

Review

Neuronal Cell Rearrangement During Aging: Antioxidant Compounds as a Potential Therapeutic Approach

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Abstract: Aging is a natural process that leads to time-related changes and a decrease in cognitive abilities, executive functions, and attention. In neuronal aging, brain cells struggle to respond to oxidative stress. The structure, function, and survival of neurons can be mediated by different pathways that are sensitive to oxidative stress and age-related low-energy states. Mitochondrial impairment is one of the most noticeable signs of brain aging. Damaged mitochondria are thought to be one of the main causes that feed the inflammation related to aging. Also, protein turnover is involved in age-related impairments. The brain, due to its high oxygen usage, is particularly susceptible to oxidative damage. This review explores the mechanisms underlying neuronal cell rearrangement during aging, focusing on morphological changes that contribute to cognitive decline and increased susceptibility to neurodegenerative diseases. Potential therapeutic approaches are discussed, including the use of antioxidants (e.g., Vitamin C, Vitamin E, glutathione, carotenoids, quercetin, resveratrol, and curcumin) to mitigate oxidative damage, enhance mitochondrial function, and maintain protein homeostasis. This comprehensive overview aims to provide insights into the cellular and molecular processes of neuronal aging and highlight promising therapeutic avenues to counteract age-related neuronal deterioration.

Keywords: neurodegeneration; brain; oxidative damage; antioxidants; ROS; mitochondrial dysfunction; organelles

1. Introduction

Aging is a natural process that affects all organs, including the nervous system. Neurons, compared to other cell types, are more likely to change in function and metabolism due to insults, diseases, and/or neurodegenerative disorders, especially those that are age-related. In neuronal aging, brain cells find it difficult to respond to oxidative stress due to the accumulation of aggregated proteins in the cytoplasm and mitochondria [\[1\]](#page-10-0). Neurons tend to shrink and morphologically change the structure of their dendrites over time. According to studies conducted by Buell and Coleman, dendritic growth remains possible even in the aging human brain [\[2\]](#page-10-1).

Aging is related to decreased executive function, cognitive abilities, and attention impairment [\[3\]](#page-10-2). Additionally, aging leads to a reduction in dopamine, serotonin (5-HT), and brain-derived neurotrophic factor (BDNF) levels, which regulate neuronal survival. Dopaminergic neurotransmitter systems are persistently adjusted during aging [\[4\]](#page-10-3). Compromised 5-HT and BDNF are central to the pathogenesis of neurodegenerative diseases such as Alzheimer's disease [\[5\]](#page-11-0). On the other hand, there is an increment in monoamine oxidase, which leads to the catabolism of the neurotransmitters releasing hydrogen peroxide (H_2O_2) which can trigger the formation of hydroxyl radicals O_2^- · [\[6,](#page-11-1)[7\]](#page-11-2). Parameters such as regional cerebral blood flow and regional cerebral metabolic rates for glucose

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and oxygen are indicators of normal brain activity, and they tend to decrease remarkably during aging [\[8\]](#page-11-3). Moreover, many neurotrophic factors and cytokines may contribute to neurogenesis after injury [\[9](#page-11-4)[,10\]](#page-11-5). Dietary reductions, resulting in increased production of protein chaperones in brain cells, along with the use of folic acid, can improve cognitive abilities in mouse models of AD or PD [\[11\]](#page-11-6). Additionally, the use of antioxidants, such as saffron, can have neuroprotective effects by modulating neurotransmitters, reducing inflammation, oxidative stress, and apoptosis [\[12\]](#page-11-7).

The first part of this review aims to provide insights into the cellular and molecular processes of neuronal aging, while the second part discusses promising therapeutic interventions based on antioxidant origins to counteract age-related neuronal deterioration.

2. Materials and Methods

The literature reviewed included preclinical and clinical studies. The in vitro and in vivo studies were selected based on the following points of view: neurological aging, mitochondrial dysfunction, oxidative damage, and antioxidants. Literature search was run by Google Scholar, PubMed, and free PMC articles using results up to August 2024.

The research queries included the following terms: brain aging, neuronal cell rearrangement, and proteostasis, combined with terms to determine the outcomes of interest such as neuroprotection, therapeutical approaches in aging, and antioxidants effects.

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From the literature search, we obtained more than 16,000 reviews, from which 221 studies were identified and analyzed, and 179 were taken in consideration as suitable for this article review.

3. Age-Related Alterations

3.1. Age-Related Changes in the Brain

Aging goes along with progressive neuron loss and cognitive and behavioral decline [\[13,](#page-11-8)[14\]](#page-11-9). The cognitive deficits include not only memory loss but also decreased motor performance [\[15,](#page-11-10)[16\]](#page-11-11). The aging process is related to impairment of the central nervous system [\[17\]](#page-11-12). Brain aging is also associated with a reduction in neurophysiological functions and neuro-inflammation, neuron cell shrinking, dendritic degradation, demyelination, and microglial activation [\[18\]](#page-11-13). One of the main characteristics of an aging brain is neuroinflammation induced by the glial cells [\[19\]](#page-11-14). Cellular senescence can play a crucial role in the aging brain [\[20\]](#page-11-15). Studies in recent years, conducted on genetic mouse models, have revealed the effects of anti-inflammatory agents such as ibuprofen, which defend against cognitive decline induced by chronic inflammation by reducing markers of senescence in neurons [\[21\]](#page-11-16). Normally, starting from mid-life, the volume and the cortical thickness begin to decline with the passing of years. What we notice more frequently is that injuries such as white matter hyperintensities start showing with age [\[22\]](#page-11-17). In the first stages, the aging mechanisms include mostly the cells, but then, slowly, tissue and organ alterations begin to happen [\[23\]](#page-11-18). The age-dependent brain structure differences in the frontal lobe and medial temporal regions are related to changes in cognitive functions of the same regions. What is observed is also a relation between degeneration of age-related white matter segments and cognitive performance [\[24\]](#page-11-19). To achieve a particular target in an aging brain, the adaptive brain uses a normal mechanism called scaffolding, through which different and replacement neural circuits are used [\[25\]](#page-11-20). Neurofibrillary tangles (NFTs) and senile plaques (SPs) are found in physiological conditions during aging but also in pathological conditions such as Alzheimer's disease (AD). The lesions tend to worsen with age and progression of the disease [\[26\]](#page-11-21). Neuronal loss might be the cause of cognitive disabilities and structural changes. In an aged brain, the most commonly reported are accumulation of $A\beta$ protein and α -synuclein [\[27\]](#page-11-22).

During aging, neurons experience morphological changes such as dendritic length, decrease in spine numbers, and as a result changes in synaptic densities. All the studies suggest that neuronal dysfunction in physiological age-related cases is due to changes

inside the cortex resulting in myelin dystrophy and variations in synaptic transmission [\[28\]](#page-11-23). Intracellular accumulation of Aβ protein leads to oxidative stress and mitochondrial dysfunction, which may lead to neuronal death [\[29\]](#page-11-24). Structure, function, and survival of neurons can be mediated by different pathways that are all sensitive to oxidative stress and age-related low-energy states [\[26\]](#page-11-21).

Heat shock proteins (HSPs), also called stress proteins, are cellular components, acting as molecular chaperones, whose main function is to control the folding of proteins; additionally, the HSPs are able to prevent the cellular aggregation of amyloid peptide and hyperphosphorylated tau in neurodegenerative diseases. Stress proteins participate in protein synthesis and transport, and they are activated in stressful situations. Being highly expressed in the nervous system, these proteins protect the neurons and glia [\[30\]](#page-11-25). They also regulate the apoptosis-controlling inflammatory pathway while secreting inflammatory mediators, assist in the degradation of abnormal proteins, and modulate antigen presentation. A collapse or malfunction of these proteins contributes to the generation of these insoluble hyperphosphorylated inclusions [\[31\]](#page-11-26).

3.2. Mitochondrial Dysfunction in Brain Aging

Mitochondrial dysfunction is one of the most remarkable signs of brain aging [\[32\]](#page-11-27). According to recent studies, aging is the outcome of free radicals originally generated by cellular processes that are harmful, dangerous, and might lead to impairment [\[33\]](#page-11-28). Mitochondria are organelles exerting a crucial role in energy production, controlling the cell cycle from growth to apoptosis, regulating intracellular calcium, producing reactive oxygen species (ROS), and so on [\[34\]](#page-11-29). As we age, their efficiency in energy production declines, leading to reduced cellular function. Additionally, aging mitochondria generate more ROS, which can damage cellular components, including mitochondrial DNA (mtDNA). Over time, mutations accumulate in mtDNA, further impairing mitochondrial function and contributing to cellular aging. This dysfunction can also trigger cellular senescence, where cells stop dividing and secrete inflammatory factors, exacerbating tissue aging and age-related diseases. Also, dysfunctional mitochondria in many age-related diseases such as Alzheimer's, Parkinson's, and Huntington's diseases have been reported, as well as in inflammatory conditions [\[35\]](#page-11-30).

Specifically, one of the many main theories of aging indicates that oxidative stress, produced during the phosphorylation of mitochondrial macromolecules such as proteins, lipids, and mtDNA, is the one responsible for aging. We observe deletions of mtDNA more frequently in aged generations [\[36\]](#page-12-0). According to some studies, mtDNA deletions are more probable to happen during the adjustment of the injured mtDNA than during its replication [\[37\]](#page-12-1). It seems that ROS are the main factor responsible for mtDNA mutations during aging [\[38\]](#page-12-2). mtDNA is responsible for the encoding of crucial elements of oxidative phosphorylation and the synthesis of proteins [\[39\]](#page-12-3). Therefore, oxidative damage-induced mtDNA mutations that damage the function of the respiratory mitochondrial chain will cause the formation of more and more ROS, resulting in cell death [\[40](#page-12-4)[,41\]](#page-12-5). Aging as a physiological process reduces the efficiency of oxidative phosphorylation, resulting in increased ROS production and altered ATP production. The great amount of ROS produced oxidizes more macromolecules eventually; consequently, we observe a worsening of mitochondrial function, generating more ROS in a vicious cycle [\[42\]](#page-12-6). Mitochondria play a crucial role in controlling the apoptosis activating the permeability transition pore. This activation is intensified in the brain of aged mice [\[43\]](#page-12-7). Aging is also connected to a reduced ability to produce ATP by oxidative phosphorylation in mammalian brains [\[44\]](#page-12-8). What we observe is a decrease in ATP production capacity with time [\[45\]](#page-12-9). At this point, cells should be either able to function with less energy or they should find a way to produce energy through non-oxidative metabolism. All this will affect other mechanisms, metabolisms, and signaling pathways [\[46\]](#page-12-10).

The old mitochondria reduce the synthesis of mitochondrial proteins, lower the activity of oxidative enzymes, and minimize the mitochondrial mass, resulting in a decrease in the production of ATP [\[47\]](#page-12-11). The functions of ATP-dependent proteases, such as AFG3L2, AFG3L2/SPG7, LONP1, and YME1L, which regulate functions such as ribosome assembly, transcription and translation, adaptation to hypoxia, protein import, lipid trafficking, and mitochondrial dynamics, have been shown to lessen with aging, resulting in the inability to maintain a healthy and functional mitochondrion [\[48\]](#page-12-12). YME1L and OMA1 are proteases located in the inner membrane of the mitochondria responsible for the regulation of mitochondrial morphology [\[49\]](#page-12-13). They can degrade one another based on insults, leading to depolarization of the mitochondrial membrane and consumption of ATP; as a result, they can balance mitochondrial dynamics [\[50\]](#page-12-14). LONP1, on the other hand, supports protein folding while arranging the proteolytic activity [\[51\]](#page-12-15).

During aging, senescent cells tend to accumulate, causing a functional decline leading to morbidity and mortality [\[52\]](#page-12-16). Mitochondrial dysfunction in senescent cells can be considered as both a reduction in respiratory function and a decreased mitochondrial membrane potential (MMP). We can benefit from having the MMP decrease at a steady rate because the electron transport chain (ETC) passes in a more oxidized state, reducing ROS production, but also the ETC can become non-functional [\[53\]](#page-12-17). In elderly people, the respiratory chain complexes form super complexes, which, if not stable, lead to the formation of more ROS [\[54,](#page-12-18)[55\]](#page-12-19).

To maintain MMP and to produce ATP, it is essential for mitochondria to find the balance between *NAD*⁺ and NADH, as this ratio is very important for mitochondrial metabolism. The levels of NAD^+ within the cells are regulated by a balance between its production and its degradation [\[56\]](#page-12-20). *NAD*⁺ consumption during stress and aging and the disruption in *NAD*+/NADH will lead to mitochondrial dysfunction, mitochondrial membrane depolarization, and altered mitochondrial permeability transition, which are connected to cell death [\[57\]](#page-12-21).

