

Epigallocatechin-3-gallate therapeutic potential in human diseases: molecular mechanisms and clinical studies

Manzar Alam^{1†}, Mehak Gulzar^{1†}, Mohammad Salman Akhtar², Summya Rashid³, Zulfareen¹, Tanuja¹, Anas Shamsi^{4*} and Md. Imtaiyaz Hassan^{1*}

Abstract

Green tea has garnered increasing attention across age groups due to its numerous health benefts, largely attributed to Epigallocatechin 3-gallate (EGCG), its key polyphenol. EGCG exhibits a wide spectrum of biological activities, including antioxidant, anti-infammatory, antibacterial, anticancer, and neuroprotective properties, as well as benefts for cardiovascular and oral health. This review provides a comprehensive overview of recent fndings on the therapeutic potential of EGCG in various human diseases. Neuroprotective effects of EGCG include safeguarding neurons from damage and enhancing cognitive function, primarily through its antioxidant capacity to reduce reactive oxygen species (ROS) generated during physiological stress. Additionally, EGCG modulates key signaling pathways such as JAK/STAT, Delta-Notch, and TNF, all of which play critical roles in neuronal survival, growth, and function. Furthermore, EGCG is involved in regulating apoptosis and cell cycle progression, making it a promising candidate for the treatment of metabolic diseases, including cancer and diabetes. Despite its promising therapeutic potential, further clinical trials are essential to validate the efficacy and safety of EGCG and to optimize its delivery to target tissues. While many reviews have addressed the anticancer properties of EGCG, this review focuses on the molecular mechanisms and signaling pathways by which EGCG used in specifc human diseases, particularly cancer, neurodegenerative and metabolic diseases. It serves as a valuable resource for researchers, clinicians, and healthcare professionals, revealing the potential of EGCG in managing neurodegenerative disorders, cancer, and metabolic diseases and highlighting its broader therapeutic values.

Keywords Epigallocatechin gallate, Neurological disorders, Cancer therapy, Neuroinfammation, Targeted therapy, Natural products

† Manzar Alam and Mehak Gulzar contributed equally to this work.

*Correspondence: Anas Shamsi anas.shamsi18@gmail.com Md. Imtaiyaz Hassan mihassan@jmi.ac.in ¹ Centre for Interdisciplinary Research in Basic Sciences, Jamia Millia Islamia, Jamia Nagar, New Delhi 110025, India ² Department of Basic Medical Sciences, Faculty of Applied Medical Sciences, Albaha University, Albaha, Saudi Arabia ³ Department of Pharmacology & Toxicology, College of Pharmacy, Prince Sattam Bin Abdulaziz University, PO Box 173, 11942 Al-Kharj, Saudi Arabia

4 Center of Medical and Bio-Allied Health Sciences Research (CMBHSR), Ajman University, P.O. Box 346, Ajman, UAE

Introduction

Polyphenols are a group of bioactive compounds found in foods and beverages, mainly present in green tea. They act as an antioxidant, anti-inflammatory, and neuroprotective agent, which belongs to catechins [\[1](#page-14-0)]. Among many other compounds, the potential and major biochemical compound is Epigallocatechin-3-gallate (EGCG), the most abundant favone-3-ol polyphenol present in green tea. Structurally, EGCG contains eight free hydroxyl groups responsible for its bioactivities properties [[2\]](#page-14-1). Nowadays, the consumption of green tea among the youth has increased in folds, providing

© The Author(s) 2024. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit [http://creativecommons.org/licenses/by/4.0/.](http://creativecommons.org/licenses/by/4.0/)

numerous benefts to human health [[3](#page-14-2)[–5](#page-14-3)]. Polyphenols represent therapeutic agents against various symptoms. EGCG has inhibitory functions in multiple diseases that arise due to abnormal changes or modifcations with an efect as anti-infammatory properties and antibacterial efects that appear within its efective range. In green tea, EGCG accounts for about 59% of the total catechins and exhibits a broad spectrum of biological activities, such as oral disease linked with microbes. EGCG plays crucial roles against pathogenic microorganisms, such as several Gram-positive and Gram-negative bacteria, and a wide range of anti-infective drugs [[6,](#page-14-4) [7](#page-14-5)]. Studies have been conducted that proved its role in anti-oxidant, anticancer, anti-diabetic, anti-infammatory, anti-microbial, anti-oxidant, anti-diabetic, neuroprotective activities, anti-cancer, anti-infammatory, anti-microbial, etc. [\[8](#page-14-6)]. Thus, therapeutic advantages from EGCG utilization were observed in oral infections, cancer, obesity, diabetes, neurodegenerative, and infammatory diseases (Fig. [1\)](#page-1-0).

