RESEARCH ARTICLE

RNA mis-splicing in children with congenital myotonic dystrophy is associated with physical function

Julia M. Hartman^{1,2,3,4} , Kobe Ikegami^{2,3}, Marina Provenzano^{2,3}, Kameron Bates^{2,3}, Amanda Butler^{2,5}, Aileen S. Jones^{2,5}, Kiera N. Berggren^{2,3}, Jeanne Dekdebrun⁶, Marnee J. McKay⁷, Jennifer N. Baldwin⁷, Kayla M. D. Cornett^{7,8}, Joshua Burns^{7,8}, Michael Kiefer^{2,9}, Nicholas E. Johnson^{2,3,4}, Melissa A. Hale^{2,3,4}, & on behalf of the DMCRN Consortium

¹Medical Scientist Training Program, Virginia Commonwealth University, Richmond, Virginia, 23298, USA

²Center for Inherited Myology Research, Virginia Commonwealth University, Richmond, Virginia, 23298, USA

³Department of Neurology, Virginia Commonwealth University, Richmond, Virginia, 23298, USA

⁴Department for Human and Molecular Genetics, Virginia Commonwealth University, Richmond, Virginia, 23298, USA

⁵Children's Hospital of Richmond at Virginia Commonwealth University, Pediatric Therapy Services, Richmond, Virginia, 23220, USA

⁶Department of Neurology, University of Rochester School of Medicine and Dentistry, Rochester, New York, 14642, USA

 7 Sydney School of Health Sciences, Faculty of Medicine and Health, The University of Sydney, Sydney, New South Wales, 2006, Australia

⁸Sydney Children's Hospitals Network (Randwick and Westmead), Sydney, New South Wales, Australia

Abstract

⁹Department of Physical Therapy, Virginia Commonwealth University, Richmond, Virginia, 23298, USA

Correspondence

Melissa A. Hale, Department of Neurology, Virginia Commonwealth University, Richmond, VA 23298, USA. Tel: 804-506-3248; Fax: 804-828-8534; E-mail: melissa. hale@vcuhealth.org

Received: 11 September 2024; Accepted: 14 September 2024

Annals of Clinical and Translational Neurology 2024; 11(12): 3175–3191

doi: 10.1002/acn3.52224

Introduction

Myotonic dystrophy type 1 (DM1) is the most common form of muscular dystrophy with a prevalence of 1 in

2100.¹ DM1 is inherited in an autosomal dominant manner and is caused by a CTG trinucleotide repeat expansion (CTG_n) in the 3' untranslated region of the dystrophia myotonica protein kinase (*DMPK*) gene.^{2–4}

Objectives: Dysregulated RNA alternative splicing is the hallmark of myotonic

dystrophy type 1 (DM1). However, the association between RNA mis-splicing and

physical function in children with the most severe form of disease, congenital

myotonic dystrophy (CDM), is unknown. **Methods**: Eighty-two participants (42 adults with DM1 and 40 children with CDM) with muscle biopsies and measures of myotonia, motor function, and strength were combined from five observational

studies. Data were normalized and correlated with an aggregate measure of alter-

native splicing dysregulation, [MBNL]_{inferred}, in skeletal muscle biopsies. Multiple linear regression analysis was performed to predict [MBNL]_{inferred} using clinical

outcome measures alone. Similar analyses were performed to predict 12-month physical function using baseline metrics. **Results**: Myotonia (measured via vHOT)

was significantly correlated with RNA mis-splicing in our cross-sectional population of all DM1 individuals; CDM participants alone displayed no myotonia despite a similar range of RNA mis-splicing. Measures of motor performance and muscle strength were significantly associated with [MBNL]_{inferred} in our cohort of all DM1 individuals and when assessing children with CDM independently. Multiple linear regression analyses yielded two models capable of predicting [MBNL]_{inferred} from select clinical outcome assessments alone in all subjects (adjusted $R^2 = 0.6723$) or exclusively in children with CDM (adjusted $R^2 = 0.5875$). **Interpretation**: Our findings establish significant correlations between skeletal muscle performance and a composite measure of alternative splicing dysregulation, [MBNL]_{inferred}, in DM1. The strength of these correlations and the development of predictive models will assist in designing efficacious clini-

cal trials for individuals with DM1, particularly CDM.

Although nearly every organ system can be affected, the core clinical features of DM1 include progressive distal muscle weakness, early onset cataracts, and myotonia (i.e., delayed muscle relaxation following contraction).^{5,6}

Congenital myotonic dystrophy (CDM) is the most severe form of DM1 and results from large, intergenerational CTG_n expansion between parent and child.^{7–9} CDM presents at birth with symptoms of hypotonia, clubfoot, feeding difficulties, and respiratory distress.^{10–12} In contrast to adults with DM1 who present with a consistent, progressive decline in muscle function over time, children with CDM exhibit a natural improvement and stabilization of motor function and muscle performance in early childhood. As children with CDM progress through adolescence and into early adulthood, symptoms become more consistent with adult-onset DM1.^{12,13}

In DM1, the core pathogenesis is the sequestration of muscleblind-like (MBNL) RNA binding proteins by toxic, expanded CUG_n RNAs within nuclear aggregates termed foci. This leads to an overall reduction in the functional concentration of MBNL in affected tissues and subsequent dysregulation of RNA metabolism.^{14,15} As critical regulators of fetal to adult mRNA isoform transitions, depletion of functional MBNL levels leads to global perturbations in splicing regulation and reversion of transcripts to the fetal isoform in affected tissues.^{16–20} Numerous transcripts are mis-spliced, although only a select few have been linked to disease phenotypes in DM1 cell and animal models, including *CLCN1* and myotonia,^{21,22} *SCN5A* and cardiac arrhythmia,²³ and *BIN1* and muscle weakness.²⁴

While RNA mis-splicing of individual events has been correlated with ankle dorsiflexion performance and manual muscle testing in individuals with adult-onset DM1,^{25,26} this relationship has not been replicated using other measures of physical performance. Additionally, no such associations have been evaluated in children with CDM. Previous work by our group has characterized global splicing dysregulation in skeletalmuscle of DM1 adults and a cohort of children with CDM using a composite measure of MBNL-dependent splicing, [MBNL]_{inferred}. This representative metric of free intracellular MBNL concentration strongly correlates with global splicing dysregulation as captured by total RNA sequencing.^{27,28}

Using this data, we aimed to investigate if transcriptome-wide RNA mis-splicing as measured by [MBNL]_{inferred} correlates with a wider breadth of functional measures assessing myotonia, motor function, and strength in a cross-sectional cohort of adults with DM1 and children with CDM. Additionally, we sought to evaluate if [MBNL]_{inferred} levels in individuals with CDM correlate with the patterns of functional improvement observed throughout childhood in this affected

population. Using these correlative analyses, we developed regression models to predict [MBNL]_{inferred} using measures of physical function. This model may offer a noninvasive alternative for predicting disease-associated mis-splicing in affected individuals. Lastly, we developed regression models to predict 12-month physical function using baseline performance and baseline [MBNL]_{inferred} values.

Methods

Study design

Clinical phenotypes and associated biopsies from five multisite longitudinal observational studies were used to determine the correlation between splicing dysregulation (as measured by [MBNL]_{inferred}) and physical function in a cohort of children with CDM and adults with DM1. Muscle biopsies were collected at a baseline visit. Clinical assessments were completed at baseline and at a 3-month visit and/or at a 12-month visit depending on the study they were enrolled in. Participants provided informed consent before enrollment per the local site's Institutional Review Board. For participants under 18, written informed consent was obtained from one parent and verbal assent from children over the age of 8.