Inflammation is a physiological process that arises after an insult with the main aim of restoring cells and damaged tissues after eliminating the danger. After this phase, the inflammatory process should come to an end; if not, the inflammation might become chronic. Non-functional mitochondria are thought to be one of the main causes that nourish inflammation related to aging. Respiratory dysfunction and oxidative stress are the mechanisms through which the mitochondria get involved in the aging process [\[58\]](#page-12-22). To avoid the release of the content of the damaged or aged mitochondria in the cytoplasm or extracellular space, it is very important to sequester the defective mitochondria in a double-membraned autophagosome via mitophagy, with the consequence of having a minor risk of developing chronic inflammation [\[59\]](#page-12-23). Impaired age-related mitochondria can also unbalance calcium buffering, leading to a variation in the concentration of free calcium ions in cells [\[60\]](#page-12-24), causing obstacles with synaptic activity since Ca^{2+} participates in the transmission of the signal. Ca^{2+} homeostasis can be regulated by protein channels, which we find in both inner and outer mitochondrial membranes and by the endoplasmic reticulum [\[61\]](#page-12-25). During aging, neurons find it hard to maintain the required energy level, and this influences cytoplasmic Ca^{2+} levels. Therefore, we find Ca^{2+} signaling to be disturbed and associated with a debilitating impairment [\[62\]](#page-12-26). Ca^{2+} increments in the cytosol force the mitochondria to rapidly uptake it to avoid overloading the cytosol. This, on the other hand, can result in mitochondrial overcharge, ROS generation, and ATP depletion [\[63\]](#page-12-27).

3.3. Morphological Rearrangement in Brain Aging

Different studies have shown that interruptions in brain synaptic circuitry can contribute to cognitive decline [\[64\]](#page-12-28). During physiological aging, there is a decline in the white matter in the cerebral hemispheres and structural irregularities in myelin [\[65\]](#page-12-29). With age, the structure of myelin undergoes alterations, and the axons start showing evidence of degeneration, resulting in the loss of nerve fibers. Additionally, according to a study by Sandell conducted on monkeys, the number of myelinated nerve fibers tends to decrease over time compared to young monkeys [\[66\]](#page-13-0). The structure of the myelin sheath undergoes

age-related changes and modifications, leading to defective and weak conduction speed of neuronal signals along the axons [\[67\]](#page-13-1).

Protein turnover in the brain is a process that undergoes age-related impairments. In addition, altered proteolysis of various cell types is believed to be related to cytotoxicity. Protein turnover is thought to be related to protein oxidation and aggregation of different forms, such as Lewy bodies and lipofuscin granules [\[68\]](#page-13-2). Accumulation of lipofuscin granules is found in both senescence and various age-related and neurodegenerative diseases such as Alzheimer's disease. Lipofuscin can accumulate because cells find it hard to exocytose it [\[69\]](#page-13-3). Neurons that contain a great quantity of lipofuscin under physiological conditions are less affected by degenerative alterations [\[70\]](#page-13-4). Forming lipofuscin is understood as a form of neuronal adaptation, and those neurons that lack this capability are more vulnerable to degeneration [\[70–](#page-13-4)[72\]](#page-13-5). Nevertheless, evidence shows that accumulated lipofuscin can be dangerous, interfering with normal cellular functioning because its deposits contain toxic compounds [\[73\]](#page-13-6). Therefore, lipofuscin elimination can be a possible target in anti-aging therapies [\[74\]](#page-13-7). The study conducted on rats while being treated with melatonin or coenzyme Q10 for 4 weeks revealed a reduction in lipofuscin [\[75\]](#page-13-8). The presence of lipofuscin increases with age [\[76\]](#page-13-9).

During aging, carbohydrate-rich inclusions in the neuronal soma are found, also called glycogen inclusions or polyglucosan bodies, which can also contain proteins. These inclusions are both found in aging brains and in neurodegenerative diseases and might play a powerful role in cell cycles. However, it is not yet clear whether these carbohydrate bodies are the cause or the result of brain senescence [\[77\]](#page-13-10).

Axons tend to increase in diameter, becoming larger and thicker with age. Evidence has demonstrated that mitochondria generate more oxidative stress in axons, leading to a vulnerability to further injury because the mitochondria themselves suffer compromised functional integrity [\[78\]](#page-13-11). The selective destruction of axons is a process that occurs due to aging, and it is a characteristic of neurodegeneration because it induces neuronal deterioration [\[79\]](#page-13-12). The first process occurring is axonal degeneration; after that, neuronal death occurs [\[80\]](#page-13-13). Myelin degradation is subsequent to the breakdown of the axonal cytoskeleton [\[81\]](#page-13-14). This self-destructive process, with its molecular mechanism, can be an important target for the strategies implicated in neurodegenerative disorders. According to Lasiene, internode lengths undergo a significant reduction with age [\[82\]](#page-13-15). Axonal active destruction relies on the activation of the mitochondrial permeability transition pore (mPTP) between both mitochondrial membranes, the inner and the outer ones [\[83\]](#page-13-16). This activation, while provoking the mitochondrial permeability transition, increases the ROS in axons contributing to axonal degeneration [\[84\]](#page-13-17).

3.4. Protein Degradation in Brain Aging

Protein degradation is a crucial process that helps break down and remove the aggregated or the misfolded ones [\[85\]](#page-13-18). Protein homeostasis, also called proteostasis, promotes healthy protein synthesis, folding, and degradation. In the meantime, biological molecules that can modify the conformation or the structure of proteins can be used as a potential target to improve the symptoms of some neurodegenerative disorders [\[86\]](#page-13-19). Additionally, small molecules considered to be pharmacological chaperones can stimulate the refolding of impaired proteins [\[87\]](#page-13-20). Proteolytic enzymes, such as peptidases/proteases, are found in different intracellular and extracellular compartments, and they can degrade proteins or in some cases form a fusion with the pharmacological chaperones to prevent non-specific aggregation [\[49\]](#page-12-13). The synthesis of the chaperones is a stress-induced process that is damaged and weakened during longevity [\[88\]](#page-13-21). Aging is connected to the misregulation of protein maintenance [\[89\]](#page-13-22) and to the alteration of proteostasis [\[90\]](#page-13-23). The mechanism of proteostasis includes processes that lead to the balance of accurately folded proteins in their three-dimensional structure and processes where the proteasome or lysosome degrades the proteins [\[91\]](#page-13-24). Proteolytic systems, autophagy–lysosomal and ubiquitin–proteasome more specifically, are also age-dependent systems [\[92\]](#page-13-25). The ubiquitin–proteasome system

is the one that regulates proteostasis in the nucleus [\[49\]](#page-12-13). Additionally, the ubiquitin proteases also regulate stem cell functions since they tend to decrease with the passing of the years [\[93\]](#page-13-26). Proteases can also operate as signaling molecules since we find them in both intracellular compartments and extracellular areas [\[49\]](#page-12-13). Moreover, they can have an impact on cell senescence since this last one is a causal factor of aging and can guide cellular reprogramming [\[49\]](#page-12-13). Membrane proteases along with extracellular proteases can regulate extracellular proteolysis [\[94\]](#page-13-27). Extracellular proteases themselves eliminate misfolded proteins because the unfolded protein response can, in an indirect way, modulate extracellular proteostasis [\[95\]](#page-14-0). Secreted extracellular chaperones, which act as intermediaries for misfolded proteins, help maintain proteostasis [\[96\]](#page-14-1).

3.5. Oxidative Stress in Brain Aging

The brain, since it uses a great quantity of oxygen for energy production, is one of the organs that is at greater risk of oxidative damage. "Inflammaging" is a term used to describe the inflammation process generated by ROS and how this might affect the nervous system during aging [\[97\]](#page-14-2). "Inflammaging", compared to inflammation, is a situation where the inflammation happens without having an apparent pathogen [\[98\]](#page-14-3). The main focus is not to eliminate the inflammation but to support the equilibrium between inflammaging and processes that contrast it [\[99\]](#page-14-4). One of the first steps of aging is an increased number of senescent cells [\[100\]](#page-14-5) that accumulate in tissues while releasing pro-inflammatory cytokines and proteases [\[101\]](#page-14-6). Among the many theories of aging, the free radical theory is one of the most studied [\[102\]](#page-14-7). According to this theory, elderly people have excessive amounts of oxidized products compared to younger generations. Therefore, using antioxidants in this group is essential for increasing lifespan [\[103\]](#page-14-8) (Figure [1\)](#page-5-0). Lifespan is inversely proportional to the concentration of ROS in the mitochondria [\[104\]](#page-14-9). ROS are produced in normal cellular metabolism. The three most crucial ROS are the superoxide anion (O_2^-) $\frac{1}{2}$.), the hydroxyl radical (OH·), and hydrogen peroxide (H_2O_2) [\[105\]](#page-14-10) (Figure [1\)](#page-5-0). The hydroxyl radical is the most reactive species, and it can be the cause of damage to all macromolecules such as lipids, proteins, carbohydrates, and nucleic acids. Defects or deficiencies of enzymes that are capable of scavenging free radicals such as super oxide dismutases can be correlated to the accumulation of free radicals [\[106\]](#page-14-11). Organic compounds are thermodynamically unbalanced in oxygen surrounding the generation of oxygen radicals, which might be the cause of damaging oxidative reactions [\[107\]](#page-14-12).

OXIDANTS > ANTIOXIDANTS = OXIDATIVE STRESS

Figure 1. Balance and imbalance between ROS and antioxidants. Figure 1. Balance and imbalance between ROS and antioxidants.
 Figure 1. Balance and imbalance between ROS and antioxidants.

4. Therapeutic Approaches to Counteract Brain Aging

There are several therapeutic approaches aimed at counteracting brain aging and neuronal cell rearrangement. As we mentioned above, mitochondrial dysfunction is closely associated with brain aging due to oxidative damage. Strategies to protect mitochondria include the use of antioxidants to reduce oxidative stress and the development of drugs that enhance mitochondrial function. Antioxidants play a crucial role in compensating for the negative effects of the ROS and scavenge free radicals while multiplying the antioxidant defense [\[108\]](#page-14-13). The antioxidant properties of different compounds are mainly supplied by phenolic acids, phenolic diterpenes, flavonoids, and volatile oils [\[109](#page-14-14)[,110\]](#page-14-15). By their use, what we obtain is a dynamic but fine-tuned redox equilibrium. Additionally, these molecules can also prevent or delay the oxidation of substances that cause the formation of ROS and interrupt propagation of autoxidation [\[111\]](#page-14-16). The antioxidant action of these compounds is probably connected to their capability to adjust the activity of the redox enzymes while interconnecting with redox signaling pathways [\[112\]](#page-14-17). The activity of the antioxidants in vivo is multiplied because those that cannot be absorbed in the small intestine can be split into smaller molecules, not only increasing the possibility of better absorption but also resulting in many different potential targets [\[113\]](#page-14-18). Many dietary antioxidants get involved in different redox regulations, such as MAPK-mediated pathways that are responsible for controlling the expression of many genes. Polyphenols, while activating Nrf2, can enhance the activity of antioxidant enzymes like CAT, GPx, glutathione reductase, and paraoxonases [\[114\]](#page-14-19). Through these redox-balanced mechanisms, we obtain lifespanincreasing effects [\[115\]](#page-14-20). Another mechanism through which antioxidants can manifest their effects is by interacting with specific proteins which play a crucial role in intracellular signaling [\[116\]](#page-14-21). Playing a role as a reducing agent and interfering with metal ion chelation while eliminating free radicals are ways through which antioxidants can exert their effect [\[117\]](#page-14-22). When the protection that comes from endogenous antioxidants is not enough, we might rely on the help that we find from diet- or drug-derived antioxidants [\[118\]](#page-14-23).

In this review, we will focus on emerging evidence about Vitamin C, Vitamin E, glutathione, carotenoids, quercetin, resveratrol, and curcumin intake during aging.

