16-13). As *Pseudo-nitzschia* blooms in California coastal waters are an ongoing concern, OEHHA assesses monitoring data for DA levels in California seafood samples analyzed by the California Department of Public Health (CDPH) laboratories. In consultation with CDPH, OEHHA makes formal health-based recommendations to the California Department of Fish and Wildlife (CDFW) regarding delay of opening, closure, or re-opening actions for California recreational and commercial fisheries under the authority of California Fish and Game Code section 5523 (CDFW) [\(2017\)](#page-14-11).

OEHHA, CDPH, and CDFW coordinate DA-related regulatory actions and provide the public with information in the form of responses to frequently asked questions (CDPH [2017](#page-14-12); OEHHA [2018\)](#page-16-14). These three entities, in cooperation with the California State Water Resources Control Board, comprise the Interagency harmful algal bloom (HAB)-related Illness Workgroup which investigates and tracks potential HAB-related illnesses in humans and animals throughout California, including DA-induced ASP, which is a reportable illness in California (OEHHA [2022b,](#page-16-15) [2024](#page-16-16)). OEHHA has hosted two informational workshops on DA toxicity (2017 and 2019), which featured invited speakers from the research community (OEHHA [2017,](#page-16-17) [2019b](#page-16-18)).

In 2020, DA was one of 22 chemicals presented to the OEHHA Developmental and Reproductive Toxicant Identification Committee (DARTIC) for prioritization for consideration under California's Proposition 65 (OEHHA [2020\)](#page-16-19). Chemicals included in that document had passed a data screen followed by a preliminary toxicological evaluation of available evidence for reproductive harm.

# **1.7 | Empirical Data: Implications for DA-Related DNT Risk**

The extensive background information on DA summarized above affirms that life stage considerations are relevant to risk for DA's adverse neurological effects. Developing vertebrates are at particular risk of DA-induced DNT due to the immaturity of developing brain tissues. Developing mammals face the additional complexities of placental transfer, recirculation between the fetus and amniotic fluid with slow elimination from the amniotic fluid, and potential postnatal exposure via milk from exposed maternal animals.

An earlier review reported on experimental evidence for DNT occurring at DA doses one to two orders of magnitude below levels causing neurotoxicity in adults (Costa, Giordano, and Faustman [2010](#page-14-0)). This present effort compiles and summarizes the currently available experimental data on the developmental toxicity of DA in developing nonhuman animals. Taking a comprehensive look at dose–response data for DNT associated with DA exposure early in life for several species will facilitate considerations of whether the current action levels provide protection during the most sensitive life stages.

# **2 | Methods**

# **2.1 | Literature Identification and Selection**

## **2.1.1 | Search Strategies**

The literature search strategy for prioritization candidates is detailed in a 2020 report by OEHHA (OEHHA [2020\)](#page-16-19). The studies retrieved and reviewed for that document provided an initial nucleus for the DNT data presented in Tables.

The literature search for DA has been updated, and relevant new publications retrieved for inclusion. Additional efforts to identify relevant literature have included cross-checking reference lists from published papers, and author-based searches for papers published by researchers known to have worked on the DNT of DA.

## **2.1.2** | **Inclusion Criteria for Laboratory and Wild Animal Studies**

Laboratory studies included in the data tables met the following criteria by reporting:

- Original data.
- At least one dose level of DA plus appropriate controls.
- Single or repeated DA dosing restricted to the pre- and/or postnatal periods of brain development, with specific timing reported to differentiate between DNT and adult neurotoxicity resulting from acute or chronic DA exposures of mature subjects.
- Assessment and reporting of DNT outcomes such as behavioral, histopathological, and/or biochemical effects.

• DNT assessment must have occurred subsequent to DA dosing; while timing of dosing is restrictive to the period of neurodevelopment, assessment of DNT outcomes could have been conducted at any time following dosing, from immediately post-dosing until later in life.

For wild marine mammals, only rough estimates of dosing and timing of DA exposure could be made. Tabulated studies reported data on young animals believed to have been exposed during the prenatal and/or neonatal (nursing) periods. All of the selected literature reported on California sea lions found stranded, with DA exposure inferred from the timing and extent of known toxigenic *Pseudo-nitzschia* blooms capable of, or actively producing, DA (Smith et al. [2023](#page-16-20)). Some studies measured DA concentrations in affected animals' tissues and body fluids.

## **3 | Results**

Tables [1–7](#page-5-0) summarize methods and results from whole-animal studies of DNT in DA-exposed animals. Each table represents studies grouped by time of exposure and species tested. Emphasis is placed on identifying the lowest dose at which DNT was observed for each study, and the specific effects reported at that dose. Where necessary to facilitate comparison between studies, units have been converted to mg/kg bw. Brief synopses of the tabulated information are presented below.

# **3.1 | Prenatal Exposure**

## **3.1.1 | Monkey**

Two publications reported on a single study population of *M. fascicularis* that was exposed to DA during gestation, and fol-lowed from birth to 1–2 months of age (Burbacher et al. [2019;](#page-14-13) Grant et al. [2019](#page-15-18)). Maternal animals were given DA orally, with doses of 0, 0.075, or 0.15mg/kg-day throughout breeding and gestation. No adverse effects were noted for newborns, but testing at 1–2months postnatal age revealed impairment of recognition memory in the 0.15mg DA/kg bw-day group (Burbacher et al. [2019](#page-14-13); Grant et al. [2019](#page-15-18)). See Table [1](#page-5-0) for more details of study methods and results.

## **3.1.2** | **Rodents**

One study in rats (Levin et al. [2005\)](#page-15-19) and five in mice (Dakshinamurti et al. [1993](#page-14-14); Mills et al. [2016;](#page-16-21) Shiotani et al. [2017;](#page-16-22) Tanemura et al. [2009](#page-17-8); Zuloaga et al. [2016\)](#page-17-9) provided data on the DNT of DA following gestational exposure. See Tables [2](#page-5-1) and [3](#page-6-0) for brief summaries of study methods and outcomes.

For three rodent studies conducted by the subcutaneous (SC) route of exposure, the lowest LOAELs during gestation ranged between 0.6 to 1.5mg/kg bw (Levin et al. [2005](#page-15-19); Mills et al. [2016;](#page-16-21)

<span id="page-5-0"></span>**TABLE 1** | Gestational exposure only; Non-human primates (*Macaca fascicularis*).



*Note:* The two papers tabulated above report studies of the same experimental animals evaluated at different times for different outcomes.

<span id="page-5-2"></span>aLOAEL=lowest observed adverse effect level. Outcomes for exposed animals significant at *p*<0.05 or less.

<span id="page-5-3"></span> $bNOAEL = no observed adverse effect level.$ 

<span id="page-5-4"></span>c DA=domoic acid.

<span id="page-5-5"></span> $dNI = not identified$ .

<span id="page-5-6"></span>e PNM=postnatal month.



<span id="page-5-1"></span>**TABLE 2** | Gestational exposure only; Rats.

<span id="page-5-7"></span> ${}^a$ GD = gestation day.

<span id="page-5-8"></span> $\rm bPNW$  = postnatal week.

#### <span id="page-6-0"></span>**TABLE 3** | Gestational exposure only; Mice.



<span id="page-6-1"></span><sup>a</sup>ANOVA=analysis of variance.

<span id="page-6-2"></span> $bMRI =$ magnetic resonance imaging.

<span id="page-6-4"></span><span id="page-6-3"></span>c ASD=autism spectrum disorder. dEEG = electroencephalogram.

<span id="page-6-5"></span>eGABA=gamma-aminobutyric acid.

Zuloaga et al. [2016\)](#page-17-9). Reported outcomes included altered social behaviors and impaired maze performances.

A mouse gavage study administered DA daily on GD 10–17, at doses of 0, 1, or 3mg DA/kg maternal bw-day (Shiotani et al. [2017\)](#page-16-22). Postnatal behavioral testing of prenatally-exposed pups revealed both dose and sex influences on offspring performance. Significant changes were found in postweaning tests of motor coordination and indicators of anxiety behavior, with no identified NOAEL dose, while maternal animals showed no seizures or other effects at any tested dose.

A single IP injection of 1mg DA/kg bw to pregnant mice was associated with behaviors indicating impaired learning and memory and increased anxiety in male offspring of 11weeks postnatal age (Tanemura et al. [2009](#page-17-8)). The study also noted myelination

## <span id="page-7-4"></span>**TABLE 4** | Zebrafish fertilized eggs or embryos.



<span id="page-7-0"></span>ahpf=hours post-fertilization.

<span id="page-7-1"></span>

<span id="page-7-2"></span><sup>b</sup>dpf=days post-fertilization.<br><sup>c</sup>Dose expressed as ng/embryo.

<span id="page-7-3"></span> $dPTZ =$ pentylenetetrazole.

failure and overgrowth of neuronal processes in limbic cortex neurons at necropsy of treated animals. In another study, a single IV dose of 0.6mg DA/kg bw to pregnant dams altered postnatal electroencephalogram (EEG) results as well as histopathology and brain biochemistry (Dakshinamurti et al. [1993\)](#page-14-14).