Children with CDM between the ages of 0–17 (inclusive) were enrolled in the HELP-CDM, TREAT-CDM, or ASPIRE studies (NCT03059264, NCT05224778).¹³ Diagnostic criteria for CDM were defined as symptoms of myotonic dystrophy in the newborn period (<30 days) including hypotonia, respiratory distress, feeding difficulty, or clubfoot requiring hospitalization greater than 72 h, and a genetic test confirming an expanded trinucle-otide CTG_n repeat in the *DMPK* gene (CTG repeats greater than 200) or an affected mother. Exclusion criteria were described previously.¹³

Adults with DM1 over the age of 18 were enrolled in HELP-DM1 or END-DM1 studies (NCT03981575). Participants were enrolled if they had a clinical diagnosis of DM1 or a positive genetic test confirming an expanded CTG repeat in the *DMPK* gene. They were excluded if they had symptomatic renal or liver disease, uncontrolled diabetes mellitus or thyroid disorders, or were pregnant. Mexiletine or other anti-myotonia agents were required to be stopped at least 72 hours prior to a study visit.

Select participants provided a muscle biopsy as part of an in-house biorepository study. While clinical assessments were not available for these participants, [MBNL]_{inferred} was calculated and these values were used to further illustrate the dynamic range of mis-splicing observed across the lifespan in Figure 1.

Clinical assessments

Myotonia was assessed viaparticipant (or caregiver if under 12) report and clinical observation. Participants rated their myotonia severity on a 6-point scale from "1, I don't experience this" to "6, It affects my life severely" as part of the Myotonic Dystrophy Health Index (MDHI) or Congenital and Childhood Myotonic Dystrophy Health Index (CCMDHI).²⁹ Clinical myotonia was observed using video hand opening time (vHOT),^{30,31} in which the time to extend the thumb after 4 sec of maximal hand flexion contraction is calculated via video recording of the procedure. The assessment was modified for children to squeeze a rubber toy for improved understanding of the task and time was scored live using a stopwatch.

Motor function measures included the 9-hole peg test,³² 6-min walk,^{33,34} 4 stair climb,³⁵ and 10-meter walk/ run test³⁵ administered according to previously described methods. For adults enrolled in HELP-DM1, a 10-stair climb with railing as described in the modified dynamic gait index (mDGI) was performed.³⁶

Strength measures for knee extension,³⁷ grip,¹³ and ankle dorsiflexion¹³ were assessed via Quantitative Myometry (QMT) according to previously published methods. In adults, a fixed QMT system with a force transducer attached to a metal frame was used to provide adequate stabilization due to high force outputs. In children, handheld dynamometry was used to reduce the administration burden. Strength was measured in kilogram-force (kgf) units and converted to Newtons (N) by standard conversion of 9.80665 (e.g., 1 kgf = 9.80665 N).

Muscle biopsy collection and derivation of [MBNL]_{inferred}

Muscle biopsies from adult DM1 participants were collected with either a Bergstrom or 14-gauge argon Supercore needle of the tibialis anterior (TA) as previously described.³⁸ Muscle biopsies of the vastus lateralis from CDM children were obtained using either an open surgical technique or a 14-gauge Argon Supercore needle.²⁸ In all studies, biopsies were not performed in individuals with known bleeding disorders, a history of anti-coagulation medication use, or a platelet count less than 50,000. To participate in the muscle biopsy, ankle dorsiflexion strength had to be between 4+ and 4– on the Medical Research Council (MRC) scale for muscle strength.

Total RNA sequencing was performed on RNA extracted from all muscle biopsies, and [MBNL]_{inferred} values were derived for each individual as previously described.^{27,28} In brief, percent-spliced in (PSI) values from 9 skipped exon splicing events with high predictive

power of overall global mis-splicing in DM1 muscle were used to calculate [MBNL]_{inferred}.^{27,28} Given that we were utilizing additional samples, [MBNL]_{inferred} values were recalculated using all participants. CDM participant sub-cohort classifications were previously defined and based on age at biopsy.²⁸ The CDM sub-cohorts are defined as (i) CDM_{infant} (\leq 2 years), (ii) CDM_{child} (>2– 8 years), and (iii) CDM_{adolescent} (\geq 8 years).²⁸ Individual PSI values for *CLCN1* exon 7a and *CACNA1S* exon 29 events were derived from comparisons between each affected group (CDM sub-cohort or DM1 adults) and associated age-matched, unaffected individuals as previously reported.²⁸

Statistical analyses

Motor function and strength measures were converted to percent predicted of sex and age-matched healthy controls for analysis. Percent predicted for walk/running speed was calculated using previously published data^{39,40} and percent predicted for the 6-min walk test, stair climb speed, 9-hole peg test, grip strength, ankle dorsiflexion, and knee extension were calculated using raw data from the 1000 Norms Project.^{41,42}

Study data were collected and managed using RED-Cap® electronic data capture tools^{43,44} and exported to Microsoft Excel Version 16.75 for analysis. Statistical analyses were performed using GraphPad Prism 10.1.0, and *p*-values <0.05 were considered significant. Clinical data points inconsistent with the cohort distribution were verified with the original data sources by the clinical evaluators and subsequently reviewed by the study's principal investigator (P.I.).

Univariate correlations and multiple linear regression were performed in GraphPad Prism. Univariate Spearman correlations assumed data are not sampled from Gaussian distributions with a two-tailed p-value and a 95% confidence interval. Given that myotonia was measured using a self—/parent-reporting scale, data were assumed to be nonparametric. Responses were accumulated between the various DM1 subgroups, and statistical significance between the group medians was evaluated using a Kruskal–Wallis test. Dunn's test was used to correct for multiple comparisons. To test for the overall difference between total adult DM1 and CDM myotonia scale responses, a Mann–Whitney test was used that assumed non-Gaussian distributed data with a two-tailed *p*-value.

Multiple linear regression analysis assumed a least squares regression, and adjusted R-squared was used to quantify goodness-of-fit. D'Agostino-Pearson omnibus normality test, Anderson-Darling test, Shapiro–Wilk normality test, and the Kolmogorov–Smirnov normality test with Dallal–Wilkinson–Lillie for p-value were all performed alongside multiple linear regression analysis to ensure data normality. A 95% confidence level was used.

Results

Evaluation of [MBNL]_{inferred} in skeletal muscle biopsies from children with CDM and adults with DM1

We had previously used total RNA sequencing to characterize MBNL-dependent splicing dysregulation in skeletal muscle samples from subjects with CDM and DM1 and calculated [MBNL] inferred, an aggregate metric of RNA mis-splicing representative of estimated intracellular concentrations of free MBNL. [MBNL]_{inferred} values range from 0 to 1, with 0 indicating high MBNL depletion consistent with significant mis-splicing and 1 representative of intracellular MBNL concentrations comparable to unaffected individuals. Using this methodology, we identified a triphasic modality of RNA mis-splicing progrespediatric sion across development within our cross-sectional cohort of children with CDM which appeared to mirror the triphasic phenotypic progression observed.²⁸ In brief, individuals with CDM under the age of 2 (CDM_{infant}) presented with severe mis-splicing consistent with the severity of disease presentation at birth. This is followed by a universal and significant reversal of splicing dysregulation in early childhood (CDM_{child}, 2-8 years). This observation is supported by the longitudinal sampling of CDM-01 at 2 weeks and 8 years of age where a marked increase in [MBNL]_{inferred} was observed ([MBNL]_{inferred} = 0.015 and 0.577, respectively). RNA mis-splicing universally improved in early childhood (CDM_{child}, 2-8 years) with some individuals approaching

splicing patterns like that of unaffected age-matched individuals. Sampled adolescent children displayed a spectrum of [MBNL]_{inferred} values, indicating a wide range of global splicing dysregulation. Post 8 years of age, a gradient of [MBNL]_{inferred} in CDM_{adolescent} was observed and mirrors that of individuals with adult-onset DM1, for which a full range of splicing dysregulation is observed. This triphasic pattern of RNA mis-splicing throughout pediatric development in children with CDM is visualized in Figure 1.