Vitamin C, also known as ascorbic acid, plays an antioxidant role by scavenging oxygen free radicals [\[119\]](#page-14-24) (Table [1\)](#page-9-0). The concentration of ascorbic acid in the body is considered to be the highest compared to other types of vitamins [\[120\]](#page-14-25). An absence of Vitamin C in the brain of mouse models, affected by Alzheimer's disease, indicates that there is an imbalance in the redox processes stimulating the generation of $β$ -amyloid [\[121\]](#page-14-26). According to the study conducted by El-Gendy et al., Vitamin C might be able to reduce the oxidative damage caused by imidacloprid. The protective effect of the pre-treatment is better compared to the post-treatment [\[122\]](#page-14-27) (Table [1\)](#page-9-0). One of the influencing factors of healthy aging is the daily intake of Vitamin C, reducing the risk of mortality [\[123\]](#page-15-0) (Table [1\)](#page-9-0). As specified by Brubacher et al., a lower mean Vitamin C status appears in a group of older people compared to a group of young ones after an intake of 60 mg/day. Thus, the demand for Vitamin C in the older generations is more elevated [\[124\]](#page-15-1). On the other hand, the study by Murata and colleagues compared healthy and young adults with older adult groups, who were chronically ill and hospitalized. The group of elderly people following long-term supplements showed decreased Vitamin C status [\[125\]](#page-15-2).

Vitamin C is not the only one to have a lower concentration in AD patients; Vitamin E also shows reduced concentration in contrast to subjects with normal cognitive functions [\[126\]](#page-15-3). Vitamin E, also known as α -tocopherol, is concentrated mostly in the interior part, the hydrophobic one, of the cell membrane and plays a role as a shelter in case of membrane injury. Additionally, Vitamin E slows down and interferes with the production of ROS and stimulates the apoptosis of cancer cells [\[105\]](#page-14-10) (Table [1\)](#page-9-0). Vitamin E is an antioxidant that acts also as a neuroprotective agent and can be used to prevent neurodegenerative diseases [\[127](#page-15-4)[,128\]](#page-15-5) (Table [1\)](#page-9-0). Different studies revealed that the combination of Vitamin C and Vitamin E prevents and blocks age-associated impairments of cognitive abilities [\[129\]](#page-15-6). In the meantime, the studies conducted on old rats, while combining both vitamins, show

a decrease in the oxidative stress and protection of the hypothalamus and cortex in case of cold exposure-induced oxidative stress resulting in better protection against oxidative stress [\[130–](#page-15-7)[132\]](#page-15-8).

Glutathione (GSH) is the most soluble antioxidant. Glutathione purifies hydrogen peroxide and preserves the membrane lipids from oxidative stress. In addition, GSH is able to keep the cells safe while working alongside proapoptotic and antiapoptotic signaling pathways [\[133\]](#page-15-9) (Table [1\)](#page-9-0). GSH is a tripeptide containing cysteine, glutamic acid, and glycine, found mostly in the glial cells, and it is necessary for cell proliferation and ROS neutralization [\[134\]](#page-15-10) (Table [1\)](#page-9-0). Additionally, GSH is related to the impediment of mitochondrial damage and cell death. GSH might function as a signaling molecule in the CNS as well [\[135\]](#page-15-11). During aging and neurodegenerative diseases, what we observe is a debilitation of GSH functions, and this is due to a loss of neurons [\[136\]](#page-15-12). At this elderly stage of life, GSH plays a very important role in defending neurons in case of damage caused by ROS [\[137\]](#page-15-13) (Table [1\)](#page-9-0). An age-dependent reduction in GSH levels can be the cause of the outbreak of age-dependent neurodegenerative disorders [\[138,](#page-15-14)[139\]](#page-15-15).

Carotenoids such as β-carotene tend to react with oxygen radicals. The antioxidant effect of carotenoids is mainly demonstrated in low oxygen partial pressure [\[140\]](#page-15-16). In patients suffering from neurodegenerative diseases, especially those affected by AD, the blood concentrations of carotenoids were identified to be low [\[141\]](#page-15-17). One of the mechanisms through which β-carotene performs its antioxidant property is the interaction with biologi-cal membranes [\[142\]](#page-15-18) (Table [1\)](#page-9-0). Studies demonstrate that (Z)-isomers of β-carotene have antioxidant activity in vitro [\[143\]](#page-15-19). The β -carotene activity should also be seen from another point of view. As we all know, a telomere's length shortens with age, and that might result in apoptosis and senescence. It looks like, according to the study by Boccardi et al., that β-carotene can adjust the telomerase activity while aging, protecting the telomers from oxidative stress $[141]$ (Table [1\)](#page-9-0). β-carotene is an antioxidant with a potential association to the maintenance of cognitive functions, and apparently it works better in synergy with other supplementations such as Vitamin E, Vitamin C, zinc, and selenium [\[144\]](#page-15-20). This was proved by a study conducted on men treated with 50 mg of β-carotene for 18 years. Their cognitive memory was better compared to the men treated with a placebo. The benefits can be attributed to the early age of starting to consume it and/or the long period of using it [\[145\]](#page-15-21). β-carotene along with Vitamin E can prevent lipid peroxidation [\[146\]](#page-15-22). In animal models, increased levels of β-carotene are associated with higher levels of the neurotrophin brain-derived neurotrophic factor (BDNF), which has a crucial role in synaptic plasticity [\[147\]](#page-15-23).

Quercetin is a flavonoid that is able to contrast the age-related increase in oxidative stress in the mitochondria, ameliorating the mitochondrial dynamics [\[148\]](#page-15-24) (Table [1\)](#page-9-0). It is also capable of slowing down cellular aging, boosting cell proliferation [\[149\]](#page-15-25) (Table [1\)](#page-9-0). Quercetin is not only an antioxidant, but it is involved in antiapoptotic and anti-inflammatory processes as well, so it can be thought of as a possible nutraceutical to be used in agerelated disorders [\[150\]](#page-15-26). The accumulation of free radicals, while increasing oxidative stress, contributes to the production and release of pro-inflammatory cytokines such as interleukin 1β, tumor necrosis factor, and interferon γ [\[151\]](#page-15-27). Sirtuin 1 (SIRT1), which is a member of the NAD+-dependent deacetylase enzyme family, while adjusting the expression of pro-inflammatory cytokines, can diminish the degeneration of dopaminergic neurons [\[152\]](#page-16-0). Therefore, SIRT1 may be a promising therapeutical agent against agerelated diseases [\[153\]](#page-16-1). Quercetin stimulates the activity of SIRT1, modulating the functions of inflammatory cytokines and therefore reducing ROS production and neuronal cell death [\[150,](#page-15-26)[154\]](#page-16-2) (Table [1\)](#page-9-0). In the study conducted on aged mice, quercetin dampened neuro-inflammation through the SIRT1/NLRP3 pathway [\[155\]](#page-16-3).

Several studies, based on cognitive functions revealed that the polyphenol resveratrol can protect cells from neuronal damage [\[156\]](#page-16-4). According to studies by Caldeira et al., resveratrol's antioxidant mechanisms change depending on how old the cell is. The study by Caldeira conducted on the human mononuclear cells of two different kinds of donors,

middle-aged and elderly donors, reflects how there is an increment of the activity of the superoxide dismutase enzyme in the cells of the elderly group after being exposed to resveratrol, the group where the highest levels of ROS production were also found [\[157\]](#page-16-5). Mitochondrial function can be preserved using resveratrol [\[158\]](#page-16-6) (Table [1\)](#page-9-0). The brain can benefit from resveratrol treatments because they may result in improved memory while reducing oxidative damage [\[159,](#page-16-7)[160\]](#page-16-8) (Table [1\)](#page-9-0). Resveratrol's main functions, based on antiaging mechanisms, are enhancing oxidative stress while improving mitochondrial function and regulating cell apoptosis; therefore, it could be a bioactive compound to be used to target age-dependent diseases [\[161\]](#page-16-9). Resveratrol is considered a powerful antioxidant because it can not only clean free radicals, but it can also influence the increase in the activity of other antioxidant enzymes [\[162\]](#page-16-10) (Table [1\)](#page-9-0). While acting as a purifier of ROS, resveratrol can prevent lipid peroxidation [\[163\]](#page-16-11) (Table [1\)](#page-9-0). The higher the concentration of this polyphenol, the more antioxidant properties it reveals [\[164\]](#page-16-12). In cases of neurological disorders, resveratrol can inhibit the polymerization of the β-amyloid peptide [\[165\]](#page-16-13). Since resveratrol, after entering the bloodstream, can pass the blood–brain barrier, it can be used as a possible compound targeting neurodegenerative diseases [\[166](#page-16-14)[–168\]](#page-16-15). According to the study by Fiorillo et al., resveratrol inhibits the mitochondrial ATP synthase acting as an anti-aging compound [\[169\]](#page-16-16).

and aging composite [109]. mitochondrial dysfunction related to brain aging [\[170\]](#page-16-17) (Table [1\)](#page-9-0). The effects of curcumin are also dependent on cell age, and this is proved by administering it to middle-aged monkeys for a period from 14 to 18 months. The results of the study conducted on middle-aged monkeys, whose cognitive decline had just begun, showed that curcumin can enhance cognitive performance. However, more research is needed in older monkeys, who also present a rise in inflammatory markers and serious cognitive disabilities [$171,172$]. Furthermore, curcumin can be defensive towards rat hippocampus cells in case of homocysteine (Hcy) neurotoxicity to enhance memory deficiency [\[173\]](#page-16-20). The clinical trials demonstrated that since curcumin can be effective in reducing oxidative stress and inflammation, this Indian spice can be used in cases of age-related cognitive decline [174] (Table 1). In th[e i](#page-9-0)n vivo studies using neonatal Sprague-Dawley rats, where curcumin was administered at a dose of 100 mg/kg, we observed not only benefits in the improvement of white matter injury but an inhibition of the expression of iNOS and NOX in the microglia as well [175]. The fact that the nanoformulation of curcumin can increase NF-kB expression in fibroblast cells should encourage more studies to use curcumin as an anti-aging formulation [\[176](#page-16-23)[,177\]](#page-16-24). Curcumin can exert a protective role during inflammaging by blocking cognitive malfunction in **transgenic AD mice [178,179].**

Table 1. Main antioxidant compounds acting on brain aging.

[105,127,128]

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5. Discussion 5. Discussion

factors, including metabolism, oxidative stress, and mitochondrial function. As neurons susceptibility to neurodegenerative diseases. Neuronal cell rearrangement during aging is a complex process influenced by Neuronal cell rearrangement during aging is a complex process influenced by various age, they undergo significant changes that can lead to cognitive decline and increased

Aged neurons often exhibit glucose hypometabolism, which affects their energy supply and can lead to impaired synaptic function and reduced neuronal plasticity. Increased oxidative stress is another hallmark of aging neurons, where the accumulation of ROS can damage cellular components, including DNA, proteins, and lipids. This oxidative damage is intensified in neurodegenerative diseases. Mitochondria, the powerhouses of the cell, become less efficient with age, contributing to reduced ATP production and increased ROS generation, further impairing neuronal function. Additionally, aging neurons often accumulate misfolded and aggregated proteins, which can disrupt cellular processes and contribute to neurodegenerative conditions like Alzheimer's and Parkinson's diseases.

Mitochondrial dysfunction is one of the most remarkable signs of brain aging. Aging is the outcome of free radicals originally generated by cellular processes that are harmful, dangerous, and might lead to impairment. We find dysfunctional mitochondria in many age-related diseases such as Alzheimer's, Parkinson's, and Huntington's neurodegenerative diseases but also in inflammatory conditions. Free radicals can damage lipids, proteins, and carbohydrates. Peroxisomes are also organelles that go through morphological and structural age-related changes. Proteostasis promotes healthy protein synthesis, folding, and degradation, and this process is related to misregulation of protein maintenance during aging.

6. Conclusions

Aging is a natural and universal process that leads to time-related changes and a decrease in cognitive abilities, executive functions, and attention impairment. In neuronal aging, brain cells find difficulty in responding to oxidative stress due to accumulation of aggregated proteins in cytoplasm and mitochondria. Potential therapeutic approaches include antioxidants, which aim to reduce oxidative stress. The use of antioxidants can have neuroprotective effects while modulating neurotransmitters and reducing inflammation, oxidative stress, and apoptosis. Intracellular accumulation of $A\beta$ protein leads to oxidative stress and mitochondrial dysfunction, which may lead to neuronal death. Structure, function, and survival of neurons can be mediated by different pathways that are all sensitive to oxidative stress and age-related low-energy states. Agents that protect mitochondrial function, such as mitochondrial-targeted antioxidants and peptides, can help maintain ATP production and reduce oxidative stress. Since most of the damage is caused by oxidative stress and ROS, the use of antioxidants such as Vitamin C, Vitamin E, glutathione, carotenoids, quercetin, resveratrol, and curcumin is essential in increasing lifespan. Furthermore, antioxidants play an important role in the prevention of mitochondrial damage and cell death, boosting cell proliferation and improving memory.