# **3.1.3** | **Zebrafish**

Six studies used zebrafish as the test species for DA-induced DNT (Panlilio, Aluru, and Hahn [2020;](#page-16-23) Panlilio et al. [2023,](#page-16-0) [2021](#page-16-24); Tiedeken and Ramsdell [2007](#page-17-10), [2009;](#page-17-11) Tiedeken, Ramsdell,

# <span id="page-8-0"></span>**TABLE 5** | Neonatal exposure; Rats.



(Continues)

# **TABLE 5** | (Continued)



#### **TABLE 5** | (Continued)



and Ramsdell [2005](#page-17-12)). Routes of exposure included "bath" exposure, microinjection, and IV injection. LOAELs by route were: 0.09E-6 mg DA/embryo by IV, 0.13E-6 mg/kg DA by microinjection, and 0.25 mM DA in the bath solution for larvae. Effects reported included seizures, reduced startle response, altered gene expression, reduced presence of reticulospinal neurons and primarily motor neuron axon collaterals, and disrupted myelination. See Table [4](#page-7-4) for more details of study methods and findings.

## **3.2 | Neonatal/Postnatal Exposure**

#### **3.2.1 | Rodents**

Young pups were dosed with DA in 18 rodent studies, and later assessed using behavioral and/or physiological studies at hours to weeks post-dosing. Only one study was conducted in mice, which was also the only study to use the gavage route (Sasaki et al. [2021](#page-16-27)). The other 17 studies used the rat model, with dosing by IP (Doucette et al. [2000](#page-15-25); Xi, Peng, and Ramsdell [1997](#page-17-15)) and/or SC injection (Adams, Doucette, and Ryan [2008](#page-14-16); Adams et al. [2009](#page-14-15); Bernard et al. [2007](#page-14-19); Burt, Ryan, and Doucette [2008a,](#page-14-17) [2008b;](#page-14-18) Doucette et al. [2004](#page-15-24), [2000;](#page-15-25) Gill et al. [2009](#page-15-22), [2012;](#page-15-21) Jandová et al. [2014;](#page-15-20) Levin et al. [2006](#page-15-23); Marriott et al. [2016](#page-16-25); Perry, Ryan, and Tasker [2009;](#page-16-26) Tasker et al. [2005;](#page-17-14) Thomsen et al. [2016;](#page-17-13) Wang et al. [2000\)](#page-17-16). Twelve of the 18 studies used a 0.02mg DA/kg bwday dose on each of postnatal days (PND) 8–14. Once established as reliably resulting in DNT, in the absence of general offspring or maternal toxicity, this dosing protocol was used for all but one study published from 2005 through 2016. Dose and timing procedures were selected to ensure detectable DNT.

Studies varied in the evaluations performed and in age(s) at testing. Significant adverse effects included alterations in perceptual processing, altered social behavior and spontaneous activity patterns, increased stress-related behavior patterns and seizure activity, and altered maze performance with impaired responses to reversal challenges to previously learned mazes. Measurable effects were reported on open-field behaviors as late as PND 150 following dosing with 0.02mg DA/kg-day on PND 8–14 (Burt, Ryan, and Doucette [2008b](#page-14-18)). Tables [5](#page-8-0) and [6](#page-11-0) provide more details of methods and outcomes for these studies, including histopathology and biochemical results.

# **3.2.2** | **Wild California Sea Lions (***Zalophus californianus***)**

DA toxicosis has been identified and studied in stranded marine mammals, most extensively in the California sea lion. Four studies of young sea lions ranged in type from a case study of a single individual to observations of larger populations of, or including, immature animals (Goldstein et al. [2008](#page-15-3), [2009](#page-15-15); Krucik et al. [2023](#page-15-26); Simeone et al. [2019\)](#page-16-28).

These wild animals were exposed to DA in utero and/or via their mother's milk due to maternal consumption of a natural diet including DA-contaminated marine organisms. Older immature animals may also have been exposed by direct consumption of a DA-contaminated diet. Exact dosing is unknown, but exposure was inferred from the presence of *Pseudo-nitzschia* blooms of identified toxigenic size class and environmental monitoring in ocean waters near the sea lions' breeding and pupping locations (Smith et al. [2023\)](#page-16-20). One study measured DA in bodily fluid samples from fetuses or pups (Goldstein et al. [2009\)](#page-15-15). The others presumed DA-exposure based on documented environmental sampling and field observations, along with pathological symptoms specifically characteristic of DA toxicosis (Krucik et al. [2023;](#page-15-26) Simeone et al. [2019\)](#page-16-28) and/or from identification of DA in tissues collected from other animals rescued at the same time and location (Goldstein et al. [2008\)](#page-15-3).

<span id="page-11-0"></span>

#### <span id="page-11-1"></span>**TABLE 7** | Wildlife; Marine mammals.



Neurological effects were so profound as to cause death or preclude release of surviving animals back to the wild after poststranding veterinary care. Effects included seizures (Goldstein et al. [2008](#page-15-3); Simeone et al. [2019](#page-16-28)), adult-onset medial temporal lobe epilepsy (Krucik et al. [2023\)](#page-15-26), brain edema (Goldstein et al. [2008](#page-15-3), [2009](#page-15-15)), and hippocampal lesions including one animal with hippocampal-focused encephalitis (Simeone et al. [2019\)](#page-16-28). See Table [7](#page-11-1) for more information on study populations and observations.

#### **4 | Discussion**

All the animal studies following gestational or larval exposure to DA (Tables [1–4](#page-5-0)) reported DNT. While adverse effects were not detected at birth for prenatally-exposed monkeys (Burbacher et al. [2019\)](#page-14-13), the same cohort of exposed infants demonstrated DNT at 1–2months postnatal age (Grant et al. [2019](#page-15-18)). All but one study (Sasaki et al. [2021\)](#page-16-27) of DA exposure in early postnatal life (Tables [5–7](#page-8-0)) reported DNT effects of DA. While different studies evaluated various specific effects, all of the observed outcomes are consistent with consequences of the known action of DA as an agonist of glutamate kainate receptors. Precise comparisons among studies for NOAEL and LOAEL doses are complicated by speciesdifferences in DA TK parameters, which in turn influenced choices for experimental route of exposure. Most of the experimental animal studies were designed to investigate specific DNT outcomes, rather than to establish a dose–response relationship.

## **4.1 | Gestational or Larval Experimental Exposures**

The similarities between macaque monkeys and humans in TK models for oral DA exposure support experimental non-human primates as a particularly relevant model for gestational exposure linked to DNT (Jing et al. [2018](#page-15-14); Shum et al. [2020](#page-16-2)). Data from other laboratory and wild species provide considerable supporting evidence for DNT following developmental exposure to similar dose levels of DA.

Impairment of recognition memory was found in 1–2monthold *M. fascicularis* following repeated daily prenatal exposure to DA (Grant et al. [2019\)](#page-15-18). The NOAEL from this study was 0.075mg/kg bw-day, which is 10 times lower than the adult NOAEL of 0.75mg/kg bw used to establish the TDI by the State of Washington (Marien [1996\)](#page-16-12).

The prevalence of parenteral routes of exposure in the studies of gestational exposure to DA in laboratory rats and mice may reflect the findings of reduced DA absorption from the rodent gut as compared to humans and other primates (Iverson and Truelove [1994\)](#page-15-5). Only one of five rodent injection studies administered a range of DA doses to pregnant rats (Levin et al. [2005](#page-15-19)). The study reported a NOAEL dose for DA of 0.3mg/kg on GD 13, with a LOAEL of 0.6mg/kg. Other investigators chose a single dose, selected to induce DNT outcomes in the absence of other observed toxic effects in offspring or maternal animals, and given to mice on a single gestation day (Dakshinamurti et al. [1993](#page-14-14); Mills et al. [2016](#page-16-21); Tanemura et al. [2009](#page-17-8); Zuloaga et al. [2016](#page-17-9)). Results were generally consistent across these studies, even including the one oral mouse study (Shiotani et al. [2017](#page-16-22)), with LOAEL doses ranging from 0.6–1.5mg/ kg bw. Observed effects of DA included altered maze performance, locomotor impairments, reduced social behaviors, altered functional connectivity patterns and sensorimotor gating, effects on

learning and anxiety behaviors, reduced seizure thresholds, and altered histopathology including myelination failure.

All but one zebrafish study was conducted by injection, either into a fertilized egg or embryo (Tiedeken and Ramsdell [2007;](#page-17-10) Tiedeken, Ramsdell, and Ramsdell [2005\)](#page-17-12) or into a larval vein (Panlilio, Aluru, and Hahn [2020;](#page-16-23) Panlilio et al. [2023,](#page-16-0) [2021\)](#page-16-24). While TK data for DA in zebrafish are not available, observations of excitotoxicity in adult anchovies given DA by intracoelomic injection suggested that fish may differ from mammals in distribution of absorbed DA, rather than in resistance to DA neurotoxicity (Lefebvre [2001\)](#page-15-13). Zebrafish demonstrated similar symptoms and sensitivity to DA as other species studied (Panlilio, Aluru, and Hahn [2020;](#page-16-23) Tiedeken, Ramsdell, and Ramsdell [2005\)](#page-17-12). (Panlilio, Aluru, and Hahn [2020\)](#page-16-23) expressly chose test doses "similar to those causing behavioral effects in developing [neonatal] rodents," while also below levels "associated with acute toxicity in adult humans."

