

Review

The Key Nutrients in the Mediterranean Diet and Their Effects in Inflammatory Bowel Disease: A Narrative Review

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Abstract: The gut microbiome, a collection of gut microorganisms, is crucial in the development and progression of inflammatory bowel diseases (IBD). Therefore, diet and dietary interventions are promising strategies to shape the gut microbiota for IBD management. Of all the diets studied in the IBD field, the Mediterranean diet has the least restrictive nature, promoting long-term adherence. The Mediterranean diet is rich in plants, with a high daily intake of fruits and vegetables (high in fiber, antioxidants, and vitamins), olive oil, whole grains, legumes, and nuts. It includes the moderate consumption of animal products such as oily fish (rich in mono- and polyunsaturated fatty acids), dairy products, and poultry, with a limited intake of red meat and processed foods. This diet is associated with a decreased risk of chronic diseases, including IBD. However, the mechanisms of specific nutrients behind these effects in the Mediterranean diet remain under investigation. Therefore, in this review, we aim to provide an overview of the nutrients that are abundant in the Mediterranean diet and their effects on IBD, with a main focus on preclinical evidence. While several nutrients like fructo-oligosaccharide, chitosan, plant-derived protein, polyphenols, omega-3 polyunsaturated fatty acids, and resveratrol have shown potential beneficial effects in preclinical models, clinical evidence is often limited. However, understanding the complex interactions between specific nutrients and IBD is essential to developing a tailored, multidisciplinary, and personalized approach for disease management; therefore, further research is required.

Keywords: Mediterranean diet; nutrients; inflammatory bowel disease; IBD

1. Introduction

The gut microbiome, which is a collection of microorganisms in the human gut, plays a critical role in the pathogenesis of inflammatory bowel disease (IBD), including ulcerative colitis (UC) and Crohn's disease (CD) [\[1](#page-11-0)[,2\]](#page-11-1). Alterations in the composition and function of the microbiome might lead to inflammation and damage to the intestinal wall [\[3\]](#page-11-2), which can trigger symptoms such as abdominal pain, bloody diarrhea, and weight loss [\[4\]](#page-11-3). Diet and dietary interventions have, therefore, emerged as a promising strategy to modulate the gut microbiota in IBD [\[5–](#page-12-0)[7\]](#page-12-1), especially since many nutritional deficiencies (e.g., folate/vitamin B9) have been observed in IBD [\[8\]](#page-12-2).

The traditional Mediterranean diet is defined as a diet high in fiber and low in saturated/animal fat, with a high intake of fruits, vegetables, whole grains, legumes, nuts, olive oil, and limited consumption of meat and ultra-processed foods [\[9\]](#page-12-3). This type of diet has been shown to be associated with the pro- or anti-inflammatory characteristics of the gut microbiome [\[10\]](#page-12-4) and has been associated with a decreased risk of late-onset

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CD [\[11\]](#page-12-5). In this regard, the consumption of nuts, oily fish, fruit, vegetables, and cereals was associated with a higher abundance of beneficial short-chain fatty acid (SCFA)-producing gut microbiota [\[10\]](#page-12-4). Conversely, Western diets high in ultra-processed foods have been associated with negative impacts on IBD risk by affecting the intestinal barrier and altering the composition of the gut microbiota, as detailed by Vissers et al. (2022) [\[12\]](#page-12-6). A recent review on the clinical effects of the Mediterranean diet in IBD patients concluded that most clinical evidence supports this diet as an approach for IBD prevention and management [\[9\]](#page-12-3). In comparison to other diets investigated for IBD, the Mediterranean diet is less restrictive and may, therefore, allow greater long-term adherence in patients [\[13](#page-12-7)[,14\]](#page-12-8). Additionally, the Mediterranean diet is hypothesized to be beneficial by improving the lipid profile, protecting against oxidative stress, inflammation, and platelet aggregation, modifying the hormones and growth factors involved in diseases such as cancer, inhibiting the nutrientsensing pathways, and finally, modifying the gut microbiota composition [\[15\]](#page-12-9). For example, adherence to a Mediterranean diet was shown to improve clinical scores and inflammatory markers in children [\[16\]](#page-12-10). Therefore, the latest AGA Clinical Practice Update on diet and nutritional therapies in IBD patients states that unless there is a contraindication, all patients should be advised to follow a Mediterranean diet [\[8\]](#page-12-2). Importantly, the traditional Mediterranean diet is more than just a dietary regimen; it is a lifestyle that emphasizes balanced nutrition, regular physical activity, and social connections, which might, together, contribute to its health benefits.

Nevertheless, the exact mechanisms behind the effects of the Mediterranean diet on the gut microbiome in IBD remain under investigation [\[9\]](#page-12-3). Therefore, in this review, we aim to provide an overview of the nutrients present in the Mediterranean diet and their effects on IBD, with a main focus on recently obtained preclinical data; when available, clinical evidence is also pinpointed.

2. Methods

Data for this review were identified by searches of PubMed and references from relevant articles using the search terms "Mediterranean diet", "carbohydrates", "fatty acids", "olive oil", "wine", "protein", "fat", "nuts", "polyphenols", "spices", "herbs", "food additives", "gut microbiome", "preclinical", "in vitro", "in vivo", "inflammatory bowel disease" ("IBD"), and any combinations of these terms. Recent articles published in peerreviewed journals in English between 2019 and 2024 were included, and earlier articles were also considered when highly relevant (>100 citations), e.g., meta-analyses. Very limited clinical evidence is currently available on the effects of nutrients in the Mediterranean diet; therefore, we choose to focus on preclinical data here, although, when available, clinical data are discussed for translational reasons.

3. Carbohydrates

Carbohydrates, including inulin, galacto-oligosaccharides (GOS), fructo-oligosaccharides (FOS), lactulose, derivatives of galactose, and β -glucans [\[17\]](#page-12-11) have been studied for their prebiotic capacity in IBD [\[18\]](#page-12-12). Prebiotics are non-digestible food components that selectively stimulate the growth or activity of desirable microorganisms, like *Bifidobacteria* spp. and *Lactobacilli* spp., and are hypothesized to positively impact the gut microbiome and alleviate IBD symptoms [\[17](#page-12-11)[,19\]](#page-12-13). Despite these promising prospects, a well-powered placebocontrolled trial of FOS (15 g/day) showed no clinical effect in active CD patients [\[20\]](#page-12-14). However, a recent pilot trial evaluating oligofructose/inulin supplementation (15 g/day) in inactive CD patients and their healthy siblings [\[21\]](#page-12-15) showed more pronounced effects in the healthy siblings, including decreased blood $CD³⁺$ T cells. Nonetheless, improved intestinal permeability was observed in the patients [\[21\]](#page-12-15). Furthermore, both patients and siblings showed increased levels of total *Bifidobacteria* spp. and *Bifidobacterium longum* [\[21\]](#page-12-15). In siblings, there were additional increases in *Bifidobacterium adolescentis* and *Roseburia* spp., which were not observed in the patients [\[21\]](#page-12-15). Conversely, a pilot trial evaluating different levels of carbohydrate-restrictive diets positively impacted symptoms and reduced inflammation in pediatric CD patients, with more restrictive diets showing a greater improvement in resolving inflammation [\[22\]](#page-12-16). Nevertheless, a specific carbohydrate diet was not more effective than the Mediterranean diet in achieving symptomatic remission, fecal calprotectin, and the CRP (C-reactive protein) response in adult CD patients [\[13\]](#page-12-7). In the case of UC, prebiotic GOS administration (2.8 g/day) did not result in improved clinical outcomes. However, it did normalize the gut microbiota in milder patients by increasing the levels of *Bifidobacterium* spp. and *Christensenellaceae* spp. [\[23\]](#page-12-17). Nevertheless, these findings require validation in randomized controlled trials.

Furthermore, FOS (2 g per kg/day) has been shown to alleviate the pathological immune response and prevent intestinal barrier impairment in an acute mouse model of colitis [\[24\]](#page-12-18). Additionally, various native starches derived from, e.g., potatoes, peas [\[25\]](#page-12-19), and polysaccharides from the fruits of *Lycium barbarum* L. (200 mg/kg body weight/day) have been demonstrated to reduce disease activity and histologic damage in mouse models of colitis, as well as to support gut microbiota homeostasis by inhibiting rather pathogenic bacteria [\[26\]](#page-12-20). Pectic polysaccharides from guavira pomace, a co-product from the fruit pulp industry, have been shown to reverse increased permeability and transepithelial electrical resistance, potentially offering therapeutic benefits for treating abdominal pain and UC [\[27\]](#page-13-0). Furthermore, fruits like cranberries (30 g/day of freeze-dried cranberry powder) have been shown to attenuate changes in microbiota composition and functionality induced by an animal-based diet in healthy adults [\[28\]](#page-13-1). Glycolipids derived from tilapia heads have also been shown to reduce inflammation, similar to sulfasalazine controls, while regulating the gut microbiota composition by increasing beneficial bacteria and decreasing harmful ones [\[29\]](#page-13-2).

Chitosan, an amino-polysaccharide derived from chitin, which can be obtained from the shells of crustaceans, fish scales, insects, and fungi, has attracted specific interest [\[30\]](#page-13-3). Its non-toxic nature, mucoadhesive properties, and ability to load and transport drugs make it an ideal biopolymer for developing transmucosal drug delivery systems [\[30\]](#page-13-3). Indeed, chitosan has been shown to have intestinal barrier-enhancing properties, as well as microbiota-modulatory properties, both in vivo (150 mg/kg) and in vitro $[31,32]$ $[31,32]$. Unlike commercially available gastro-resistant polymers, chitosan possesses the "generally regarded as safe" (GRAS) status, appointing it as a potentially safe option for long-term therapy in chronic diseases like IBD [\[33–](#page-13-6)[35\]](#page-13-7).

While dietary fiber is beneficial for those with normal microbial fermentative capacity, due to the production of short-chain fatty acids, certain patients with active IBD may experience negative effects from certain dietary fibers, due to decreased fermentative microbial activity [\[36\]](#page-13-8). The response to dietary fiber in IBD varies significantly and is influenced by individual differences in baseline gut microbiome composition and disease activity, which are shaped by dietary habits [\[20](#page-12-14)[,37–](#page-13-9)[39\]](#page-13-10). Therefore, a combination of dietary carbohydrates or prebiotics with probiotics (synbiotics) provides a potential solution to this issue [\[17](#page-12-11)[,40\]](#page-13-11).

4. Proteins

Historically, proteins have been evaluated primarily for their ability to supply essential amino acids and support protein synthesis [\[41\]](#page-13-12). Animal-derived proteins, particularly those found in red and processed meats, have been associated with increased inflammation and the exacerbation of IBD symptoms in animal models, whereas plant-derived proteins exhibit anti-inflammatory properties that can help mitigate disease activity [\[42,](#page-13-13)[43\]](#page-13-14). In humans, a diet low in red and processed meat may reduce flare-ups of UC but has not shown effectiveness in preventing relapses of CD [\[44,](#page-13-15)[45\]](#page-13-16). Other protein-rich components of the Mediterranean diet are dairy products, including yogurt and, increasingly, also kefir. The latter, being a fermented product rich in *Lactobacillus* spp., has especially gained increasing attention in IBD. In a rat model, a dose-dependent effect has been observed, where moderate kefir administration (PBS/10% kefir) provided better outcomes than higher doses (PBS/30% kefir) [\[46\]](#page-13-17). In humans, a randomized clinical trial found that

kefir supplementation (400 mL/day) modulated the gut microbiota and improved shortterm quality of life for IBD patients [\[47\]](#page-13-18). Probiotic Lactobacilli from kefir were shown to downregulate T cells in the inflammatory lamina propria of active IBD patients, suggesting that the microbial content of kefir may play a significant role [\[48\]](#page-13-19). However, animalderived proteins from fish, poultry, and dairy products are essential components of the Mediterranean diet, providing necessary amino acids, vitamin B12, and heme iron, which are crucial for metabolic processes and maintaining muscle mass [\[15\]](#page-12-9).

In addition to animal-derived proteins, plant-derived proteins are well-represented in the Mediterranean diet and are found in legumes, nuts, and seeds, which are consumed daily [\[15\]](#page-12-9). These plant proteins can significantly increase anti-inflammatory butyrateproducing bacteria, enhance bacterial diversity, and reduce pro-inflammatory bacteria [\[49\]](#page-13-20). Specifically, a meta-analysis identified high protein intake as a risk factor for IBD in an Asian population, potentially due to a dietary shift away from traditional plant-based patterns [\[43\]](#page-13-14).

Certain amino acids, such as leucine, isoleucine, valine, glutamine, and glutamate (all at 3 g/L), have been selectively utilized in colonic microbiota studies in piglets, resulting in the production of health-promoting short-chain fatty acids (acetate and propionate), highlighting their prebiotic potential [\[50\]](#page-13-21). Specific amino acids, particularly tryptophan, are being intensively studied for their role in IBD pathophysiology [\[51\]](#page-13-22). Alterations in tryptophan metabolism have been linked to dysregulated immune responses in chronic inflammatory conditions like IBD [\[51\]](#page-13-22). Tryptophan can be converted into indole metabolites by the gut microbiota; these promote epithelial barrier functions and exert anti-inflammatory effects [\[52\]](#page-13-23). For example, a diet low in fat and high in fiber has been shown to be associated with a significant increase in tryptophan concentrations in UC patients [\[53\]](#page-14-0). Likewise, a pilot study demonstrated shifts in microbiota composition and increased tryptophan metabolites after four days of the Mediterranean diet [\[54\]](#page-14-1). Although the exact mechanisms remain unclear, it is hypothesized to be mediated by metabolic interactions in the gut microbiota [\[55\]](#page-14-2). In addition, both in vivo work and human data have indicated a role for tryptophan metabolite supplementation, as xanthurenic (400 mg/kg daily) and kynurenic acids (300 mg/kg daily) could decrease colitis severity [\[56\]](#page-14-3). Additionally, manipulating the endogenous tryptophan metabolism with recombinant aminoadipate aminotransferase showed protective effects [\[56\]](#page-14-3). However, a clinical trial on 5-hydroxytryptophan supplementation (50 to 3000 mg/day) showed no improvement in fatigue for patients in remission, indicating that further research is needed [\[57\]](#page-14-4). Targeting tryptophan metabolism remains a promising strategy for managing inflammation and restoring the immune balance in IBD patients.

