Preplanned Studies

Hidden Spread of Rare Salmonella Serovars Isolated from Healthy Individuals — Yulin City, Guangxi Zhuang Autonomous Region, China, 2013–2022

Yao Peng¹; Mengyu Wang¹.²; Changyu Zhou³; Zhenpeng Li¹; Zhe Li¹; Yuan Zhang¹; Biao Kan¹; Ming Luo³.*; Xin Lu¹.*

Summary

What is already known about this topic?

The emergence of rare multidrug-resistant *Salmonella* serovars is increasingly common in developing countries, posing a significant public health risk.

What is added by this report?

This report identified five rare *Salmonella* serovars in China for the first time and using whole-genome sequencing, identified their hidden spread among healthy individuals. Additionally, antibiotic resistance genes and virulence genes were identified.

What are the implications for public health practice?

Healthy individuals, especially food handlers, may be sources of hidden *Salmonella* dissemination. Therefore, monitoring *Salmonella* prevalence in healthy individuals is important for understanding and managing the risks of foodborne illness.

Gastrointestinal infections, predominantly foodborne, are among the most common illnesses worldwide, affecting an estimated 550 million people annually, including 220 million children younger than five years of age (1). Salmonella enterica is a major pathogen implicated in these infections. Salmonella comprises more than 2,500 serovars. Salmonella and Salmonella Typhimurium Enteritidis frequently associated with diarrheal illnesses (2). Recent reports indicate that outbreaks can also be attributed to rare Salmonella serovars. For example, S. Ealing, S. Welikade, S. Isangi, and S. Coeln have been linked to outbreaks in the UK, France, China, and Norway, respectively (3–6).

One Health is an integrated approach to balancing and optimizing the health of people, animals, and ecosystems. It leverages the close, interdependent links among these fields to create new surveillance and disease control methods (7). Healthy individuals,

particularly food handlers, could be potential sources of hidden *Salmonella* dissemination. Therefore, monitoring *Salmonella* prevalence among healthy individuals is essential for comprehensively understanding and managing the associated risks.

Monitoring the prevalence of multidrug-resistant (MDR) *Salmonella* isolates is a common method for assessing the risk of salmonellosis. This approach facilitates timely detection and intervention in cases of salmonellosis, thereby limiting further pathogen dissemination. Whole-genome sequencing (WGS) is widely used to predict and identify serovars, virulence factors, and antimicrobial resistance genes (ARGs) in *Salmonella* surveillance.

Using resistance profiling and genomic analysis of *Salmonella* isolates collected from healthy individuals during a decade-long, large-scale, laboratory-based investigation, we identified five rare *Salmonella* serovars reported for the first time in China and revealed the prevalence of MDR isolates and the hidden dissemination of these serovars.

Between 2013 and 2020, fecal samples were systematically collected from individuals undergoing occupational health examinations in Yulin City, Guangxi Zhuang Autonomous Region, China. Salmonella spp. were cultured from these samples according to a previously established protocol (8). Serovars were determined using slide agglutination and by predicting genome sequences using Salmonella In Silico Typing Resource (SISTR). A systematic literature review using keyword searches for "Salmonella" and "serotype" was performed. Corresponding genomes were retrieved from the EnteroBase and GenBank databases to verify the novelty of the identified serovars within China.

Two commercial antimicrobial susceptibility testing panels (BD Phoenix™ NMIC-413 and BD Phoenix™ RUONMIC-801) were used with the microbroth dilution method, following the manufacturer's

instructions. For clarity, only the common panel of 11 antibiotics is presented, namely: Amikacin (AMK), Ceftazidime (CAZ), Chloramphenicol (CHL), Ciprofloxacin (CIP), Colistin (COL), Ertapenem (ETP), Meropenem (MEM), Piperacillin-Tazobacta (TZP), Tetracycline (TET), Tigecycline (TIG), Trimethoprim-Sulfamethoxazole (SXT). Results were interpreted in accordance with the Clinical and Laboratory Standards Institute (CLSI) 2022 guidelines. *Escherichia coli* ATCC 25922 served as the quality control strain.

Genomic DNA was extracted from 15 bacterial isolates using the Wizard Genomic DNA Purification Kit (Promega) after incubation. Paired-end libraries with insert sizes of 300-500 bp were constructed and sequenced on the MGI200 platform according to the manufacturer's protocol. The sequencing reads for each isolate were assembled using SPAdes v3.5.0. ARGs, plasmid profiles, and virulence factors were identified using Abricate software and databases, including CARD, PlasmidFinder, and VFDB.