#### **4.2 | Postnatal Experimental Exposures**

Sasaki and colleagues administered a single oral dose of DA (3.0mg/kg) to mice either on PND 14 or at 10weeks postnatal age (adult), and conducted behavioral testing at 12–13weeks postnatal age (Sasaki et al. [2021](#page-16-27)). In seeming contradiction to the generally greater sensitivity of immature individuals, effects on test performance were noted only in animals treated as adults. Treatment on PND 14, however, may have missed the critical window for irreversible developmental DA effects, while at the same time leaving 10weeks for possible repair of transitory effects before testing at 12weeks. The adult animals treated at 10weeks had only two weeks recovery time before testing.

All of the early life experimental studies performed in rats used an injection method of DA exposure, with most giving repeated doses of 0 or 0.02mg/kg-day on each of PNDs 8–14 (Adams, Doucette, and Ryan [2008;](#page-14-16) Adams et al. [2009;](#page-14-15) Bernard et al. [2007;](#page-14-19) Burt, Ryan, and Doucette [2008a](#page-14-17), [2008b;](#page-14-18) Doucette et al. [2004;](#page-15-24) Gill et al. [2009](#page-15-22), [2012](#page-15-21); Marriott et al. [2016](#page-16-25); Perry, Ryan, and Tasker [2009;](#page-16-26) Tasker et al. [2005](#page-17-14); Thomsen et al. [2016\)](#page-17-13). The lowest identified adverse effect dose for postnatal rat studies found seizure-like activity and altered hippocampal histopathology in adult animals following neonatal exposure to 0.005mg/ kg-day on PND 8–14 (Doucette et al. [2004\)](#page-15-24).

## **4.3 | Wild Sea Lions, Pre- and Post-Gestational Environmental Exposures**

Wild California sea lions are large placental mammals, whose natural diet contains many of the same seafood species humans also enjoy eating. The Channel Islands off the coast of Santa Barbara, California, and surrounding waters are a primary breeding and pupping ground for these animals. Recurring *Pseudo-nitzschia* blooms have affected the area in recent decades, causing serious harm to sea lions as well as other wild species (Brodie et al. [2006;](#page-14-10) Goldstein et al. [2008](#page-15-3)).

A survey of pregnant female sea lions stranded during California's *Pseudo-nitzschia* blooms of 1998 and 2002 found 209 cases of reproductive failure attributed to DA exposure (Brodie et al. [2006](#page-14-10)). Types of reproductive failure included spontaneous abortion, premature live birth, fetal death, and maternal death prior to parturition. Another study specifically assessed DA contents in fetuses and fetal membranes from stranded pregnant females (Lefebvre et al. [2018\)](#page-15-16). Although these studies did not assess DNT, they do provide context for evidence from surviving pups. In particular, the analysis of fetal body fluids demonstrated detectable levels of DA up to 8days following maternal rescue (i.e., post cessation of environmental exposure).

Sea lion pups believed to have been exposed to DA, prenatally and/or neonatally, exhibited clinical symptoms and postmortem brain pathology consistent with DA toxicosis (Goldstein et al. [2008](#page-15-3), [2009;](#page-15-15) Krucik et al. [2023](#page-15-26); Simeone et al. [2019](#page-16-28)). Beyond evidence that DA exposure occurred or was likely, quantitative exposure assessment for environmentally-exposed sea lions is not possible. Thus, direct comparisons cannot be made to adverse effect doses of DA identified for humans or experimental animal species. Additional complicating factors for assessment of sea lion data are the frequency of concurrent infections (e.g., bacterial, protozoal, viral), and contamination of the same habitat with chemicals such as DDTs and PCBs (Goldstein et al. [2009](#page-15-15)).

# **4.4 | Life Course Considerations for Sensitivity to DA**

The importance of a specific developmental stage to outcomes resulting from toxic exposures is a foundational principle of developmental toxicology (NRC [2000](#page-16-29)). The evidence summarized in this paper (Tables [1–7\)](#page-5-0) supports a peak of susceptibility to DA during prenatal and/or early postnatal development, when developing vertebrate nervous tissues are most vulnerable.

Gestation and early infancy are not the only life stages with enhanced sensitivity to the neurological effects of DA. Exposures to DA late in life are also of elevated concern (Hendrix et al. [2023\)](#page-15-27). An association between increased severity of effects and advanced age was noted in the initial sample of acutely poisoned adult humans (Perl, Bedard, Kosatsky, Hockin, Todd, Remis, et al. [1990](#page-16-6); Wekell, Hurst, and Lefebvre [2004\)](#page-17-6). All three patients who died in the hospital were over the age of 70, while only four of 19 hospitalized patients were under the age of 65. The younger hospitalized patients all had preexisting medical conditions, notably compromised kidney function. Experiments in mice compared sex and age for influence on sensitivity to effects of DA (Hendrix et al. [2023\)](#page-15-27). Aged mice were found to be more susceptible to acute DA neurotoxicity than young adult mice, and females more sensitive than males. The older animals also accumulated higher concentrations of DA in serum and tissues compared to younger animals.

Concern has also been expressed for consequences of repeated, chronic exposures of healthy adult human populations to DA at levels below regulatory limits. A minimal, but detectable, memory decline was associated with repetitive sub-acute, dietary exposures to DA at levels below established allowable concentrations in the seafood consumed (Grattan et al. [2021;](#page-15-28) Stuchal et al. [2020](#page-17-17)). These effects were observed in a cohort of 500 adult Native American tribal members residing in coastal areas of the Pacific Northwest, who were studied over a period of 8 years, and who otherwise exhibited stable cognition.

# **5 | Conclusions**

For any toxin, adverse effect dose levels can vary with exposure patterns, the sensitivity of target tissues, as well as with TK factors such as rates of absorption and elimination. Apart from genetic variation within and between populations, life course considerations are critical for determining the sensitive windows for exposure to toxins (Halfon et al. [2018](#page-15-29); Schaffer, Smith, and Faustman [2017](#page-16-30)).

Internationally, government agencies as well as individual researchers have opined that current regulatory standards for DA in seafood, which have not been revisited since initial implementation, may be insufficient to protect exposed individuals at all life stages and/or patterns of consumption (COT [2001;](#page-14-20) EFSA [2009;](#page-15-11) Grattan et al. [2021](#page-15-28); Wekell, Hurst, and Lefebvre [2004\)](#page-17-6).

Current US federal action levels for DA are based on a LOAEL of 1.0mg DA/kg bw, adjusted by a 10-fold UF for within-species variation to a tolerable dose of 0.1mg DA/ kg bw (DHHS [1993\)](#page-15-17). The European Union adopted the Health Canada and US FDA regulatory limit of 20ppm for DA in bivalve shellfish in 1997 (COT [2001;](#page-14-20) EU. [1997\)](#page-15-30). The United Kingdom Food Standard Agency Committee on Toxicity (COT) (COT [2001\)](#page-14-20), and the European Food Safety Authority Panel on Contaminants in the Food Chain (EFSA) (EFSA [2009](#page-15-11)), reviewed the regulatory limit for DA. Both groups considered an UF of 10 to be inadequate. The COT concluded that, while the action level "may protect against major outbreaks," it provides a "pragmatic guideline rather than a toxicologically based limit." EFSA noted that sensitive indicators of adverse neurological effects were not assessed in affected people during the 1987 ASP outbreak. EFSA recommended lowering the action limit to 4.5ppm in shellfish meat, based on a 60 kg body weight, a 400 g meal size, and an additional UF of three to extrapolate from a LOAEL.

In addition, or alternatively, to changing regulatory action levels, public health outreach to communities and groups at risk of DAtoxicity has been recommended (Grattan et al. [2021](#page-15-28)). Clinicians caring for patients sickened after consumption of seafood, particularly shellfish, could benefit from raised awareness of DA toxicity. Initial symptoms of general acute gastrointestinal illness combined with the rapid clearance of DA from the body may complicate accurate diagnosis (CPCS [2016\)](#page-14-21). An example of relevant outreach is provided by the Washington State Department of Health, which issued an interim health advisory for regular consumers of razor clams containing allowable levels of DA recommending that consumers "…eat no more than 15 razor clams each month for 12 consecutive months…." (WADOH [2023\)](#page-17-18). While the interim advisory is intended for everyone, sensitive subpopulations are specified: "…especially women who are or might become pregnant, nursing mothers, children, the elderly, and people with compromised renal function." Similar public health messaging on all potentially impacted types of seafood

could be made more widely available through relevant agencies in affected states, and through educational materials for health professionals serving sensitive populations.

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#### **Conflicts of Interest**

The authors declare no conflicts of interest.

#### **Data Availability Statement**

Data sharing is not applicable to this article as no datasets were generated or analyzed during the current study.

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#### **References**

<span id="page-14-16"></span>Adams, A., T. A. Doucette, and C. L. Ryan. 2008. "Altered Pre-Pulse Inhibition in Adult Rats Treated Neonatally With Domoic Acid." *Amino Acids* 35: 157–160.

<span id="page-14-15"></span>Adams, A. L., T. A. Doucette, R. James, and C. L. Ryan. 2009. "Persistent Changes in Learning and Memory in Rats Following Neonatal Treatment With Domoic Acid." *Physiology & Behavior* 96, no. 4–5: 505– 512. [https://doi.org/10.1016/j.physbeh.2008.11.019.](https://doi.org/10.1016/j.physbeh.2008.11.019)

<span id="page-14-8"></span>Anderson, D. M., E. Fensin, C. J. Gobler, et al. 2021. "Marine Harmful Algal Blooms (HABs) in the United States: History, Current Status and Future Trends." *Harmful Algae* 102: 101975. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.hal.2021.101975) [hal.2021.101975](https://doi.org/10.1016/j.hal.2021.101975).