5. Fats

High-fat diets, particularly those rich in saturated fats, such as the Western diet, have been linked to increased intestinal inflammation and the exacerbation of IBD symptoms in animal models by causing a dysbalanced gut microbiome, or dysbiosis and impaired intestinal barrier integrity [\[58](#page-14-5)[,59\]](#page-14-6). In contrast, the Mediterranean diet is low in saturated fat, although higher in unsaturated fats, and a fat blend based on this diet was shown to protect against colitis development in mucin-2 (MUC2)-deficient mice [\[60\]](#page-14-7). Specific dietary interventions that are low in fat and high in fiber were shown to reduce levels of inflammatory markers and increase quality of life in UC [\[53\]](#page-14-0), and to induce a rapid response in active pediatric CD patients [\[61\]](#page-14-8). These studies indicate a potential role for dietary fat in IBD, depending on its composition.

A major source of fat in the Mediterranean diet is olive oil [\[62](#page-14-9)[,63\]](#page-14-10), which has a high content of MUFA (65–83%), particularly oleic acid, along with some polyunsaturated fatty acids (PUFAs) such as linoleic acid. This lipid profile has been associated with protective effects against coronary, autoimmune, and inflammatory disorders [\[64\]](#page-14-11). Moreover, olive oil is considered to be rich in polyphenols, which contribute to its antioxidant and antiinflammatory properties, further enhancing its potential health benefits. Several studies

have investigated the supplementation of olive oil, of which an extensive overview is given by Vrdoljak et al. (2022) [\[62\]](#page-14-9). In summary, olive oil is an excellent dietary compound that may support traditional IBD drugs and assist patients in controlling their condition [\[62\]](#page-14-9). However, not all results are positive and seem to be dependent on variations in composition, especially with regard to polyphenol content (Table [1\)](#page-4-0). For example, some researchers increased the polyphenolic content by using extra-virgin olive oil that was enhanced by its unsaponifiable fraction and enriched with specific polyphenols like hydroxytyrosol, which may have contributed to its effectiveness [\[62,](#page-14-9)[65,](#page-14-12)[66\]](#page-14-13). Of note, the polyphenol content in extra-virgin olive oil can vary significantly, depending on the olive cultivar, the abundance of secoiridoids and their derivatives, and the oil production and storage processes [\[67\]](#page-14-14). Indeed, a recent preclinical study investigated the effects of using extra virgin olive oils from four Italian cultivars known for their high polyphenolic content on mouse models. The study found that using these oils helped to reduce rectal bleeding and improve intestinal permeability. Additionally, the study observed a decrease in the expression of pro-inflammatory cytokines and an improvement in the histopathological characteristics of inflammation [\[68\]](#page-14-15). However, differences between cultivars were observed; therefore, a detailed characterization of monocultivar extra-virgin olive oils might be useful for a more targeted nutraceutical choice [\[68\]](#page-14-15). Additionally, oleuropein appears to be capable of downregulating inflammatory cytokines such as interleukin-6 (IL-6), metalloprotease secretion, and COX-2 [\[69\]](#page-14-16). More recently, protective effects were observed with synbiotic treatment in an in vivo UC model comprising *Lactobacillus plantarum SC-5* and tyrosol extracted from olive oil [\[70\]](#page-14-17). In addition, hydroxytyrosol was shown to counteract ERK1/2 and mTOR activation, pro-inflammatory cytokine release, and autophagy in primary human colonic cells, and was, therefore, hypothesized to be the most important anti-inflammatory and antioxidant compound present in olive oil [\[71\]](#page-14-18). Nevertheless, very limited human studies are available; thus, large, multicentric, randomized control trials evaluating the effects of olive oil in modulating the course of IBD are required.

Table 1. Use of extra-virgin olive oil or its components in preclinical IBD models over the past five years.

UC: ulcerative colitis; DAI: disease activity index; EVO: Extra Virgin olive; NA: Not applicable.

Furthermore, certain fatty acids, such as omega-3 polyunsaturated fatty acids (PUFAs) originating from the consumption of fish [\[72\]](#page-14-19) and from plant sources, including seeds, have been shown to possess anti-inflammatory properties; therefore, they may alleviate IBD symptoms by modulating immune responses and reducing pro-inflammatory cytokine production [\[73\]](#page-14-20). The rectal administration of fish oil (2 mL), known to be high in omega-3 PUFAs, has been shown to have beneficial effects that were mutualistic to

mesalamine treatment [\[74\]](#page-14-21). Additionally, eicosapentaenoic acid (500 mg, twice daily) was shown to reduce fecal calprotectin and prevent relapses in UC [\[75\]](#page-15-0). This was confirmed by an evaluation of tuna oil in preclinical UC models [\[76\]](#page-15-1). Moreover, marine phospholipid nanoliposomes combining omega-3 PUFAs with phosphatidylcholine have been shown to be preferentially distributed in the inflamed gut and to delay disease onset upon prophylactic administration in an in vivo colitis model [\[77\]](#page-15-2). In humans, the effects of omega 3 on IBD are still uncertain. While some studies suggest the anti-inflammatory effects of omega-3 fatty acids in IBD [\[78,](#page-15-3)[79\]](#page-15-4), a recent meta-analysis of randomized controlled trials concluded that PUFA supplementation has little or no effect on the prevention or treatment of IBD [\[80\]](#page-15-5). Furthermore, it has been shown that an excess of PUFAs in the diet may even compromise gut health by disrupting epithelial stress responses [\[81\]](#page-15-6). Consequently, more studies are needed; in the meantime, the moderate supplementation of fish oil, algae, and their derivatives could be considered [\[73\]](#page-14-20). Additionally, nuts are often consumed in the Mediterranean diet and are an important source of sterols. Sterols have previously been shown to have anti-inflammatory and antioxidant properties, which may reduce inflammation in IBD patients [\[82–](#page-15-7)[84\]](#page-15-8). For example, studies have demonstrated that in those with chronic inflammatory disorders, nut intake is linked to lower levels of inflammatory markers like CRP and IL-6 [\[85\]](#page-15-9). Additionally, the polyphenolic [\[86](#page-15-10)[–89\]](#page-15-11) (flavonoids) and phenolic acids [\[90](#page-15-12)[–93\]](#page-15-13) in nuts enhance their antioxidant activity, next to having barrier protection and microbiota modulatory potential.

Elsewhere, seafood and nuts have also been associated with IBD development in another case-control study [\[94\]](#page-15-14). Additionally, lower uptake of nuts, seeds, and yogurt was observed pre- and post-IBD diagnosis in a retrospective cohort study [\[95\]](#page-15-15). Therefore, understanding the intricate relationship between fat, its metabolism, and IBD in clinical and preclinical models is crucial for developing targeted dietary interventions and therapeutic strategies to effectively manage these chronic inflammatory disorders.

6. Red Wine

Red wine is moderately consumed in the Mediterranean diet and both clinical and preclinical studies have suggested that moderate consumption of red wine may confer protective benefits against the development and progression of IBD [\[96](#page-15-16)[–100\]](#page-16-0). Importantly, some clinical studies have reported associations between moderate red wine consumption and a reduced risk of IBD incidence and severity, possibly attributable to its anti-inflammatory and antioxidant effects [\[101\]](#page-16-1). The first compound of red wine that has previously been studied is resveratrol. Preclinical studies have demonstrated that resveratrol (1 mg/kg/day) may possess anti-inflammatory properties, modulate immune responses, and reduce oxidative stress, and would thus be able to attenuate intestinal inflammation [\[96](#page-15-16)[,97\]](#page-15-17). In addition, resveratrol administration led to increased levels of *Lactobacilli* and *Bifidobacteria*, as well as decreased levels of *Enterobacteria* in a disease model [\[97\]](#page-15-17). A second compound of interest is tannin, which is highly present in red wine. Limited clinical research has been performed in the framework of IBD, although tannins have been studied in relation to the progression of dextran sodium sulfate (DSS)-induced colitis in a preclinical setting [\[98\]](#page-16-2). In these experiments, a correlation between the quantity of tannins $(0, 10, 50, \text{ and } 250 \text{ mg/kg})$ and the outcomes of DSS-induced colitis was observed [\[98\]](#page-16-2). In addition, their effects were shown to be mediated by tannin-induced enrichment of the microbial metabolite, gallic acid [\[98\]](#page-16-2). Although resveratrol and tannins might have beneficial effects, it is essential to note that excessive alcohol consumption, including red wine, may exacerbate IBD symptoms and increase the disease severity [\[101\]](#page-16-1). Furthermore, it is uncertain whether these positive effects can be achieved by drinking wine alone, as the beverage only contains limited concentrations of these compounds [\[102\]](#page-16-3). As a result, further research is required to elucidate the precise mechanisms and to establish optimal and potential health-benefitting consumption patterns.

7. Other Specific Nutrients and Vitamins

Consumption of a wide variety of plant-derived components in the Mediterranean diet promotes exposure to bioactive molecules that can modify inflammation (Table [2\)](#page-6-0).

Quercetin is a plant pigment and a potent antioxidative flavonoid that is found mostly in onions, grapes, berries, cherries, broccoli, and citrus fruits [\[103\]](#page-16-4). Quercetin intake in a typical Mediterranean diet may vary, but estimates suggest that it could range from 10 to 20 mg per day from foods alone [\[104\]](#page-16-5). It is a versatile antioxidant that has been shown to possess protective abilities against the tissue injury induced by various drug toxicities, together with anti-cancer, anti-ulcer, anti-diabetic, and anti-hypertensive properties [\[105\]](#page-16-6). From animal studies, it emerges that quercetin (100–1500 ppm in diet) helps to protect the enterocytes from apoptosis caused by oxidative stress, which, in turn, helps provide protection against colitis [\[106,](#page-16-7)[107\]](#page-16-8). Moreover, it increases intestinal microbial diversity by promoting the growth of *Bacteroides* spp., *Bifidobacterium* spp., *Lactobacillus* spp., and *Clostridium* spp., while reducing the levels of *Fusobacterium* spp. and *Enterococcus* spp. [\[108\]](#page-16-9). Additionally, quercetin seems to reduce intestinal permeability by enhancing the expression of tight junctions [\[107](#page-16-8)[,109\]](#page-16-10). It also promotes the proliferation of intestinal cells and supports the regenerative capacity of intestinal mesenchymal stem cells from animals [\[107,](#page-16-8)[109\]](#page-16-10). In addition, this component also intervenes in the regulation of the immune system, reducing the infiltration of neutrophils and macrophages into the colonic tissue of mice. Furthermore, it inhibits chronic intestinal inflammation by reducing macrophage infiltration by several pro-inflammatory cytokines, such as TNF-α, IL-1β, IL-6, and IL-17. Finally, it promotes antiinflammatory IL-10 secretion in the colonic tissues [\[110\]](#page-16-11). In summary, quercetin might be useful in IBD management [\[111](#page-16-12)[,112\]](#page-16-13). However, its low water solubility and bioavailability have been a significant challenge [\[113\]](#page-16-14). To address this issue, various strategies are actively being investigated to improve its absorption. For example, recent studies have demonstrated remarkable success in using quercetin nanoparticles [\[114](#page-16-15)[–116\]](#page-16-16). These nanoparticles have shown regulatory effects on the gut microbiota and their metabolites like SCFAs, resulting in a significant reduction of inflammatory infiltration of the colon [\[114](#page-16-15)[–116\]](#page-16-16).

Table 2. Overview of the discussed nutrients or compounds, their subtype, and in which foods they are present.

Another pigment, astaxanthin, is a potent antioxidant belonging to the carotenoid family. It is primarily sourced from microalgae, salmon, shrimp, and krill, and possesses remarkable anti-inflammatory properties attributable to its ability to scavenge free radicals, inhibit oxidative stress, and modulate immune responses [\[117\]](#page-16-17). The Mediterranean diet encourages the consumption of about three servings of fish or seafood per week, which could translate into an estimated intake of 1–5 mg of astaxanthin per week [\[122\]](#page-16-22). In preclinical in vivo studies [\[123](#page-17-0)[–129\]](#page-17-1), astaxanthin has demonstrated its efficacy in attenuating inflammation, preserving mucosal integrity, and regulating the expression of pro-inflammatory cytokines and mediators implicated in IBD pathogenesis. For example, the decreased expression of IL-1β, IL-6, TNF- α , IL-36 α , and IL-36 γ has been observed after treatment with 4 mg astaxanthin in nanocarriers [\[123\]](#page-17-0). Clinical investigations evaluating astaxanthin supplementation in other diseases like type 2 diabetes, arthritis, and endometriosis have shown promising results in modulating the observed inflammation and reducing disease severity [\[130–](#page-17-2)[132\]](#page-17-3). Although further research in the framework of IBD is still required, these findings may support the potential application of diets rich in astaxanthin, like the Mediterranean diet.