To assess potential correlations between skeletal muscle performance measures and alternative splicing dysregulation, we aggregated CDM and DM1 subjects that (i) received a muscle biopsy, (ii) had a [MBNL]_{inferred} score derived from matched skeletal muscle RNA, and (iii) possessed clinical outcomes from the same associated visit. While a total of 116 muscle biopsies were available from 82 unique participants across the five natural history studies and an in-house biorepository study (Fig. 1, Table S1), only 101 of the biopsies had associated clinical outcome data (n = 31 CDM & 70 DM1, Table S2). The finalized cross-sectional cohort that we used for the analyses herein contains participants with time point matched [MBNL]_{inferred} scores and clinical outcomes from either baseline visits (children and adults), 3-month visits (adults only), or sporadic longitudinal sampling (three children with CDM only, indicated in Fig. 1 and Table S1). Additional clinical information from participants at 12-month visits was collected for predictive modeling even though there was no associated muscle biopsy, and therefore no available [MBNL]inferred score, from that timepoint (Tables S1 and S2). One sample (CDM-38) was excluded from data analysis as the biopsy was collected from the soleus. Baseline visit characteristics including mean [MBNL]_{inferred} and measures of myotonia, motor



Figure 1. [MBNL]_{inferred} values calculated from DM1 and CDM skeletal muscle total RNA sequencing across pediatric development and adulthood. [MBNL]_{inferred} values range from 0 to 1 and inversely correlate with global mis-splicing severity. CDM infants (CDM_{infant}; \leq 2 years) are shown in yellow. CDM children (CDM_{child}; 2–8 years) are shown in green; CDM adolescent (CDM_{adolescent}; 8–16 years) individuals are shown in blue. Adult DM1 (20–69 years) individuals are shown in purple. CDM-01 longitudinal biopsies shown as black-outlined squares. CDM-30 longitudinal biopsies are shown as black-outlined circles. CDM-37 longitudinal biopsies are shown as black-outlined.

		CDM		DM1	
Baseline characteristics	Infant	Child	Adolescent	Adults	
Sample size (n)	7	15	20	42	
Biological sex (n)					
Male	6	11	9	17	
Female	1	4	11	25	
Age (years)	0.76 ± 0.7	5.2 ± 1.5	10.9 ± 2.9	39.3 ± 10.2	
CTG repeat length	1600 ± 141.4	1035 ± 334.6	1191 ± 540.6	426 ± 253.5	
[MBNL] _{inferred}	0.29 ± 0.3	0.69 ± 0.2	0.61 ± 0.3	0.47 ± 0.2	
Biopsy site	Vastus lateralis	Vastus lateralis	Vastus lateralis	Tibialis anterior	
Clinical outcome measures					
Myotonia Scale	1	2.1 ± 1.4	2.4 ± 1.5	3.2 ± 1.5	
Myotonia vHOT _{thumb} (s)	NA	0.5 ± 0.2	1.1 ± 1.1	7.5 ± 8.4	
6-minute walk (m)	NA	338 ± 91	365± 140	377 ± 103	
Walk/running speed (m/s)	NA	1.9 ± 0.6	2.1 ± 0.74	1.99 ± 0.9	
Stair climb speed (stairs/s)	NA	1.2 ± 0.4	1.2 ± 0.7	1.7 ± 0.7	
9-hole peg test (s)	NA	47.7 ± 14.7	44.6± 31.9	20.1 ± 6.6	
Grip strength (kgf)	NA	4.1 ± 1.5	7.1 ± 5	13.9 ± 8.5	
Ankle dorsiflexion (kgf)	NA	5.0 ± 2.9	6 ± 3.7	8 ± 5.4	
Knee extension (kgf)	NA	8.2 ± 2.6	9.6 ± 3.8	22.1 ± 9.0	

Table 1.	Characteristics	of	CDM	and DM1	participants	at	baseline	visit
----------	-----------------	----	-----	---------	--------------	----	----------	-------

Mean \pm one SD reported unless otherwise indicated.

kgf, kilogram-force; m, meters; n, number of samples; s, seconds.

function, and strength are presented in Table 1 for all disease subcohorts. Raw clinical outcome data for all participants at all biopsy-matched timepoint sampling can be found in Table S2.

RNA mis-splicing correlates with myotonia measures in all participants with DM1 but not in children with CDM alone

Myotonia was assessed via two methods - a qualitative participant/caregiver reported score of impact on daily living (from MDHI/CCHDMI) and quantitatively via video of hand opening time (vHOT). The impact of myotonia on a participant's life via participant/caregiver reported scale was found to be significantly different between all groups (p = 0.0079) (Fig. 2A). Given the large observed differences in mean [MBNL]inferred values between CDM subcohorts (Table 1), it is reasonable to infer that [MBNL]_{inferred} values correlate minimally with the impact of myotonia on a participant's life (Fig. 2A). Multiple comparisons testing revealed a significant difference between CDM_{child} responses (median response $\cong 1$) and adult DM1 responses (median response \cong 3) to the myotonia scale (p = 0.0498) (Fig. 2A). When comparing the values of the self-reported scale in adult DM1 individuals to total CDM responses (CDM_{total}), a significant increase in perceived impact of myotonia was observed for adults with DM1 (Fig. 2B).

vHOT was used to quantitatively evaluate clinical myotonia between the DM1 subgroups. vHOT_{thumb} was found to correlate moderately with [MBNL]inferred in all individuals with DM1 (Spearman r = -0.54) (Fig. 2C, left). However, there was no significant correlation observed when examining just participants with CDM despite the comparable range of spliceopathy observed (Spearman r = 0.08) (Fig. 2C, right). To further investigate this phenomenon, we directly examined the relationship between vHOT and CLCN1 exon 7a inclusion (percent spliced in, PSI). Defects in skeletal muscle chloride conductance due to mis-splicing of CLCN1 are postulated to be causative of myotonia and restoration of aberrant exon 7a inclusion corrects this phenotype in DM1 mouse models.⁴⁵ Consistent with this pathogenic mechanism, CLCN1 PSI was significantly correlated with vHOT_{thumb} times in all individuals with DM1 (Spearman r = -0.48). In contrast, this relationship was not observed in participants with CDM alone (Fig. 2D, right). While select CDM_{adolescent} individuals have nearly 100% CLCN1 exon 7a inclusion, like that of their adult DM1 counterparts with the highest vHOT_{thumb} times, minimal myotonia was observed. Mis-splicing of CACNA1S (CaV1.1), a calcium channel that controls skeletal muscle excitation-contraction coupling, has also been associated with exacerbated myopathy and myotonia in DM1.46,47 In our complete cross-sectional cohort, reduced CACNA1S exon 29 inclusion was significantly negatively correlated with longer vHOT_{thumb} times,



Figure 2. Myotonia measures in adult DM1 and CDM participants correlate disparately with skeletal muscle spliceopathy. (A) Myotonia MDH// CCMDHI scale average responses across CDM subcohorts and adult DM1 participants. Results are expressed as median \pm 95% confidence interval (CI) via Kruskal–Wallis test with Dunn's multiple comparisons test. (B) Myotonia scale average value between DM1 individuals and all CDM individuals. Results are expressed as median \pm 95% CI via unpaired Mann–Whitney test. (C) Correlation between [MBNL]_{inferred} and vHOT_{thumb} times in all DM1 individuals (left) and CDM individuals alone (right panel). Adult DM1 measures are removed from the statistical analysis in the right panel but grayed out and included in the figure for visual reference. (D) Correlation between *CLCN1* exon 7a percent spliced in (PSI) and vHOT_{thumb} times in all DM1 individuals and CDM individuals. All correlations are reported from a two-tailed Spearman test.

but not in participants with CDM alone (Fig. S1). Overall, the lack of correlation between CDM vHOT_{thumb} times and either a composite measure of disease-associated spliceopathy or mis-splicing of specific, phenotype-associated events is driven by the lack of quantifiable myotonia within children with CDM throughout development. This is consistent with previous observations that individuals with CDM within the first decade of life do not experience clinical myotonia.⁴⁸ However, our analysis is the first of its kind to demonstrate that this phenomenon occurs even when phenotype-associated RNA mis-splicing occurs at levels comparable to adults with DM1 who present with severe myotonia.