<span id="page-14-7"></span>AOP-48. 2023. "Binding of Agonists to Ionotropic Glutamate Receptors in Adult Brain Causes Excitotoxicity that Mediates Neuronal Cell Death, Contributing to Learning and Memory Impairment." (AOP: 48). <https://aopwiki.org/aops/48>.

<span id="page-14-9"></span>Baj, A., E. Moro, M. Bistoletti, V. Orlandi, F. Crema, and C. Giaroni. 2019. "Glutamatergic Signaling Along the Microbiota-Gut-Brain Axis." *International Journal of Molecular Sciences* 20, no. 6: 1482. [https://](https://www.mdpi.com/1422-0067/20/6/1482) [www.mdpi.com/1422-0067/20/6/1482.](https://www.mdpi.com/1422-0067/20/6/1482)

<span id="page-14-4"></span>Bates, S., K. A. Hubbard, N. Lundholm, M. Montresor, and C. Pin Leaw. 2018. "Pseudo-Nitzschia, Nitzschia, and Domoic Acid: New Research

Since 2011." *Harmful Algae* 79: 3–43. [https://doi.org/10.1016/j.hal.2018.](https://doi.org/10.1016/j.hal.2018.06.001) [06.001.](https://doi.org/10.1016/j.hal.2018.06.001)

<span id="page-14-19"></span>Bernard, P. B., D. S. Macdonald, D. A. Gill, C. L. Ryan, and R. A. Tasker. 2007. "Hippocampal Mossy Fiber Sprouting and Elevated trkB Receptor Expression Following Systemic Administration of Low Dose Domoic Acid During Neonatal Development." *Hippocampus* 17, no. 11: 1121– 1133. <https://doi.org/10.1002/hipo.20342>.

<span id="page-14-5"></span>Bernstein, S., R. I. Ruiz-Cooley, R. Kudela, C. R. Anderson, R. Dunkin, and J. C. Field. 2021. "Stable Isotope Analysis Reveals Differences in Domoic Acid Accumulation and Feeding Strategies of Key Vectors in a California Hotspot for Outbreaks." *Harmful Algae* 110: 102117. [https://](https://doi.org/10.1016/j.hal.2021.102117) [doi.org/10.1016/j.hal.2021.102117.](https://doi.org/10.1016/j.hal.2021.102117)

<span id="page-14-6"></span>Bowen, L., S. Knowles, K. Lefebvre, et al. 2022. "Divergent Gene Expression Profiles in Alaskan Sea Otters: An Indicator of Chronic Domoic Acid Exposure?" *Oceans* 3, no. 3: 401–418. [https://www.mdpi.](https://www.mdpi.com/2673-1924/3/3/27) [com/2673-1924/3/3/27](https://www.mdpi.com/2673-1924/3/3/27).

<span id="page-14-10"></span>Brodie, E. C., F. M. Gulland, D. J. Greig, et al. 2006. "Domoic Acid Causes Reproductive Failure in California Sea Lions (*Zalophus californianus*)." *Marine Mammal Science* 22, no. 3: 700–707.

<span id="page-14-13"></span>Burbacher, T. M., K. S. Grant, R. Petroff, et al. 2019. "Effects of Oral Domoic Acid Exposure on Maternal Reproduction and Infant Birth Characteristics in a Preclinical Nonhuman Primate Model." *Neurotoxicology and Teratology* 72: 10–21. [https://doi.org/10.1016/j.ntt.](https://doi.org/10.1016/j.ntt.2019.01.001) [2019.01.001](https://doi.org/10.1016/j.ntt.2019.01.001).

<span id="page-14-17"></span>Burt, M. A., C. L. Ryan, and T. A. Doucette. 2008a. "Altered Responses to Novelty and Drug Reinforcement in Adult Rats Treated Neonatally With Domoic Acid." *Physiology & Behavior* 93, no. 1–2: 327–336. [https://](https://doi.org/10.1016/j.physbeh.2007.09.003) [doi.org/10.1016/j.physbeh.2007.09.003](https://doi.org/10.1016/j.physbeh.2007.09.003).

<span id="page-14-18"></span>Burt, M. A., C. L. Ryan, and T. A. Doucette. 2008b. "Low Dose Domoic Acid in Neonatal Rats Abolishes Nicotine Induced Conditioned Place Preference During Late Adolescence." *Amino Acids* 35, no. 1: 247–249. <https://doi.org/10.1007/s00726-007-0584-2>.

<span id="page-14-11"></span>California Fish and Game Code. 2017. "California Fish and Game Code—FGC section 5523 et seq." <https://leginfo.legislature.ca.gov/>.

<span id="page-14-12"></span>CDPH. 2017. "Domoic Acid Frequently Asked Questions." [https://www.](https://www.cdph.ca.gov/Programs/CEH/DRSEM/Pages/EMB/Shellfish/Domoic-Acid.aspx) [cdph.ca.gov/Programs/CEH/DRSEM/ Pages/ EMB/Shellfish/Domoic-](https://www.cdph.ca.gov/Programs/CEH/DRSEM/Pages/EMB/Shellfish/Domoic-Acid.aspx)[Acid.aspx](https://www.cdph.ca.gov/Programs/CEH/DRSEM/Pages/EMB/Shellfish/Domoic-Acid.aspx).

<span id="page-14-3"></span>Clancy, B., R. B. Darlington, and B. L. Finlay. 2001. "Translating Developmental Time Across Mammalian Species." *Neuroscience* 105, no. 1: 7–17. [https://www.sciencedirect.com/science/article/abs/pii/](https://www.sciencedirect.com/science/article/abs/pii/S0306452201001713) [S0306452201001713](https://www.sciencedirect.com/science/article/abs/pii/S0306452201001713).

<span id="page-14-2"></span>Clancy, B., B. L. Finlay, R. B. Darlington, and K. J. S. Anand. 2008. "Extrapolating Brain Development From Experimental Species to Humans." *Neurotoxicology* 28, no. 5: 1–15. [https://pmc.ncbi.nlm.nih.](https://pmc.ncbi.nlm.nih.gov/articles/PMC2077812/) [gov/articles/PMC2077812/](https://pmc.ncbi.nlm.nih.gov/articles/PMC2077812/).

<span id="page-14-0"></span>Costa, L. G., G. Giordano, and E. M. Faustman. 2010. "Domoic Acid as a Developmental Neurotoxin." *Neurotoxicology* 31, no. 5: 409–423. <https://doi.org/10.1016/j.neuro.2010.05.003>.

<span id="page-14-1"></span>Costa, L. G., M. Guizzetti, and A. Vitalone. 2004. "Diet-Brain Connections: Role of Neurotoxicants." *Environmental Toxicology and Pharmacology* 19, no. 3: 395–400. [https://doi.org/10.1016/j.etap.2004.](https://doi.org/10.1016/j.etap.2004.12.001) [12.001.](https://doi.org/10.1016/j.etap.2004.12.001)

<span id="page-14-20"></span>COT. 2001. "(Committee on Toxicity of Chemicals in Food, Consumer Products, and the Environment) Statement on Amnesic Shellfish Poisoning." [https://cot.food.gov.uk/sites/default/files/cot/cot](https://cot.food.gov.uk/sites/default/files/cot/cot-asp.pdf)[asp.pdf.](https://cot.food.gov.uk/sites/default/files/cot/cot-asp.pdf)

<span id="page-14-21"></span>CPCS. 2016. "Amnesic Shellfish Poisoning." California Poison Control System. <https://calpoison.org/content/amnesic-shellfish-poisoning>.

<span id="page-14-14"></span>Dakshinamurti, K., S. K. Sharma, M. Sundaram, and T. Watanabe. 1993. "Hippocampal Changes in Developing Postnatal Mice Following Intrauterine Exposure to Domoic Acid." *Journal of Neuroscience* 13, no. 10: 4486–4495. [https://doi.org/10.1523/jneurosci.13-10-04486.1993.](https://doi.org/10.1523/jneurosci.13-10-04486.1993)

<span id="page-15-17"></span>DHHS. 1993. "Memorandum—Marine Biotoxins in Dungness Crab. Memorandum—Marine Biotoxins in Dungness Crab." [http://www.](http://www.oceansciencetrust.org/wp-content/uploads/2016/07/DA_Crabs_93.pdf) oceansciencetrust.org/wp- [content/uploads/2016/07/DA\\_Crabs\\_](http://www.oceansciencetrust.org/wp-content/uploads/2016/07/DA_Crabs_93.pdf) [93.pdf.](http://www.oceansciencetrust.org/wp-content/uploads/2016/07/DA_Crabs_93.pdf)

<span id="page-15-24"></span>Doucette, T. A., P. B. Bernard, H. Husum, M. A. Perry, C. L. Ryan, and R. A. Tasker. 2004. "Low Doses of Domoic Acid During Postnatal Development Produce Permanent Changes in Rat Behaviour and Hippocampal Morphology." *Neurotoxicity Research* 6, no. 7–8: 555–563. <https://doi.org/10.1007/bf03033451>.

<span id="page-15-25"></span>Doucette, T. A., S. M. Strain, G. V. Allen, C. L. Ryan, and R. A. Tasker. 2000. "Comparative Behavioural Toxicity of Domoic Acid and Kainic Acid in Neonatal Rats." *Neurotoxicology and Teratology* 22, no. 6: 863– 869. [https://doi.org/10.1016/s0892-0362\(00\)00110-0](https://doi.org/10.1016/s0892-0362(00)00110-0).