 EGCG ameliorates oral health by blocking the deterioration of periodontitis dentin erosion and defending oral mucosa against oral tumor cells. Dose-dependent mechanisms for fora-related diseases such as caries and periodontal EGCG have been found to be highly efective. High doses of EGCG act as bactericidal, i.e., kill bacteria by destroying their structures, whereas low doses exert antibacterial efects by reducing virulence,

Fig. 1 Mechanistic Overview of Modulation of Cellular Signaling Pathways by EGCG. This fgure illustrates the multifaceted mechanism of EGCG and its interactions with key signaling and molecular pathways. EGCG is shown to regulate various target genes and proteins, including JAK/STAT, NF-κB, AKT, and Notch pathways, highlighting its critical roles in cellular processes such as apoptosis, proliferation, and survival. The diagram emphasizes the biological functions of EGCG, including its anti-cancer, anti-infammatory, and antioxidant activities. By inhibiting oxidative stress, reducing infammation, and promoting apoptosis in cancer cells, EGCG showcases its therapeutic potential in combating diseases like cancer, cardiovascular , and neurodegenerative diseases

which often leads to bacterial membrane disruption and growth inhibition [[9\]](#page-15-0).

EGCG is a promising therapeutic molecule to prevent neurodegenerative diseases (NDs). It has efective antioxidant and anti-infammatory efects and subsequent therapeutic potential [[10,](#page-15-1) [11\]](#page-15-2). EGCG can modulate several targets associated with the pathogenesis of several chronic diseases $[12-14]$ $[12-14]$. EGCG is promising for promoting healthy aging by recovering the morphologic and functional modifcations that happen in a naturally aging brain, suppressing cognitive dysfunction and diminishing oxidative injury in the brain. EGCG possesses neuroprotective efects against several neural injuries [\[15](#page-15-5)].

The neuroprotective effects of EGCG were reported in Alzheimer's (AD) and Parkinson's disease (PD) [\[10](#page-15-1), [13\]](#page-15-6). EGCG has the potential to attenuate cell death, repress ROS accumulation and free intracellular calcium, modify signalling cascades, and consequently decrease oxidative stress. However, in the case of AD, ROS accumulation leads to β-amyloid degradation and repression in the phosphorylation of tau protein $[16]$ $[16]$ $[16]$. EGCG, with its amyloid precursor protein (APP) processing ability, offers an alternative strategy for AD prevention. Studies have been performed on diferent cell lines, including MC65, EOC 13.31, SweAPP N2a, and N2a/APP695, proving its anti-neuroinfammatory capacity by inhibiting microglia-induced cytotoxicity $[17]$ $[17]$ $[17]$.

EGCG is a well-explored phytochemical with numerous health benefts that can improve the human lifestyle [[18](#page-15-9)–[21](#page-15-10)]. Its clinical application remains limited due to its poor physicochemical stability and low oral bioavailability. While EGCG exhibits favourable and strong advantages against biochemical and molecularrelated diseases, its efectiveness in clinical settings does not fully match that of frst-line chemotherapy agents. There is still a certain gap that prevents the real potential of EGCG.

This review focuses on biochemical metabolism, and physiological role in humans. Here, we aimed to compile and present the latest research on biological activities, including the antioxidant, anti-infammatory, anti-microbial, and neuroprotective properties of EGCG. We further explored the underlying mechanism by which EGCG modulates various cellular signalling pathways, such as those involved in infammation, apoptosis, and epigenetic modifcations, to exert its therapeutic efects. In addition, we summarize the recent fndings on the molecular mechanisms and therapeutic efects of EGCG in oral-related and neurological diseases. We discuss the potential of EGCG as a promising therapeutic drug for treating these conditions.