RNA mis-splicing correlates moderately with select motor function measures in all participants with DM1

We next assessed whether global mis-splicing as measured by [MBNL]_{inferred} correlated with several timed motor tests within all DM1 participants or selectively within children with CDM, as the utility of certain clinical outcome measures may vary with age and disease severity.⁴⁹

The 9-hole peg test was significantly correlated, albeit weakly, with [MBNL]_{inferred} values in all individuals with DM1 (Spearman r = -0.23). The relative association significantly improved in children with CDM alone whereby individuals with higher [MBNL]_{inferred} values were able to perform the test much quicker with results more in line with control times (i.e., resulting in a lower % predicted) (Spearman r = -0.66) (Fig. 3A). In contrast to the 9-hole peg test, weak to moderate positive correlations were observed for both 6-minute walk distance and stair climbing speed in all participants with DM1 and in children with CDM alone (Fig. 3B,C). The strongest correlation observed between any motor function measure and [MBNL]_{inferred} was 10-meter walk/running speed. Percent predicted walk/running speed had a strong positive association with [MBNL]_{inferred} levels in all individuals with DM1 that was only minimally reduced when assessing participants with CDM independently (Fig. 3D).

[MBNL]_{inferred} correlates strongly with skeletal muscle strength measures in all participants with DM1

Quantitative muscle testing of knee extension (KE), hand grip (HG), and ankle dorsiflexion (ADF) strength had the strongest associations with [MBNL]_{inferred} levels compared to all other outcome assessments utilized in these analyses. When adults with DM1 and children with CDM were evaluated in combination, higher levels of [MBNL]_{inferred} were strongly correlated with muscle strength; the highest reported association was with ADF (Spearman r = 0.68) (Fig. 4). The strength of these relationships was generally maintained in CDM individuals alone (Fig. 4). Strikingly, the relative association of [MBNL] inferred with ADF in children with CDM was identical to that observed in all participants with DM1 (Spearman r = 0.69), in part due to the range of ADF performance captured in CDM_{adolescent} participants that replicated the range observed in adults (Fig. 4C, blue and purple samples, respectively). Consistent with previous reports of correlations between RNA mis-splicing and muscle strength, we found that measures of strength correlate strongly with [MBNL]_{inferred} in this assembled DM1 cohort.

[MBNL]_{inferred} can be predicted using physical function measures alone

Given the numerous significant correlations between [MBNL]_{inferred} and many of the clinical outcomes assessed

above, we sought to determine if clinical outcome measures could accurately predict [MBNL]_{inferred} via multiple linear regression modeling. Measures significantly correlated with [MBNL]_{inferred} in the complete DM1 cohort inclusive of both adults and children (Figs. 2–4) were utilized in the "All – DM1 and CDM" model. Multiple linear regression analysis using all available sampled outcome measures led to a model that included 45 participants and had an adjusted $r^2 = 0.6723$ whereby the developed model was able to account for the majority of the variance in [MBNL]_{inferred} (Fig. 5A,C).

Next, we sought to build an equation to accurately predict [MBNL]_{inferred} in only children with CDM utilizing significantly correlated clinical outcome measures when this participant group was evaluated independently (Figs. 2-4). Multiple linear regression analysis using all available sampled CDM outcome measures led to a model that included 13 CDM participants and had an adjusted $r^2 = 0.5875$ (Fig. 5B,D). Despite the reduced sample size, the model still managed to moderately predict [MBNL]_{inferred} in children with CDM. Overall, these analyses indicate that clinical outcome measures of myotonia, motor function, and strength correlate significantly with MBNL-dependent mis-splicing within all participants with DM1 independent of age of disease onset. Additionally, these measures can be used in patients with DM1 to accurately predict disease-associated patterns of mis-splicing in skeletal muscle.

12-month physical function can be predicted using baseline physical function measures and [MBNL]_{inferred}

Given the strength of the correlations between [MBNL]_{inferred} and several of the clinical outcomes assessed above, we aimed to determine if future clinical performance could be accurately predicted using baseline [MBNL]_{inferred} values and baseline clinical performance assessments in adults with DM1 and in children with CDM. We first looked at the outcome measures that had the strongest correlations with [MBNL]_{inferred} values within the complete cross-sectional cohort and in children with CDM independently – 10-meter walk/running speed and ADF strength.

Multiple linear regression analysis for prediction of 12-month ambulation speed using baseline walk/running speed and [MBNL]_{inferred} in our cohort of adults with DM1 had an adjusted $r^2 = 0.681$ (Fig. 6A, Model 1). Modeling in children with CDM alone suggested that these baseline values were similarly predictive of 12-month performance (adjusted $r^2 = 0.646$) (Fig. 6A, Model 2). Interestingly, while baseline [MBNL]_{inferred} did not contribute significantly to the predictive power of



Figure 3. MBNL-dependent mis-splicing as measured by [MBNL]_{inferred} correlates moderately with motor function measures in DM1 individuals. Correlation between [MBNL]_{inferred} and (A) 9-hole peg test (% predicted), (B) stair climbing speed (% predicted), (C) 6-min walk distance (% predicted), and (D) walk/running speed (% predicted) in all DM1 individuals and CDM individuals. Left panels represent correlations of all DM1 subjects and right panels represent CDM subjects only. The adult DM1 measures were excluded from the CDM statistical analyses but grayed out and included in the figure on the right for visual reference. Correlations are reported from a two-tailed Spearman test.

either model, it was trending toward significance in the model for adults, but not in the model for CDM (p = 0.1675 & p = 0.7125, respectively).

Multiple linear regression analysis for predicting 12-month ADF strength using baseline values and [MBNL]_{inferred} in our cohort of adults with DM1 had an adjusted $r^2 = 0.7126$ (Fig. 6B, Model 1). The predictive power of the same model for just children with CDM was reduced (adjusted $r^2 = -0.4904$) (Fig. 6B, Model 2). Again, when we evaluated the contribution of baseline [MBNL]_{inferred} to our 12-month ADF predictive model in children with CDM, it was much less significant (p = 0.9233) as compared to that for adults with DM1 (p = 0.0895).

Overall, of the seven motor function and strength outcome measures assessed, [MBNL]_{inferred} only contributed significantly to the predictive power of our 12-month multiple linear regression models for stair climbing speed (Fig. 7). In our complete cross-sectional cohort of both adults and children, baseline performance on this outcome assessment in combination with [MBNL]_{inferred} was able to accurately predict 12-month performance (adjusted $r^2 = 0.8057$) (Fig. 7A,B, Model 1). Multiple linear regression analysis for predicting 12-month stair climbing speed using baseline performance and [MBNL]_{inferred} in our cohort of CDM children alone had an adjusted $r^2 = 0.8629$ (Fig. 7B, Model 2), whereas the model for our cohort of adults with DM1 had an adjusted $r^2 = 0.6899$ (Fig. 7B, Model 3). The predictive utility of [MBNL]_{inferred} followed the same pattern observed above, whereby this measure of mis-splicing contributed more to our adult model (p = 0.0661) as compared to the model for children with CDM (p = 0.1049).

Discussion

In this study, we have identified associations between physical function and RNA mis-splicing in skeletal muscle as captured by [MBNL]_{inferred}, a composite metric of global alternative splicing dysregulation, in adults and children with DM1. This study describes significant correlations between mis-splicing and timepoint-matched assessments of clinical performance in children with CDM. Despite the availability of a small cohort of children and the wide age range assessed, the correlations observed across outcome assessments are surprisingly strong, underscoring theshared associations between alternative splicing dysregulation and phenotypic metrics in both adults with DM1 and children with CDM (Figs. 2–4).