<span id="page-15-0"></span>Doucette, T. A., and R. A. Tasker. 2016. "Perinatal Domoic Acid as a Neuroteratogen." *Current Topics in Behavioral Neurosciences* 29: 87– 110. [https://doi.org/10.1007/7854\\_2015\\_417](https://doi.org/10.1007/7854_2015_417).

<span id="page-15-11"></span>EFSA. 2009. "Scientific Opinion of the Panel on Contaminants in the Food Chain on a Request From the European Commission on Marine Biotoxins in Shellfish—Domoic Acid." *EFSA Journal* 1181: 1–61.

<span id="page-15-30"></span>EU. 1997. "Council Directive 97/61/EC of 20 October 1997 Amending the Annex to Directive 91/492/EEC Laying Down the Health Conditions for the Production and Placing on the Market of Live Bivalve Molluscs." Official Journal of the European Communities. L 295/35.

<span id="page-15-10"></span>Fritz, L., M. A. Quilliam, J. L. C. Wright, A. M. Beale, and T. M. Work. 1992. "An Outbreak of Domoic Acid Poisoning Attributed to the Pennate Diatom *Pseudonitzschia australis*." *Journal of Phycology* 28: 439–442.

<span id="page-15-8"></span>Gibble, C. M., R. M. Kudela, S. Knowles, B. Bodenstein, and K. A. Lefebvre. 2021. "Domoic Acid and Saxitoxin in Seabirds in the United States Between 2007 and 2018." *Harmful Algae* 103: 101981. [https://doi.](https://doi.org/10.1016/j.hal.2021.101981) [org/10.1016/j.hal.2021.101981](https://doi.org/10.1016/j.hal.2021.101981).

<span id="page-15-22"></span>Gill, D. A., J. F. Bastlund, N. J. Anderson, and R. A. Tasker. 2009. "Reductions in Paradoxical Sleep Time in Adult Rats Treated Neonatally With Low Dose Domoic Acid." *Behavioural Brain Research* 205, no. 2: 564–567.<https://doi.org/10.1016/j.bbr.2009.07.018>.

<span id="page-15-21"></span>Gill, D. A., M. A. Perry, E. P. McGuire, A. Pérez-Gómez, and R. A. Tasker. 2012. "Low-Dose Neonatal Domoic Acid Causes Persistent Changes in Behavioural and Molecular Indicators of Stress Response in Rats." *Behavioural Brain Research* 230, no. 2: 409–417. [https://doi.org/](https://doi.org/10.1016/j.bbr.2012.02.036) [10.1016/j.bbr.2012.02.036](https://doi.org/10.1016/j.bbr.2012.02.036).

<span id="page-15-3"></span>Goldstein, T., J. A. K. Mazet, T. S. Zabka, et al. 2008. "Novel Symptomatology and Changing Epidemiology of Domoic Acid Toxicosis in California Sea Lions (*Zalophus californianus*): An Increasing Risk to Marine Mammal Health." *Proceedings of the Royal Society B* 275: 267–276.

<span id="page-15-15"></span>Goldstein, T., T. S. Zabka, R. L. Delong, et al. 2009. "The Role of Domoic Acid in Abortion and Premature Parturition of California Sea Lions (*Zalophus californianus*) on San Miguel Island." *California. Journal of Wildlife Diseases* 45, no. 1: 91–108. [https://doi.org/10.7589/0090-3558-](https://doi.org/10.7589/0090-3558-45.1.91) [45.1.91](https://doi.org/10.7589/0090-3558-45.1.91).

<span id="page-15-4"></span>Grant, K. S., T. M. Burbacher, E. M. Faustman, and L. Grattan. 2010. "Domoic Acid: Neurobehavioral Consequences of Exposure to a Prevalent Marine Biotoxin." *Neurotoxicology and Teratology* 32, no. 2: 132–141. <https://doi.org/10.1016/j.ntt.2009.09.005>.

<span id="page-15-18"></span>Grant, K. S., B. Crouthamel, C. Kenney, et al. 2019. "Preclinical Modeling of Exposure to a Global Marine Bio-Contaminant: Effects of In Utero Domoic Acid Exposure on Neonatal Behavior and Infant Memory." *Neurotoxicology and Teratology* 73: 1–8. [https://doi.org/10.](https://doi.org/10.1016/j.ntt.2019.01.003) [1016/j.ntt.2019.01.003](https://doi.org/10.1016/j.ntt.2019.01.003).

<span id="page-15-1"></span>Grattan, L. M. 2022. "Invited Perspective: The Relevance of Animal Models of Domoic Acid Neurotoxicity to Human Health." *Environmental Health Perspectives* 130, no. 9: 091302. [https://doi.org/](https://doi.org/10.1289/EHP11774) [10.1289/EHP11774](https://doi.org/10.1289/EHP11774).

<span id="page-15-28"></span>Grattan, L. M., L. Kaddis, J. K. Tracy, and J. G. Morris. 2021. "Long Term Memory Outcome of Repetitive, Low-Level Dietary Exposure to Domoic Acid in Native Americans." *International Journal of Environmental Research and Public Health* 18, no. 8: 3955. [https://www.](https://www.mdpi.com/1660-4601/18/8/3955) [mdpi.com/1660-4601/18/8/3955.](https://www.mdpi.com/1660-4601/18/8/3955)

<span id="page-15-29"></span>Halfon, N., C. B. Forrest, R. M. Lerner, and E. M. Faustman. 2018. *Handbook of Life Course Health Development*, edited by C. B. F. N. Halfon, R. M. Lerner, and E. M. Faustman, 1st ed. Cham, Switzerland: Springer Cham.<https://doi.org/10.1007/978-3-319-47143-3>.

<span id="page-15-27"></span>Hendrix, A. M., K. A. Lefebvre, E. K. Bowers, R. Stuppard, T. Burbacher, and D. J. Marcinek. 2023. "Age and Sex as Determinants of Acute Domoic Acid Toxicity in a Mouse Model." *Toxins (Basel)* 15, no. 4: 259. [https://www.mdpi.com/2072-6651/15/4/259.](https://www.mdpi.com/2072-6651/15/4/259)

<span id="page-15-5"></span>Iverson, F., and J. Truelove. 1994. "Toxicology and Seafood Toxins: Domoic Acid." *Natural Toxins* 2: 334–339. [https://doi.org/10.1002/nt.](https://doi.org/10.1002/nt.2620020514) [2620020514](https://doi.org/10.1002/nt.2620020514).

<span id="page-15-20"></span>Jandová, K., P. Kozler, M. Langmeier, D. Marešová, J. Pokorný, and V. Riljak. 2014. "Influence of Low-Dose Neonatal Domoic Acid on the Spontaneous Behavior of Rats in Early Adulthood." *Physiological Research* 63, no. Suppl 4: S521–S528.

<span id="page-15-14"></span>Jing, J., R. Petroff, S. Shum, et al. 2018. "Toxicokinetics and Physiologically Based Pharmacokinetic Modeling of the Shellfish Toxin Domoic Acid in Nonhuman Primates." *Drug Metabolism and Disposition* 46, no. 2: 155–165. [https://doi.org/10.1124/dmd.117.078485.](https://doi.org/10.1124/dmd.117.078485)

<span id="page-15-9"></span>Kreuder, C. M., M. A. Miller, L. J. Lowenstine, et al. 2005. "Evaluations of Cardiac Lesions and Risk Factors Associated With Myocarditis and Dilated Cardiomyopathy in Southern Sea Otters (*Enhydra lutris nereis*)." *American Journal of Veterinary Research* 66, no. 2: 289–299.

<span id="page-15-26"></span>Krucik, D. D. R., P. Cook, M. Cathey, et al. 2023. "Adult-Onset Epilepsy and Hippocampal Pathology in a California Sea Lion (*Zalophus californianus*): A Case Study of Suspected In Utero Exposure to Domoic Acid." *Neurotoxicology* 96: 13–18. [https://doi.org/10.1016/j.neuro.2023.02.010.](https://doi.org/10.1016/j.neuro.2023.02.010)

<span id="page-15-2"></span>Lanphear, B. P. 2015. "The Impact of Toxins on the Developing Brain." *Annual Review of Public Health* 36: 211–230.

<span id="page-15-12"></span>Larm, J. A., P. M. Beart, and N. S. Cheung. 1997. "Neurotoxin Domoic Acid Produces Cytotoxicity via Kainate- and AMPA-Sensitive Receptors in Cultured Cortical Neurones." *Neurochemistry International* 31, no. 5: 677–682. [https://doi.org/10.1016/s0197-0186\(97\)00030-2](https://doi.org/10.1016/s0197-0186(97)00030-2).

<span id="page-15-6"></span>Lefebvre, K. A., E. R. Frame, and P. S. Kendrick. 2012. "Domoic Acid and Fish Behavior: A Review." *Harmful Algae* 13: 126–130. [https://doi.](https://doi.org/10.1016/j.hal.2011.09.011) [org/10.1016/j.hal.2011.09.011](https://doi.org/10.1016/j.hal.2011.09.011).