7. Limitations and Future Studies

Overall, neuronal cell rearrangement during aging is a multifaceted process that significantly impacts brain function. Understanding the underlying mechanisms can help develop targeted therapeutic strategies to mitigate age-related neuronal decline and improve quality of life for the elderly. To test the antioxidants' efficiency further, clinical studies should be conducted. Also, it would be interesting to test different dosages of antioxidants, alone or in combination, to better understand the antioxidants' effects and the scavenging free oxygen radicals' properties. Future research should focus on combining pharmacological and lifestyle interventions to create comprehensive treatment plans for aging-related neuronal disorders.

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References

- 1. Castelli, V.; Benedetti, E.; Antonosante, A.; Catanesi, M.; Pitari, G.; Ippoliti, R.; Cimini, A.; D'angelo, M. Neuronal Cells Rearrangement During Aging and Neurodegenerative Disease: Metabolism, Oxidative Stress and Organelles Dynamic. *Front. Mol. Neurosci.* **2019**, *12*, 132. [\[CrossRef\]](https://doi.org/10.3389/fnmol.2019.00132) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31191244)
- 2. Buell, S.J.; Coleman, P.D. Dendritic growth in the aged human brain and failure of growth in senile dementia. *Science* **1979**, *206*, 854–856. [\[CrossRef\]](https://doi.org/10.1126/science.493989)
- 3. Alexander, G.E.; Ryan, L.; Bowers, D.; Foster, T.C.; Bizon, J.L.; Geldmacher, D.S.; Glisky, E.L. Characterizing cognitive aging in humans with links to animal models. *Front. Aging Neurosci.* **2012**, *4*, 21. [\[CrossRef\]](https://doi.org/10.3389/fnagi.2012.00021) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/22988439)
- 4. Roth, G.S.; Joseph, J.A. Cellular and molecular mechanisms of impaired dopaminergic function during aging. *Ann. N. Y. Acad. Sci.* **1994**, *719*, 129–135. [\[CrossRef\]](https://doi.org/10.1111/j.1749-6632.1994.tb56824.x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/8010587)
- 5. Mattson, M.P.; Maudsley, S.; Martin, B. BDNF and 5-HT: A dynamic duo in age-related neuronal plasticity and neurodegenerative disorders. *Trends Neurosci.* **2004**, *27*, 589–594. [\[CrossRef\]](https://doi.org/10.1016/j.tins.2004.08.001)
- 6. Bej, E.; Cesare, P.; Volpe, A.R.; D'angelo, M.; Castelli, V. Oxidative Stress and Neurodegeneration: Insights and Therapeutic Strategies for Parkinson's Disease. *Neurol. Int.* **2024**, *16*, 502–517. [\[CrossRef\]](https://doi.org/10.3390/neurolint16030037)
- 7. Volchegorskii, I.A.; Shemyakov, S.E.; Turygin, V.V.; Malinovskaya, N.V. The age dynamics of monoamine oxidase activity and levels of lipid peroxidation products in the human brain. *Neurosci. Behav. Physiol.* **2004**, *34*, 303–305. [\[CrossRef\]](https://doi.org/10.1023/B:NEAB.0000018736.84877.4f)
- 8. Ibáñez, V.; Pietrini, P.; Furey, M.L.; Alexander, G.E.; Millet, P.; Bokde, A.L.; Teichberg, D.; Schapiro, M.B.; Horwitz, B.; Rapoport, S.I. Resting state brain glucose metabolism is not reduced in normotensive healthy men during aging, after correction for brain atrophy. *Brain Res. Bull.* **2004**, *63*, 147–154. [\[CrossRef\]](https://doi.org/10.1016/j.brainresbull.2004.02.003)
- 9. Mattson, M.P.; Chan, S.L.; Duan, W. Modification of brain aging and neurodegenerative disorders by genes, diet, and behavior. *Physiol. Rev.* **2002**, *82*, 637–672. [\[CrossRef\]](https://doi.org/10.1152/physrev.00004.2002)
- 10. Bodles, A.M.; Barger, S.W. Cytokines and the aging brain—What we don't know might help us. *Trends Neurosci.* **2004**, *27*, 621–626. [\[CrossRef\]](https://doi.org/10.1016/j.tins.2004.07.011)
- 11. Mattson, M.P. Will caloric restriction and folate protect against AD and PD? *Neurology* **2003**, *60*, 690–695. [\[CrossRef\]](https://doi.org/10.1212/01.WNL.0000042785.02850.11) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/12601113)
- 12. Bej, E.; Volpe, A.R.; Cesare, P.; Cimini, A.; D'Angelo, M.; Castelli, V. Therapeutic potential of saffron in brain disorders: From bench to bedside. *Phytotherapy Res.* **2024**, *38*, 2482–2495. [\[CrossRef\]](https://doi.org/10.1002/ptr.8169) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38446350)
- 13. Yankner, B.A.; Lu, T.; Loerch, P. The aging brain. *Annu. Rev. Pathol.* **2008**, *3*, 41–66. [\[CrossRef\]](https://doi.org/10.1146/annurev.pathmechdis.2.010506.092044) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/18039130)
- 14. Reuter-Lorenz, P.A.; Lustig, C. Brain aging: Reorganizing discoveries about the aging mind. *Curr. Opin. Neurobiol.* **2005**, *15*, 245–251. [\[CrossRef\]](https://doi.org/10.1016/j.conb.2005.03.016)
- 15. Seidler, R.D.; Bernard, J.A.; Burutolu, T.B.; Fling, B.W.; Gordon, M.T.; Gwin, J.T.; Kwak, Y.; Lipps, D.B. Motor control and aging: Links to age-related brain structural, functional, and biochemical effects. *Neurosci. Biobehav. Rev.* **2010**, *34*, 721–733. [\[CrossRef\]](https://doi.org/10.1016/j.neubiorev.2009.10.005)
- 16. Fjell, A.M.; Sneve, M.H.; Storsve, A.B.; Grydeland, H.; Yendiki, A.; Walhovd, K.B. Brain Events Underlying Episodic Memory Changes in Aging: A Longitudinal Investigation of Structural and Functional Connectivity. *Cereb. Cortex* **2016**, *26*, 1272–1286. [\[CrossRef\]](https://doi.org/10.1093/cercor/bhv102)
- 17. Poon, H.; Calabrese, V.; Scapagnini, G.; Butterfield, D. Free radicals and brain aging. *Clin. Geriatr. Med.* **2004**, *20*, 329–359. [\[CrossRef\]](https://doi.org/10.1016/j.cger.2004.02.005)
- 18. Blinkouskaya, Y.; Caçoilo, A.; Gollamudi, T.; Jalalian, S.; Weickenmeier, J. Brain aging mechanisms with mechanical manifestations. *Mech. Ageing Dev.* **2021**, *200*, 111575. [\[CrossRef\]](https://doi.org/10.1016/j.mad.2021.111575)
- 19. Lupo, G.; Gaetani, S.; Cacci, E.; Biagioni, S.; Negri, R. Molecular Signatures of the Aging Brain: Finding the Links Between Genes and Phenotypes. *Neurother. J. Am. Soc. Exp. Neurother.* **2019**, *16*, 543–553. [\[CrossRef\]](https://doi.org/10.1007/s13311-019-00743-2)
- 20. Baker, D.J.; Petersen, R.C. Cellular senescence in brain aging and neurodegenerative diseases: Evidence and perspectives. *J. Clin. Investig.* **2018**, *128*, 1208–1216. [\[CrossRef\]](https://doi.org/10.1172/JCI95145)
- 21. Fielder, E.; Tweedy, C.; Wilson, C.; Oakley, F.; LeBeau, F.E.N.; Passos, J.F.; Mann, D.A.; von Zglinicki, T.; Jurk, D. Anti-inflammatory treatment rescues memory deficits during aging in *nfkb1*−/[−] mice. *Aging Cell* **2020**, *19*, e13188. [\[CrossRef\]](https://doi.org/10.1111/acel.13188) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32915495)
- 22. MacDonald, M.E.; Pike, G.B. MRI of healthy brain aging: A review. *NMR Biomed.* **2021**, *34*, e4564. [\[CrossRef\]](https://doi.org/10.1002/nbm.4564) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34096114)
- 23. Apostolova, L.G.; Green, A.E.; Babakchanian, S.; Hwang, K.S.; Chou, Y.-Y.; Toga, A.W.; Thompson, P.M. Hippocampal atrophy and ventricular enlargement in normal aging, mild cognitive impairment (mci), and alzheimer disease. *Alzheimer Dis. Assoc. Disord.* **2012**, *26*, 17–27. [\[CrossRef\]](https://doi.org/10.1097/WAD.0b013e3182163b62) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/22343374)
- 24. Lockhart, S.N.; DeCarli, C. Structural imaging measures of brain aging. *Neuropsychol. Rev.* **2014**, *24*, 271–289. [\[CrossRef\]](https://doi.org/10.1007/s11065-014-9268-3)
- 25. Park, D.C.; Reuter-Lorenz, P. The adaptive brain: Aging and neurocognitive scaffolding. *Annu. Rev. Psychol.* **2009**, *60*, 173–196. [\[CrossRef\]](https://doi.org/10.1146/annurev.psych.59.103006.093656)
- 26. Shankar, S. Biology of aging brain. *Indian J. Pathol. Microbiol.* **2010**, *53*, 595–604. [\[CrossRef\]](https://doi.org/10.4103/0377-4929.71995)
- 27. Schmitt, H.P. Epsilon-Glycation, APP and Abeta in ageing and Alzheimer disease: A hypothesis. *Med. Hypotheses* **2006**, *66*, 898–906. [\[CrossRef\]](https://doi.org/10.1016/j.mehy.2005.11.016)
- 28. Dickstein, D.L.; Kabaso, D.; Rocher, A.B.; Luebke, J.I.; Wearne, S.L.; Hof, P.R. Changes in the structural complexity of the aged brain. *Aging Cell* **2007**, *6*, 275–284. [\[CrossRef\]](https://doi.org/10.1111/j.1474-9726.2007.00289.x)
- 29. Mattson, M.P. Apoptosis in neurodegenerative disorders. *Nat. Rev. Mol. Cell Biol.* **2000**, *1*, 120–129. [\[CrossRef\]](https://doi.org/10.1038/35040009)
- 30. Richter-Landberg, C.; Goldbaum, O. Stress proteins in neural cells: Functional roles in health and disease. *Cell. Mol. Life Sci.* **2003**, *60*, 337–349. [\[CrossRef\]](https://doi.org/10.1007/s000180300028)
- 31. van Noort, J. Stress proteins in CNS inflammation. *J. Pathol.* **2008**, *214*, 267–275. [\[CrossRef\]](https://doi.org/10.1002/path.2273) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/18161755)
- 32. Hekimi, S.; Lapointe, J.; Wen, Y. Taking a "good" look at free radicals in the aging process. *Trends Cell Biol.* **2011**, *21*, 569–576. [\[CrossRef\]](https://doi.org/10.1016/j.tcb.2011.06.008) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21824781)
- 33. Bartman, S.; Coppotelli, G.; Ross, J.M. Mitochondrial Dysfunction: A Key Player in Brain Aging and Diseases. *Curr. Issues Mol. Biol.* **2024**, *46*, 1987–2026. [\[CrossRef\]](https://doi.org/10.3390/cimb46030130) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38534746)
- 34. Coppotelli, G.; Ross, J.M. Mitochondria in Ageing and Diseases: The Super Trouper of the Cell. *Int. J. Mol. Sci.* **2016**, *17*, 711. [\[CrossRef\]](https://doi.org/10.3390/ijms17050711) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27187361)
- 35. Fortini, P.; Pascucci, B. Special Issue "Mitochondrial Dysfunction: A Common Trigger in Neurodegenerative and Metabolic Non-Communicable Diseases". *Int. J. Mol. Sci.* **2024**, *25*, 4004. [\[CrossRef\]](https://doi.org/10.3390/ijms25074004)
- 36. Cui, H.; Kong, Y.; Zhang, H. Oxidative stress, mitochondrial dysfunction, and aging. *J. Signal Transduct.* **2012**, *2012*, 646354. [\[CrossRef\]](https://doi.org/10.1155/2012/646354)