The carotenoid lycopene, which is mostly present in tomatoes and other red fruits, has previously been shown to be involved in reducing inflammation [\[118\]](#page-16-18). Lycopene may scavenge reactive oxygen species, suppress pro-inflammatory cytokines, and modify the signaling pathways linked to immune responses [\[118\]](#page-16-18). Furthermore, lycopene shows promise in lowering oxidative stress and maintaining intestinal barrier integrity [\[118\]](#page-16-18), two important aspects of IBD pathogenesis and the reason it gained interest as a dietary supplement. The average dose of lycopene in the Mediterranean diet is difficult to quantify, considering the high variability of plant products; however, based on typical daily consumption patterns, the Mediterranean diet may provide approximately 6–15 mg of lycopene [\[133\]](#page-17-4). Research employing animal models has shown positive findings regarding lycopene's (12 mg/kg) ability to reduce inflammation and disease severity by improving epithelial barrier functions and inhibiting *E. coli* adhesion [\[134](#page-17-5)[,135\]](#page-17-6). However, further research is mandatory; Mendelian randomization studies have suggested that certain circulating antioxidants, minerals, and vitamins might also be associated with IBD development [\[136\]](#page-17-7).

Although not part of the traditional Mediterranean diet, turmeric's bioactive ingredient, curcumin, is increasingly used in the modern Mediterranean kitchen and has gained interest due to its possible beneficial effects for different diseases, including IBD [\[137\]](#page-17-8). Curcumin's anti-inflammatory effects are mediated via a number of pathways, including immune cell function regulation, the inhibition of inflammatory cytokines, and suppression of the NF-κB signaling pathway [\[138,](#page-17-9)[139\]](#page-17-10). Furthermore, curcumin possesses antioxidant properties by scavenging free radicals and lowering oxidative stress—two important aspects of the pathophysiology of IBD [\[138\]](#page-17-9). Preclinical research employing animal models of IBD has shown encouraging findings, suggesting that curcumin supplementation (100 mg/kg) may alleviate colitis and improve dysbiosis [\[137](#page-17-8)[,140\]](#page-17-11). Curcumin's clinical translation is hampered by certain issues, including limited bioavailability and fast metabolism, despite its promising medicinal promise [\[141\]](#page-17-12). Several strategies, including combination therapy and nano-formulations, are being investigated to improve curcumin's bioavailability and effectiveness for improved IBD management [\[141\]](#page-17-12). For example, a pilot trial showed positive results, including clinical and endoscopic efficacy and safety for IBD, when assessing Theracurmin[®], a nanoparticle-based drug delivery system with increased bioavailability [\[141\]](#page-17-12). However, larger studies are required to confirm these findings.

Green tea is often added to the Mediterranean diet and contains high levels of the polyphenol epigallocatechin gallate (ECGC) [\[119\]](#page-16-19). ECGC has been shown to be able to improve lipid metabolism and to reduce inflammation and oxidative stress in an in vivo model of obesity [\[119\]](#page-16-19). In addition, decreased inflammation has also been observed in multiple sclerosis and type 2 diabetes upon supplementation with ECGC [\[142,](#page-17-13)[143\]](#page-18-0). Nevertheless, these effects remain to be evaluated in IBD.

Vitamin D or calciferol deficiency is a common issue in IBD due to several reasons, such as immunosuppressive treatments affecting sun exposure, dietary restrictions, and impaired absorption [\[8](#page-12-2)[,144\]](#page-18-1). Besides playing a critical role in bone and muscle health, vitamin D also supports the integrity of the gut epithelium, innate immune barrier function, and eubiosis [\[144,](#page-18-1)[145\]](#page-18-2). Vitamin D intake was positively associated with greater adherence to the Mediterranean diet [\[146\]](#page-18-3). This result may seem surprising, given that vitamin Drich foods, like cheese, beef liver, egg yolk, tuna, mackerel, and salmon, are not typical compounds of the Mediterranean diet per se, except for some specific cheeses and fish [\[147\]](#page-18-4). Indeed, the Mediterranean diet does not inherently provide high levels of vitamin D [\[121\]](#page-16-21). However, this diet enhances the body's ability to absorb vitamin D effectively, which could be mediated by the gut microbiome [\[121\]](#page-16-21). This has been confirmed in a cross-sectional study in which adherence to the Mediterranean diet for a year could increase vitamin D levels [\[148\]](#page-18-5). Specifically, a Mediterranean diet intervention appeared to have a greater impact on the bacterial populations of the main phyla of Firmicutes and Proteobacteria, with Firmicutes increasing and Proteobacteria decreasing [\[148\]](#page-18-5). Unfortunately, the majority of clinical intervention studies supplementing vitamin D have failed [\[149\]](#page-18-6), indicating that there might be other interactions of importance; therefore, further research is required.

8. Multiple Nutrients United in One Diet

A Mediterranean dietary pattern is rich in various potential beneficial compounds, which collectively exert anti-inflammatory, antioxidant, and gut microbiota-modulating effects in preclinical research [\[9\]](#page-12-3) and are discussed throughout this review. Nevertheless, it has to be noted that understanding the role of a specific class of compounds in foods that promote health is challenging, due to chemically complex food matrices. Often, the individual compounds tested in clinical trials lack sufficient effectiveness or yield negative results. This could potentially be explained by the lack of preclinical dose optimization and potential discrepancies between the evaluated doses and their actual presence in the Mediterranean diet, leading to over- or underdosing. Nevertheless, all these compounds are united in the Mediterranean diet, in which the different nutrients might exert synergistic effects. Therefore, several clinical studies have been performed, aiming to evaluate the effects of this diet in clinical trials, which have extensively been reviewed by Godny and Dotan (2024) [\[9\]](#page-12-3). In summary, implementation of the Mediterranean diet has been associated with beneficial results in IBD, especially in contrast to Western diets. Epidemiological evidence suggests a potential association between the Mediterranean diet and reduced CD risk. Furthermore, adherence to this diet has shown moderate improvements in clinical and inflammatory markers among active CD patients and has helped maintain low fecal calprotectin levels in patients with quiescent UC [\[13\]](#page-12-7). Mechanistically, the Mediterranean diet fosters the proliferation of beneficial gut bacteria and enhances microbial community diversity.

9. Concluding Discussion and Future Perspectives

Adherence to the Mediterranean diet may offer a multifaceted approach to managing inflammation in IBD through its diverse array of beneficial compounds, not only by shaping the body's microbial community but also by changing the micro-environment. Therefore, recently published American guidelines suggest that all patients in remission or with mild to moderate symptoms should be advised to follow a Mediterranean diet unless there would be any contraindication [\[8\]](#page-12-2). Moreover, additives such as emulsifiers are relatively limited in abundance in the Mediterranean diet, which, in turn, is beneficial since emulsifiers have been associated with IBD progression [\[12](#page-12-6)[,150\]](#page-18-7). Thus, along with many potentially beneficial compounds and nutrients, the Mediterranean diet contains limited amounts of harmful ones. Overall, strong clinical evidence provided by randomized controlled trials is lacking, and there is also a scarcity of information on the small intestines. Moreover, although the observed preclinical effects might be influenced by changes in microbial composition, these changes may subsequently exert indirect effects via SCFA by the inhibition of histone deacetylation (HDAC) [\[151\]](#page-18-8) (Figure [1\)](#page-9-0). The precise mechanisms for most nutrients remain to be elucidated. To bridge these gaps, it is essential to conduct further research to understand the interactions between nutrients and the microbiome in both the small and large intestines, paving the way for more effective clinical interventions.

Figure 1. Overview of the potential mechanisms of the nutrients present in the Mediterranean diet, **Figure 1.** Overview of the potential mechanisms of the nutrients present in the Mediterranean diet, including fiber, essential amino acids such as tryptophan, polyunsaturated fatty acids, and resveratrol. SCFA: short-chain fatty acids; HDAC: histone deacetylation.

 $N_{\rm eff}$ specific dietary compounds such as chief for the en-Nevertheless, specific dietary compounds such as chitosan could be used for the encapsulation of drugs [\[31,](#page-13-4)[35\]](#page-13-7), which might serve as an add-on, for example, for probiotic or oral fecal microbiota delivery, in order to increase its efficacy [\[35,](#page-13-7)[152,](#page-18-9)[153\]](#page-18-10). Moreover, there might be a synergistic effect of IBD therapies and adherence to the Mediterranean diet, which could break the currently observed therapeutic ceiling. Although not part of the daily IBD clinical recommendations, several diets have been evaluated in the framework of fecal microbiota transplantation (FMT). For example, an exclusion diet alone achieved higher remission rates and mucosal healing compared to a single dose of FMT, with or without a supportive exclusion diet [\[6\]](#page-12-21). Additionally, seven weekly FMT sessions, supported with an anti-inflammatory diet, were shown to effectively induce and maintain deep remission compared to standard medical therapy [\[5\]](#page-12-0), which highlights the potential of diet as an add-on strategy in IBD. Contrastingly, diets low in fiber [\[154\]](#page-18-11), like exclusive enteral nutrition (EEN), have been shown to be efficacious in inducing clinical remission in CD, albeit with stronger evidence in children [\[8\]](#page-12-2), and are hypothesized to change the microbial composition [\[155\]](#page-18-12). Although this change is not always observed as being healthier, it may disrupt the established dysbiotic microbial communities, thereby allowing for the re-colonization and establishment of communities that promote a more balanced interaction with the host. In this regard, a Mediterranean lifestyle might be beneficial as a maintenance approach. However, more research is required to evaluate the interactions of a Mediterranean diet or lifestyle and treatment options for the management of IBD.

A major problem in human dietary research is the evaluation of adherence to a specific diet, due to a multitude of available scales, as well as the cumbersome implementation of a true control group, which leads to very heterogeneous results [\[156\]](#page-18-13) and is, therefore, concerning. While the Mediterranean diet offers potential benefits for IBD patients and is relatively less restrictive than other diets, adherence might remain a challenge due to gastrointestinal symptoms [\[9\]](#page-12-3), psychological factors, and a lack of tailored support. Addressing these barriers through education, individualized dietary plans, and supportive interventions is crucial for enhancing adherence and ultimately improving health outcomes for IBD patients. Hence, animal models are perfectly suited for performing dietary intervention studies (Figure [2\)](#page-10-0). In this regard, many in vivo studies have been performed involving Western and regular chow in mice; however, research investigating the Mediterranean diet is rather limited in availability. Nevertheless, to avoid results getting lost in translation from obtained in vivo data to real-world patient information, dietary effects could be evaluated in patient-derived ex vivo models, such as biopsies [\[157\]](#page-18-14) and organoid-derived monolayer cultures [\[158\]](#page-18-15); humanized in vivo models [\[159\]](#page-18-16) or even studies with healthy individuals could be considered for evaluating specific compounds, prior to progressing to trials involving patients. Further research could include diets versus single food-component supplementation to evaluate their specific effects. Specifically, in clinical trials, some foods might be high in a specific compound, but, for standardization purposes, it might be favorable to use a prebiotic compound.

Figure 2. Overview of future perspectives. A major issue associated with dietary interventions is **Figure 2.** Overview of future perspectives. A major issue associated with dietary interventions is adherence to the diet. Therefore, preclinical in vivo trials can aid in evaluating the effects of specific adherence to the diet. Therefore, preclinical in vivo trials can aid in evaluating the effects of specific diets and nutrients. Other approaches include well-designed human trials in which diets and nutrients ents are evaluated, as well as capsules that can be derived from and/or contain beneficial or existing are evaluated, as well as capsules that can be derived from and/or contain beneficial or existing of the intervention, the intervention, the intervention of distribution of α compounds. In addition, the interaction of dietary interventions or nutrients with existing treatments and those in development should be investigated. Finally, the effects of physical activity should also be integrated in future work. The integration of genetic factors, the gut microbiome, clinical parameters, diet, and physical activity using machine learning techniques will lead to personalized nutrition or lifestyle advice in both health and disease.

Beyond dietary composition, the Mediterranean lifestyle is characterized by communal meals, leisurely dining, and physical activity. This active lifestyle has been associated with a reduced risk of chronic diseases such as cardiovascular ailments, metabolic disorders, and certain cancers, as well as improved longevity and overall well-being. A recent clinical trial in pediatric IBD patients has shown that a 12-week lifestyle intervention including physical activity improved fatigue perception, quality of life, and bowel symptoms [\[160\]](#page-18-17). Moreover, a multicenter cross-sectional study identified that IBD and its symptoms severely affect physical activity levels, due to fatigue and the fear of diarrhea [\[161\]](#page-18-18). In addition, physicians reported a lack of knowledge in this field; therefore, work remains to achieve optimization. However, more research is still required to evaluate the effects of physical activity on disease-specific quality of life, biomarkers, and general health status. Overall, applying the Mediterranean lifestyle in IBD serves as a model for promoting health and longevity through a balanced integration of dietary, social, and physical activities, thereby meriting a multidisciplinary management approach.

The Mediterranean lifestyle, especially its diet, is commonly embraced for its health benefits, but these effects may not be suitable for all IBD patients due to the highly variable course of the disease. Given that IBD patients frequently experience dysbiosis [\[2\]](#page-11-1), and the Mediterranean diet's benefits are often mediated through changes in the gut microbiota with a certain baseline presence, a one-size-fits-all approach is inadequate. Specifically, personalized dietary strategies (Figure [2\)](#page-10-0) will be essential in the future [\[161\]](#page-18-18). This approach entails a multi-disciplinary evaluation of patients' genetics, the unique gut microbiome, disease variability, physical activity, and other environmental factors, combined via machine learning techniques to personalize dietary interventions. These interventions aim to treat inflammation, symptoms, and dysbiosis. It is of the utmost importance to identify the effects of food compounds on both the host and microbiome to develop effective treatments. Investigating the complex interactions between lifestyle, nutrients, and IBD will provide an individualized yet multidisciplinary strategy for disease management. Therefore, further research is mandatory, including preclinical work to identify potential beneficial nutrients and optimal doses, followed by robust clinical trials.