<span id="page-15-16"></span>Lefebvre, K. A., A. Hendrix, B. Halaska, et al. 2018. "Domoic Acid in California Sea Lion Fetal Fluids Indicates Continuous Exposure to a Neuroteratogen Poses Risks to Mammals." *Harmful Algae* 79: 53–57. [https://doi.org/10.1016/j.hal.2018.06.003.](https://doi.org/10.1016/j.hal.2018.06.003)

<span id="page-15-7"></span>Lefebvre, K. A., D. P. Noren, I. R. Schultz, S. M. Bogard, J. Wilson, and B.-T. L. Eberhart. 2007. "Uptake, Tissue Distribution and Excretion of Domoic Acid After Oral Exposure in Coho Salmon (*Oncorhynchus kisutch*)." *Aquatic Toxicology* 81, no. 3: 266–274. [https://doi.org/10.](https://doi.org/10.1016/j.aquatox.2006.12.009) [1016/j.aquatox.2006.12.009.](https://doi.org/10.1016/j.aquatox.2006.12.009)

<span id="page-15-13"></span>Lefebvre, K. A. D., S. L. Dovel, and M. W. Silver. 2001. "Tissue Distribution and Neurotoxic Effects of Domoic Acid in a Prominent Vector Species, the Northern Anchove *Engraulis mordax*." *Marine Biology* 138: 693–700. [https://doi.org/10.1007/s002270000509.](https://doi.org/10.1007/s002270000509)

<span id="page-15-23"></span>Levin, E. D., W. G. Pang, J. Harrison, P. Williams, A. Petro, and J. S. Ramsdell. 2006. "Persistent Neurobehavioral Effects of Early Postnatal Domoic Acid Exposure in Rats." *Neurotoxicology and Teratology* 28, no. 6: 673–680. [https://doi.org/10.1016/j.ntt.2006.08.005.](https://doi.org/10.1016/j.ntt.2006.08.005)

<span id="page-15-19"></span>Levin, E. D., K. Pizarro, W. G. Pang, J. Harrison, and J. S. Ramsdell. 2005. "Persisting Behavioral Consequences of Prenatal Domoic Acid Exposure in Rats." *Neurotoxicology and Teratology* 27, no. 5: 719–725. [https://doi.org/10.1016/j.ntt.2005.06.017.](https://doi.org/10.1016/j.ntt.2005.06.017)

<span id="page-16-12"></span>Marien, K. 1996. "Establishing Tolerable Dungeness Crab (*Cancer magister*) and Razor Clam (*Siliqua patula*) Domoic Acid Contaminant Levels." *Environmental Health Perspectives* 104, no. 11: 1230–1236.

<span id="page-16-25"></span>Marriott, A. L., R. A. Tasker, C. L. Ryan, and T. A. Doucette. 2016. "Alterations to Prepulse Inhibition Magnitude and Latency in Adult Rats Following Neonatal Treatment With Domoic Acid and Social Isolation Rearing." *Behavioural Brain Research* 298: 310–317. [https://](https://doi.org/10.1016/j.bbr.2015.11.009) [doi.org/10.1016/j.bbr.2015.11.009](https://doi.org/10.1016/j.bbr.2015.11.009).

<span id="page-16-8"></span>Maucher Fuquay, J., N. Muha, Z. Wang, and J. S. Ramsdell. 2012a. "Elimination Kinetics of Domoic Acid From the Brain and Cerebrospinal Fluid of the Pregnant Rat." *Chemical Research in Toxicology* 25, no. 12: 2805–2809. [https://doi.org/10.1021/tx300434s.](https://doi.org/10.1021/tx300434s)

<span id="page-16-9"></span>Maucher Fuquay, J., N. Muha, Z. Wang, and J. S. Ramsdell. 2012b. "Toxicokinetics of Domoic Acid in the Fetal Rat." *Toxicology* 294, no. 1: 36–41. <https://doi.org/10.1016/j.tox.2012.01.012>.

<span id="page-16-10"></span>Maucher, J. M., and J. S. Ramsdell. 2005. "Domoic Acid Transfer to Milk: Evaluation of a Potential Route of Neonatal Exposure." *Environmental Health Perspectives* 113, no. 4: 461–464. [https://doi.org/](https://doi.org/10.1289/ehp.7649) [10.1289/ehp.7649](https://doi.org/10.1289/ehp.7649).

<span id="page-16-21"></span>Mills, B. D., H. L. Pearce, O. Khan, B. R. Jarrett, D. A. Fair, and G. P. Lahvis. 2016. "Prenatal Domoic Acid Exposure Disrupts Mouse Pro-Social Behavior and Functional Connectivity MRI." *Behavioural Brain Research* 308: 14–23. <https://doi.org/10.1016/j.bbr.2016.03.039>.

<span id="page-16-5"></span>Moriarty, M. E., M. T. Tinker, M. A. Miller, et al. 2021. "Exposure to Domoic Acid Is an Ecological Driver of Cardiac Disease in Southern Sea Otters☆." *Harmful Algae* 101: 101973.<https://doi.org/10.1016/j.hal.2020.101973>.

<span id="page-16-29"></span>NRC. 2000. *Scientific Frontiers in Developmental Toxicology and Risk Assessment*, edited by N. R. C. C. o. D. Toxicology. Washington, DC: National Academies Press (US). [https://www.ncbi.nlm.nih.gov/books/](https://www.ncbi.nlm.nih.gov/books/NBK225674/) [NBK225674/](https://www.ncbi.nlm.nih.gov/books/NBK225674/).

<span id="page-16-17"></span>OEHHA. 2017. "Domoic Acid Workshop: Evaluating the State of the Science and Implications for Human Toxicity." [https://oehha.ca.gov/](https://oehha.ca.gov/fish/events/domoic-acid-workshop) [fish/events/domoic-acid-workshop](https://oehha.ca.gov/fish/events/domoic-acid-workshop).

<span id="page-16-14"></span>OEHHA. 2018. "Frequently Asked Questions about Domoic Acid in Seafood." [https://oehha.ca.gov/fish/fact-sheet/frequently-asked-quest](https://oehha.ca.gov/fish/fact-sheet/frequently-asked-questions-about-domoic-acid-seafood) [ions-about-domoic-acid-seafood](https://oehha.ca.gov/fish/fact-sheet/frequently-asked-questions-about-domoic-acid-seafood).

<span id="page-16-13"></span>OEHHA. 2019a. "Domoic Acid (a Marine Biotoxin) in Fish and Shellfish." [https://oehha.ca.gov/fish/general-info/domoic-acid-marine](https://oehha.ca.gov/fish/general-info/domoic-acid-marine-biotoxin-fish-and-shellfish)[biotoxin-fish-and-shellfish.](https://oehha.ca.gov/fish/general-info/domoic-acid-marine-biotoxin-fish-and-shellfish)

<span id="page-16-18"></span>OEHHA. 2019b. "Domoic Acid Webinar: Research on Effects of Repeat Low-Level Exposures and its Implications for Human Toxicity." [https://](https://oehha.ca.gov/fish/events/domoic-acid-webinar-research-effects-repeat-low-level-exposures-and-its-implications) [oehha.ca.gov/fish/events/domoic-acid-webinar-research-effects-repea](https://oehha.ca.gov/fish/events/domoic-acid-webinar-research-effects-repeat-low-level-exposures-and-its-implications) [t-low-level-exposures-and-its-implications.](https://oehha.ca.gov/fish/events/domoic-acid-webinar-research-effects-repeat-low-level-exposures-and-its-implications)

<span id="page-16-19"></span>OEHHA. 2020. "Prioritization: Chemicals Identified for Consultation with the Developmental and Reproductive Toxicant Identification Committee." [https://www.google.com/url?client=internal-element](https://www.google.com/url?client=internal-element-cse&cx=001779225245372747843:v3sx-oyt7xc&q=https://oehha.ca.gov/media/downloads/crnr/dartprioritization100120.pdf&sa=U&ved=2ahUKEwiL5tv7j-z_AhWOhu4BHWPpB2YQFnoECAQQAg&usg=AOvVaw1Thlijw0lbAK4uFt48HsAq)[cse&cx=001779225245372747843:v3sx-oyt7xc&q=https://oehha.ca.](https://www.google.com/url?client=internal-element-cse&cx=001779225245372747843:v3sx-oyt7xc&q=https://oehha.ca.gov/media/downloads/crnr/dartprioritization100120.pdf&sa=U&ved=2ahUKEwiL5tv7j-z_AhWOhu4BHWPpB2YQFnoECAQQAg&usg=AOvVaw1Thlijw0lbAK4uFt48HsAq) [gov/media/downloads/crnr/dartprioritization100120.pdf&sa=U&ved=](https://www.google.com/url?client=internal-element-cse&cx=001779225245372747843:v3sx-oyt7xc&q=https://oehha.ca.gov/media/downloads/crnr/dartprioritization100120.pdf&sa=U&ved=2ahUKEwiL5tv7j-z_AhWOhu4BHWPpB2YQFnoECAQQAg&usg=AOvVaw1Thlijw0lbAK4uFt48HsAq) [2ahUKEwiL5tv7j-z\\_AhWOhu4BHWPpB2YQFnoECAQQAg&usg=](https://www.google.com/url?client=internal-element-cse&cx=001779225245372747843:v3sx-oyt7xc&q=https://oehha.ca.gov/media/downloads/crnr/dartprioritization100120.pdf&sa=U&ved=2ahUKEwiL5tv7j-z_AhWOhu4BHWPpB2YQFnoECAQQAg&usg=AOvVaw1Thlijw0lbAK4uFt48HsAq) [AOvVaw1Thlijw0lbAK4uFt48HsAq](https://www.google.com/url?client=internal-element-cse&cx=001779225245372747843:v3sx-oyt7xc&q=https://oehha.ca.gov/media/downloads/crnr/dartprioritization100120.pdf&sa=U&ved=2ahUKEwiL5tv7j-z_AhWOhu4BHWPpB2YQFnoECAQQAg&usg=AOvVaw1Thlijw0lbAK4uFt48HsAq).