To trace the origins of the *S*. Ouakam and *S*. Ealing serotype epidemics, 79 *S*. Ouakam and 114 *S*. Ealing isolates were sourced from the EnteroBase database. These isolates were analyzed using single-nucleotide polymorphisms (SNPs) and hierarchical clustering of core genome multilocus sequence typing (HierCC). The analyses were performed using the online analytical pipeline within EnteroBase. The resulting data were used to generate a phylogenetic tree and

visualized using the iTOL v6 platform.

A total of 7,044 Salmonella isolates were recovered from 372,708 individual samples in a retrospective investigation conducted from 2013 to 2022, with an isolation rate of 1.89% among healthy individuals. Among these, 119 distinct Salmonella enterica serovars were identified in healthy individuals from Yulin. Utilizing an extensive literature review, we report the first identification of five rare Salmonella serovars in China. A total of 14 isolates of rare serovars were sourced from individuals ranging in age from 17 to 60 years, comprising 86% (12/14) males and 14% (2/14) females. Notably, 93% (13/14) of the individuals were engaged in food handling, while the remaining 7% (1/14) were not. The isolates comprised five rare serovars: S. Welikade (n=1), S. Mountpleasant (n=1), S. Coeln (n=1), S. Ealing (n=2), and S. Ouakam (n=9).

S. Welikade, S. Mountpleasant, and S. Ealing isolates were susceptible to all 11 antibiotics tested. In contrast, the S. Coeln isolate and nine S. Ouakam isolates were MDR. Both exhibited resistance to CHL, SXT, and TET. Notably, the S. Coeln isolate was resistant to TIG, an antibiotic typically reserved for severe Gram-negative infections. Meanwhile, all nine S. Ouakam isolates were resistant to CAZ and CIP (Table 1).

Analysis of the 15 isolates' genomes revealed five distinct plasmid types. The two *S.* Ealing isolates lacked plasmids, whereas the *S.* Mountpleasant isolate harbored an IncQ1 plasmid. The *S.* Welikade isolate

TABLE 1. Antimicrobial resistance profile of rare serovar isolates from Yulin City, Guangxi Zhuang Autonomous Region, China.

Strain	Serovars	Year of isolation	Antimicrobial resistance profile
Sa13427	Salmonella Welikade	2013	-
Sa15171	Salmonella Mountpleasant	2016	-
Sa17519	Salmonella Ouakam	2020	CHL-SXT-TET-CAZ-CIP
Sa17528	Salmonella Ouakam	2020	CHL-SXT-TET-CAZ-CIP
Sa17531	Salmonella Ouakam	2020	CHL-SXT-TET-CAZ-CIP
Sa17537	Salmonella Ouakam	2020	CHL-SXT-TET-CAZ-CIP
Sa17538	Salmonela Ouakam	2020	CHL-SXT-TET-CAZ-CIP
Sa17539	Salmonella Ouakam	2020	CHL-SXT-TET-CAZ-CIP
Sa17541	Salmonella Ouakam	2020	CHL-SXT-TET-CAZ-CIP
Sa17542	Salmonella Ouakam	2020	CHL-SXT-TET-CAZ-CIP
Sa17544	Salmonella Ouakam	2020	CHL-SXT-TET-CAZ-CIP
Sa20877	Salmonella Coeln	2022	CHL-SXT-TET-TIG
Sa21032	Salmonella Ealing	2022	-
Sa21220	Salmonella Ealing	2022	_

Note: "-" indicates isolates with no antimicrobial resistance profile.

contained both Col440I and p0111 plasmids. All *S.* Ouakam and *S.* Coeln isolates carried the IncHI2/IncHI2A plasmid (Figure 1).

The *S*. Ouakam isolates possessed diverse ARGs, conferring resistance to aminoglycosides, sulfonamides, lincosamides, tetracyclines, beta-lactams, chloramphenicol, quinolones, and rifamycins. The *S*. Coeln isolates exhibited resistance mediated by a broader spectrum of 14 ARGs, attributed to the presence of IncHI2/IncHI2A plasmids. Without these plasmids, the *S*. Welikade, *S*. Mountpleasant, and *S*. Ealing isolates displayed a more restricted resistance profile, predominantly featuring a single ARG, *aac*(*6*')-

Iaa (Figure 1).

Our investigation revealed 147 distinct virulence genes within the *Salmonella* isolates, highlighting multiple mechanisms underlying virulence and pathogenicity. The *cdtB* gene for typhoid toxin production was found in *S.* Welikade, while *astA* (EAST1 toxin) was detected in the Sa17631 isolate of *S.* Ouakam. Furthermore, all rare serotype isolates harbored gene clusters involved in iron uptake (*entA*, *entB*, *entC*, *entD*, *entE*, *entF*, *entS*) and the *mig-14* gene, associated with resistance to antimicrobial peptides (Figure 1).