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References

- 1. Manichanh, C.; Borruel, N.; Casellas, F.; Guarner, F. The gut microbiota in IBD. *Nat. Rev. Gastroenterol. Hepatol.* **2012**, *9*, 599–608. [\[CrossRef\]](https://doi.org/10.1038/nrgastro.2012.152) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/22907164)
- 2. Sabino, J.; Valles-Colomer, M.; Falony, G.; Kathagen, G.; Caenepeel, C.; Cleynen, I.; van der Merwe, S.; Vermeire, S. Quantitative microbiome profiling disentangles inflammation- and bile duct obstruction-associated microbiota alterations across PSC/IBD diagnoses. *Nat. Microbiol.* **2019**, *4*, 1826–1831.
- 3. Arnauts, K.; Sudhakar, P.; Verstockt, S.; Lapierre, C.; Potche, S.; Caenepeel, C.; Raes, J.; Vermeire, S.; Sabino, J.; Verfaillie, C. Microbiota, not host origin drives ex vivo intestinal epithelial responses. *Gut Microbes* **2022**, *14*, 2089003. [\[CrossRef\]](https://doi.org/10.1080/19490976.2022.2089003) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35758256)
- 4. Ni, J.; Wu, G.D.; Albenberg, L.; Tomov, V.T. Gut microbiota and IBD: Causation or correlation? *Nat. Rev. Gastroenterol. Hepatol.* **2017**, *14*, 573–584. [\[CrossRef\]](https://doi.org/10.1038/nrgastro.2017.88)
- 5. Kedia, S.; Virmani, S.; Vuyyuru, S.K.; Kumar, P.; Kante, B.; Sahu, P.; Kaushal, K.; Farooqui, M.; Singh, M.; Verma, M.; et al. Faecal microbiota transplantation with anti-inflammatory diet (FMT-AID) followed by anti-inflammatory diet alone is effective in inducing and maintaining remission over 1 year in mild to moderate ulcerative colitis: A randomised controlled trial. *Gut* **2022**, *71*, 2401–2413. [\[CrossRef\]](https://doi.org/10.1136/gutjnl-2022-327811)
- 6. Shabat, C.S.; Scaldaferri, F.; Zittan, E.; Hirsch, A.; Mentella, M.C.; Musca, T.; Cohen, N.A.; Ron, Y.; Isakov, N.F.; Pfeffer, J.; et al. Use of Faecal Transplantation with a Novel Diet for Mild to Moderate Active Ulcerative Colitis: The CRAFT UC Randomised Controlled Trial. *J. Crohns Colitis* **2022**, *16*, 369–378. [\[CrossRef\]](https://doi.org/10.1093/ecco-jcc/jjab165)
- 7. Duan, L.; Cheng, S.; Li, L.; Liu, Y.; Wang, D.; Liu, G. Natural Anti-Inflammatory Compounds as Drug Candidates for Inflammatory Bowel Disease. *Front. Pharmacol.* **2021**, *12*, 684486. [\[CrossRef\]](https://doi.org/10.3389/fphar.2021.684486)
- 8. Hashash, J.G.; Elkins, J.; Lewis, J.D.; Binion, D.G. AGA Clinical Practice Update on Diet and Nutritional Therapies in Patients with Inflammatory Bowel Disease: Expert Review. *Gastroenterology* **2024**, *166*, 521–532. [\[CrossRef\]](https://doi.org/10.1053/j.gastro.2023.11.303)
- 9. Godny, L.; Dotan, I. Is the Mediterranean Diet in Inflammatory Bowel Diseases Ready for Prime Time? *J. Can. Assoc. Gastroenterol.* **2024**, *7*, 97–103. [\[CrossRef\]](https://doi.org/10.1093/jcag/gwad041)
- 10. Bolte, L.A.; Vich Vila, A.; Imhann, F.; Collij, V.; Gacesa, R.; Peters, V.; Wijmenga, C.; Kurilshikov, A.; Campmans-Kuijpers, M.J.E.; Fu, J.; et al. Long-term dietary patterns are associated with pro-inflammatory and anti-inflammatory features of the gut microbiome. *Gut* **2021**, *70*, 1287–1298. [\[CrossRef\]](https://doi.org/10.1136/gutjnl-2020-322670)
- 11. Khalili, H.; Håkansson, N.; Chan, S.S.; Chen, Y.; Lochhead, P.; Ludvigsson, J.F.; Chan, A.T.; Hart, A.R.; Olén, O.; Wolk, A. Adherence to a Mediterranean diet is associated with a lower risk of later-onset Crohn's disease: Results from two large prospective cohort studies. *Gut* **2020**, *69*, 1637–1644. [\[CrossRef\]](https://doi.org/10.1136/gutjnl-2019-319505) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31900290)
- 12. Vissers, E.; Wellens, J.; Sabino, J. Ultra-processed foods as a possible culprit for the rising prevalence of inflammatory bowel diseases. *Front. Med.* **2022**, *9*, 1058373. [\[CrossRef\]](https://doi.org/10.3389/fmed.2022.1058373) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36419796)
- 13. Lewis, J.D.; Sandler, R.S.; Brotherton, C.; Brensinger, C.; Li, H.; Kappelman, M.D.; Daniel, S.G.; Bittinger, K.; Albenberg, L.; Valentine, J.F.; et al. A Randomized Trial Comparing the Specific Carbohydrate Diet to a Mediterranean Diet in Adults with Crohn's Disease. *Gastroenterology* **2021**, *161*, 837–852.e9. [\[CrossRef\]](https://doi.org/10.1053/j.gastro.2021.05.047) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34052278)
- 14. Sabino, J.; Lewis, J.D.; Colombel, J.-F. Treating Inflammatory Bowel Disease with Diet: A Taste Test. *Gastroenterology* **2019**, *157*, 295–297. [\[CrossRef\]](https://doi.org/10.1053/j.gastro.2019.06.027) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31254503)
- 15. Guasch-Ferré, M.; Willett, W.C. The Mediterranean diet and health: A comprehensive overview. *J. Intern. Med.* **2021**, *290*, 549–566. [\[CrossRef\]](https://doi.org/10.1111/joim.13333) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34423871)
- 16. El Amrousy, D.; Elashry, H.; Salamah, A.; Maher, S.; Abd-Elsalam, S.M.; Hasan, S. Adherence to the Mediterranean Diet Improved Clinical Scores and Inflammatory Markers in Children with Active Inflammatory Bowel Disease: A Randomized Trial. *J. Inflamm. Res.* **2022**, *15*, 2075–2086. [\[CrossRef\]](https://doi.org/10.2147/JIR.S349502)
- 17. Martyniak, A.; Medyńska-Przęczek, A.; Wędrychowicz, A.; Skoczeń, S.; Tomasik, P.J. Prebiotics, Probiotics, Synbiotics, Paraprobiotics and Postbiotic Compounds in IBD. *Biomolecules* **2021**, *11*, 1903. [\[CrossRef\]](https://doi.org/10.3390/biom11121903)
- 18. Wedlake, L.; Slack, N.; Andreyev, H.J.N.; Whelan, K. Fiber in the Treatment and Maintenance of Inflammatory Bowel Disease. *Inflamm. Bowel Dis.* **2014**, *20*, 576–586. [\[CrossRef\]](https://doi.org/10.1097/01.MIB.0000437984.92565.31)
- 19. Hill, C.; Guarner, F.; Reid, G.; Gibson, G.R.; Merenstein, D.J.; Pot, B.; Morelli, L.; Canani, R.B.; Flint, H.J.; Salminen, S.; et al. Expert consensus document: The international scientific association for probiotics and prebiotics consensus statement on the scope and appropriate use of the term probiotic. *Nat. Rev. Gastroenterol. Hepatol.* **2014**, *11*, 506–514. [\[CrossRef\]](https://doi.org/10.1038/nrgastro.2014.66)
- 20. Benjamin, J.L.; Hedin, C.R.H.; Koutsoumpas, A.; Ng, S.C.; McCarthy, N.E.; Hart, A.L.; Kamm, M.A.; Sanderson, J.D.; Knight, S.C.; Forbes, A.; et al. Randomised, double-blind, placebo-controlled trial of fructo-oligosaccharides in active Crohn's disease. *Gut* **2011**, *60*, 923–929. [\[CrossRef\]](https://doi.org/10.1136/gut.2010.232025)
- 21. Hedin, C.R.; McCarthy, N.E.; Louis, P.; Farquharson, F.M.; McCartney, S.; Stagg, A.J.; Lindsay, J.O.; Whelan, K. Prebiotic fructans have greater impact on luminal microbiology and CD3+ T cells in healthy siblings than patients with Crohn's disease: A pilot study investigating the potential for primary prevention of inflammatory bowel disease. *Clin. Nutr.* **2021**, *40*, 5009–5019. [\[CrossRef\]](https://doi.org/10.1016/j.clnu.2021.05.033) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34364241)
- 22. Suskind, D.L.; Lee, D.; Kim, Y.-M.; Wahbeh, G.; Singh, N.; Braly, K.; Nuding, M.; Nicora, C.D.; Purvine, S.O.; Lipton, M.S.; et al. The Specific Carbohydrate Diet and Diet Modification as Induction Therapy for Pediatric Crohn's Disease: A Randomized Diet Controlled Trial. *Nutrients* **2020**, *12*, 3749. [\[CrossRef\]](https://doi.org/10.3390/nu12123749) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33291229)
- 23. Wilson, B.; Eyice, Ö.; Koumoutsos, I.; Lomer, M.C.; Irving, P.M.; Lindsay, J.O.; Whelan, K. Prebiotic Galactooligosaccharide Supplementation in Adults with Ulcerative Colitis: Exploring the Impact on Peripheral Blood Gene Expression, Gut Microbiota, and Clinical Symptoms. *Nutrients* **2021**, *13*, 3598. [\[CrossRef\]](https://doi.org/10.3390/nu13103598) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34684597)
- 24. Liao, M.; Zhang, Y.; Qiu, Y.; Wu, Z.; Zhong, Z.; Zeng, X.; Zeng, Y.; Xiong, L.; Wen, Y.; Liu, R. Fructooligosaccharide supplementation alleviated the pathological immune response and prevented the impairment of intestinal barrier in DSS-induced acute colitis mice. *Food Funct.* **2021**, *12*, 9844–9854. [\[CrossRef\]](https://doi.org/10.1039/D1FO01147B)
- 25. Xu, Z.; Liu, W.; Zhang, Y.; Zhang, D.; Qiu, B.; Wang, X.; Liu, J.; Liu, L. Therapeutic and Prebiotic Effects of Five Different Native Starches on Dextran Sulfate Sodium-Induced Mice Model of Colonic Colitis. *Mol. Nutr. Food Res.* **2021**, *65*, 2000922. [\[CrossRef\]](https://doi.org/10.1002/mnfr.202000922)
- 26. Zhou, W.; Kan, X.; Chen, G.; Sun, Y.; Ran, L.; Yan, Y.; Mi, J.; Lu, L.; Zeng, X.; Cao, Y. The polysaccharides from the fruits of Lycium barbarum L. modify the gut community profile and alleviate dextran sulfate sodium-induced colitis in mice. *Int. J. Biol. Macromol.* **2022**, *222*, 2244–2257. [\[CrossRef\]](https://doi.org/10.1016/j.ijbiomac.2022.10.015)
- 27. de Oliveira, N.M.T.; da Luz, B.B.; Schneider, V.S.; da Costa Filho, H.B.; de Araujo Sousa, P.S.; de Paula Werner, M.F.; de Souza, M.H.L.P.; Rocha, J.A.; Nicolau, L.A.D.; Cordeiro, L.M.C.; et al. Dietary polysaccharides from guavira pomace, a co-product from the fruit pulp industry, display therapeutic application in gut disorders. *Food Res. Int.* **2022**, *156*, 111291. [\[CrossRef\]](https://doi.org/10.1016/j.foodres.2022.111291)
- 28. Rodríguez-Morató, J.; Matthan, N.R.; Liu, J.; de la Torre, R.; Chen, C.-Y.O. Cranberries attenuate animal-based diet-induced changes in microbiota composition and functionality: A randomized crossover controlled feeding trial. *J. Nutr. Biochem.* **2018**, *62*, 76–86. [\[CrossRef\]](https://doi.org/10.1016/j.jnutbio.2018.08.019)
- 29. Gu, Z.; Zhu, Y.; Jiang, S.; Xia, G.; Li, C.; Zhang, X.; Zhang, J.; Shen, X. Tilapia head glycolipids reduce inflammation by regulating the gut microbiota in dextran sulphate sodium-induced colitis mice. *Food Funct.* **2020**, *11*, 3245–3255. [\[CrossRef\]](https://doi.org/10.1039/D0FO00116C)
- 30. Collado-González, M.; González Espinosa, Y.; Goycoolea, F.M. Interaction Between Chitosan and Mucin: Fundamentals and Applications. *Biomimetics* **2019**, *4*, 32. [\[CrossRef\]](https://doi.org/10.3390/biomimetics4020032)
- 31. Wang, J.; Zhang, C.; Guo, C.; Li, X. Chitosan ameliorates DSS-induced ulcerative colitis mice by enhancing intestinal barrier function and improving microflora. *Int. J. Mol. Sci.* **2019**, *20*, 5751. [\[CrossRef\]](https://doi.org/10.3390/ijms20225751) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31731793)