<span id="page-16-4"></span>OEHHA. 2022a. "Marine Harmful Algal Blooms." [https://oehha.ca.](https://oehha.ca.gov/media/epic/downloads/04marinehabs.pdf) [gov/media/epic/downloads/04marinehabs.pdf](https://oehha.ca.gov/media/epic/downloads/04marinehabs.pdf).

<span id="page-16-15"></span>OEHHA. 2022b. "Title 17, California Code of Regulations (CCR) §2500, §2593, §2641.5-2643.20, and §2800-2812 Reportable Diseases and Conditions." [https://www.cdph.ca.gov/Programs/CID/DCDC/CDPH%](https://www.cdph.ca.gov/Programs/CID/DCDC/CDPH Document Library/ReportableDiseases.pdf) [20Document%20Library/ReportableDiseases.pdf](https://www.cdph.ca.gov/Programs/CID/DCDC/CDPH Document Library/ReportableDiseases.pdf).

<span id="page-16-16"></span>OEHHA. 2024. "Marine Harmful Algal Bloom (HAB)-Related Illness Tracking." [https://oehha.ca.gov/fish/general-info/marine-harmful](https://oehha.ca.gov/fish/general-info/marine-harmful-algal-bloom-hab-related-illness-tracking)[algal-bloom-hab-related-illness-tracking](https://oehha.ca.gov/fish/general-info/marine-harmful-algal-bloom-hab-related-illness-tracking).

<span id="page-16-23"></span>Panlilio, J. M., N. Aluru, and M. E. Hahn. 2020. "Developmental Neurotoxicity of the Harmful Algal Bloom Toxin Domoic Acid: Cellular and Molecular Mechanisms Underlying Altered Behavior in the Zebrafish Model." *Environmental Health Perspectives* 128, no. 11: 117002. [https://doi.org/10.1289/ehp6652.](https://doi.org/10.1289/ehp6652)

<span id="page-16-0"></span>Panlilio, J. M., K. M. Hammar, N. Aluru, and M. E. Hahn. 2023. "Developmental Exposure to Domoic Acid Targets Reticulospinal Neurons and Leads to Aberrant Myelination in the Spinal Cord." *Scientific Reports* 13, no. 1: 2587. [https://doi.org/10.1038/s41598-023-](https://doi.org/10.1038/s41598-023-28166-2) [28166-2](https://doi.org/10.1038/s41598-023-28166-2).

<span id="page-16-24"></span>Panlilio, J. M., I. T. Jones, M. C. Salanga, N. Aluru, and M. E. Hahn. 2021. "Developmental Exposure to Domoic Acid Disrupts Startle Response Behavior and Circuitry in Zebrafish." *Toxicological Sciences* 182, no. 2: 310–326. [https://doi.org/10.1093/toxsci/kfab066.](https://doi.org/10.1093/toxsci/kfab066)

<span id="page-16-11"></span>Perl, T. M., L. Bédard, T. Kosatsky, et al. 1990. "Amnesic Shellfish Poisoning: A New Clinical Syndrome Due to Domoic Acid." *Canada Diseases Weekly Report* 16, no. Suppl 1E: 7–8.

<span id="page-16-6"></span>Perl, T. M., L. Bédard, T. Kosatsky, J. C. Hockin, E. C. Todd, and R. S. Remis. 1990. "An Outbreak of Toxic Encephalopathy Caused by Eating Mussels Contaminated With Domoic Acid." *New England Journal of Medicine* 322, no. 25: 1775–1780.

<span id="page-16-7"></span>Perl, T. M., J. Teitelbaum, J. Hockin, and E. C. Todd. 1990. "Domoic Acid Toxicity. Panel Discussion: Definition of the Syndrome." *Canada Diseases Weekly Report* 16 Suppl 1E: 41–45.

<span id="page-16-26"></span>Perry, M. A., C. L. Ryan, and R. A. Tasker. 2009. "Effects of Low Dose Neonatal Domoic Acid Administration on Behavioural and Physiological Response to Mild Stress in Adult Rats." *Physiology & Behavior* 98, no. 1–2: 53–59. [https://doi.org/10.1016/j.physbeh.2009.](https://doi.org/10.1016/j.physbeh.2009.04.009) [04.009](https://doi.org/10.1016/j.physbeh.2009.04.009).

<span id="page-16-1"></span>Petroff, R., A. Hendrix, S. Shum, K. S. Grant, K. A. Lefebvre, and T. M. Burbacher. 2021. "Public Health Risks Associated With Chronic, Low-Level Domoic Acid Exposure: A Review of the Evidence." *Pharmacology & Therapeutics* 227: 107865. [https://doi.org/10.1016/j.pharmthera.2021.](https://doi.org/10.1016/j.pharmthera.2021.107865) [107865](https://doi.org/10.1016/j.pharmthera.2021.107865).

<span id="page-16-3"></span>Rodier, P. M. 1995. "Developing Brain as a Target of Toxicity." *Environmental Health Perspectives* 103: 73–76.

<span id="page-16-27"></span>Sasaki, T., H. Saito, Y. Hiradate, K. Hara, and K. Tanemura. 2021. "Behavioural Effects in Mice Orally Exposed to Domoic Acid or Ibotenic Acid Are Influenced by Developmental Stages and Sex Differences." *Biochemical and Biophysical Research Communications* 558: 175–182. [https://doi.org/10.1016/j.bbrc.2021.04.080.](https://doi.org/10.1016/j.bbrc.2021.04.080)

<span id="page-16-30"></span>Schaffer, R. M., M. N. Smith, and E. M. Faustman. 2017. "Developing the Regulatory Utility of the Exposome: Mapping Exposures for Risk Assessment Through Lifestage Exposome Snapshots (LEnS)." *Environmental Health Perspectives* 125, no. 8: 085003. [https://doi.org/](https://doi.org/10.1289/EHP1250) [10.1289/EHP1250](https://doi.org/10.1289/EHP1250).

<span id="page-16-22"></span>Shiotani, M., T. B. Cole, S. Hong, et al. 2017. "Neurobehavioral Assessment of Mice Following Repeated Oral Exposures to Domoic Acid During Prenatal Development." *Neurotoxicology and Teratology* 64: 8–19. <https://doi.org/10.1016/j.ntt.2017.09.002>.

<span id="page-16-2"></span>Shum, S., J. Jing, R. Petroff, et al. 2020. "Maternal-Fetal Disposition of Domoic Acid Following Repeated Oral Dosing During Pregnancy in Nonhuman Primate." *Toxicology and Applied Pharmacology* 398: 115027. <https://doi.org/10.1016/j.taap.2020.115027>.

<span id="page-16-28"></span>Simeone, C., D. Fauquier, J. Skidmore, et al. 2019. "Clinical Signs and Mortality of Non-Released Stranded California Sea Lions Housed in Display Facilities: The Suspected Role of Prior Exposure to Algal Toxins." *Veterinary Record* 185, no. 10: 304. [https://doi.org/10.1136/vr.](https://doi.org/10.1136/vr.105371) [105371.](https://doi.org/10.1136/vr.105371)

<span id="page-16-20"></span>Smith, J., J. A. Cram, M. P. Berndt, V. Hoard, D. Shultz, and A. C. Deming. 2023. "Quantifying the Linkages Between California Sea Lion (*Zalophus californianus*) Strandings and Particulate Domoic Acid Concentrations at Piers Across Southern California." *Frontiers in Marine Science* 10: 1278293. [https://doi.org/10.3389/fmars.2023.](https://doi.org/10.3389/fmars.2023.1278293) [1278293.](https://doi.org/10.3389/fmars.2023.1278293)

<span id="page-17-17"></span>Stuchal, L. D., L. M. Grattan, K. M. Portier, et al. 2020. "Dose-Response Assessment for Impaired Memory From Chronic Exposure to Domoic Acid Among Native American Consumers of Razor Clams." *Regulatory Toxicology and Pharmacology* 117: 104759. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.yrtph.2020.104759) [yrtph.2020.104759.](https://doi.org/10.1016/j.yrtph.2020.104759)

<span id="page-17-2"></span>Suzuki, C. A., and S. L. Hierlihy. 1993. "Renal Clearance of Domoic Acid in the Rat." *Food and Chemical Toxicology* 31, no. 10: 701–706.

<span id="page-17-8"></span>Tanemura, K., K. Igarashi, T. R. Matsugami, K. Aisaki, S. Kitajima, and J. Kanno. 2009. "Intrauterine Environment-Genome Interaction and children's Development (2): Brain Structure Impairment and Behavioral Disturbance Induced in Male Mice Offspring by a Single Intraperitoneal Administration of Domoic Acid (DA) to Their Dams." *Journal of Toxicological Sciences* 34: Sp279–Sp286. [https://doi.org/10.](https://doi.org/10.2131/jts.34.sp279) [2131/jts.34.sp279](https://doi.org/10.2131/jts.34.sp279).