Biological and pharmacological efects of EGCG

The biological action of EGCG lies in its phenolic hydroxyl groups with molecular structures that are implicated in food production. EGCG has a protecive efect on inflammation due to its antioxidant properties [\[22](#page-15-11), [23](#page-15-12)]. It prevents cellular oxidation and inhibits cell-free radical damage [\[24](#page-15-13)–[26\]](#page-15-14). EGCG attenuates oxidative breakdown under elevated temperatures and alkali states and accelerates the degradation rate with the increase in temperature and oxidation concentration $[27]$ $[27]$. The effects of EGCG could be highly synergistic when combined with other catechins and thus metabolically stimulated to form stronger and more efficient bioactive compounds [[28,](#page-15-16) [29](#page-15-17)].

A detailed analysis of the biological efects of polyphenolic compounds present in green tea has shown the diference in pharmacokinetics activity in the individual compounds. It was experimentally proven that intake of 1.5 mM of green tea elevates the level of both ((-)-epicatechin-3-gallate) EGC and EGCG, but the variation is observed in the duration of half-life. For instance, EGC plasma level rises quickly with a low elimination halftime of 1.7 h, whereas EGCG levels rise slowly with a higher half-time of 3.9 h [\[30,](#page-15-18) [31](#page-15-19)].

EGCG is absorbed in the intestine when ingested orally, but its bioavailability has always been a question. Due to its oxidation, efflux, and metabolism, there is a low availability in the gut area. Microbiota present in the gut region deconjugate and degrade EGCG. Studies have even found that EGCG undergoes metabolism before absorption. With the help of gut microbes such as *Enterobacter aerogenes*, *Klebsiella pneumoniae* subspecies EGCG breaks down to EGC and gallic acid [\[32](#page-15-20)]. Later, EGC degrades to 5-(3,5-dihydroxyphenyl)−4-hydroxyvaleric acid, which is the main metabolite of EGCG. The absorbed compound is 5-(3',5'-dihydroxyphenyl)-γ-valerolactone and its glucuronide form is the major urinary metabolite.

The consumption of EGCG has shown various physiological and pharmacological health benefts [[33](#page-15-21)]. EGCG is one of the oldest polyphenols that have been experimented with to test its efficiency on bacteria. EGCG has a potential efect on the growth of *Staphylococcus aureus*, especially methicillin-resistant *Streptococcus* and *E. coli* [[34\]](#page-15-22). EGCG has shown anti bacterial - activity against a heterogeneity of Gram-positive and Gram-negative pathogens, viruses, fungi, and prions. Hence, it is known to be a prospective anti-infective agent. It has antifungal action against human-pathogenic yeast [[35,](#page-15-23) [36](#page-15-24)].

 EGCG inhibits enoyl-acyl carrier protein reductase (ENR) in *Plasmodium falciparum*. The mechanism of inhibition is believed to involve a slow-tight binding mechanism. This means that EGCG binds to the enzyme slowly but forms a very stable complex,

efectively inhibiting its activity. Inhibiting ENR would disrupt the fatty acid biosynthesis pathway, which is essential for the survival of parasite. This could lead to reduced parasite viability and potentially prevent or treat malaria infections. Targeting ENR represents a novel approach to combating malaria, which is becoming increasingly resistant to traditional antimalarial drugs [[37\]](#page-15-25). Studies have revealed the mechanism by which EGCG inhibits the occurrence of bacterial outbursts in the body. EGCG interacts with σ, thus modulating the activity of RybB and CsgD genes. This interaction downstream disrupts the bioflm found by the bacteria. This is achieved by alternating curli subunit and c-di-GMP required for membrane formation [[38](#page-15-26)], as described in Fig. [2.](#page-3-0)

 EGCG induces changes in the proteome of *Microcystis aeruginosa* under stress conditions. It inhibits carbon and nitrogen assimilation along with chlorophyll synthesis and cell division. In contrast, stress response proteins such as superoxide dismutase and glutaredoxin were found to be upregulated. The observed proteomic changes provide insight into the molecular mechanism of the inhibitory activity of EGCG on *M. aeruginosa* [\[39](#page-15-27)].