- 32. Moutaharrik, S.; Maroni, A.; Neut, C.; Dubuquoy, C.; Dubuquoy, L.; Foppoli, A.; Cerea, M.; Palugan, L.; Siepmann, F.; Siepmann, J.; et al. In vitro and in vivo evaluation of a pH-, microbiota- and time-based oral delivery platform for colonic release. *Eur. J. Pharm. Biopharm.* **2023**, *183*, 13–23. [\[CrossRef\]](https://doi.org/10.1016/j.ejpb.2022.12.013) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36563887)
- 33. Kulkarni, N.; Jain, P.; Shindikar, A.; Suryawanshi, P.; Thorat, N. Advances in the colon-targeted chitosan based multiunit drug delivery systems for the treatment of inflammatory bowel disease. *Carbohydr. Polym.* **2022**, *288*, 119351. [\[CrossRef\]](https://doi.org/10.1016/j.carbpol.2022.119351) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35450623)
- 34. Helmy, A.M.; Elsabahy, M.; Soliman, G.M.; Mahmoud, M.A.; Ibrahim, E.A. Development and in vivo evaluation of chitosan beads for the colonic delivery of azathioprine for treatment of inflammatory bowel disease. *Eur. J. Pharm. Sci.* **2017**, *109*, 269–279. [\[CrossRef\]](https://doi.org/10.1016/j.ejps.2017.08.025)
- 35. Xu, J.; Tam, M.; Samaei, S.; Lerouge, S.; Barralet, J.; Stevenson, M.M.; Cerruti, M. Mucoadhesive chitosan hydrogels as rectal drug delivery vessels to treat ulcerative colitis. *Acta Biomater.* **2017**, *48*, 247–257. [\[CrossRef\]](https://doi.org/10.1016/j.actbio.2016.10.026)
- 36. Armstrong, H.K.; Bording-Jorgensen, M.; Santer, D.M.; Zhang, Z.; Valcheva, R.; Rieger, A.M.; Kim, J.S.-H.; Dijk, S.I.; Mahmood, R.; Ogungbola, O.; et al. Unfermented β-fructan Fibers Fuel Inflammation in Select Inflammatory Bowel Disease Patients. *Gastroenterology* **2023**, *164*, 228–240. [\[CrossRef\]](https://doi.org/10.1053/j.gastro.2022.09.034)
- 37. Armstrong, H.; Mander, I.; Zhang, Z.; Armstrong, D.; Wine, E. Not All Fibers Are Born Equal; Variable Response to Dietary Fiber Subtypes in IBD. *Front. Pediatr.* **2021**, *8*, 620189. [\[CrossRef\]](https://doi.org/10.3389/fped.2020.620189)
- 38. James, S.L.; Christophersen, C.T.; Bird, A.R.; A Conlon, M.; Rosella, O.; Gibson, P.R.; Muir, J.G. Abnormal fibre usage in UC in remission. *Gut* **2015**, *64*, 562–570. [\[CrossRef\]](https://doi.org/10.1136/gutjnl-2014-307198)
- 39. Venkataraman, A.; Sieber, J.R.; Schmidt, A.W.; Waldron, C.; Theis, K.R.; Schmidt, T.M. Variable responses of human microbiomes to dietary supplementation with resistant starch. *Microbiome* **2016**, *4*, 33. [\[CrossRef\]](https://doi.org/10.1186/s40168-016-0178-x)
- 40. Roselli, M.; Finamore, A. Use of Synbiotics for Ulcerative Colitis Treatment. *Curr. Clin. Pharmacol.* **2019**, *15*, 174–182.
- 41. Jahan-Mihan, A.; Luhovyy, B.L.; El Khoury, D.; Anderson, G.H. Dietary proteins as determinants of metabolic and physiologic functions of the gastrointestinal tract. *Nutrients* **2011**, *3*, 574–603. [\[CrossRef\]](https://doi.org/10.3390/nu3050574) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/22254112)
- 42. Raffner Basson, A.; Adrian, G.-N.; Alexandria, L.; Ludovica, B.; Danielle, K.; Alexandra, W.; Luca, D.M.; Gina, P.; Abdullah, O.; Alexander, R.-P.; et al. Replacing Animal Protein with Soy-Pea Protein in an 'American Diet' Controls Murine Crohn Disease-Like Ileitis Regardless of Firmicutes: Bacteroidetes Ratio. *J. Nutr.* **2021**, *151*, 579–590. [\[CrossRef\]](https://doi.org/10.1093/jn/nxaa386) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33484150)
- 43. Zhou, X.-L.; Zhao, Q.-Q.; Li, X.-F.; Li, Z.; Zhao, S.-X.; Li, Y.-M. Protein intake and risk of inflammatory bowel disease: A meta-analysis. *Asia Pac. J. Clin. Nutr.* **2022**, *31*, 443–449. [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36173216)
- 44. Jowett, S.L. Influence of dietary factors on the clinical course of ulcerative colitis: A prospective cohort study. *Gut* **2004**, *53*, 1479–1484. [\[CrossRef\]](https://doi.org/10.1136/gut.2003.024828) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/15361498)
- 45. Albenberg, L.; Brensinger, C.M.; Wu, Q.; Gilroy, E.; Kappelman, M.D.; Sandler, R.S.; Lewis, J.D. A Diet Low in Red and Processed Meat Does Not Reduce Rate of Crohn's Disease Flares. *Gastroenterology* **2019**, *157*, 128–136.e5. [\[CrossRef\]](https://doi.org/10.1053/j.gastro.2019.03.015)
- 46. Sevencan, N.O.; Isler, M.; Kapucuoglu, F.N.; Senol, A.; Kayhan, B.; Kiztanir, S.; Kockar, M.C. Dose-dependent effects of kefir on colitis induced by trinitrobenzene sulfonic acid in rats. *Food Sci. Nutr.* **2019**, *7*, 3110–3118. [\[CrossRef\]](https://doi.org/10.1002/fsn3.1174)
- 47. Yilmaz, I.; Dolar, M.E.; Ozpinar, H. Effect of administering kefir on the changes in fecal microbiota and symptoms of inflammatory bowel disease: A randomized controlled trial. *Turk. J. Gastroenterol.* **2019**, *30*, 242–253. [\[CrossRef\]](https://doi.org/10.5152/tjg.2018.18227)
- 48. Curciarello, R.; Canziani, K.E.; Salto, I.; Romero, E.B.; Rocca, A.; Doldan, I.; Peton, E.; Brayer, S.; Sambuelli, A.M.; Goncalves, S.; et al. Probiotic Lactobacilli Isolated from Kefir Promote Down-Regulation of Inflammatory Lamina Propria T Cells from Patients with Active IBD. *Front. Pharmacol.* **2021**, *12*, 658026. [\[CrossRef\]](https://doi.org/10.3389/fphar.2021.658026)
- 49. Di Rosa, C.; Di Francesco, L.; Spiezia, C.; Khazrai, Y.M. Effects of Animal and Vegetable Proteins on Gut Microbiota in Subjects with Overweight or Obesity. *Nutrients* **2023**, *15*, 2675. [\[CrossRef\]](https://doi.org/10.3390/nu15122675)
- 50. Abbeele, P.V.D.; Ghyselinck, J.; Marzorati, M.; Koch, A.-M.; Lambert, W.; Michiels, J.; Chalvon-Demersay, T. The Effect of Amino Acids on Production of SCFA and bCFA by Members of the Porcine Colonic Microbiota. *Microorganisms* **2022**, *10*, 762. [\[CrossRef\]](https://doi.org/10.3390/microorganisms10040762)
- 51. Benech, N.; Rolhion, N.; Sokol, H. Gut Microbiota Reprogramming of Tryptophan Metabolism During Pregnancy Shapes Host Insulin Resistance. *Gastroenterology* **2022**, *162*, 1587–1589. [\[CrossRef\]](https://doi.org/10.1053/j.gastro.2022.01.059) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35247461)
- 52. Agus, A.; Planchais, J.; Sokol, H. Gut Microbiota Regulation of Tryptophan Metabolism in Health and Disease. *Cell Host Microbe* **2018**, *23*, 716–724. [\[CrossRef\]](https://doi.org/10.1016/j.chom.2018.05.003) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29902437)
- 53. Fritsch, J.; Garces, L.; Quintero, M.A.; Pignac-Kobinger, J.; Santander, A.M.; Fernández, I.; Ban, Y.J.; Kwon, D.; Phillips, M.C.; Knight, K.; et al. Low-Fat, High-Fiber Diet Reduces Markers of Inflammation and Dysbiosis and Improves Quality of Life in Patients with Ulcerative Colitis. *Clin. Gastroenterol. Hepatol.* **2020**, *19*, 1189–1199.e30. [\[CrossRef\]](https://doi.org/10.1016/j.cgh.2020.05.026)
- 54. Zhu, C.; Sawrey-Kubicek, L.; Beals, E.; Rhodes, C.H.; Houts, H.E.; Sacchi, R.; Zivkovic, A.M. Human gut microbiome composition and tryptophan metabolites were changed differently by fast food and Mediterranean diet in 4 days: A pilot study. *Nutr. Res.* **2020**, *77*, 62–72. [\[CrossRef\]](https://doi.org/10.1016/j.nutres.2020.03.005)
- 55. Sinha, A.K.; Laursen, M.F.; Brinck, J.E.; Rybtke, M.L.; Hjørne, A.P.; Procházková, N.; Pedersen, M.; Roager, H.M.; Licht, T.R. Dietary fibre directs microbial tryptophan metabolism via metabolic interactions in the gut microbiota. *Nat. Microbiol.* **2024**, *9*, 1964–1978. [\[CrossRef\]](https://doi.org/10.1038/s41564-024-01737-3)
- 56. Michaudel, C.; Danne, C.; Agus, A.; Magniez, A.; Aucouturier, A.; Spatz, M.; Lefevre, A.; Kirchgesner, J.; Rolhion, N.; Wang, Y.; et al. Rewiring the altered tryptophan metabolism as a novel therapeutic strategy in inflammatory bowel diseases. *Gut* **2023**, *72*, 1296–1307. [\[CrossRef\]](https://doi.org/10.1136/gutjnl-2022-327337)
- 57. Truyens, M.; Lobatón, T.; Ferrante, M.; Bossuyt, P.; Vermeire, S.; Pouillon, L.; Dewint, P.; Cremer, A.; Peeters, H.; Lambrecht, G.; et al. Effect of 5-Hydroxytryptophan on Fatigue in Quiescent Inflammatory Bowel Disease: A Randomized Controlled Trial. *Gastroenterology* **2022**, *163*, 1294–1305.e3. [\[CrossRef\]](https://doi.org/10.1053/j.gastro.2022.07.052)
- 58. Tian, Q.B.; Chen, S.J.; Xiao, L.J.; Xie, J.Q.; Zhao, H.B.; Zhang, X. Potential effects of nutrition-induced alteration of gut microbiota on inflammatory bowel disease: A review. *J. Dig. Dis.* **2024**, *25*, 78–90. [\[CrossRef\]](https://doi.org/10.1111/1751-2980.13256)
- 59. Arnone, D.; Vallier, M.; Hergalant, S.; Chabot, C.; Ndiaye, N.C.; Moulin, D.; Aignatoaei, A.-M.; Alberto, J.-M.; Louis, H.; Boulard, O.; et al. Long-Term Overconsumption of Fat and Sugar Causes a Partially Reversible Pre-inflammatory Bowel Disease State. *Front. Nutr.* **2021**, *8*, 758518. [\[CrossRef\]](https://doi.org/10.3389/fnut.2021.758518)
- 60. Haskey, N.; Ye, J.; Estaki, M.; Meza, A.A.V.; Barnett, J.A.; Yousefi, M.; Birnie, B.W.; Gruenheid, S.; Ghosh, S.; Gibson, D.L. A Mediterranean-like fat blend protects against the development of severe colitis in the mucin-2 deficient murine model. *Gut Microbes* **2022**, *14*, 2055441. [\[CrossRef\]](https://doi.org/10.1080/19490976.2022.2055441)
- 61. Boneh, R.S.; Van Limbergen, J.; Wine, E.; Assa, A.; Shaoul, R.; Milman, P.; Cohen, S.; Kori, M.; Peleg, S.; On, A.; et al. Dietary Therapies Induce Rapid Response and Remission in Pediatric Patients with Active Crohn's Disease. *Clin. Gastroenterol. Hepatol.* **2020**, *19*, 752–759. [\[CrossRef\]](https://doi.org/10.1016/j.cgh.2020.04.006) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32302709)
- 62. Vrdoljak, J.; Kumric, M.; Vilovic, M.; Martinovic, D.; Tomic, I.J.; Krnic, M.; Kurir, T.T.; Bozic, J. Effects of Olive Oil and Its Components on Intestinal Inflammation and Inflammatory Bowel Disease. *Nutrients* **2022**, *14*, 757. [\[CrossRef\]](https://doi.org/10.3390/nu14040757)
- 63. Vaghari-Tabari, M.; Alemi, F.; Zokaei, M.; Moein, S.; Qujeq, D.; Yousefi, B.; Farzami, P.; Hosseininasab, S.S. Polyphenols and inflammatory bowel disease: Natural products with therapeutic effects? *Crit. Rev. Food Sci. Nutr.* **2022**, *64*, 4155–4178. [\[CrossRef\]](https://doi.org/10.1080/10408398.2022.2139222)
- 64. Jimenez-Lopez, C.; Carpena, M.; Lourenço-Lopes, C.; Gallardo-Gomez, M.; Lorenzo, J.M.; Barba, F.J.; Prieto, M.A.; Simal-Gandara, J. Bioactive Compounds and Quality of Extra Virgin Olive Oil. *Foods* **2020**, *9*, 1014. [\[CrossRef\]](https://doi.org/10.3390/foods9081014)
- 65. Sánchez-Fidalgo, S.; Sanchez De Ibargüen, L.; Cárdeno, A.; Alarcon De La Lastra, C. Influence of extra virgin olive oil diet enriched with hydroxytyrosol in a chronic DSS colitis model. *Eur. J. Nutr.* **2012**, *51*, 497–506. [\[CrossRef\]](https://doi.org/10.1007/s00394-011-0235-y)