<span id="page-17-14"></span>Tasker, R. A., M. A. Perry, T. A. Doucette, and C. L. Ryan. 2005. "NMDA Receptor Involvement in the Effects of Low Dose Domoic Acid in Neonatal Rats." *Amino Acids* 28, no. 2: 193–196. [https://doi.org/10.1007/](https://doi.org/10.1007/s00726-005-0167-z) [s00726-005-0167-z](https://doi.org/10.1007/s00726-005-0167-z).

<span id="page-17-0"></span>Teitelbaum, J. S., R. J. Zatorre, S. Carpenter, et al. 1990. "Neurologic Sequelae of Domoic Acid Intoxication Due to the Ingestion of Contaminated Mussels." *New England Journal of Medicine* 322, no. 25: 1781–1787. <https://doi.org/10.1056/NEJM199006213222505>.

<span id="page-17-13"></span>Thomsen, M. B., T. P. Lillethorup, S. Jakobsen, et al. 2016. "Neonatal Domoic Acid Alters In Vivo Binding of  $[(11)C]$ Yohimbine to  $\alpha(2)$ adrenoceptors in Adult Rat Brain." *Psychopharmacology* 233, no. 21–22: 3779–3785. <https://doi.org/10.1007/s00213-016-4416-5>.

<span id="page-17-10"></span>Tiedeken, J. A., and J. S. Ramsdell. 2007. "Embryonic Exposure to Domoic Acid Increases the Susceptibility of Zebrafish Larvae to the Chemical Convulsant Pentylenetetrazole." *Environmental Health Perspectives* 115, no. 11: 1547–1552. [https://doi.org/10.1289/ehp.10344.](https://doi.org/10.1289/ehp.10344)

<span id="page-17-11"></span>Tiedeken, J. A., and J. S. Ramsdell. 2009. "DDT Exposure of Zebrafish Embryos Enhances Seizure Susceptibility: Relationship to Fetal p,p'- DDE Burden and Domoic Acid Exposure of California Sea Lions." *Environmental Health Perspectives* 117, no. 1: 68–73. [https://doi.org/10.](https://doi.org/10.1289/ehp.11685) [1289/ehp.11685](https://doi.org/10.1289/ehp.11685).

<span id="page-17-12"></span>Tiedeken, J. A., J. S. Ramsdell, and A. F. Ramsdell. 2005. "Developmental Toxicity of Domoic Acid in Zebrafish (*Danio rerio*)." *Neurotoxicology and Teratology* 27, no. 5: 711–717. [https://doi.org/10.1016/j.ntt.2005.](https://doi.org/10.1016/j.ntt.2005.06.013) [06.013.](https://doi.org/10.1016/j.ntt.2005.06.013)

<span id="page-17-5"></span>Todd, E. C. 1990. "Chronology of the Toxic Mussels Outbreak." *Canada Diseases Weekly Report* 16, no. Suppl 1E: 3–4.

<span id="page-17-3"></span>Truelove, J., and F. Iverson. 1994. "Serum Domoic Acid Clearance and Clinical Observations in the Cynomolgus Monkey and Sprague-Dawley Rat Following a Single i.v. Dose." *Bulletin of Environmental Contamination and Toxicology* 52, no. 4: 479–486.

<span id="page-17-4"></span>Truelove, J., R. Mueller, O. Pulido, L. Martin, S. Fernie, and F. Iverson. 1997. "30-Day Oral Toxicity Study of Domoic Acid in Cynomolgus Monkeys: Lack of Overt Toxicity at Doses Approaching the Acute Toxic Dose." *Natural Toxins* 5, no. 3: 111–114. [https://doi.org/10.1002/1522-](https://doi.org/10.1002/1522-7189(1997)5:3%3C111::aid-nt5%3E3.0.co;2-6) [7189\(1997\)5:3<111::aid-nt5>3.0.co;2-6](https://doi.org/10.1002/1522-7189(1997)5:3%3C111::aid-nt5%3E3.0.co;2-6).

<span id="page-17-1"></span>USEPA. 2020. "Use of New Approach Methodologies to Derive Extrapolation Factors and Evaluate Developmental Neurotoxicity for Human Health Risk Assessment." Agency Issue Paper.

<span id="page-17-7"></span>USFDA. 2021. "Appendix 5: FDA and EPA Safety Levels and Regulations and Guidance." [https://www.google.com/url?sa=t&rct=j&q=&esrc=](https://www.google.com/url?sa=t&rct=j&q=&esrc=s&source=web&cd=&cad=rja&uact=8&ved=2ahUKEwipi7jf_6zrAhWFtp4KHdQOCTMQFjAAegQIBRAB&url=https%3A%2F%2Fwww.fda.gov%2Fmedia%2F80400%2Fdownload&usg=AOvVaw3gZChY_lOemec-gay1fSuD) [s&source=web&cd=&cad=rja&uact=8&ved=2ahUKEwipi7jf\\_6zrAh](https://www.google.com/url?sa=t&rct=j&q=&esrc=s&source=web&cd=&cad=rja&uact=8&ved=2ahUKEwipi7jf_6zrAhWFtp4KHdQOCTMQFjAAegQIBRAB&url=https%3A%2F%2Fwww.fda.gov%2Fmedia%2F80400%2Fdownload&usg=AOvVaw3gZChY_lOemec-gay1fSuD) [WFtp4KHdQOCTMQFjAAegQIBRAB&url=https%3A%2F%2Fwww.](https://www.google.com/url?sa=t&rct=j&q=&esrc=s&source=web&cd=&cad=rja&uact=8&ved=2ahUKEwipi7jf_6zrAhWFtp4KHdQOCTMQFjAAegQIBRAB&url=https%3A%2F%2Fwww.fda.gov%2Fmedia%2F80400%2Fdownload&usg=AOvVaw3gZChY_lOemec-gay1fSuD) [fda.gov%2Fmedia%2F80400%2Fdownload&usg=AOvVaw3gZChY\\_](https://www.google.com/url?sa=t&rct=j&q=&esrc=s&source=web&cd=&cad=rja&uact=8&ved=2ahUKEwipi7jf_6zrAhWFtp4KHdQOCTMQFjAAegQIBRAB&url=https%3A%2F%2Fwww.fda.gov%2Fmedia%2F80400%2Fdownload&usg=AOvVaw3gZChY_lOemec-gay1fSuD) [lOemec-gay1fSuD](https://www.google.com/url?sa=t&rct=j&q=&esrc=s&source=web&cd=&cad=rja&uact=8&ved=2ahUKEwipi7jf_6zrAhWFtp4KHdQOCTMQFjAAegQIBRAB&url=https%3A%2F%2Fwww.fda.gov%2Fmedia%2F80400%2Fdownload&usg=AOvVaw3gZChY_lOemec-gay1fSuD).

<span id="page-17-18"></span>WADOH. 2023. "Domoic Acid in Razor Clams." Interim Health Advisory on Eating Razor Clams, Washington State Department of Health. [https://doh.wa.gov/community-and-environment/shellfish/](https://doh.wa.gov/community-and-environment/shellfish/recreational-shellfish/illnesses/biotoxins/domoic-acid-razor-clams) [recreational-shellfish/illnesses/biotoxins/domoic-acid-razor-clams](https://doh.wa.gov/community-and-environment/shellfish/recreational-shellfish/illnesses/biotoxins/domoic-acid-razor-clams).

<span id="page-17-16"></span>Wang, G. J., L. C. Schmued, A. M. Andrews, A. C. Scallet, W. Slikker Jr., and Z. Binienda. 2000. "Systemic Administration of Domoic Acid-Induced Spinal Cord Lesions in Neonatal Rats." *Journal of Spinal Cord Medicine* 23, no. 1: 31–39. [https://doi.org/10.1080/10790268.2000.](https://doi.org/10.1080/10790268.2000.11753506) [11753506.](https://doi.org/10.1080/10790268.2000.11753506)

<span id="page-17-6"></span>Wekell, J. C., J. Hurst, and K. A. Lefebvre. 2004. "The Origin of the Regulatory Limits for PSP and ASP Toxins in Shellfish." *Journal of Shellfish Research* 23, no. 3: 927–930.

<span id="page-17-15"></span>Xi, D., Y. G. Peng, and J. S. Ramsdell. 1997. "Domoic Acid Is a Potent Neurotoxin to Neonatal Rats." *Natural Toxins* 5, no. 2: 74–79. [https://](https://doi.org/10.1002/(sici)(1997)5:2%3C74::Aid-nt4%3E3.0.Co;2-i) [doi.org/10.1002/\(sici\)\(1997\)5:2<74::Aid-nt4>3.0.Co;2-i.](https://doi.org/10.1002/(sici)(1997)5:2%3C74::Aid-nt4%3E3.0.Co;2-i)

<span id="page-17-9"></span>Zuloaga, D. G., G. P. Lahvis, B. Mills, H. L. Pearce, J. Turner, and J. Raber. 2016. "Fetal Domoic Acid Exposure Affects Lateral Amygdala Neurons, Diminishes Social Investigation and Alters Sensory-Motor Gating." *Neurotoxicology* 53: 132–140. [https://doi.org/10.1016/j.neuro.](https://doi.org/10.1016/j.neuro.2016.01.007) [2016.01.007](https://doi.org/10.1016/j.neuro.2016.01.007).