Importantly, the regression model equations developed indicate that there are pathways forward in being able to predict levels of disease-associated spliceopathy in skeletal muscle within individuals with DM1 using clinical outcome measures alone (Fig. 5). Given the variable degree of mis-splicing in CDM, the ability to detect those children optimally suited for clinical trials by clinical outcome assessments alone may lessen the need for muscle biopsies in the future. The 12-month predictive multiple linear regression modeling analyses using baseline measures of alternative splicing dysregulation and baseline functional outcome performance points to the prognostic utility of measures such as [MBNL]_{inferred} (Figs. 6 and 7). [MBNL]_{inferred} consistently trended toward significance in the 12-month predictive models for adults with DM1; the small sample sizes $(n \le 15)$ likely contributed to the lack of statistical significance observed. Contrary to adults, it appears that [MBNL]_{inferred} is unlikely to contribute to 12-month predictive models in children with CDM due to the dynamic and variable pattern of splicing dysregulation occurring throughout development (Fig. 1). This is critical to keep in mind as we move forward in designing clinical trials for children with CDM, as RNA mis-splicing in this patient population is dynamic and might confound results depending on age at assessment and the natural trajectory of CDM disease progression (CDM_{infant} vs. CDM_{child} vs. CDM_{adolescent}).

Overall, there was a positive correlation between [MBNL]_{inferred} and motor/strength performance in all individuals with DM1 regardless of age of onset. Individuals with lower [MBNL]_{inferred} levels reflective of extensive dysregulated RNA splicing were not able to walk as far, run as fast, and took longer to climb stairs. In children with CDM, those with lower [MBNL]_{inferred} levels also exhibited reduced dexterity on the 9-hole peg test, although this pattern was not seen in adults with DM1. Given the severe phenotype in infancy, it is reasonable to



Figure 4. MBNL-dependent mis-splicing as measured by [MBNL]_{inferred} correlates with muscle strength independent of age. Correlation between [MBNL]_{inferred} and (A) knee extension (% predicted), (B) hand grip strength (% predicted), and (C) ankle dorsiflexion (% predicted) in all DM1 and CDM individuals. Left panels represent correlations of all DM1 subjects and right panels represent CDM subjects only. The adult DM1 measures were excluded from the CDM statistical analyses but grayed out and included in the figure on the right for visual reference. Correlations are reported from a two-tailed Spearman test.

expect that this severe motor impairment may allow for detection of change on the 9-hole peg test compared to the less severely affected adults with DM1. Most strikingly, individuals with higher [MBNL]_{inferred} levels performed more similarly to unaffected individuals as measured by quantitative muscle testing (knee extension, ankle dorsiflexors, grip strength). Importantly, these associations were not substantially impacted by choice of muscle biopsied or QMT testing method; [MBNL]_{inferred} was derived from vastus lateralis biopsies in children with CDM whereas distal tibialis anterior biopsies were utilized for adult-onset DM1 participants. Despite this difference



[MBNL]_{inferred} Linear Regression Intercept Table

Outcome n F (DFn, DFd) P value Adjusted	R ²
(C) Model 1 [MBNL] _{inferred} - All DM1 and CDM 45 F (4, 40) = 23.57 P<0.0001 0.6723	
Variable Estimate SE 95% Cl t	P value
β0 Intercept -0.1162 0.1085 (-0.3355 to 0.1031) 1.071	0.2908
β1 6-minute walk (% Predicted) 0.0003441 0.002155 (-0.0040 to 0.0047) 0.1597	0.8740
β2 Fast Walk/Running Speed 0.004033 0.0007990 (0.0024 to 0.0056) 5.048 (% Predicted)	<0.0001
β3 Knee Extension (% Predicted) 0.0008911 0.0004359 (1.01e-005 to 0.0018) 2.044	0.0475
β4 Ankle Dorsiflexion (% Predicted) 0.002795 0.001371 (2.35e-005 to 0.0056) 2.038	0.0482
Outcome n F (DFn, DFd) P value Adjusted	R ²
(D) Model 2 [MBNL] _{inferred} - CDM 13 F (3, 9) = 6.697 P=0.0114 0.5875	
Variable Estimate SE 95% CI t	P value
β0 Intercept 0.3704 0.2742 (-0.2499 to 0.9908) 1.351	0.2097
β1 Fast Walk/Running Speed 0.001678 0.001171 (-0.00097 to 0.0043) 1.433 (% Predicted)	0.1856
β2 9-Hole Peg Test (% Predicted) -0.000576 0.0007203 (-0.0022 to 0.0011) 0.7994	0.4446

Figure 5. Multiple linear regression modeling to predict [MBNL]_{inferred} in DM1 participants using functional outcomes. (A) Multiple linear regression plot showing observed [MBNL]_{inferred} versus predicted [MBNL]_{inferred} for all DM1 individuals. (B) Multiple linear regression plot showing observed [MBNL]_{inferred} versus predicted [MBNL]_{inferred} for CDM individuals. Intercept table of multiple linear regression models for (C) complete DM1 cohort and (D) CDM cohort alone using clinical outcome measures to predict [MBNL]_{inferred}. Multiple statistical elements are reported (|t|, *t*-test statistic; 95% CI, 95% confidence interval; DFd, degrees of freedom denominator; DFn, degrees of freedom numerator; *F*, F-distribution; SE, standard error). Extended intercept tables for the complete DM1 cohort model are provided in Table S3 and for the CDM cohort in Table S4.

in muscle groups used to measure RNA mis-splicing, children with CDM alone had a similar magnitude, if not stronger, association between [MBNL]_{inferred} and measures of distal strength (hand grip and ankle dorsiflexion) when compared to our analyses that included all DM1 participants. Altogether, the results suggest that these associations are more indicative of global alterations in muscle function and strength rather than these patterns being reflective of one specific muscle group. Perhaps the most surprising finding of these analyses was the observed lack of correlation between [MBNL]_{inferred} or *CLCN1/CACNA1S* mis-splicing levels with quantitative measures of myotonia in children with CDM. Previous work has shown that increased inclusion of *CLCN1* exon 7a leads to reduced density of CLCN1 within muscle fibers and myotonic discharges. Correction of this mis-splicing via targeted antisense oligonucleotides is sufficient to rescue myotonia in DM1 mouse models.⁴⁵

		Outcome	n	F (DFn, DFd)	P value	Adjusted R ²	_
(A)	Model 1	Fast Walk/Running Speed (% Predicted) - Adults	12	F (2, 9) = 12.73	P=0.0024	0.6809	
		Variable	Estimate	SE	95% CI	t	P value
-	β0 β1 β2	Intercept Baseline [MBNL] _{inferred} Baseline Fast Walk/Running Speed (% Predicted)	39.66 -31.03 0.8320	20.29 20.67 0.1660	(-6.250 to 85.56) (-77.78 to 15.73) (0.4566 to 1.207)	1.954 1.501 5.013	0.0824 0.1675 0.0007
		Outcome	n	F (DFn, DFd)	P value	Adjusted R ²	
	Model 2	Fast Walk/Running Speed (% Predicted) - Children	14	F (2, 11) = 12.84	P=0.0013	0.6456	
		Variable	Estimate	SE	95% CI	t	P value
·	β0 β1 β2	Intercept Baseline [MBNL] _{intered} Baseline Fast Walk/Running Speed (% Predicted)	-8.508 13.24 1.035	25.66 35 0.2357	(-64.98 to 47.97) (-63.80 to 90.27) (0.5160 to 1.553)	0.3316 0.3782 4.390	0.7464 0.7125 0.0011
		Outcome	n	F (DFn, DFd)	P value	Adjusted R ²	
(B)	Model 1	Ankle Dorsiflexon (% Predicted) - Adults	15	F (2, 12) = 18.44	P=0.0002	0.7136	_
		Variable	Estimate	SE	95% CI	t	P value
	β0 β1 β2	Intercept Baseline [MBNL] _{inferred} Baseline Ankle Dorsiflexion (% Predicted)	8.159 38.58 0.4888	12.25 20.89 0.1028	(-18.54 to 34.86) (-6.921 to 84.09) (0.2647 to 0.7128)	0.6658 1.847 4.753	0.5181 0.0895 0.0005
		Outcome	n	F (DFn, DFd)	P value	Adjusted R ²	
	Model 2	Ankle Dorsiflexion (% Predicted) - Children	5	F (2, 2) = 0.3419	P=0.7452	-0.4904	
		Variable	Estimate	SE	95% CI	t	P value
-	β0 β1 β2	Intercept Baseline [MBNL] _{interred} Baseline Ankle Dorsiflexion (% Predicted)	16.73 12.40 0.6690	94.52 113.9 0.8446	(-390.0 to 423.4) (-477.9 to 502.7) (-2.965 to 4.303)	0.1770 0.1089 0.7920	0.8758 0.9233 0.5114