We analyzed a dataset of 34,408 SNPs to elucidate

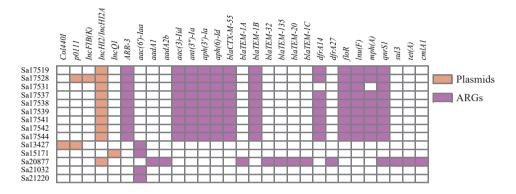


FIGURE 1. Distribution of plasmids and antimicrobial resistance genes (ARGs) among the studied isolates.

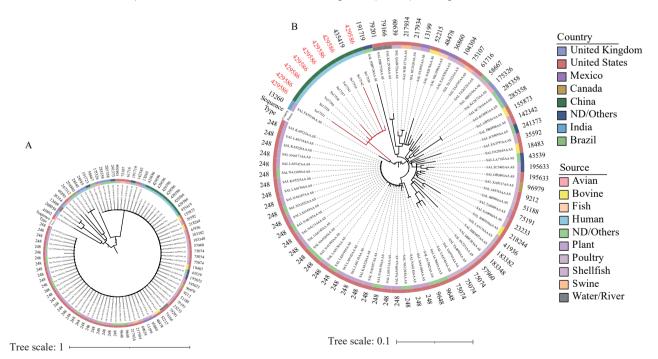


FIGURE 2. Phylogenetic tree of *Salmonella Ouakam* isolates. (A) Phylogenetic structure of 88 *Salmonella* Ouakam isolates, including 79 isolates from EnteroBase and 9 isolates from Yulin. (B) Phylogenetic tree of isolates belonging to group 2 in Figure 2A.

Note: The isolates in red font represent those collected in this study.

the phylogenetic relationships among 88 S. Ouakam genomes. The resulting phylogenetic tree revealed two distinct groups (Figure 2A). To facilitate further analysis, we reconstructed the phylogenetic tree specifically for the isolates of group 2 (Figure 2B). Nine S. Ouakam isolates, derived from healthy individuals, formed a cluster within clade demonstrating SNP distances ranging from 0 to 58. To define outbreak clusters accurately, we employed the internationally recognized HierCC HC5 standard, which is widely used in infectious disease outbreaks (9). Seven S. Ouakam isolates classified under the same HC5 cluster (type 428586) were obtained from individuals employed by the same company, indicating the potential for hidden transmission of Salmonella spp. among this population.

Analysis of 116 isolates allowed for phylogenetic tree reconstruction for S. Ealing, revealing two distinct groups. Within group B, two clades, designated α and

 β , were identified. Isolates recovered in this study, characterized by HC5 type 435357, clustered within clade α . Two additional isolates within clade α , originating from plants in Paraguay and the Netherlands, corresponded to HC5 types 268261 and 214748, respectively. The remaining four isolates within clade α were all characterized by HC5 type 353819 (Figure 3).

DISCUSSION

From 2014 to 2021, the top 10 serovars among sporadic clinical *Salmonella* isolates in China were Typhimurium, Enteritidis, I 4,[5],12:i:-, London, Stanley, Derby, Rissen, Thompson, Agona, and Goldcoast. In southern China, Typhimurium and I 4,[5],12:i:- were the most prevalent serovars (10). Although none of the five rare serovars identified in

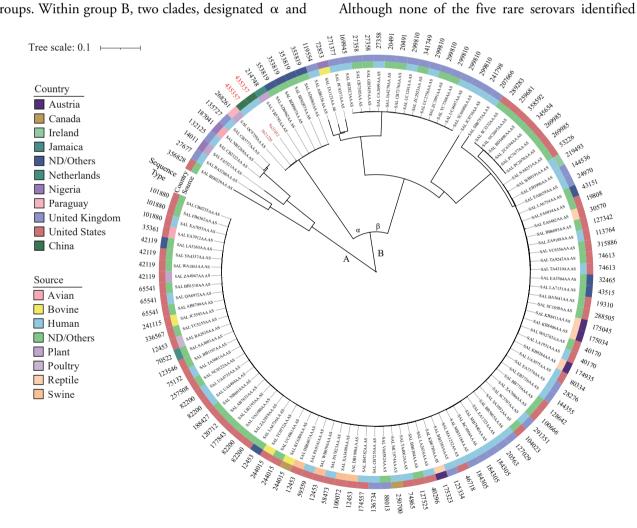


FIGURE 3. The phylogenetic tree of 116 *S*. Ealing isolates, including 114 isolates from EnteroBase and 2 isolates from Yulin City, Guangxi Zhuang Autonomous Region, China.