- 66. Sánchez-Fidalgo, S.; Cárdeno, A.; Sánchez-Hidalgo, M.; Aparicio-Soto, M.; Villegas, I.; Rosillo, M.; de la Lastra, C.A. Dietary unsaponifiable fraction from extra virgin olive oil supplementation attenuates acute ulcerative colitis in mice. *Eur. J. Pharm. Sci.* **2013**, *48*, 572–581. [\[CrossRef\]](https://doi.org/10.1016/j.ejps.2012.12.004)
- 67. Negro, C.; Aprile, A.; Luvisi, A.; Nicolì, F.; Nutricati, E.; Vergine, M.; Miceli, A.; Blando, F.; Sabella, E.; De Bellis, L. Phenolic Profile and Antioxidant Activity of Italian Monovarietal Extra Virgin Olive Oils. *Antioxidants* **2019**, *8*, 161. [\[CrossRef\]](https://doi.org/10.3390/antiox8060161)
- 68. Cariello, M.; Contursi, A.; Gadaleta, R.M.; Piccinin, E.; De Santis, S.; Piglionica, M.; Spaziante, A.F.; Sabbà, C.; Villani, G.; Moschetta, A. Extra-Virgin Olive Oil from Apulian Cultivars and Intestinal Inflammation. *Nutrients* **2020**, *12*, 1084. [\[CrossRef\]](https://doi.org/10.3390/nu12041084)
- 69. Motawea, M.H.; Elmaksoud, H.A.A.; Elharrif, M.G.; Desoky, A.A.E.; Ibrahimi, A. Evaluation of Anti-Inflammatory and Antioxidant Profile of Oleuropein in Experimentally Induced Ulcerative Colitis. *Int. J. Mol. Cell Med.* **2020**, *9*, 224–233. [\[CrossRef\]](https://doi.org/10.22088/IJMCM.BUMS.9.3.224)
- 70. Yu, F.; Hu, X.; Ren, H.; Wang, X.; Shi, R.; Guo, J.; Chang, J.; Zhou, X.; Jin, Y.; Li, Y.; et al. Protective effect of synbiotic combination of Lactobacillus plantarum SC-5 and olive oil extract tyrosol in a murine model of ulcerative colitis. *J. Transl. Med.* **2024**, *22*, 308. [\[CrossRef\]](https://doi.org/10.1186/s12967-024-05026-9)
- 71. Santarelli, R.; Pompili, C.; Montani, M.S.G.; Evangelista, L.; Gonnella, R.; Cirone, M. 3,4-Dihydroxyphenylethanol (DPE or Hydroxytyrosol) Counteracts ERK1/2 and mTOR Activation, Pro-Inflammatory Cytokine Release, Autophagy and Mitophagy Reduction Mediated by Benzo[a]pyrene in Primary Human Colonic Epithelial Cells. *Pharmaceutics* **2022**, *14*, 663. [\[CrossRef\]](https://doi.org/10.3390/pharmaceutics14030663) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35336037)
- 72. Abedi, E.; Sahari, M.A. Long-chain polyunsaturated fatty acid sources and evaluation of their nutritional and functional properties. *Food Sci. Nutr.* **2014**, *2*, 443–463. [\[CrossRef\]](https://doi.org/10.1002/fsn3.121) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25473503)
- 73. Smyth, M.; Lunken, G.; Jacobson, K. Insights into Inflammatory Bowel Disease and Effects of Dietary Fatty Acid Intake With a Focus on Polyunsaturated Fatty Acids Using Preclinical Models. *J. Can. Assoc. Gastroenterol.* **2024**, *7*, 104–114. [\[CrossRef\]](https://doi.org/10.1093/jcag/gwad058) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38314173)
- 74. Yorulmaz, E.; Yorulmaz, H.; Gökmen, E.S.; Altınay, S.; Küçük, S.H.; Zengi, O.; Çelik, D.S.; ¸Sit, D. Therapeutic effectiveness of rectally administered fish oil and mesalazine in trinitrobenzenesulfonic acid-induced colitis. *Biomed. Pharmacother.* **2019**, *118*, 109247. [\[CrossRef\]](https://doi.org/10.1016/j.biopha.2019.109247) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31351432)
- 75. Scaioli, E.; Sartini, A.; Bellanova, M.; Campieri, M.; Festi, D.; Bazzoli, F.; Belluzzi, A. Eicosapentaenoic Acid Reduces Fecal Levels of Calprotectin and Prevents Relapse in Patients with Ulcerative Colitis. *Clin. Gastroenterol. Hepatol.* **2018**, *16*, 1268–1275.e2. [\[CrossRef\]](https://doi.org/10.1016/j.cgh.2018.01.036)
- 76. Bouhend, A.; Keddari, S.; Yahla, I.; Sadouki, O.; Bououdina, M. Therapeutic Benefits of Tuna Oil by In Vitro and In Vivo Studies Using a Rat Model of Acetic Acid-Induced Ulcerative Colitis. *Appl. Biochem. Biotechnol.* **2023**, *196*, 3817–3843. [\[CrossRef\]](https://doi.org/10.1007/s12010-023-04736-y)
- 77. Shabana, S.; Hamouda, H.I.; Hamadou, A.H.; Ahmed, B.; Chi, Z.; Liu, C. Marine phospholipid nanoliposomes: A promising therapeutic approach for inflammatory bowel disease: Preparation, safety, and efficacy evaluation. *Colloids Surf. B Biointerfaces* **2023**, *234*, 113702. [\[CrossRef\]](https://doi.org/10.1016/j.colsurfb.2023.113702)
- 78. Fard, N.A.; Bording-Jorgensen, M.; Wine, E. A Potential Role for Gut Microbes in Mediating Effects of Omega-3 Fatty Acids in Inflammatory Bowel Diseases: A Comprehensive Review. *Curr. Microbiol.* **2023**, *80*, 363.
- 79. Astore, C.; Nagpal, S.; Gibson, G. Mendelian Randomization Indicates a Causal Role for Omega-3 Fatty Acids in Inflammatory Bowel Disease. *Int. J. Mol. Sci.* **2022**, *23*, 14380. [\[CrossRef\]](https://doi.org/10.3390/ijms232214380)
- 80. Ajabnoor, S.M.; Thorpe, G.; Abdelhamid, A.; Hooper, L. Long-term effects of increasing omega-3, omega-6 and total polyunsaturated fats on inflammatory bowel disease and markers of inflammation: A systematic review and meta-analysis of randomized controlled trials. *Eur. J. Nutr.* **2021**, *60*, 2293–2316. [\[CrossRef\]](https://doi.org/10.1007/s00394-020-02413-y)
- 81. Schwärzler, J.; Mayr, L.; Vila, A.V.; Grabherr, F.; Niederreiter, L.; Philipp, M.; Grander, C.; Meyer, M.; Jukic, A.; Tröger, S.; et al. PUFA-Induced Metabolic Enteritis as a Fuel for Crohn's Disease. *Gastroenterology* **2022**, *162*, 1690–1704. [\[CrossRef\]](https://doi.org/10.1053/j.gastro.2022.01.004) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35031299)
- 82. Aldini, R.; Micucci, M.; Cevenini, M.; Fato, R.; Bergamini, C.; Nanni, C.; Cont, M.; Camborata, C.; Spinozzi, S.; Montagnani, M.; et al. Antiinflammatory Effect of Phytosterols in Experimental Murine Colitis Model: Prevention, Induction, Remission Study. *PLoS ONE* **2014**, *9*, e108112. [\[CrossRef\]](https://doi.org/10.1371/journal.pone.0108112) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25268769)
- 83. Cheon, J.H.; Kim, J.S.; Kim, J.M.; Kim, N.; Jung, H.C.; Song, I.S. Plant sterol guggulsterone inhibits nuclear factor-κB signaling in intestinal epithelial cells by blocking IκB kinase and ameliorates acute murine colitis. *Inflamm. Bowel Dis.* **2006**, *12*, 1152–1161. [\[CrossRef\]](https://doi.org/10.1097/01.mib.0000235830.94057.c6) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/17119390)
- 84. Velde, A.A.T.; Brüll, F.; Heinsbroek, S.E.M.; Meijer, S.L.; Lütjohann, D.; Vreugdenhil, A.; Plat, J. Effects of Dietary Plant Sterols and Stanol Esters with Low- and High-Fat Diets in Chronic and Acute Models for Experimental Colitis. *Nutrients* **2015**, *7*, 8518–8531. [\[CrossRef\]](https://doi.org/10.3390/nu7105412)
- 85. Rajaram, S.; Damasceno, N.R.T.; Braga, R.A.M.; Martinez, R.; Kris-Etherton, P.; Sala-Vila, A. Effect of Nuts on Markers of Inflammation and Oxidative Stress: A Narrative Review. *Nutrients* **2023**, *15*, 1099. [\[CrossRef\]](https://doi.org/10.3390/nu15051099)
- 86. Yeon, N.-R.; Cho, J.S.; Yoo, H.-S.; Jeon, S.H.; Yi, C.-M.; Jung, M.-J.; Lee, Y.-S.; Shin, E.-B.; Kim, N.; Kim, H.; et al. Dextran sodium sulfate (DSS)-induced colitis is alleviated in mice after administration of flavone-derived NRF2-activating molecules. *Life Sci.* **2024**, *340*, 122424. [\[CrossRef\]](https://doi.org/10.1016/j.lfs.2024.122424)
- 87. Wu, B.; Cox, A.D.; Chang, H.; Kennett, M.; Rosa, C.; Chopra, S.; Li, S.; Reddivari, L. Maize near-isogenic lines with enhanced flavonoids alleviated dextran sodium sulfate-induced murine colitis via modulation of the gut microbiota. *Food Funct.* **2023**, *14*, 9606–9616. [\[CrossRef\]](https://doi.org/10.1039/D3FO02953K)
- 88. Li, M.; Weigmann, B. Effect of a Flavonoid Combination of Apigenin and Epigallocatechin-3-Gallate on Alleviating Intestinal Inflammation in Experimental Colitis Models. *Int. J. Mol. Sci.* **2023**, *24*, 16031. [\[CrossRef\]](https://doi.org/10.3390/ijms242216031)
- 89. Li, M.; Liu, Y.; Weigmann, B. Biodegradable Polymeric Nanoparticles Loaded with Flavonoids: A Promising Therapy for Inflammatory Bowel Disease. *Int. J. Mol. Sci.* **2023**, *24*, 4454. [\[CrossRef\]](https://doi.org/10.3390/ijms24054454)
- 90. Li, K.; Wu, J.; Xu, S.; Li, X.; Zhang, Y.; Gao, X.-J. Rosmarinic acid alleviates intestinal inflammatory damage and inhibits endoplasmic reticulum stress and smooth muscle contraction abnormalities in intestinal tissues by regulating gut microbiota. *Microbiol. Spectr.* **2023**, *11*, e0191423. [\[CrossRef\]](https://doi.org/10.1128/spectrum.01914-23)
- 91. Han, X.; Li, M.; Sun, L.; Liu, X.; Yin, Y.; Hao, J.; Zhang, W. p-Hydroxybenzoic Acid Ameliorates Colitis by Improving the Mucosal Barrier in a Gut Microbiota-Dependent Manner. *Nutrients* **2022**, *14*, 5383. [\[CrossRef\]](https://doi.org/10.3390/nu14245383) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36558542)
- 92. Han, D.; Wu, Y.; Lu, D.; Pang, J.; Hu, J.; Zhang, X.; Wang, Z.; Zhang, G.; Wang, J. Polyphenol-rich diet mediates interplay between macrophage-neutrophil and gut microbiota to alleviate intestinal inflammation. *Cell Death Dis.* **2023**, *14*, 656. [\[CrossRef\]](https://doi.org/10.1038/s41419-023-06190-4) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37813835)
- 93. Tian, Y.; Jian, T.; Li, J.; Huang, L.; Li, S.; Lu, H.; Niu, G.; Meng, X.; Ren, B.; Liao, H.; et al. Phenolic acids from Chicory roots ameliorate dextran sulfate sodium–induced colitis in mice by targeting TRP signaling pathways and the gut microbiota. *Phytomedicine* **2024**, *128*, 155378. [\[CrossRef\]](https://doi.org/10.1016/j.phymed.2024.155378) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38507851)
- 94. Farsi, F.; Tahvilian, N.; Heydarian, A.; Karimi, S.; Ebrahimi, S.; Ebrahimi-Daryani, N.; Tabataba-Vakili, S.; Heshmati, J.; Mokhtare, M. Evaluating macro- and micronutrients and food groups intake with the risk of developing inflammatory bowel disease: Is there any association? *Food Sci. Nutr.* **2022**, *10*, 3920–3930. [\[CrossRef\]](https://doi.org/10.1002/fsn3.2988)
- 95. Preda, C.M.; Istratescu, D.; Nitescu, M.; Manuc, T.; Manuc, M.; Stroie, T.; Catrinoiu, M.; Tieranu, C.; Meianu, C.G.; Tugui, L.; et al. Impact of Dietary Patterns in Inflammatory Bowel Disease Subtypes. *Maedica A J. Clin. Med.* **2023**, *18*, 174–181. [\[CrossRef\]](https://doi.org/10.26574/maedica.2023.18.2.174)
- 96. Nunes, S.; Danesi, F.; Del Rio, D.; Silva, P. Resveratrol and inflammatory bowel disease: The evidence so far. *Nutr. Res. Rev.* **2018**, *31*, 85–97. [\[CrossRef\]](https://doi.org/10.1017/S095442241700021X)