- 37. Krishnan, K.J.; Reeve, A.K.; Samuels, D.C.; Chinnery, P.F.; Blackwood, J.K.; Taylor, R.W.; Wanrooij, S.; Spelbrink, J.N.; Lightowlers, R.N.; Turnbull, D.M. What causes mitochondrial DNA deletions in human cells? *Nat. Genet.* **2008**, *40*, 275–279. [\[CrossRef\]](https://doi.org/10.1038/ng.f.94)
- 38. Payne, B.A.I.; Chinnery, P.F. Mitochondrial dysfunction in aging: Much progress but many unresolved questions. *Biochim. Biophys. Acta* **2015**, *1847*, 1347–1353. [\[CrossRef\]](https://doi.org/10.1016/j.bbabio.2015.05.022)
- 39. Falkenberg, M.; Larsson, N.-G.; Gustafsson, C.M. DNA replication and transcription in mammalian mitochondria. *Annu. Rev. Biochem.* **2007**, *76*, 679–699. [\[CrossRef\]](https://doi.org/10.1146/annurev.biochem.76.060305.152028)
- 40. Chomyn, A.; Attardi, G. MtDNA mutations in aging and apoptosis. *Biochem. Biophys. Res. Commun.* **2003**, *304*, 519–529. [\[CrossRef\]](https://doi.org/10.1016/S0006-291X(03)00625-9)
- 41. Hiona, A.; Leeuwenburgh, C. The role of mitochondrial DNA mutations in aging and sarcopenia: Implications for the mitochondrial vicious cycle theory of aging. *Exp. Gerontol.* **2008**, *43*, 24–33. [\[CrossRef\]](https://doi.org/10.1016/j.exger.2007.10.001) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/17997255)
- 42. Guo, Y.; Guan, T.; Shafiq, K.; Yu, Q.; Jiao, X.; Na, D.; Li, M.; Zhang, G.; Kong, J. Mitochondrial dysfunction in aging. *Ageing Res. Rev.* **2023**, *88*, 101955. [\[CrossRef\]](https://doi.org/10.1016/j.arr.2023.101955)
- 43. Mather, M.; Rottenberg, H. Aging Enhances the Activation of the Permeability Transition Pore in Mitochondria. *Biochem. Biophys. Res. Commun.* **2000**, *273*, 603–608. [\[CrossRef\]](https://doi.org/10.1006/bbrc.2000.2994)
- 44. Boveris, A.; Navarro, A. Brain mitochondrial dysfunction in aging. *IUBMB Life* **2008**, *60*, 308–314. [\[CrossRef\]](https://doi.org/10.1002/iub.46)
- 45. Short, K.R.; Bigelow, M.L.; Kahl, J.; Singh, R.; Coenen-Schimke, J.; Raghavakaimal, S.; Nair, K.S. Decline in skeletal muscle mitochondrial function with aging in humans. *Proc. Natl. Acad. Sci. USA* **2005**, *102*, 5618–5623. [\[CrossRef\]](https://doi.org/10.1073/pnas.0501559102)
- 46. Houtkooper, R.H.; Argmann, C.; Houten, S.M.; Cantó, C.; Jeninga, E.H.; Andreux, P.A.; Thomas, C.; Doenlen, R.; Schoonjans, K.; Auwerx, J. The metabolic footprint of aging in mice. *Sci. Rep.* **2011**, *1*, 134. [\[CrossRef\]](https://doi.org/10.1038/srep00134) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/22355651)
- 47. Petersen, K.F.; Befroy, D.; Dufour, S.; Dziura, J.; Ariyan, C.; Rothman, D.L.; DiPietro, L.; Cline, G.W.; Shulman, G.I. Mitochondrial Dysfunction in the Elderly: Possible Role in Insulin Resistance. *Science* **2003**, *300*, 1140–1142. [\[CrossRef\]](https://doi.org/10.1126/science.1082889) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/12750520)
- 48. Deshwal, S.; Fiedler, K.U.; Langer, T. Mitochondrial Proteases: Multifaceted Regulators of Mitochondrial Plasticity. *Annu. Rev. Biochem.* **2020**, *89*, 501–528. [\[CrossRef\]](https://doi.org/10.1146/annurev-biochem-062917-012739)
- 49. Rai, M.; Curley, M.; Coleman, Z.; Demontis, F. Contribution of proteases to the hallmarks of aging and to age-related neurodegeneration. *Aging Cell* **2022**, *21*, e13603. [\[CrossRef\]](https://doi.org/10.1111/acel.13603)
- 50. Anand, R.; Wai, T.; Baker, M.J.; Kladt, N.; Schauss, A.C.; Rugarli, E.; Langer, T. The *i*-AAA protease YME1L and OMA1 cleave OPA1 to balance mitochondrial fusion and fission. *J. Cell Biol.* **2014**, *204*, 919–929. [\[CrossRef\]](https://doi.org/10.1083/jcb.201308006)
- 51. Shin, C.-S.; Meng, S.; Garbis, S.D.; Moradian, A.; Taylor, R.W.; Sweredoski, M.J.; Lomenick, B.; Chan, D.C. LONP1 and mtHSP70 cooperate to promote mitochondrial protein folding. *Nat. Commun.* **2021**, *12*, 265. [\[CrossRef\]](https://doi.org/10.1038/s41467-020-20597-z) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33431889)
- 52. Kirkland, J.L.; Tchkonia, T. Senolytic drugs: From discovery to translation. *J. Intern. Med.* **2020**, *288*, 518–536. [\[CrossRef\]](https://doi.org/10.1111/joim.13141) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32686219)
- 53. Miwa, S.; Kashyap, S.; Chini, E.; von Zglinicki, T. Mitochondrial dysfunction in cell senescence and aging. *J. Clin. Investig.* **2022**, *132*, e158447. [\[CrossRef\]](https://doi.org/10.1172/JCI158447)
- 54. Ramírez-Camacho, I.; Flores-Herrera, O.; Zazueta, C. The relevance of the supramolecular arrangements of the respiratory chain complexes in human diseases and aging. *Mitochondrion* **2019**, *47*, 266–272. [\[CrossRef\]](https://doi.org/10.1016/j.mito.2019.01.001)
- 55. Dencher, N.A.; Frenzel, M.; Reifschneider, N.H.; Sugawa, M.; Krause, F. Proteome Alterations in Rat Mitochondria Caused by Aging. *Ann. N. Y. Acad. Sci.* **2007**, *1100*, 291–298. [\[CrossRef\]](https://doi.org/10.1196/annals.1395.030)
- 56. Liu, L.; Su, X.; Quinn, W.J.; Hui, S.; Krukenberg, K.; Frederick, D.W.; Redpath, P.; Zhan, L.; Chellappa, K.; White, E.; et al. Quantitative Analysis of NAD Synthesis-Breakdown Fluxes. *Cell Metab.* **2018**, *27*, 1067–1080.e5. [\[CrossRef\]](https://doi.org/10.1016/j.cmet.2018.03.018)
- 57. Alano, C.C.; Ying, W.; Swanson, R.A. Poly (ADP-ribose) polymerase-1-mediated cell death in astrocytes requires NAD+ depletion and mitochondrial permeability transition. *J. Biol. Chem.* **2004**, *279*, 18895–18902. [\[CrossRef\]](https://doi.org/10.1074/jbc.M313329200) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/14960594)
- 58. Picca, A.; Lezza, A.M.S.; Leeuwenburgh, C.; Pesce, V.; Calvani, R.; Landi, F.; Bernabei, R.; Marzetti, E. Fueling Inflamm-Aging through Mitochondrial Dysfunction: Mechanisms and Molecular Targets. *Int. J. Mol. Sci.* **2017**, *18*, 933. [\[CrossRef\]](https://doi.org/10.3390/ijms18050933)
- 59. Montava-Garriga, L.; Ganley, I.G. Outstanding Questions in Mitophagy: What We Do and Do Not Know. *J. Mol. Biol.* **2020**, *432*, 206–230. [\[CrossRef\]](https://doi.org/10.1016/j.jmb.2019.06.032)
- 60. Brown, M.R.; Geddes, J.W.; Sullivan, P.G. Brain region-specific, age-related, alterations in mitochondrial responses to elevated calcium. *J. Bioenerg. Biomembr.* **2004**, *36*, 401–406. [\[CrossRef\]](https://doi.org/10.1023/B:JOBB.0000041775.10388.23)
- 61. Rowland, A.A.; Voeltz, G.K. Endoplasmic reticulum–mitochondria contacts: Function of the junction. *Nat. Rev. Mol. Cell Biol.* **2012**, *13*, 607–615. [\[CrossRef\]](https://doi.org/10.1038/nrm3440) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/22992592)
- 62. Gant, J.C.; Chen, K.-C.; Kadish, I.; Blalock, E.M.; Thibault, O.; Porter, N.M.; Landfield, P.W. Reversal of Aging-Related Neuronal Ca2+ Dysregulation and Cognitive Impairment by Delivery of a Transgene Encoding FK506-Binding Protein 12.6/1b to the Hippocampus. *J. Neurosci.* **2015**, *35*, 10878–10887. [\[CrossRef\]](https://doi.org/10.1523/JNEUROSCI.1248-15.2015) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26224869)
- 63. Brookes, P.S.; Yoon, Y.; Robotham, J.L.; Anders, M.W.; Sheu, S.-S. Calcium, ATP, and ROS: A mitochondrial love-hate triangle. *Am. J. Physiol. Physiol.* **2004**, *287*, C817–C833. [\[CrossRef\]](https://doi.org/10.1152/ajpcell.00139.2004) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/15355853)
- 64. Samson, R.D.; Barnes, C.A. Impact of aging brain circuits on cognition. *Eur. J. Neurosci.* **2013**, *37*, 1903–1915. [\[CrossRef\]](https://doi.org/10.1111/ejn.12183)
- 65. Sandell, J.H.; Peters, A. Effects of age on nerve fibers in the rhesus monkey optic nerve. *J. Comp. Neurol.* **2001**, *429*, 541–553. [\[CrossRef\]](https://doi.org/10.1002/1096-9861(20010122)429:4%3C541::AID-CNE3%3E3.0.CO;2-5)
- 66. Sandell, J.H.; Peters, A. Disrupted myelin and axon loss in the anterior commissure of the aged rhesus monkey. *J. Comp. Neurol.* **2003**, *466*, 14–30. [\[CrossRef\]](https://doi.org/10.1002/cne.10859)
- 67. Bowley, M.P.; Cabral, H.; Rosene, D.L.; Peters, A. Age changes in myelinated nerve fibers of the cingulate bundle and corpus callosum in the rhesus monkey. *J. Comp. Neurol.* **2010**, *518*, 3046–3064. [\[CrossRef\]](https://doi.org/10.1002/cne.22379)
- 68. Keller, J.N.; Dimayuga, E.; Chen, Q.; Thorpe, J.; Gee, J.; Ding, Q. Autophagy, proteasomes, lipofuscin, and oxidative stress in the aging brain. *Int. J. Biochem. Cell Biol.* **2004**, *36*, 2376–2391. [\[CrossRef\]](https://doi.org/10.1016/j.biocel.2004.05.003)
- 69. Fonseca, D.B.; Sheehy, M.R.J.; Blackman, N.; Shelton, P.M.J.; Prior, A.E. Reversal of a hallmark of brain ageing: Lipofuscin accumulation. *Neurobiol. Aging* **2005**, *26*, 69–76. [\[CrossRef\]](https://doi.org/10.1016/j.neurobiolaging.2004.02.013)
- 70. Gray, D.A.; Woulfe, J. Lipofuscin and aging: A matter of toxic waste. *Sci. Aging Knowl. Environ.* **2005**, *2005*, re1. [\[CrossRef\]](https://doi.org/10.1126/sageke.2005.5.re1)
- 71. Stojanovic, A.; Roher, A.E.; Ball, M.J. Quantitative analysis of lipofuscin and neurofibrillary tangles in the hippocampal neurons of alzheimer disease brains. *Dement. Geriatr. Cogn. Disord.* **1994**, *5*, 229–233. [\[CrossRef\]](https://doi.org/10.1159/000106728) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/7951677)
- 72. Zhao, L.; Spassieva, S.D.; Jucius, T.J.; Shultz, L.D.; Shick, H.E.; Macklin, W.B.; Hannun, Y.A.; Obeid, L.M.; Ackerman, S.L. A Deficiency of Ceramide Biosynthesis Causes Cerebellar Purkinje Cell Neurodegeneration and Lipofuscin Accumulation. *PLOS Genet.* **2011**, *7*, e1002063. [\[CrossRef\]](https://doi.org/10.1371/journal.pgen.1002063) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21625621)
- 73. Brunk, U.T.; Terman, A. Lipofuscin: Mechanisms of age-related accumulation and influence on cell function. *Free Radic. Biol. Med.* **2002**, *33*, 611–619. [\[CrossRef\]](https://doi.org/10.1016/S0891-5849(02)00959-0) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/12208347)