Note: The isolates collected from Yulin are highlighted in red.

this study have been found in clinical isolates in China, previous reports have documented the presence and dissemination of rare Salmonella serovars across various geographic locations, impacting both agricultural and public health sectors. S. Ealing has been identified in turkey production in the USA (11), within food samples in Nigeria (12), and among poultry in Brazil (13). In 1985, an outbreak of S. Ealing affecting infants was reported in the UK (3). Outbreaks of S. Coeln linked to minced meat and salad consumption were recorded in France in 1998 and Norway from 2013 to 2014, respectively (6). Data on S. Welikade are scarce, with isolated reports from France, Sri Lanka, Australia, and Sweden (4). No reports concerning S. Mountpleasant have been identified. Furthermore, S. Ouakam isolates have been sourced from a variety of contexts, including ground turkey, turkey breakfast sausage (14), retail poultry, pigs, the swine environment (15), and chicken meat (16) from children under five with diarrhea in rural Burkina Faso (17). This study is the first to elucidate the genomic characteristics linked to virulence and resistance in five Salmonella serovars isolated from healthy individuals in China. Through analysis of these genomic features, we aim to contribute evidence to improve risk assessment and deepen understanding of the determinants of antimicrobial resistance and pathogenicity among these emerging pathogens.

Seven S. Ouakam isolates were collected from healthy individuals, all employees at the same enterprise, within one week. Furthermore, classification within the same HC5 cluster suggests potential hidden, asymptomatic Salmonella transmission. The isolation of S. Ealing strains of the same HC5 type nearly one month apart suggests that this serovar may exhibit relative stability over a short period, indicating the potential for sustained transmission and the presence of asymptomatic carriers, which increases the risk of spread. Therefore, monitoring transmission dynamics and detecting asymptomatic carriers are crucial for controlling its spread and preventing disease.

This study was subject to at least two limitations. First, the food strains, which asymptomatic carriers may come into contact with, were not collected, so the transmission of strains from food to humans could not be determined. Second, there were no detailed epidemiological data to confirm an outbreak of *S.* Ouakam among the asymptomatic employees. Over a 10-year surveillance period, we documented the first domestic identification of five rare *Salmonella* serovars.

Genomic sequencing and phenotypic resistance profiling revealed that these rare serovar isolates were MDR and harbored ARGs. Phylogenetic analysis further revealed the undetected dissemination of *S.* Ouakam. This study underscores the critical importance of active surveillance among healthy individuals, especially within key occupational groups such as food workers, to monitor foodborne pathogens.

Conflicts of interest: No conflicts of interest.

Funding: Supported by the National Key Research and Development Program of China (2022YFC2303900), and the Major Projects of the National Natural Science Foundation of China (22193064).

doi: 10.46234/ccdcw2024.248

* Corresponding authors: Xin Lu, luxin@icdc.cn; Ming Luo, ylcdc_lm@163.com.

Submitted: July 29, 2024; Accepted: November 14, 2024

REFERENCES

- WHO. Salmonella (non-typhoidal). World Health Organization. 2018. https://www.who.int/news-room/fact-sheets/detail/salmonella-(non-typhoidal).[2024-10-20].
- Hendriksen RS, Vieira AR, Karlsmose S, Lo Fo Wong DMA, Jensen AB, Wegener HC, et al. Global monitoring of Salmonella serovar distribution from the World Health Organization global foodborne infections network country data bank: results of quality assured laboratories from 2001 to 2007. Foodborne Pathog Dis 2011;8(8):887 900. https://doi.org/10.1089/fpd.2010.0787.
- Gunn L, Finn S, Hurley D, Bai L, Wall E, Iversen C, et al. Molecular characterization of *Salmonella* serovars anatum and ealing associated with two historical outbreaks, linked to contaminated powdered infant formula. Front Microbiol 2016;7:1664. https://doi.org/10.3389/fmicb. 2016.01664.
- Cherchame E, Guillier L, Lailler R, Vignaud ML, Jourdan-Da Silva N, Le Hello S, et al. *Salmonella enterica* subsp. enterica Welikade: guideline for phylogenetic analysis of serovars rarely involved in foodborne outbreaks. BMC Genomics 2022;23(1):217. https://doi.org/10.1186/ s12864-022-08439-2.
- 5. Li XP, Gao RH, Hou PB, Ren YY, Zhang HN, Jiang KY, et al. Characterization of the *Salmonella enterica* serotype Isangi isolated from patients for the first time in China. Foodborne Pathog Dis 2017;14(8): 427 31. https://doi.org/10.1089/fpd.2016.2269.
- Vestrheim DF, Lange H, Nygård K, Borgen K, Wester AL, Kvarme ML, et al. Are ready-to-eat salads ready to eat? An outbreak of Salmonella Coeln linked to imported, mixed, pre-washed and bagged salad, Norway, November 2013. Epidemiol Infect 2016;144(8):1756 60. https://doi.org/10.1017/S0950268815002769.
- 7. WHO. One health. World Health Organization. 2023. https://www.who.int/news-room/fact-sheets/detail/one-health.[2024-10-23].
- 8. Lu X, Zeng M, Xu JL, Zhou HJ, Gu BK, Li ZP, et al. Epidemiologic