12-month Physical Function Linear Regression Intercept Table

Figure 6. Multiple linear regression predictive models of 12-month ankle dorsiflexion strength and 10-meter walk/running speed in DM1 participants. Intercept table of multiple linear regression models for (A) 12-month walk/running speed (% predicted) and (B) 12-month ankle dorsiflexion strength (% predicted) using baseline [MBNL]_{inferred} values and baseline physical function values. Multiple statistical elements are reported (|t|, t-test statistic; 95% CI, 95% confidence interval; DFd, degrees of freedom denominator; DFn, degrees of freedom numerator; *F*, *F*-distribution; SE, standard error). Extended intercept tables for the 12-month walk/running speed models and 12-month ankle dorsiflexion strength are in Tables S5 and S6, respectively.

Mis-splicing of the CaV1.1 calcium channel encoded by *CACNA1S* has also been linked with exacerbated myotonia in these model systems.⁴⁶ Consistent with these reports, increased inclusion of *CLCN1* exon7a or exclusion of *CACNA1S* exon29 was associated moderately with longer vHOT_{thumb} times in all DM1 participants; no such associations of myotonia with causative RNA mis-splicing events have been previously reported in human DM1 studies. However, these associations did not hold true in children with CDM. All CDM participants, regardless of sub-cohort classification, [MBNL]_{inferred}, or *CLCN1/CAC-NA1S* PSI values, had low vHOT_{thumb} times (<3.2 s). In fact, CDM_{adolescent} individuals who had nearly 100% *CLCN1* exon inclusion, like the most severely affected adults with DM1, possessed minimal quantitative myotonia. Similar patterns were observed for *CACNA1S* mis-

Predicted 12-month Stair S Climbing Speed (% Predicted)	$100 - r^2$ 80 - 60 - 60 - 60 - 60 - 60 - 60 - 60 -	= 0.805	7	•	•
	20 1 20	40	60	80	100
	Cli	Actua imbing S	l 12-mor Speed (%	nth Stai ⁄₀ Predi	r cted)

All - DN	/11 aı	nd C	DM
----------	--------	------	----

(B)	Outcome	n	F (DFn, DFd)	P value	Adjusted R ²	_
Model 1	Stair Climbing Speed (% Predicted) - Adults/Children	28	F (2, 25) = 56.98	P<0.0001	0.8057	
	Variable	Estimate	SE	95% CI	t	P value
β0 β1 β2	Intercept Baseline [MBNL] _{inferred} Baseline Stair Climbing Speed (% Predicted)	-0.5239 23.11 0.7103	6.848 8.759 0.07049	(-14.63 to 13.58) (5.070 to 41.15) (0.5652 to 0.8555)	0.07651 2.638 10.08	0.9396 0.0141 <0.0001
	Outcome	n	F (DFn, DFd)	P value	Adjusted R ²	
Model 2	Stair Climbing Speed (% Predicted) - Children	14	F (2, 11) = 41.90	P<0.0001	0.8629	_
	Variable	Estimate	SE	95% CI	t	P value
β0 β1 β2	Intercept Baseline [MBNL] _{interred} Baseline Stair Climbing Speed (% Predicted)	-6.398 17.26 0.9154	7.724 9.768 0.1075	(-23.40 to 10.60) (-4.238 to 38.76) (0.6788 to 1.152)	0.8284 1.767 8.516	0.4251 0.1049 <0.0001
	Outcome	n	F (DFn, DFd)	P value	Adjusted R ²	
Model 3	Stair Climbing Speed (% Predicted) - Adults	14	F (2, 11) = 15.46	P=0.0006	0.6899	_
	Variable	Estimate	SE	95% CI	t	P value
β0 β1 β2	Intercept Baseline [MBNL] _{interred} Baseline Stair Climbing Speed (% Predicted)	5.637 30.98 0.5539	11.35 15.19 0.1207	(-19.35 to 30.62) (-2.443 to 64.41) (0.2883 to 0.8195)	0.4966 2.040 4.590	0.6293 0.0661 0.0008

Figure 7. Multiple linear regression predictive modeling of 12-month stair climbing speed in DM1 participants. (A) Multiple linear regression plot showing observed 12-month stair climbing speed (% predicted) versus predicted 12-month stair climbing speed (% predicted) for all DM1 individuals (adults and children). (B) Intercept table of multiple linear regression models for complete DM1 cohort (Model 1), CDM cohort alone (Model 2), and adult cohort alone (Model 3). Multiple statistical elements are reported (*Jt*), *t*-test statistic; 95% CI, 95% confidence interval; DFd, degrees of freedom denominator; DFn, degrees of freedom numerator; *F*, *F*-distribution; SE, standard error). Extended intercept tables for all three models are provided in Table S7.

splicing. Altogether, these data suggest that while patterns of DM1 spliceopathy previously linked to myotonia are preserved in CDM skeletal muscle, these pediatric patients are not experiencing clinical myotonia in the same manner as adults with DM1. Future work will be required to evaluate the molecular mechanisms behind this lack of observable myotonia in the CDM population. In total, these data suggest that myotonia measures appear to be a poor outcome assessment for children with CDM, especially as correction of myotonia and the associated RNA splicing patterns have been proposed as early markers of therapeutic efficacy in DM1 clinical trials.

Previous work by our group has shown that children with CDM display a triphasic pattern of disease progression that is mirrored by changes in RNA mis-splicing.^{13,28} While the CDM participants evaluated here were sampled cross-sectionally, the results captured in this study begin to verify that the triphasic pattern of alternative splicing dysregulation in CDM matches the observed changes in physical function. However, we were unable to fully validate this proposed pattern given the limited sample size and data availability within select CDM sub-cohorts, notably CDM infants (<2 years). Few clinical measures were able to be assessed for these individuals due to the developmental constraints of this age group. Future directions include improving our clinical outcome measures to include infant-specific strength/motor testing. Another challenge in obtaining larger sample sizes lies in the difficulty in obtaining reliable outcomes from patients with physical, cognitive, and behavioral impairments. Due to these challenges, it is unclear whether clinical outcome measures reflect true maximum physical ability, or rather a child's ability to understand and follow instructions. This could be especially relevant in measures such as the 9-hole peg test. Future studies with expanded cohort sizes will need to be performed to assess the impacts of cognitive impairment as a potential confounding variable in physical outcome measure performance and identification of representative biomarkers that correlate with measures of cognitive function.

The development of disease-modifying therapies for adults with DM1 has rapidly accelerated in recent years with many in early-phase clinical trials. Given the shared genetic basis of disease, there is significant interest in extending these trials into the CDM population. However, a major limitation of these efforts has been insufficient understanding of the natural history of disease progression in children with CDM and how the molecular pathogenesis, particularly alternative splicing dysregulation, correlates with clinical outcome measures. This analysis connects measures of splicing dysregulation to a range of clinical outcome measures in a cross-sectional cohort including both adult DM1 and CDM participants.