- 74. Riga, S.; Riga, D.; Schneider, F.; Halalau, F. Processing, lysis, and elimination of brain lipopigments in rejuvenation therapies. *Ann. New York Acad. Sci.* **2006**, *1067*, 383–387. [\[CrossRef\]](https://doi.org/10.1196/annals.1354.054)
- 75. El Mohsen, M.M.A.; Iravani, M.M.; Spencer, J.P.; Rose, S.; Fahim, A.T.; Motawi, T.M.; Ismail, N.A.; Jenner, P. Age-associated changes in protein oxidation and proteasome activities in rat brain: Modulation by antioxidants. *Biochem. Biophys. Res. Commun.* **2005**, *336*, 386–391. [\[CrossRef\]](https://doi.org/10.1016/j.bbrc.2005.07.201)
- 76. Braak, H.; Thal, D.R.; Ghebremedhin, E.; Del Tredici, K. Stages of the pathologic process in Alzheimer disease: Age categories from 1 to 100 years. *J. Neuropathol. Exp. Neurol.* **2011**, *70*, 960–969. [\[CrossRef\]](https://doi.org/10.1097/NEN.0b013e318232a379)
- 77. Rai, A.; Subramaniam, G. Polyglucosan Bodies in Aged Brain and Neurodegeneration: Cause or Consequence? In *Models, Molecules and Mechanisms in Biogerontology*; Rath, P.C., Ed.; Springer: Singapore, 2019; pp. 57–89. [\[CrossRef\]](https://doi.org/10.1007/978-981-13-3585-3_4)
- 78. Stahon, K.E.; Bastian, C.; Griffith, S.; Kidd, G.J.; Brunet, S.; Baltan, S. Age-Related Changes in Axonal and Mitochondrial Ultrastructure and Function in White Matter. *J. Neurosci.* **2016**, *36*, 9990–10001. [\[CrossRef\]](https://doi.org/10.1523/JNEUROSCI.1316-16.2016)
- 79. Neukomm, L.J.; Freeman, M.R. Diverse cellular and molecular modes of axon degeneration. *Trends Cell Biol.* **2014**, *24*, 515–523. [\[CrossRef\]](https://doi.org/10.1016/j.tcb.2014.04.003)
- 80. Adalbert, R.; Coleman, M.P. Review: Axon pathology in age-related neurodegenerative disorders. *Neuropathol. Appl. Neurobiol.* **2013**, *39*, 90–108. [\[CrossRef\]](https://doi.org/10.1111/j.1365-2990.2012.01308.x)
- 81. Coleman, M.P.; Freeman, M.R. Wallerian degeneration, wld(s), and nmnat. *Annu. Rev. Neurosci.* **2010**, *33*, 245–267. [\[CrossRef\]](https://doi.org/10.1146/annurev-neuro-060909-153248)
- 82. Lasiene, J.; Matsui, A.; Sawa, Y.; Wong, F.; Horner, P.J. Age-related myelin dynamics revealed by increased oligodendrogenesis and short internodes. *Aging Cell* **2009**, *8*, 201–213. [\[CrossRef\]](https://doi.org/10.1111/j.1474-9726.2009.00462.x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/19338498)
- 83. Barrientos, S.A.; Martinez, N.W.; Yoo, S.; Jara, J.S.; Zamorano, S.; Hetz, C.; Twiss, J.L.; Alvarez, J.; Court, F.A. Axonal degeneration is mediated by the mitochondrial permeability transition pore. *J. Neurosci.* **2011**, *31*, 966–978. [\[CrossRef\]](https://doi.org/10.1523/JNEUROSCI.4065-10.2011)
- 84. Villegas, R.; Martinez, N.W.; Lillo, J.; Pihan, P.; Hernandez, D.; Twiss, J.L.; Court, F.A. Calcium release from intra-axonal endoplasmic reticulum leads to axon degeneration through mitochondrial dysfunction. *J. Neurosci.* **2014**, *34*, 7179–7189. [\[CrossRef\]](https://doi.org/10.1523/JNEUROSCI.4784-13.2014) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24849352)
- 85. Douglas, P.M.; Dillin, A. Protein homeostasis and aging in neurodegeneration. *J. Cell Biol.* **2010**, *190*, 719–729. [\[CrossRef\]](https://doi.org/10.1083/jcb.201005144)
- 86. Balch, W.E.; Morimoto, R.I.; Dillin, A.; Kelly, J.W. Adapting proteostasis for disease intervention. *Science* **2008**, *319*, 916–919. [\[CrossRef\]](https://doi.org/10.1126/science.1141448) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/18276881)
- 87. Calamini, B.; Silva, M.C.; Madoux, F.; Hutt, D.M.; Khanna, S.; Chalfant, M.A.; Saldanha, S.A.; Hodder, P.; Tait, B.D.; Garza, D.; et al. Small-molecule proteostasis regulators for protein conformational diseases. *Nat. Chem. Biol.* **2011**, *8*, 185–196. [\[CrossRef\]](https://doi.org/10.1038/nchembio.763)
- 88. Calderwood, S.K.; Murshid, A.; Prince, T. The shock of aging: Molecular chaperones and the heat shock response in longevity and aging—A mini-review. *Gerontology* **2009**, *55*, 550–558. [\[CrossRef\]](https://doi.org/10.1159/000225957)
- 89. Powers, E.T.; Morimoto, R.I.; Dillin, A.; Kelly, J.W.; Balch, W.E. Biological and chemical approaches to diseases of proteostasis deficiency. *Annu. Rev. Biochem.* **2009**, *78*, 959–991. [\[CrossRef\]](https://doi.org/10.1146/annurev.biochem.052308.114844)
- 90. Koga, H.; Kaushik, S.; Cuervo, A.M. Protein homeostasis and aging: The importance of exquisite quality control. *Ageing Res. Rev.* **2011**, *10*, 205–215. [\[CrossRef\]](https://doi.org/10.1016/j.arr.2010.02.001)
- 91. Hartl, F.U.; Bracher, A.; Hayer-Hartl, M. Molecular chaperones in protein folding and proteostasis. *Nature* **2011**, *475*, 324–332. [\[CrossRef\]](https://doi.org/10.1038/nature10317)
- 92. Rubinsztein, D.C.; Marino, G.; Kroemer, G. Autophagy and aging. *Cell* **2011**, *146*, 682–695. [\[CrossRef\]](https://doi.org/10.1016/j.cell.2011.07.030) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21884931)
- 93. Kwon, S.-K.; Lee, D.-H.; Kim, S.-Y.; Park, J.-H.; Choi, J.; Baek, K.-H. Ubiquitin-specific protease 21 regulating the K48-linked polyubiquitination of NANOG. *Biochem. Biophys. Res. Commun.* **2017**, *482*, 1443–1448. [\[CrossRef\]](https://doi.org/10.1016/j.bbrc.2016.12.055) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27956178)
- 94. Lopes-Paciencia, S.; Saint-Germain, E.; Rowell, M.-C.; Ruiz, A.F.; Kalegari, P.; Ferbeyre, G. The senescence-associated secretory phenotype and its regulation. *Cytokine* **2019**, *117*, 15–22. [\[CrossRef\]](https://doi.org/10.1016/j.cyto.2019.01.013) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30776684)
- 95. Genereux, J.C.; Qu, S.; Zhou, M.; Ryno, L.M.; Wang, S.; Shoulders, M.D.; Kaufman, R.J.; Lasmézas, C.I.; Kelly, J.W.; Wiseman, R.L. Unfolded protein response-induced ER dj3 secretion links ER stress to extracellular proteostasis. *EMBO J.* **2015**, *34*, 4–19. [\[CrossRef\]](https://doi.org/10.15252/embj.201488896)
- 96. Wyatt, A.R.; Yerbury, J.J.; Ecroyd, H.; Wilson, M.R. Extracellular chaperones and proteostasis. *Annu. Rev. Biochem.* **2013**, *82*, 295–322. [\[CrossRef\]](https://doi.org/10.1146/annurev-biochem-072711-163904)
- 97. Santoro, A.; Martucci, M.; Conte, M.; Capri, M.; Franceschi, C.; Salvioli, S. Inflammaging, hormesis and the rationale for anti-aging strategies. *Ageing Res. Rev.* **2020**, *64*, 101142. [\[CrossRef\]](https://doi.org/10.1016/j.arr.2020.101142)
- 98. Fulop, T.; Larbi, A.; Dupuis, G.; Le Page, A.; Frost, E.H.; Cohen, A.A.; Witkowski, J.M.; Franceschi, C. Immunosenescence and Inflamm-Aging as Two Sides of the Same Coin: Friends or Foes? *Front. Immunol.* **2017**, *8*, 1960. [\[CrossRef\]](https://doi.org/10.3389/fimmu.2017.01960) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29375577)
- 99. Franceschi, C.; Capri, M.; Monti, D.; Giunta, S.; Olivieri, F.; Sevini, F.; Panourgia, M.P.; Invidia, L.; Celani, L.; Scurti, M.; et al. Inflammaging and anti-inflammaging: A systemic perspective on aging and longevity emerged from studies in humans. *Mech. Ageing Dev.* **2007**, *128*, 92–105. [\[CrossRef\]](https://doi.org/10.1016/j.mad.2006.11.016)
- 100. López-Otín, C.; Blasco, M.A.; Partridge, L.; Serrano, M.; Kroemer, G. The hallmarks of aging. *Cell* **2013**, *153*, 1194–1217. [\[CrossRef\]](https://doi.org/10.1016/j.cell.2013.05.039)
- 101. Tchkonia, T.; Zhu, Y.; van Deursen, J.; Campisi, J.; Kirkland, J.L. Cellular senescence and the senescent secretory phenotype: Therapeutic opportunities. *J. Clin. Investig.* **2013**, *123*, 966–972. [\[CrossRef\]](https://doi.org/10.1172/JCI64098)
- 102. Jose, V.; Consuelo, B.; Miquel, J. Theories of ageing. *IUBMB Life* **2007**, *59*, 249–254. [\[CrossRef\]](https://doi.org/10.1080/15216540601178067)
- 103. Miyazawa, T. Lipid hydroperoxides in nutrition, health, and diseases. *Proc. Jpn. Acad. Ser. B* **2021**, *97*, 161–196. [\[CrossRef\]](https://doi.org/10.2183/pjab.97.010) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33840675)
- 104. Cadenas, E.; Davies, K.J. Mitochondrial free radical generation, oxidative stress, and aging. *Free. Radic. Biol. Med.* **2000**, *29*, 222–230. [\[CrossRef\]](https://doi.org/10.1016/S0891-5849(00)00317-8)
- 105. Birben, E.; Sahiner, U.M.; Sackesen, C.; Erzurum, S.; Kalayci, O. Oxidative stress and antioxidant defense. *World Allergy Organ. J.* **2012**, *5*, 9–19. [\[CrossRef\]](https://doi.org/10.1097/WOX.0b013e3182439613)
- 106. Herken, H.; Uz, E.; Özyurt, H.; Söğüt, S.; Virit, O.; Akyol, O. Evidence that the activities of erythrocyte free radical scavenging enzymes and the products of lipid peroxidation are increased in different forms of schizophrenia. *Mol. Psychiatry* **2001**, *6*, 66–73. [\[CrossRef\]](https://doi.org/10.1038/sj.mp.4000789)
- 107. Zimniak, P. Relationship of electrophilic stress to aging. *Free. Radic. Biol. Med.* **2011**, *51*, 1087–1105. [\[CrossRef\]](https://doi.org/10.1016/j.freeradbiomed.2011.05.039)
- 108. Kelsey, N.A.; Wilkins, H.M.; Linseman, D.A. Nutraceutical antioxidants as novel neuroprotective agents. *Molecules* **2010**, *15*, 7792–7814. [\[CrossRef\]](https://doi.org/10.3390/molecules15117792) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21060289)
- 109. Lima, G.P.P.; Vianello, F.; Corrêa, C.R.; da Silva Campos, R.A.; Borguini, M.G. Polyphenols in Fruits and Vegetables and Its Effect on Human Health. *Food Nutr. Sci.* **2014**, *5*, 1065–1082. [\[CrossRef\]](https://doi.org/10.4236/fns.2014.511117)
- 110. Brewer, M.S. Natural Antioxidants: Sources, Compounds, Mechanisms of Action, and Potential Applications. *Compr. Rev. Food Sci. Food Saf.* **2011**, *10*, 221–247. [\[CrossRef\]](https://doi.org/10.1111/j.1541-4337.2011.00156.x)
- 111. Parcheta, M.; Świsłocka, R.; Orzechowska, S.; Akimowicz, M.; Choińska, R.; Lewandowski, W. Recent Developments in Effective Antioxidants: The Structure and Antioxidant Properties. *Materials* **2021**, *14*, 1984. [\[CrossRef\]](https://doi.org/10.3390/ma14081984)
- 112. Sies, H. Oxidative stress: A concept in redox biology and medicine. *Redox Biol.* **2015**, *4*, 180–183. [\[CrossRef\]](https://doi.org/10.1016/j.redox.2015.01.002) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25588755)