- 97. Larrosa, M.; Yañéz-Gascón, M.J.; Selma, M.V.; González-Sarrías, A.; Toti, S.; Cerón, J.J.; Tomás-Barberán, F.; Dolara, P.; Espín, J.C. Effect of a low dose of dietary resveratrol on colon microbiota, inflammation and tissue damage in a DSS-induced colitis rat model. *J. Agric. Food Chem.* **2009**, *57*, 2211–2220. [\[CrossRef\]](https://doi.org/10.1021/jf803638d)
- 98. He, Q.; Guo, K.; Wang, L.; Xie, F.; Zhao, Q.; Jiang, X.; He, Z.; Wang, P.; Li, S.; Huang, Y.; et al. Tannins amount determines whether tannase-containing bacteria are probiotic or pathogenic in IBD. *Life Sci. Alliance* **2023**, *6*, e202201702. [\[CrossRef\]](https://doi.org/10.26508/lsa.202201702)
- 99. Baur, J.A.; Sinclair, D.A. Therapeutic potential of resveratrol: The in vivo evidence. *Nat. Rev. Drug Discov.* **2006**, *5*, 493–506. [\[CrossRef\]](https://doi.org/10.1038/nrd2060)
- 100. Martín, A.R.; Villegas, I.; Sánchez-Hidalgo, M.; de la Lastra, C.A. The effects of resveratrol, a phytoalexin derived from red wines, on chronic inflammation induced in an experimentally induced colitis model. *Br. J. Pharmacol.* **2006**, *147*, 873–885. [\[CrossRef\]](https://doi.org/10.1038/sj.bjp.0706469)
- 101. White, B.A.; Ramos, G.P.; Kane, S. The Impact of Alcohol in Inflammatory Bowel Diseases. *Inflamm. Bowel Dis.* **2021**, *28*, 466–473. [\[CrossRef\]](https://doi.org/10.1093/ibd/izab089) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33988227)
- 102. Vrdoljak, J.; Kumric, M.; Kurir, T.T.; Males, I.; Martinovic, D.; Vilovic, M.; Bozic, J. Effects of Wine Components in Inflammatory Bowel Diseases. *Molecules* **2021**, *26*, 5891. [\[CrossRef\]](https://doi.org/10.3390/molecules26195891) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34641434)
- 103. Batiha, G.E.-S.; Beshbishy, A.M.; Ikram, M.; Mulla, Z.S.; El-Hack, M.E.A.; Taha, A.E.; Algammal, A.M.; Elewa, Y.H.A. The Pharmacological Activity, Biochemical Properties, and Pharmacokinetics of the Major Natural Polyphenolic Flavonoid: Quercetin. *Foods* **2020**, *9*, 374. [\[CrossRef\]](https://doi.org/10.3390/foods9030374) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32210182)
- 104. Alemayohu, M.A.; Zanolin, E.; Cazzoletti, L.; Chamitava, L.; Mattioli, V.; Plumb, J.; Burney, P.G.J.; Garcia-Larsen, V. Flavonoid Content of Selected Foods—A Comparison of Four International Composition Tables. *Curr. Dev. Nutr.* **2020**, *4*, 1155. [\[CrossRef\]](https://doi.org/10.1093/cdn/nzaa056_002)
- 105. Dodda, D.; Chhajed, R.; Mishra, J.; Padhy, M. Targeting oxidative stress attenuates trinitrobenzene sulphonic acid induced inflammatory bowel disease like symptoms in rats: Role of quercetin. *Indian J. Pharmacol.* **2014**, *46*, 286–291. [\[CrossRef\]](https://doi.org/10.4103/0253-7613.132160)
- 106. Jia, H.; Zhang, Y.; Si, X.; Jin, Y.; Jiang, D.; Dai, Z.; Wu, Z. Quercetin Alleviates Oxidative Damage by Activating Nuclear Factor Erythroid 2-Related Factor 2 Signaling in Porcine Enterocytes. *Nutrients* **2021**, *13*, 375. [\[CrossRef\]](https://doi.org/10.3390/nu13020375)
- 107. Dong, Y.; Hou, Q.; Lei, J.; Wolf, P.G.; Ayansola, H.; Zhang, B. Quercetin Alleviates Intestinal Oxidative Damage Induced by H2O² via Modulation of GSH: In Vitro Screening and In Vivo Evaluation in a Colitis Model of Mice. *ACS Omega* **2020**, *5*, 8334–8346. [\[CrossRef\]](https://doi.org/10.1021/acsomega.0c00804)
- 108. Lin, R.; Piao, M.; Song, Y. Dietary Quercetin Increases Colonic Microbial Diversity and Attenuates Colitis Severity in Citrobacter rodentium-Infected Mice. *Front. Microbiol.* **2019**, *10*, 1092. [\[CrossRef\]](https://doi.org/10.3389/fmicb.2019.01092)
- 109. Gardi, C.; Bauerova, K.; Stringa, B.; Kuncirova, V.; Slovak, L.; Ponist, S.; Drafi, F.; Bezakova, L.; Tedesco, I.; Acquaviva, A.; et al. Quercetin reduced inflammation and increased antioxidant defense in rat adjuvant arthritis. *Arch. Biochem. Biophys.* **2015**, *583*, 150–157. [\[CrossRef\]](https://doi.org/10.1016/j.abb.2015.08.008)
- 110. Riemschneider, S.; Hoffmann, M.; Slanina, U.; Weber, K.; Hauschildt, S.; Lehmann, J. Indol-3-Carbinol and Quercetin Ameliorate Chronic DSS-Induced Colitis in C57BL/6 Mice by AhR-Mediated Anti-Inflammatory Mechanisms. *Int. J. Env. Res. Public Health* **2021**, *18*, 2262. [\[CrossRef\]](https://doi.org/10.3390/ijerph18052262)
- 111. Topçu-Tarladaçalışır, Y.; Sapmaz-Metin, M.; Mercan, Z.; Erçetin, D. Quercetin Attenuates Endoplasmic Reticulum Stress and Apoptosis in TNBS-Induced Colitis by Inhibiting the Glucose Regulatory Protein 78 Activation. *Balk. Med. J.* **2024**, *41*, 30–37. [\[CrossRef\]](https://doi.org/10.4274/balkanmedj.galenos.2023.2023-10-9) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38173174)
- 112. Hu, S.; Zhao, M.; Li, W.; Wei, P.; Liu, Q.; Chen, S.; Zeng, J.; Ma, X.; Tang, J. Preclinical evidence for quercetin against inflammatory bowel disease: A meta-analysis and systematic review. *Inflammopharmacology* **2022**, *30*, 2035–2050. [\[CrossRef\]](https://doi.org/10.1007/s10787-022-01079-8) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36227442)
- 113. Enayat, S.; Ceyhan, M.; Taşkoparan, B.; Stefek, M.; Banerjee, S. CHNQ, a novel 2-Chloro-1,4-naphthoquinone derivative of quercetin, induces oxidative stress and autophagy both in vitro and in vivo. *Arch. Biochem. Biophys.* **2016**, *596*, 84–98. [\[CrossRef\]](https://doi.org/10.1016/j.abb.2016.03.004) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26946942)
- 114. Diez-Echave, P.; Ruiz-Malagón, A.J.; Molina-Tijeras, J.A.; Hidalgo-García, L.; Vezza, T.; Cenis-Cifuentes, L.; Rodríguez-Sojo, M.J.; Cenis, J.L.; Rodríguez-Cabezas, M.E.; Rodríguez-Nogales, A.; et al. Silk fibroin nanoparticles enhance quercetin immunomodulatory properties in DSS-induced mouse colitis. *Int. J. Pharm.* **2021**, *606*, 120935. [\[CrossRef\]](https://doi.org/10.1016/j.ijpharm.2021.120935)
- 115. Wang, L.; Fu, R.; Meng, Y.; Liang, J.; Xue, W.; Hu, H.; Meng, J.; Zhang, M. pH Sensitive Quercetin Nanoparticles Ameliorate DSS-Induced Colitis in Mice by Colon-Specific Delivery. *Mol. Nutr. Food Res.* **2023**, *68*, e2300051. [\[CrossRef\]](https://doi.org/10.1002/mnfr.202300051)
- 116. Shen, C.; Zhao, L.; Du, X.; Tian, J.; Yuan, Y.; Jia, M.; He, Y.; Zeng, R.; Qiao, R.; Li, C. Smart Responsive Quercetin-Conjugated Glycol Chitosan Prodrug Micelles for Treatment of Inflammatory Bowel Diseases. *Mol. Pharm.* **2021**, *18*, 1419–1430. [\[CrossRef\]](https://doi.org/10.1021/acs.molpharmaceut.0c01245)
- 117. Zhang, X.; Su, W.; Chen, Y.; Xing, S.; El-Aty, A.A.; Song, Y.; Tan, M. Bi-functional astaxanthin macromolecular nanocarriers to alleviate dextran sodium sulfate-induced inflammatory bowel disease. *Int. J. Biol. Macromol.* **2023**, *256*, 128494. [\[CrossRef\]](https://doi.org/10.1016/j.ijbiomac.2023.128494)
- 118. Story, E.N.; Kopec, R.E.; Schwartz, S.J.; Harris, G.K. An Update on the Health Effects of Tomato Lycopene. *Annu. Rev. Food Sci. Technol.* **2010**, *1*, 189–210. [\[CrossRef\]](https://doi.org/10.1146/annurev.food.102308.124120)
- 119. Yuan, H.; Li, Y.; Ling, F.; Guan, Y.; Zhang, D.; Zhu, Q.; Liu, J.; Wu, Y.; Niu, Y. The phytochemical epigallocatechin gallate prolongs the lifespan by improving lipid metabolism, reducing inflammation and oxidative stress in high-fat diet-fed obese rats. *Aging Cell* **2020**, *19*, e13199. [\[CrossRef\]](https://doi.org/10.1111/acel.13199)
- 120. Ross, A.C.; Caballero, B.H.; Cousins, R.J.; Tucker, K.L.; Ziegler, T.R. *Modern Nutrition in Health and Disease*, 11th ed.; Wolters Kluwer Health Adis (ESP): Philadelphia, PA, USA, 2012.
- 121. Zupo, R.; Lampignano, L.; Lattanzio, A.; Mariano, F.; Osella, A.R.; Bonfiglio, C.; Giannelli, G.; De Pergola, G. Association between adherence to the Mediterranean Diet and circulating Vitamin D levels. *Int. J. Food Sci. Nutr.* **2020**, *71*, 884–890. [\[CrossRef\]](https://doi.org/10.1080/09637486.2020.1744533)
- 122. Ambati, R.R.; Phang, S.-M.; Ravi, S.; Aswathanarayana, R.G. Astaxanthin: Sources, Extraction, Stability, Biological Activities and Its Commercial Applications—A Review. *Mar. Drugs* **2014**, *12*, 128–152. [\[CrossRef\]](https://doi.org/10.3390/md12010128) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24402174)
- 123. Sakai, S.; Nishida, A.; Ohno, M.; Inatomi, O.; Bamba, S.; Sugimoto, M.; Kawahara, M.; Andoh, A. Astaxanthin, a xanthophyll carotenoid, prevents development of dextran sulphate sodium-induced murine colitis. *J. Clin. Biochem. Nutr.* **2019**, *64*, 66–72. [\[CrossRef\]](https://doi.org/10.3164/jcbn.18-47) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30705514)
- 124. Zhang, C.; Xu, Y.; Wu, S.; Zheng, W.; Song, S.; Ai, C. Fabrication of astaxanthin-enriched colon-targeted alginate microspheres and its beneficial effect on dextran sulfate sodium-induced ulcerative colitis in mice. *Int. J. Biol. Macromol.* **2022**, *205*, 396–409. [\[CrossRef\]](https://doi.org/10.1016/j.ijbiomac.2022.02.057)
- 125. Chen, Y.; Su, W.; Tie, S.; Cui, W.; Yu, X.; Zhang, L.; Hua, Z.; Tan, M. Orally deliverable sequence-targeted astaxanthin nanoparticles for colitis alleviation. *Biomaterials* **2022**, *293*, 121976. [\[CrossRef\]](https://doi.org/10.1016/j.biomaterials.2022.121976)
- 126. Luo, M.; Yuan, Q.; Liu, M.; Song, X.; Xu, Y.; Zhang, T.; Zeng, X.; Wu, Z.; Pan, D.; Guo, Y. Astaxanthin nanoparticles ameliorate dextran sulfate sodium-induced colitis by alleviating oxidative stress, regulating intestinal flora, and protecting the intestinal barrier. *Food Funct.* **2023**, *14*, 9567–9579. [\[CrossRef\]](https://doi.org/10.1039/D3FO03331G)
- 127. Liu, F.; Smith, A.D.; Wang, T.T.Y.; Pham, Q.; Hou, P.; Cheung, L.; Yang, H.; Li, R.W. Phospholipid-rich krill oil promotes intestinal health by strengthening beneficial gut microbial interactions in an infectious colitis model. *Food Funct.* **2024**, *15*, 2604–2615. [\[CrossRef\]](https://doi.org/10.1039/D3FO04980A)
- 128. Liu, F.; Smith, A.D.; Solano-Aguilar, G.; Wang, T.T.Y.; Pham, Q.; Beshah, E.; Tang, Q.; Urban, J.F.; Xue, C.; Li, R.W. Mechanistic insights into the attenuation of intestinal inflammation and modulation of the gut microbiome by krill oil using in vitro and in vivo models. *Microbiome* **2020**, *8*, 83. [\[CrossRef\]](https://doi.org/10.1186/s40168-020-00843-8)
- 129. Li, F.; Cai, M.; Lin, M.; Huang, X.; Wang, J.; Ke, H.; Wang, C.; Zheng, X.; Chen, D.; Yang, S. Enhanced Biomass and Astaxanthin Production of Haematococcus pluvialis by a Cell Transformation Strategy with Optimized Initial Biomass Density. *Mar. Drugs* **2020**, *18*, 341. [\[CrossRef\]](https://doi.org/10.3390/md18070341)