Furthermore, the analyses here provide the foundation for determination of high-performing clinical outcome assessments (COAs) for use in CDM clinical trials in older pediatric patients (>6 years). This work, in combination with our characterization of CDM spliceopathy across pediatric development,²⁸ is vital in identifying the effective timing of therapeutic administration to children with CDM for clinical trial success whereby therapeutic benefit outpaces the natural dynamic changes in RNA missplicing, especially given the rapid improvement observed in the first years of life (Fig. 1). The regression models developed in this study may assist in the refinement of clinical trial design and offer a noninvasive methodology for prediction of spliceopathy using COAs alone when screening for trial inclusion. They may also reduce reliance on muscle biopsies to assess target engagement of therapeutic agents. The strength of these predictive equations can be improved through replication and validation in a secondary DM1 cohort. Additionally, these predicative equations are based on inferences of free [MBNL] in skeletal muscle and may not be reflective of disease severity and clinical performance for non-musculoskeletal metrics. Overall, these results provide the framework for both the utility of a muscle-based biomarker and clinical trial design in children with CDM.

Acknowledgments

Sources of Support: This study is supported by NINDS (R01NS104010), FDA (7R01FD006071), Muscular Dystrophy Association, Myotonic Dystrophy Foundation, Novartis Pharmaceuticals, Dyne Therapeutics, Avidity Biosciences, PepGen, and Takeda Pharmaceuticals.

Author Contributions

Conception and design of study: JMH, MAH, and NEJ. Acquisition and analysis of data: JMH, MP, KB, KI, AB, ASJ, KB, JD, MJM, JNB, KMC, JB, MK, NEJ, and MAH. Drafting a significant portion of manuscript: JMH, MK, NEJ, and MAH. Please also see the supplementary file labeled "DMCRN Consortium Members."

Conflict of Interest

Julia M. Hartman, Marina Provenzano, Kameron Bates, Kobe Ikegami, Amanda Butler, Aileen S. Jones, Kiera N. Berggren, Marnee J. McKay, Jennifer N. Baldwin, and Kayla M.D. Cornett – None. Jeanne Dekdebrun – Consultation for Avidity Biosciences, Dyne Therapeutics, Vertex, Lupin, Arthex, PepGen and Trins. Joshua Burns – Research Support from the University of Sydney, Sydney Children's Hospitals Network, Australian Government

(NHMRC#2015970, MRFF#1152226), United States Government (NIH NINDS#1U01NS109403, NIH NCATS/ NINDS# U54NS065712), Muscular Dystrophy Association, American Orthotic and Prosthetic Association, Charcot Marie Tooth Association and Charcot Marie Tooth Australia. Scientific Advisory Board fees from Faculty of Medicine Siriraj Hospital Mahidol University Thailand; Department of Rehabilitation Sciences, The Hong Kong Polytechnic University; Hereditary Neuropathy Foundation. Consulted for DTx Pharma, Applied Therapeutics, Pharnext. Michael Kiefer - Has provided consultation for Aspa therapeutics. Nicholas E. Johnson grant funding from NINDS He has received (R01NS104010, U01NS124974), NCATS (R21TR003184), CDC (U01DD001242) and the FDA (7R01FD006071). He receives royalties from the CCMDHI and the CMTHI. He receives research funds from Novartis, Takeda, PepGen, Sanofi Genzyme, Dyne, Vertex Pharmaceuticals, Fulcrum Therapeutics, AskBio, ML Bio, and Sarepta. He has provided consultation for Arthex, Angle Therapeutics, Juvena, Rgenta, PepGen, AMO Pharma, Takeda, Design, Dyne, AskBio, Avidity, and Vertex Pharmaceuticals. Melissa A. Hale - She has provided consultation for Juvena and Arrakis Therapeutics.

Data Availability Statement

RNA sequencing data used to define [MBNL]_{inferred} are available in Sequence Read Archive at https://www.ncbi. nlm.nih.gov/sra in the following BioProjects: [PRJNA1079722], [PRJNA830511], and [PRJNA1151618]. Associated BioProject and SRA accession numbers for each sample are listed in Table S1.

References

- Johnson NE, Butterfield RJ, Mayne K, et al. Populationbased prevalence of myotonic dystrophy type 1 using genetic analysis of statewide blood screening program. Neurology. 2021;96(7):e1045-e1053.
- Mahadevan M, Tsilfidis C, Sabourin L, et al. Myotonic dystrophy mutation: an unstable CTG repeat in the 3' untranslated region of the gene. Science. 1992;255 (5049):1253-1255.
- 3. Fu YH, Pizzuti A, Fenwick RG, et al. An unstable triplet repeat in a gene related to myotonic muscular dystrophy. Science. 1992;255(5049):1256-1258.
- 4. Brook JD, McCurrach ME, Harley HG, et al. Molecular basis of myotonic dystrophy: expansion of a trinucleotide (CTG) repeat at the 3' end of a transcript encoding a protein kinase family member. Cell. 1992;68(4):799-808.
- Hartman J, Patki T, Johnson NE. Diagnosis and Management of Myotonic Dystrophy Type 1. JAMA. 2024;331(14):1227-1228. doi:10.1001/jama.2024.2511

- 6. Harper PS. In: Saunders WB, ed. Myotonic Dystrophy. 3rd ed.; 2001. W. B. Saunders.
- Tsilfidis C, MacKenzie AE, Mettler G, et al. Correlation between CTG trinucleotide repeat length and frequency of severe congenital myotonic dystrophy. Nat Genet. 1992;1(3):192-195. https://www.nature.com/articles/ ng0692-192
- Harley HG, Rundle SA, MacMillan JC, et al. Size of the unstable CTG repeat sequence in relation to phenotype and parental transmission in myotonic dystrophy. Am J Hum Genet. 1993;52(6):1164-1174. https://www.ncbi.nlm. nih.gov/pmc/articles/PMC1682262/
- Barbé L, Lanni S, López-Castel A, et al. CpG methylation, a parent-of-origin effect for maternal-biased transmission of congenital myotonic dystrophy. Am J Hum Genet. 2017;100(3):488-505.
- Pucillo EM, DiBella DL, Hung M, et al. Physical function and mobility in children with congenital myotonic dystrophy. Muscle Nerve. 2017;56(2):224-229. https:// www.ncbi.nlm.nih.gov/pmc/articles/PMC5436951/
- Johnson NE, Butterfield R, Berggren K, et al. Disease burden and functional outcomes in congenital myotonic dystrophy: a cross-sectional study. Neurology. 2016;87 (2):160-167.
- 12. Johnson NE, Ekstrom A-B, Campbell C, et al. Parentreported multi-national study of the impact of congenital and childhood onset myotonic dystrophy. Dev Med Child Neurol. 2016;58(7):698-705.
- Quigg KH, Berggren KN, McIntyre M, et al. 12-month progression of motor and functional outcomes in congenital myotonic dystrophy. Muscle Nerve. 2021;63 (3):384-391. https://www.ncbi.nlm.nih.gov/pmc/articles/ PMC8038871/
- Miller JW, Urbinati CR, Teng-Umnuay P, et al. Recruitment of human muscleblind proteins to (CUG)(n) expansions associated with myotonic dystrophy. EMBO J. 2000;19(17):4439-4448.
- Lee K-Y, Li M, Manchanda M, et al. Compound loss of muscleblind-like function in myotonic dystrophy. EMBO Mol Med. 2013;5(12):1887-1900.
- Lin X, Miller JW, Mankodi A, et al. Failure of MBNL1dependent post-natal splicing transitions in myotonic dystrophy. Hum Mol Genet. 2006;15(13):2087-2097.
- 17. Wang ET, Cody NAL, Jog S, et al. Transcriptome-wide regulation of pre-mRNA splicing and mRNA localization by muscleblind proteins. Cell. 2012;150(4):710-724.
- López-Martínez A, Soblechero-Martín P, de-la-Puente-Ovejero L, et al. An overview of alternative splicing defects implicated in myotonic dystrophy type I. Genes (Basel). 2020;11(9):1109.
- 19. Degener MJF, van Cruchten RTP, Otero BA, et al. A comprehensive atlas of fetal splicing patterns in the brain of adult myotonic dystrophy type 1 patients. NAR Genom Bioinform. 2022;4(1):lqac016.