- 113. Teng, H.; Chen, L. Polyphenols and bioavailability: An update. *Crit. Rev. Food Sci. Nutr.* **2019**, *59*, 2040–2051. [\[CrossRef\]](https://doi.org/10.1080/10408398.2018.1437023) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29405736)
- 114. Barrajón-Catalán, E.; Herranz-López, M.; Joven, J.; Segura-Carretero, A.; Alonso-Villaverde, C.; Menendez, J.A.; Micol, V. Molecular promiscuity of plant polyphenols in the management of age-related diseases: Far beyond their antioxidant properties. *Adv. Exp. Med. Biol.* **2014**, *824*, 141–159. [\[CrossRef\]](https://doi.org/10.1007/978-3-319-07320-0_11)
- 115. Cherniack, E.P. The potential influence of plant polyphenols on the aging process. *Forsch. Komplementarmedizin* **2010**, *17*, 181–187. [\[CrossRef\]](https://doi.org/10.1159/000319143)
- 116. Al-Rasheed, N.M.; Fadda, L.M.; Ali, H.M.; Baky, N.A.A.; El-Orabi, N.F.; Al-Rasheed, N.M.; Yacoub, H.I. New mechanism in the modulation of carbon tetrachloride hepatotoxicity in rats using different natural antioxidants. *Toxicol. Mech. Methods* **2016**, *26*, 243–250. [\[CrossRef\]](https://doi.org/10.3109/15376516.2016.1159769)
- 117. Vaya, J.; Aviram, M. Nutritional Antioxidants Mechanisms of Action, Analyses of Activities and Medical Applications. *Curr. Med. Chem. Immunol. Endocr. Metab. Agents* **2001**, *1*, 99–117. [\[CrossRef\]](https://doi.org/10.2174/1568013013359168)
- 118. Veurink, G.; Perry, G.; Singh, S.K. Role of antioxidants and a nutrient rich diet in Alzheimer's disease. *Open Biol.* **2020**, *10*, 200084. [\[CrossRef\]](https://doi.org/10.1098/rsob.200084)
- 119. Mezzetti, A.; Lapenna, D.; Romano, F.; Costantini, F.; Pierdomenico, S.D.; De Cesare, D.; Cuccurullo, F.; Riario-Sforza, G.; Zuliani, G.; Fellin, R.; et al. Systemic oxidative stress and its relationship with age and illness. *J. Am. Geriatr. Soc.* **1996**, *44*, 823–827. [\[CrossRef\]](https://doi.org/10.1111/j.1532-5415.1996.tb03741.x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/8675932)
- 120. Abe, C.; Higuchi, O.; Matsumoto, A.; Miyazawa, T. Determination of intracellular ascorbic acid using tandem mass spectrometry. *Anal.* **2022**, *147*, 2640–2643. [\[CrossRef\]](https://doi.org/10.1039/D1AN02160E)
- 121. Dixit, S.; Bernardo, A.; Walker, J.M.; Kennard, J.A.; Kim, G.Y.; Kessler, E.S.; Harrison, F.E. Vitamin C deficiency in the brain impairs cognition, increases amyloid accumulation and deposition, and oxidative stress in app/psen1 and normally aging mice. *ACS Chem. Neurosci.* **2015**, *6*, 570–581. [\[CrossRef\]](https://doi.org/10.1021/cn500308h)
- 122. El-Gendy, K.S.; Aly, N.M.; Mahmoud, F.H.; Kenawy, A.; El-Sebae, A.K.H. The role of vitamin C as antioxidant in protection of oxidative stress induced by imidacloprid. *Food Chem. Toxicol.* **2009**, *48*, 215–221. [\[CrossRef\]](https://doi.org/10.1016/j.fct.2009.10.003) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/19833166)
- 123. Aune, D.; Keum, N.; Giovannucci, E.; Fadnes, L.T.; Boffetta, P.; Greenwood, D.C.; Tonstad, S.; Vatten, L.J.; Riboli, E.; Norat, T. Dietary intake and blood concentrations of antioxidants and the risk of cardiovascular disease, total cancer, and all-cause mortality: A systematic review and dose-response meta-analysis of prospective studies. *Am. J. Clin. Nutr.* **2018**, *108*, 1069–1091. [\[CrossRef\]](https://doi.org/10.1093/ajcn/nqy097) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30475962)
- 124. Brubacher, D.; Moser, U.; Jordan, P. Vitamin C concentrations in plasma as a function of intake: A meta-analysis. *Int. J. Vitam. Nutr. Res.* **2000**, *70*, 226–237. [\[CrossRef\]](https://doi.org/10.1024/0300-9831.70.5.226) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/11068703)
- 125. Carr, A.C.; Zawari, M. Does Aging Have an Impact on Vitamin C Status and Requirements? A Scoping Review of Comparative Studies of Aging and Institutionalisation. *Nutrients* **2023**, *15*, 915. [\[CrossRef\]](https://doi.org/10.3390/nu15040915) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36839273)
- 126. Mangialasche, F.; Xu, W.; Kivipelto, M.; Costanzi, E.; Ercolani, S.; Pigliautile, M.; Cecchetti, R.; Baglioni, M.; Simmons, A.; Soininen, H.; et al. Tocopherols and tocotrienols plasma levels are associated with cognitive impairment. *Neurobiol. Aging* **2012**, *33*, 2282–2290. [\[CrossRef\]](https://doi.org/10.1016/j.neurobiolaging.2011.11.019)
- 127. Browne, D.; McGuinness, B.; Woodside, J.V.; McKay, G.J. Vitamin E and Alzheimer's disease: What do we know so far? *Clin. Interv. Aging* **2019**, *14*, 1303–1317. [\[CrossRef\]](https://doi.org/10.2147/CIA.S186760) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31409980)
- 128. Zhao, R.; Han, X.; Zhang, H.; Liu, J.; Zhang, M.; Zhao, W.; Jiang, S.; Li, R.; Cai, H.; You, H. Association of vitamin E intake in diet and supplements with risk of dementia: A meta-analysis. *Front. Aging Neurosci.* **2022**, *14*, 955878. [\[CrossRef\]](https://doi.org/10.3389/fnagi.2022.955878)
- 129. Arzi, A.; Hemmati, A.; Razian, A. Effect of vitamins C and E on cognitive function in mouse. *Pharmacol. Res.* **2004**, *49*, 249–252. [\[CrossRef\]](https://doi.org/10.1016/j.phrs.2003.10.004)
- 130. Nazıroglu, M.; Butterworth, J.; Sonmez, T. Dietary vitamin c and e modulates antioxidant levels in blood, brain, liver, muscle, and testes in diabetic aged rats. *Int. J. Vitam. Nutr. Res.* **2011**, *81*, 347–357. [\[CrossRef\]](https://doi.org/10.1024/0300-9831/a000083)
- 131. Devi, S.A.; Manjula, K. Intermittent cold-induced hippocampal oxidative stress is associated with changes in the plasma lipid composition and is modifiable by vitamins C and E in old rats. *Neurochem. Int.* **2014**, *74*, 46–52. [\[CrossRef\]](https://doi.org/10.1016/j.neuint.2014.05.001)
- 132. Devi, S.A.; Manjula, K.; Subramanyam, M. Protective role of vitamins E and C against oxidative stress caused by intermittent cold exposure in aging rat's frontoparietal cortex. *Neurosci. Lett.* **2012**, *529*, 155–160. [\[CrossRef\]](https://doi.org/10.1016/j.neulet.2012.09.041) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/23026027)
- 133. Masella, R.; Di Benedetto, R.; Varì, R.; Filesi, C.; Giovannini, C. Novel mechanisms of natural antioxidant compounds in biological systems: Involvement of glutathione and glutathione-related enzymes. *J. Nutr. Biochem.* **2005**, *16*, 577–586. [\[CrossRef\]](https://doi.org/10.1016/j.jnutbio.2005.05.013) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/16111877)
- 134. Dringen, R. Metabolism and functions of glutathione in brain. *Prog. Neurobiol.* **2000**, *62*, 649–671. [\[CrossRef\]](https://doi.org/10.1016/S0301-0082(99)00060-X) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/10880854) 135. Freitas, H.R.; Ferreira, G.D.; Trevenzoli, I.H.; Oliveira, K.D.; de Melo Reis, R.A. Fatty Acids, Antioxidants and Physical Activity in Brain Aging. *Nutrients* **2017**, *9*, 1263. [\[CrossRef\]](https://doi.org/10.3390/nu9111263)
- 136. Iskusnykh, I.Y.; Zakharova, A.A.; Pathak, D. Glutathione in Brain Disorders and Aging. *Molecules* **2022**, *27*, 324. [\[CrossRef\]](https://doi.org/10.3390/molecules27010324)
- 137. Belrose, J.C.; Xie, Y.-F.; Gierszewski, L.J.; MacDonald, J.F.; Jackson, M.F. Loss of glutathione homeostasis associated with neuronal senescence facilitates TRPM2 channel activation in cultured hippocampal pyramidal neurons. *Mol. Brain* **2012**, *5*, 11. [\[CrossRef\]](https://doi.org/10.1186/1756-6606-5-11)
- 138. Lu, S.C. Regulation of glutathione synthesis. *Mol. Asp. Med.* **2008**, *30*, 42–59. [\[CrossRef\]](https://doi.org/10.1016/j.mam.2008.05.005)
- 139. Detcheverry, F.; Senthil, S.; Narayanan, S.; Badhwar, A. Changes in levels of the antioxidant glutathione in brain and blood across the age span of healthy adults: A systematic review. *NeuroImage: Clin.* **2023**, *40*, 103503. [\[CrossRef\]](https://doi.org/10.1016/j.nicl.2023.103503)
- 140. El-Agamey, A.; Lowe, G.M.; McGarvey, D.J.; Mortensen, A.; Phillip, D.M.; Truscott, T.; Young, A.J. Carotenoid radical chemistry and antioxidant/pro-oxidant properties. *Arch. Biochem. Biophys.* **2004**, *430*, 37–48. [\[CrossRef\]](https://doi.org/10.1016/j.abb.2004.03.007)
- 141. Boccardi, V.; Arosio, B.; Cari, L.; Bastiani, P.; Scamosci, M.; Casati, M.; Ferri, E.; Bertagnoli, L.; Ciccone, S.; Rossi, P.D.; et al. Beta-carotene, telomerase activity and Alzheimer's disease in old age subjects. *Eur. J. Nutr.* **2020**, *59*, 119–126. [\[CrossRef\]](https://doi.org/10.1007/s00394-019-01892-y)
- 142. Riccioni, G. Carotenoids and cardiovascular disease. *Curr. Atheroscler. Rep.* **2009**, *11*, 434–439. [\[CrossRef\]](https://doi.org/10.1007/s11883-009-0065-z) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/19852884)
- 143. Böhm, V.; Puspitasari-Nienaber, N.L.; Ferruzzi, M.G.; Schwartz, S.J. Trolox equivalent antioxidant capacity of different geometrical isomers of α-carotene, β-carotene, lycopene, and zeaxanthin. *J. Agric. Food Chem.* **2002**, *50*, 221–226. [\[CrossRef\]](https://doi.org/10.1021/jf010888q) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/11754571)
- 144. Abrego-Guandique, D.M.; Bonet, M.L.; Caroleo, M.C.; Cannataro, R.; Tucci, P.; Ribot, J.; Cione, E. The Effect of Beta-Carotene on Cognitive Function: A Systematic Review. *Brain Sci.* **2023**, *13*, 1468. [\[CrossRef\]](https://doi.org/10.3390/brainsci13101468)
- 145. Grodstein, F.; Kang, J.H.; Glynn, R.J.; Cook, N.R.; Gaziano, J.M. A randomized trial of beta carotene supplementation and cognitive function in menthe physicians' health study II. *Arch. Intern. Med.* **2007**, *167*, 2184–2190. [\[CrossRef\]](https://doi.org/10.1001/archinte.167.20.2184) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/17998490)
- 146. Fiedor, J.; Burda, K. Potential role of carotenoids as antioxidants in human health and disease. *Nutrients* **2014**, *6*, 466–488. [\[CrossRef\]](https://doi.org/10.3390/nu6020466)