¹ National Key Laboratory of Intelligent Tracking and Forecasting for Infectious Diseases, National Institute for Communicable Disease Control and Prevention, Chinese Center for Disease Control and Prevention, Beijing, China; ² School of Public Health, Cheeloo College of Medicine, Shandong University, Jinan City, Shandong Province, China; ³ Yulin Center for Disease Control and Prevention, Yulin City, Guangxi Zhuang Autonomous Region, China.

- and genomic insights on *mcr-1*-harbouring *Salmonella* from diarrhoeal outpatients in Shanghai, China, 2006-2016. EBioMedicine 2019;42: 133 44. https://doi.org/10.1016/j.ebiom.2019.03.006.
- Leeper MM, Tolar BM, Griswold T, Vidyaprakash E, Hise KB, Williams GM, et al. Evaluation of whole and core genome multilocus sequence typing allele schemes for Salmonella enterica outbreak detection in a national surveillance network, PulseNet USA. Front Microbiol 2023;14:1254777. https://doi.org/10.3389/fmicb.2023. 1254777.
- Li WW, Han HH, Liu JK, Ke BX, Zhan L, Yang XR, et al. Antimicrobial resistance profiles of *Salmonella* isolates from human diarrhea cases in China: an eight-year surveilance study. One Health Adv 2023;1(1):2. https://doi.org/10.1186/s44280-023-00001-3.
- Guo L, Killefer J, Kenney P, Amick-Morris J. Use of arbitrarily primed polymerase chain reaction to study Salmonella ecology in a turkey production environment. Poult Sci 1999;78(1):24 – 31. https://doi.org/ 10.1093/ps/78.1.24.
- Smith S, Braun S, Akintimehin F, Fesobi T, Bamidele M, Coker A, et al. Serogenotyping and antimicrobial susceptibility testing of *Salmonella* spp. isolated from retail meat samples in Lagos, Nigeria. Mol Cell Probes 2016;30(4):189 94. https://doi.org/10.1016/j.mcp.2016.04.001.

- Rodrigues IBBE, Silva RL, Menezes J, Machado SCA, Rodrigues DP, Pomba C, et al. High prevalence of multidrug-resistant nontyphoidal salmonella recovered from broiler chickens and chicken carcasses in Brazil. Braz J Poult Sci 2020;22(1):eRBCA – 2019-1206. https://doi. org/10.1590/1806-9061-2019-1206.
- 14. Marasini D, Abo-Shama UH, Fakhr MK. Whole-genome sequencing of *Salmonella enterica* subsp. enterica serovar Ouakam isolated from ground turkey. Genome Announc 2016;4(1):e01618 15. https://doi.org/10.1128/genomeA.01618-15.
- Keelara S, Thakur S. Dissemination of plasmid-encoded AmpC β-lactamases in antimicrobial resistant Salmonella serotypes originating from humans, pigs and the swine environment. Vet Microbiol 2014;173(1-2):76 83. https://doi.org/10.1016/j.vetmic.2014.07.018.
- Gad AH, Abo-Shama UH, Harclerode KK, Fakhr MK. Prevalence, serotyping, molecular typing, and antimicrobial resistance of *Salmonella* isolated from conventional and organic retail ground poultry. Front Microbiol 2018;9:2653. https://doi.org/10.3389/fmicb.2018.02653.
- 17. Dembélé R, Konaté A, Bonkoungou IJO, Kagambèga A, Konaté K, Serge BT, et al. Serotyping and antimicrobial susceptibility of Salmonella isolated from children under five years of age with diarrhea in rural Burkina Faso. Afr J Microbiol Res 2014;8(34):3157 63. https://doi.org/10.5897/AJMR2014.7002.