- Konieczny P, Stepniak-Konieczna E, Sobczak K. MBNL proteins and their target RNAs, interaction and splicing regulation. Nucleic Acids Res. 2014;42(17):10873-10887.
- 21. Charlet-B N, Savkur RS, Singh G, et al. Loss of the muscle-specific chloride channel in type 1 myotonic dystrophy due to misregulated alternative splicing. Mol Cell. 2002;10(1):45-53.
- 22. Mankodi A, Takahashi MP, Jiang H, et al. Expanded CUG repeats trigger aberrant splicing of ClC-1 chloride channel pre-mRNA and hyperexcitability of skeletal muscle in myotonic dystrophy. Mol Cell. 2002;10(1):35-44.
- 23. Freyermuth F, Rau F, Kokunai Y, et al. Splicing misregulation of SCN5A contributes to cardiac-conduction delay and heart arrhythmia in myotonic dystrophy. Nat Commun. 2016;7:11067. https://www.ncbi.nlm.nih.gov/ pmc/articles/PMC4831019/
- 24. Fugier C, Klein AF, Hammer C, et al. Misregulated alternative splicing of BIN1 is associated with T tubule alterations and muscle weakness in myotonic dystrophy. Nat Med. 2011;17(6):720-725.
- 25. Nakamori M, Sobczak K, Puwanant A, et al. Splicing biomarkers of disease severity in myotonic dystrophy. Ann Neurol. 2013;74(6):862-872.
- 26. Wang ET, Treacy D, Eichinger K, et al. Transcriptome alterations in myotonic dystrophy skeletal muscle and heart. Hum Mol Genet. 2019;28(8):1312-1321. https:// www.ncbi.nlm.nih.gov/pmc/articles/PMC6452195/
- Wagner SD, Struck AJ, Gupta R, et al. Dose-dependent regulation of alternative splicing by MBNL proteins reveals biomarkers for myotonic dystrophy. PLoS Genet. 2016;12 (9):e1006316. doi:10.1371/journal.pgen.1006316
- Hale MA, Bates K, Provenzano M, Johnson NE. Dynamics and variability of transcriptomic dysregulation in congenital myotonic dystrophy during pediatric development. Hum Mol Genet. 2022;32(9):1413-1428. https://www.ncbi.nlm.nih.gov/pmc/articles/PMC10117163/
- Heatwole C, Bode R, Johnson N, et al. The myotonic dystrophy health index: initial evaluation of a new outcome measure. Muscle Nerve. 2014;49(6):906-914. https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5551891/
- Puwanant A, Statland J, Dilek N, et al. Video hand opening time (vHOT) in myotonic dystrophy type 1 (DM1) (P05.188). Neurology. 2012;78(1 Supplement): P05.188. http://n.neurology.org/content/78/1_Supplement/ P05.188.abstract
- Heatwole C, Luebbe E, Rosero S, et al. Mexiletine in myotonic dystrophy type 1. Neurology. 2021;96(2): e228-e240. https://www.ncbi.nlm.nih.gov/pmc/articles/ PMC7905778/
- 32. Feys P, Lamers I, Francis G, et al. The nine-hole peg test as a manual dexterity performance measure for multiple sclerosis. Mult Scler. 2017;23(5):711-720. https://www.ncbi. nlm.nih.gov/pmc/articles/PMC5405844/

- Bushby K, Connor E. Clinical outcome measures for trials in Duchenne muscular dystrophy: report from international working group meetings. Clin Investig (Lond). 2011;1(9):1217-1235.
- 34. McDonald CM, Henricson EK, Abresch RT, et al. The 6-minute walk test and other endpoints in Duchenne muscular dystrophy: longitudinal natural history observations over 48 weeks from a multicenter study. Muscle Nerve. 2013;48(3):343-356.
- 35. McDonald CM, Henricson EK, Abresch RT, et al. The 6-minute walk test and other clinical endpoints in duchenne muscular dystrophy: reliability, concurrent validity, and minimal clinically important differences from a multicenter study. Muscle Nerve. 2013;48(3):357-368.
- 36. Shumway-Cook A, Taylor CS, Matsuda PN, et al. Expanding the scoring system for the dynamic gait index. Phys Ther. 2013;93(11):1493-1506. doi:10.2522/ptj. 20130035
- Tawil R, Forrester J, Griggs RC, et al. Evidence for anticipation and association of deletion size with severity in facioscapulohumerd muscular dystrophy. Ann Neurol. 1996;39(6):744-748. doi:10.1002/ana.410390610
- Thornton CA, Moxley RT, Eichinger K, et al. Antisense oligonucleotide targeting DMPK in patients with myotonic dystrophy type 1: a multicentre, randomised, doseescalation, placebo-controlled, phase 1/2a trial. Lancet Neurol. 2023;22(3):218-228.
- Pereira AC, Ribeiro MG, Araújo AP, de QC. Timed motor function tests capacity in healthy children. Arch Dis Child. 2016;101(2):147-151.
- Bohannon RW. Comfortable and maximum walking speed of adults aged 20—79 years: reference values and determinants. Age Ageing. 1997;26(1):15-19. doi:10.1093/ ageing/26.1.15
- 41. McKay MJ, Baldwin JN, Ferreira P, et al. Normative reference values for strength and flexibility of 1,000 children and adults. Neurology. 2017;88(1):36-43. https:// www.ncbi.nlm.nih.gov/pmc/articles/PMC5200854/
- 42. McKay MJ, Baldwin JN, Ferreira P, et al. 1000 norms project: protocol of a cross-sectional study cataloging human variation. Physiotherapy. 2016;102(1):50-56.
- 43. Harris PA, Taylor R, Thielke R, Payne J, Gonzalez N, Conde JG. Research electronic data capture (REDCap)—a metadata-driven methodology and workflow process for providing translational research informatics support. J Biomed Inform. 2009;42(2):377-381.
- Harris PA, Taylor R, Minor BL, et al. The REDCap consortium: building an international community of software platform partners. J Biomed Inform. 2019;95:103208.
- 45. Wheeler TM, Lueck JD, Swanson MS, Dirksen RT, Thornton CA. Correction of ClC-1 splicing eliminates chloride channelopathy and myotonia in mouse models of

myotonic dystrophy. J Clin Invest. 2007;117 (12):3952-3957.

- Cisco LA, Sipple MT, Edwards KM, et al. Verapamil mitigates chloride and calcium bi-channelopathy in a myotonic dystrophy mouse model. J Clin Invest. 2024; 134 (1):e173576.202=. https://www.jci.org/articles/view/173576
- Tang ZZ, Yarotskyy V, Wei L, et al. Muscle weakness in myotonic dystrophy associated with misregulated splicing and altered gating of CaV1.1 calcium channel. Hum Mol Genet. 2012;21(6):1312-1324. doi:10.1093/hmg/ddr568
- 48. Echenne B, Bassez G. Congenital and infantile myotonic dystrophy. Handb Clin Neurol. 2013;113:1387-1393.
- 49. Kroksmark A-K, Stridh M-L, Ekström A-B. Long-term follow-up of motor function and muscle strength in the

congenital and childhood forms of myotonic dystrophy type 1. Neuromuscul Disord. 2017;27(9):826-835. https:// www.sciencedirect.com/science/article/pii/S0960896616 311968

Supporting Information

Additional supporting information may be found online in the Supporting Information section at the end of the article.

Data S1. Appendix S1.