- 147. Lu, B.; Nagappan, G.; Lu, Y. BDNF and synaptic plasticity, cognitive function, and dysfunction. In *Handbook of Experimental Pharmacology*; Springer: Berlin/Heidelberg, Germany, 2014; Volume 220, pp. 223–250. [\[CrossRef\]](https://doi.org/10.1007/978-3-642-45106-5_9)
- 148. Wang, H.; Jo, Y.-J.; Oh, J.S.; Kim, N.-H. Quercetin delays postovulatory aging of mouse oocytes by regulating SIRT expression and MPF activity. *Oncotarget* **2017**, *8*, 38631–38641. [\[CrossRef\]](https://doi.org/10.18632/oncotarget.16219) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28418847)
- 149. Geng, L.; Liu, Z.; Zhang, W.; Li, W.; Wu, Z.; Wang, W.; Ren, R.; Su, Y.; Wang, P.; Sun, L.; et al. Chemical screen identifies a geroprotective role of quercetin in premature aging. *Protein Cell* **2019**, *10*, 417–435. [\[CrossRef\]](https://doi.org/10.1007/s13238-018-0567-y)
- 150. Cui, Z.; Zhao, X.; Amevor, F.K.; Du, X.; Wang, Y.; Li, D.; Shu, G.; Tian, Y.; Zhao, X. Therapeutic application of quercetin in aging-related diseases: SIRT1 as a potential mechanism. *Front. Immunol.* **2022**, *13*, 943321. [\[CrossRef\]](https://doi.org/10.3389/fimmu.2022.943321)
- 151. Ferrucci, L.; Fabbri, E. Inflammageing: Chronic inflammation in ageing, cardiovascular disease, and frailty. *Nat. Rev. Cardiol.* **2018**, *15*, 505–522. [\[CrossRef\]](https://doi.org/10.1038/s41569-018-0064-2)
- 152. Singh, V.; Ubaid, S. Role of Silent Information Regulator 1 (SIRT1) in Regulating Oxidative Stress and Inflammation. *Inflammation* **2020**, *43*, 1589–1598. [\[CrossRef\]](https://doi.org/10.1007/s10753-020-01242-9)
- 153. Shahgaldi, S.; Kahmini, F.R. A comprehensive review of Sirtuins: With a major focus on redox homeostasis and metabolism. *Life Sci.* **2021**, *282*, 119803. [\[CrossRef\]](https://doi.org/10.1016/j.lfs.2021.119803) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34237310)
- 154. Leyton, L.; Hott, M.; Acuña, F.; Caroca, J.; Nuñez, M.; Martin, C.; Zambrano, A.; Concha, M.I.; Otth, C. Nutraceutical activators of AMPK/Sirt1 axis inhibit viral production and protect neurons from neurodegenerative events triggered during HSV-1 infection. *Virus Res.* **2015**, *205*, 63–72. [\[CrossRef\]](https://doi.org/10.1016/j.virusres.2015.05.015)
- 155. Li, H.; Chen, F.-J.; Yang, W.-L.; Qiao, H.-Z.; Zhang, S.-J. Quercetin improves cognitive disorder in aging mice by inhibiting NLRP3 inflammasome activation. *Food Funct.* **2021**, *12*, 717–725. [\[CrossRef\]](https://doi.org/10.1039/D0FO01900C) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33338087)
- 156. Navarro-Cruz, A.R.; Ayala, R.R.Y.; Ochoa-Velasco, C.; Brambila, E.; Avila-Sosa, R.; Pérez-Fernández, S.; Morales-Medina, J.C.; Aguilar-Alonso, P. Effect of Chronic Administration of Resveratrol on Cognitive Performance during Aging Process in Rats. *Oxidative Med. Cell. Longev.* **2017**, *2017*, 8510761. [\[CrossRef\]](https://doi.org/10.1155/2017/8510761) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29163756)
- 157. Caldeira, C.A.; Santos, M.A.; Araújo, G.R.; Lara, R.C.; Franco, F.N.; Chaves, M.M. Resveratrol: Change of SIRT 1 and AMPK signaling pattern during the aging process. *Exp. Gerontol.* **2021**, *146*, 111226. [\[CrossRef\]](https://doi.org/10.1016/j.exger.2021.111226)
- 158. Lagouge, M.; Argmann, C.; Gerhart-Hines, Z.; Meziane, H.; Lerin, C.; Daussin, F.; Messadeq, N.; Milne, J.; Lambert, P.; Elliott, P.; et al. Resveratrol improves mitochondrial function and protects against metabolic disease by activating SIRT1 and PGC-1α. *Cell* **2006**, *127*, 1109–1122. [\[CrossRef\]](https://doi.org/10.1016/j.cell.2006.11.013)
- 159. Moussa, C.; Hebron, M.; Huang, X.; Ahn, J.; Rissman, R.A.; Aisen, P.S.; Turner, R.S. Resveratrol regulates neuro-inflammation and induces adaptive immunity in Alzheimer's disease. *J. Neuroinflam.* **2017**, *14*, 1. [\[CrossRef\]](https://doi.org/10.1186/s12974-016-0779-0)
- 160. Khan, M.M.; Ahmad, A.; Ishrat, T.; Khan, M.B.; Hoda, M.N.; Khuwaja, G.; Raza, S.S.; Khan, A.; Javed, H.; Vaibhav, K.; et al. Resveratrol attenuates 6-hydroxydopamine-induced oxidative damage and dopamine depletion in rat model of Parkinson's disease. *Brain Res.* **2010**, *1328*, 139–151. [\[CrossRef\]](https://doi.org/10.1016/j.brainres.2010.02.031)
- 161. Zhou, D.-D.; Luo, M.; Huang, S.-Y.; Saimaiti, A.; Shang, A.; Gan, R.-Y.; Li, H.-B. Effects and Mechanisms of Resveratrol on Aging and Age-Related Diseases. *Oxidative Med. Cell. Longev.* **2021**, *2021*, 9932218. [\[CrossRef\]](https://doi.org/10.1155/2021/9932218)
- 162. Izzo, C.; Annunziata, M.; Melara, G.; Sciorio, R.; Dallio, M.; Masarone, M.; Federico, A.; Persico, M. The Role of Resveratrol in Liver Disease: A Comprehensive Review from In Vitro to Clinical Trials. *Nutrients* **2021**, *13*, 933. [\[CrossRef\]](https://doi.org/10.3390/nu13030933)
- 163. Goswami, S.K.; Das, D.K. Resveratrol and chemoprevention. *Cancer Lett.* **2009**, *284*, 1–6. [\[CrossRef\]](https://doi.org/10.1016/j.canlet.2009.01.041) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/19261378)
- 164. Acquaviva, R.; Russo, A.; Campisi, A.; Sorrenti, V.; Di Giacomo, C.; Barcellona, M.; Avitabile, M.; Vanella, A. Antioxidant Activity and Protective Effect on DNA Cleavage of Resveratrol. *J. Food Sci.* **2002**, *67*, 137–141. [\[CrossRef\]](https://doi.org/10.1111/j.1365-2621.2002.tb11373.x)
- 165. Marambaud, P.; Zhao, H.; Davies, P. Resveratrol promotes clearance of Alzheimer's disease amyloid-β peptides. *J. Biol. Chem.* **2005**, *280*, 37377–37382. [\[CrossRef\]](https://doi.org/10.1074/jbc.M508246200) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/16162502)
- 166. Sun, A.Y.; Wang, Q.; Simonyi, A.; Sun, G.Y. Resveratrol as a therapeutic agent for neurodegenerative diseases. *Mol. Neurobiol.* **2010**, *41*, 375–383. [\[CrossRef\]](https://doi.org/10.1007/s12035-010-8111-y)
- 167. Richard, T.; Pawlus, A.D.; Iglésias, M.; Pedrot, E.; Waffo-Teguo, P.; Mérillon, J.; Monti, J. Neuroprotective properties of resveratrol and derivatives. *Ann. New York Acad. Sci.* **2011**, *1215*, 103–108. [\[CrossRef\]](https://doi.org/10.1111/j.1749-6632.2010.05865.x)
- 168. Tellone, E.; Galtieri, A.; Russo, A.; Giardina, B.; Ficarra, S. Resveratrol: A Focus on Several Neurodegenerative Diseases. *Oxidative Med. Cell. Longev.* **2015**, *2015*, 392169. [\[CrossRef\]](https://doi.org/10.1155/2015/392169)
- 169. Fiorillo, M.; Lamb, R.; Tanowitz, H.B.; Cappello, A.R.; Martinez-Outschoorn, U.E.; Sotgia, F.; Lisanti, M.P. Bedaquiline, an FDA-approved antibiotic, inhibits mitochondrial function and potently blocks the proliferative expansion of stem-like cancer cells (CSCs). *Aging* **2016**, *8*, 1593–1607. [\[CrossRef\]](https://doi.org/10.18632/aging.100983)
- 170. Hagl, S.; Kocher, A.; Schiborr, C.; Kolesova, N.; Frank, J.; Eckert, G.P. Curcumin micelles improve mitochondrial function in neuronal PC12 cells and brains of NMRI mice—Impact on bioavailability. *Neurochem. Int.* **2015**, *89*, 234–242. [\[CrossRef\]](https://doi.org/10.1016/j.neuint.2015.07.026)
- 171. Moore, T.L.; Bowley, B.; Shultz, P.; Calderazzo, S.; Shobin, E.; Killiany, R.J.; Rosene, D.L.; Moss, M.B. Chronic curcumin treatment improves spatial working memory but not recognition memory in middle-aged rhesus monkeys. *GeroScience* **2017**, *39*, 571–584. [\[CrossRef\]](https://doi.org/10.1007/s11357-017-9998-2)
- 172. Riddle, D.R. (Ed.) *Brain Aging: Models, Methods, and Mechanisms*, 1st ed.; CRC Press: Boca Raton, FL, USA, 2007. [\[CrossRef\]](https://doi.org/10.1201/9781420005523)
- 173. Ataie, A.; Sabetkasaei, M.; Haghparast, A.; Moghaddam, A.H.; Kazeminejad, B. Neuroprotective effects of the polyphenolic antioxidant agent, Curcumin, against homocysteine-induced cognitive impairment and oxidative stress in the rat. *Pharmacol. Biochem. Behav.* **2010**, *96*, 378–385. [\[CrossRef\]](https://doi.org/10.1016/j.pbb.2010.06.009)
- 174. Sarker, M.R.; Franks, S.F. Efficacy of curcumin for age-associated cognitive decline: A narrative review of preclinical and clinical studies. *GeroScience* **2018**, *40*, 73–95. [\[CrossRef\]](https://doi.org/10.1007/s11357-018-0017-z) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29679204)
- 175. He, L.-F.; Chen, H.-J.; Qian, L.-H.; Chen, G.-Y.; Buzby, J.S. Curcumin protects pre-oligodendrocytes from activated microglia in vitro and in vivo. *Brain Res.* **2010**, *1339*, 60–69. [\[CrossRef\]](https://doi.org/10.1016/j.brainres.2010.04.014) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/20403340)
- 176. Tavakol, S.; Zare, S.; Hoveizi, E.; Tavakol, B.; Rezayat, S.M. The impact of the particle size of curcumin nanocarriers and the ethanol on beta_1-integrin overexpression in fibroblasts: A regenerative pharmaceutical approach in skin repair and anti-aging formulations. *DARU J. Pharm. Sci.* **2019**, *27*, 159–168. [\[CrossRef\]](https://doi.org/10.1007/s40199-019-00258-3) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30875026)
- 177. Cianciulli, A.; Calvello, R.; Ruggiero, M.; Panaro, M.A. Inflammaging and Brain: Curcumin and Its Beneficial Potential as Regulator of Microglia Activation. *Molecules* **2022**, *27*, 341. [\[CrossRef\]](https://doi.org/10.3390/molecules27020341)
- 178. Parikh, A.; Kathawala, K.; Li, J.; Chen, C.; Shan, Z.; Cao, X.; Zhou, X.-F.; Garg, S.; Parikh, A.; Kathawala, K.; et al. Curcumin-loaded self-nanomicellizing solid dispersion system: Part II: In vivo safety and efficacy assessment against behavior deficit in Alzheimer disease. *Drug Deliv. Transl. Res.* **2018**, *8*, 1406–1420. [\[CrossRef\]](https://doi.org/10.1007/s13346-018-0570-0)
- 179. Calabrese, V.; Santoro, A.; Monti, D.; Crupi, R.; di Paola, R.; Latteri, S.; Cuzzocrea, S.; Zappia, M.; Giordano, J.; Calabrese, E.J.; et al. Aging and Parkinson's Disease: Inflammaging, neuroinflammation and biological remodeling as key factors in pathogenesis. *Free Radic. Biol. Med.* **2018**, *115*, 80–91. [\[CrossRef\]](https://doi.org/10.1016/j.freeradbiomed.2017.10.379)

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