- 130. Shokri-mashhadi, N.; Tahmasebi, M.; Mohammadi-asl, J.; Zakerkish, M.; Mohammadshahi, M. The antioxidant and antiinflammatory effects of astaxanthin supplementation on the expression of miR-146a and miR-126 in patients with type 2 diabetes mellitus: A randomised, double-blind, placebo-controlled clinical trial. *Int. J. Clin. Pract.* **2021**, *75*, e14022. [\[CrossRef\]](https://doi.org/10.1111/ijcp.14022)
- 131. Narayanaswam, N.K.; Caston, E.; Kumar, R.C.S.; Vijayakumar, T.M.; Vanangamudi, V.S.; Pankaj, N.; Sukkur, A. A randomized interventional clinical trial assessing the safety and effectiveness of PeaNoc XL tablets in managing joint pain and inflammation in arthritis patients. *F1000Research* **2023**, *12*, 895. [\[CrossRef\]](https://doi.org/10.12688/f1000research.138477.1)
- 132. Rostami, S.; Alyasin, A.; Saedi, M.; Nekoonam, S.; Khodarahmian, M.; Moeini, A.; Amidi, F. Astaxanthin ameliorates inflammation, oxidative stress, and reproductive outcomes in endometriosis patients undergoing assisted reproduction: A randomized, tripleblind placebo-controlled clinical trial. *Front. Endocrinol.* **2023**, *14*, 1144323. [\[CrossRef\]](https://doi.org/10.3389/fendo.2023.1144323) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37020589)
- 133. Sesso, H.D.; Liu, S.; Gaziano, J.M.; Buring, J.E. Dietary Lycopene, Tomato-Based Food Products and Cardiovascular Disease in Women. *J. Nutr.* **2003**, *133*, 2336–2341. [\[CrossRef\]](https://doi.org/10.1093/jn/133.7.2336) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/12840203)
- 134. Yue, Y.; Shi, M.; Song, X.; Ma, C.; Li, D.; Hu, X.; Chen, F. Lycopene Ameliorated DSS-Induced Colitis by Improving Epithelial Barrier Functions and Inhibiting the Escherichia coli Adhesion in Mice. *J. Agric. Food Chem.* **2024**, *72*, 5784–5796. [\[CrossRef\]](https://doi.org/10.1021/acs.jafc.3c09717) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38447175)
- 135. Yin, Z.; Wang, Q.; Cheng, H. Synergistic Protective Effect of Interactions of Quercetin with Lycopene Against Ochratoxin A-Induced Ulcerative Colitis. *Appl. Biochem. Biotechnol.* **2023**, *195*, 5253–5266. [\[CrossRef\]](https://doi.org/10.1007/s12010-022-04287-8)
- 136. Chen, J.; Ruan, X.; Yuan, S.; Deng, M.; Zhang, H.; Sun, J.; Yu, L.; Satsangi, J.; Larsson, S.C.; Therdoratou, E.; et al. Antioxidants, minerals and vitamins in relation to Crohn's disease and ulcerative colitis: A Mendelian randomization study. *Aliment. Pharmacol. Ther.* **2023**, *57*, 399–408. [\[CrossRef\]](https://doi.org/10.1111/apt.17392)
- 137. Zhou, F.; Mai, T.; Wang, Z.; Zeng, Z.; Shi, J.; Zhang, F.; Kong, N.; Jiang, H.; Guo, L.; Xu, M.; et al. The improvement of intestinal dysbiosis and hepatic metabolic dysfunction in dextran sulfate sodium-induced colitis mice: Effects of curcumin. *J. Gastroenterol. Hepatol.* **2023**, *38*, 1333–1345. [\[CrossRef\]](https://doi.org/10.1111/jgh.16205)
- 138. Meng, Z.-W.; Chang, B.; Sang, L.-X. Use of curcumin and its nanopreparations in the treatment of inflammatory bowel disease. *World J. Gastroenterol.* **2024**, *30*, 280–282. [\[CrossRef\]](https://doi.org/10.3748/wjg.v30.i3.280)
- 139. Thavorn, K.; Wolfe, D.; Faust, L.; Shorr, R.; Akkawi, M.; Isaranuwatchai, W.; Klinger, C.; Chai-Adisaksopa, C.; Tanvejsilp, P.; Nochaiwong, S.; et al. A systematic review of the efficacy and safety of turmeric in the treatment of digestive disorders. *Phytother. Res.* **2024**, *38*, 2687–2706. [\[CrossRef\]](https://doi.org/10.1002/ptr.8189)
- 140. Wei, S.-Y.; Wu, T.-T.; Huang, J.-Q.; Kang, Z.-P.; Wang, M.-X.; Zhong, Y.-B.; Ge, W.; Zhou, B.-G.; Zhao, H.-M.; Wang, H.-Y.; et al. Curcumin alleviates experimental colitis via a potential mechanism involving memory B cells and Bcl-6-Syk-BLNK signaling. *World J. Gastroenterol.* **2022**, *28*, 5865–5880. [\[CrossRef\]](https://doi.org/10.3748/wjg.v28.i40.5865)
- 141. Sugimoto, K.; Ikeya, K.; Bamba, S.; Andoh, A.; Yamasaki, H.; Mitsuyama, K.; Nasuno, M.; Tanaka, H.; Matsuura, A.; Kato, M.; et al. Highly Bioavailable Curcumin Derivative Ameliorates Crohn's Disease Symptoms: A Randomized, Double-Blind, Multicenter Study. *J. Crohns Colitis* **2020**, *14*, 1693–1701. [\[CrossRef\]](https://doi.org/10.1093/ecco-jcc/jjaa097)
- 142. de la Rubia Ortí, J.E.; Platero, J.L.; Yang, I.H.; Ceron, J.J.; Tvarijonaviciute, A.; Sabater, P.S.; Benlloch, M.; Sancho-Cantus, D.; Sancho, S. Possible Role of Butyrylcholinesterase in Fat Loss and Decreases in Inflammatory Levels in Patients with Multiple Sclerosis after Treatment with Epigallocatechin Gallate and Coconut Oil: A Pilot Study. *Nutrients* **2021**, *13*, 3230. [\[CrossRef\]](https://doi.org/10.3390/nu13093230) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34579104)
- 143. Bazyar, H.; Hosseini, S.A.; Saradar, S.; Mombaini, D.; Allivand, M.; Labibzadeh, M.; Alipour, M. Effects of epigallocatechin-3-gallate of *Camellia sinensis* leaves on blood pressure, lipid profile, atherogenic index of plasma and some inflammatory and antioxidant markers in type 2 diabetes mellitus patients: A clinical trial. *J. Complement. Integr. Med.* **2020**, *18*, 405–411. [\[CrossRef\]](https://doi.org/10.1515/jcim-2020-0090) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34187117)
- 144. Charoenngam, N.; Shirvani, A.; Kalajian, T.A.; Song, A.; Holick, M.F. The Effect of Various Doses of Oral Vitamin D₃ Supplementation on Gut Microbiota in Healthy Adults: A Randomized, Double-blinded, Dose-response Study. *Anticancer. Res.* **2020**, *40*, 551–556. [\[CrossRef\]](https://doi.org/10.21873/anticanres.13984)
- 145. Taylor, L.; Almutairdi, A.; Shommu, N.; Fedorak, R.; Ghosh, S.; Reimer, R.A.; Panaccione, R.; Raman, M. Cross-Sectional Analysis of Overall Dietary Intake and Mediterranean Dietary Pattern in Patients with Crohn's Disease. *Nutrients* **2018**, *10*, 1761. [\[CrossRef\]](https://doi.org/10.3390/nu10111761)
- 146. Castro-Quezada, I.; Román-Viñas, B.; Serra-Majem, L. The Mediterranean Diet and Nutritional Adequacy: A Review. *Nutrients* **2014**, *6*, 231–248. [\[CrossRef\]](https://doi.org/10.3390/nu6010231)
- 147. Grosso, G.; Marventano, S.; Buscemi, S.; Scuderi, A.; Matalone, M.; Platania, A.; Giorgianni, G.; Rametta, S.; Nolfo, F.; Galvano, F.; et al. Factors Associated with Adherence to the Mediterranean Diet among Adolescents Living in Sicily, Southern Italy. *Nutrients* **2013**, *5*, 4908–4923. [\[CrossRef\]](https://doi.org/10.3390/nu5124908)
- 148. Boughanem, H.; Ruiz-Limón, P.; Pilo, J.; Lisbona-Montañez, J.M.; Tinahones, F.J.; Indias, I.M.; Macías-González, M. Linking serum vitamin D levels with gut microbiota after 1-year lifestyle intervention with Mediterranean diet in patients with obesity and metabolic syndrome: A nested cross-sectional and prospective study. *Gut Microbes* **2023**, *15*, 2249150. [\[CrossRef\]](https://doi.org/10.1080/19490976.2023.2249150)
- 149. Nicholson, I.; Dalzell, A.M.; El-Matary, W. Vitamin D as a therapy for colitis: A systematic review. *J. Crohns Colitis* **2012**, *6*, 405–411. [\[CrossRef\]](https://doi.org/10.1016/j.crohns.2012.01.007)
- 150. Wellens, J.; Vermeire, S.; Sabino, J. The role of carboxymethylcellulose in health and disease: Is the plot thickening? *Gastroenterology* **2022**, *163*, 780–781. [\[CrossRef\]](https://doi.org/10.1053/j.gastro.2022.01.007)
- 151. Deleu, S.; Machiels, K.; Raes, J.; Verbeke, K. Short chain fatty acids and its producing organisms: An overlooked therapy for IBD? *EBioMedicine* **2021**, *66*, 8. [\[CrossRef\]](https://doi.org/10.1016/j.ebiom.2021.103293)
- 152. Crothers, J.W.; Chu, N.D.; Nguyen, L.T.T.; Phillips, M.; Collins, C.; Fortner, K.; Del Rio-Guerra, R.; Lavoie, B.; Callas, P.; Velez, M.; et al. Daily, oral FMT for long-term maintenance therapy in ulcerative colitis: Results of a single-center, prospective, randomized pilot study. *BMC Gastroenterol.* **2021**, *21*, 281. [\[CrossRef\]](https://doi.org/10.1186/s12876-021-01856-9) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34238227)
- 153. Haifer, C.; Paramsothy, S.; Borody, T.J.; Clancy, A.; Leong, R.W.; Kaakoush, N.O. Long-Term Bacterial and Fungal Dynamics following Oral Lyophilized Fecal Microbiota Transplantation in Clostridioides difficile Infection. *mSystems* **2021**, *6*, 10–1128. [\[CrossRef\]](https://doi.org/10.1128/mSystems.00905-20) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33531405)
- 154. Logan, M.; Svolos, V.; Nichols, B.; Milling, S.; Gaya, D.R.; Seenan, J.P.; Macdonald, J.; Hansen, R.; Ijaz, U.Z.; Russell, R.K.; et al. Analysis of 61 exclusive enteral nutrition formulas used in the management of active Crohn's disease—New insights into dietary disease triggers. *Aliment. Pharmacol. Ther.* **2020**, *51*, 935–947. [\[CrossRef\]](https://doi.org/10.1111/apt.15695) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32249975)
- 155. MacLellan, A.; Connors, J.; Grant, S.; Cahill, L.; Langille, M.G.I.; Van Limbergen, J. The Impact of Exclusive Enteral Nutrition (EEN) on the Gut Microbiome in Crohn's Disease: A Review. *Nutrients* **2017**, *9*, 447. [\[CrossRef\]](https://doi.org/10.3390/nu9050447)
- 156. Wellens, J.; Vermeire, S.; Sabino, J. Let Food Be Thy Medicine—Its Role in Crohn's Disease. *Nutrients* **2021**, *13*, 832. [\[CrossRef\]](https://doi.org/10.3390/nu13030832)
- 157. Vanhove, W.; Nys, K.; Arijs, I.; Cleynen, I.; Noben, M.; De Schepper, S.; Van Assche, G.; Ferrante, M.; Vermeire, S. Biopsy-derived intestinal epithelial cell cultures for pathway based stratification of patients with inflammatory bowel disease. *J. Crohn's Colitis* **2017**, *12*, 178. [\[CrossRef\]](https://doi.org/10.1093/ecco-jcc/jjx122)
- 158. Poletti, M.; Arnauts, K.; Ferrante, M.; Korcsmaros, T. Organoid-based Models to Study the Role of Host-microbiota Interactions in IBD. *J. Crohn's Colitis* **2020**, *15*, 1222. [\[CrossRef\]](https://doi.org/10.1093/ecco-jcc/jjaa257)
- 159. Park, J.C.; Im, S.-H. Of men in mice: The development and application of a humanized gnotobiotic mouse model for microbiome therapeutics. *Park. Im. Exp. Mol. Med.* **2020**, *52*, 1383–1396. [\[CrossRef\]](https://doi.org/10.1038/s12276-020-0473-2)
- 160. Scheffers, L.E.; Vos, I.K.; Utens, E.; Dieleman, G.; Walet, S.; Escher, J.; Berg, L.v.D. Physical Training and Healthy Diet Improved Bowel Symptoms, Quality of Life, and Fatigue in Children with Inflammatory Bowel Disease. *J. Pediatr. Gastroenterol. Nutr.* **2023**, *77*, 214–221. [\[CrossRef\]](https://doi.org/10.1097/MPG.0000000000003816)
- 161. Wellens, J.; Vissers, E.; Matthys, C.; Vermeire, S.; Sabino, J. Personalized Dietary Regimens for Inflammatory Bowel Disease: Current Knowledge and Future Perspectives. *Pharmacogenomics Pers. Med.* **2023**, *16*, 15–27. [\[CrossRef\]](https://doi.org/10.2147/PGPM.S359365)

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