Experimental evidence has proven the relationship between EGCG and exercise and their effect on the physiology of AD. EGCG and exercise, either separately or in combination, are applied to a 2-month mouse, with the impact of a 4-month wheel-running exercise attenuating or lowering the formation of soluble Aβ1–42 levels in the cortex and hippocampus [\[40](#page-15-28)]. Also, EGCG enhanced the efects when formulated as dual-drug loaded PEGylated PLGA nanoparticles [\[41](#page-15-29), [42\]](#page-15-30). However, oral administration of EGCG/AA NPs in mice afected EGCG gathering in all main organs, such as the brain [[43](#page-15-31)]. EGCG might prevent specifc biomedical vital molecular targets, including antiapoptotic Bcl-2 proteins and vascular endothelial growth factor (VEGFR) signaling [[28](#page-15-16), [44](#page-15-32)[–47](#page-16-0)]. Pharmacokinetic examination accomplished in human exhibits shows that the physiologically pertinent serum concentrations of EGCG could be in the elevated nanomolar range.

EGCG protects against Aβ-mediated cytotoxicity via the Akt pathway activation $[16]$ $[16]$. The neuroprotective results against Aβ-mediated neuronal cell death have been generated via its capability to scavenge ROS efficiently. The oral administration of EGCG has identified considerable development in spatial cognitive learning capability. EGCG can reduce Aβ and tau toxicity, explaining its promise for the obstacle of AD. EGCG has shown considerable inhibitory results against oxidative stressmediated cell death. Hence, EGCG is a therapeutic drug in the management of PD [\[48](#page-16-1), [49\]](#page-16-2).

Fig. 2 Antimicrobial Action of EGCG through Bioflm Disruption. Showing the antimicrobial mechanism of EGCG, focusing on its ability to disrupt biofilm formation. EGCG targets the σε regulatory pathway, which plays a crucial role in bacterial membrane adhesion. By interfering with the production of curli subunits and cyclic di-GMP (c-di-GMP), both essential for bioflm stability and bacterial adherence, EGCG effectively impairs biofilm formation. This disruption weakens bacterial colonization and enhances susceptibility to antimicrobial treatments, highlighting the potential of EGCG as a powerful agent in combating bacterial infections and bioflm-associated resistance

The anti-oxidation process by EGCG is of great importance to human health care [\[50](#page-16-3)]. EGCG mitigates the harmful efects of locally administered bupivacaine, a commonly used for epidural anaesthesia and nerve blocks, by suppressing bupivacaine-induced ROS production in neuroblastoma cells. Briefy, EGCG attenuates bupivacaine-induced ROS generation in neuroblastoma cells, protecting against its toxicity [\[51](#page-16-4)]. Structurally, the presence of phenol rings in the EGCG acts as a hunter of free radicals and traps the electrons that inhibit the formation of ROS. This causes a decrease in the damage caused by oxidative stress [\[52](#page-16-5)–[56\]](#page-16-6). EGCG is an essential compound that received much recognition for its numerous health benefts because it reduces infammation and helps to prevent heart and brain disease, assisting in weight loss [\[52](#page-16-5)].

EGCG prevents stimulated oxidative stress and neurotoxicity, and EC diminishes Aβ-fbril creation. EGCG can control the proteolytic processing of APP, suggesting that green tea polyphenols may be potential therapeutic agents for PD and AD [[16](#page-15-7)]. A marked reduction in β- and γ-secretase function and BACE1 and APP expression was observed [[57\]](#page-16-7). Despite the encouraging efects of preclinical examines, a translational gap exists between fundamental invention and clinical application [\[58\]](#page-16-8). Multiple studies in the brains of AD patients revealed a decline of PKC ϵ action in the membrane fraction.

 EGCG acts as a potential inhibitor for the dual-specifcity tyrosine phosphorylation-regulated kinase 1A (DYRK1A). DYRK1A overexpression induces morphological defects and impairment in the individuals, leading to Down syndrome, Pick's disease, and AD. EGCG displays appreciable inhibition of DYRK1A even at high concentrations (360 mg/kg). Since the stability is low, structurally modifed EGCG shows non-competitive properties. Difering from other published inhibitors, EGCG acts as a non-competitive inhibitor [\[59\]](#page-16-9). It revealed that prenatal exposure prevents the over-expression of DYRK1A in the brain. EGCG acts as a competitive inhibitor at the ATP-binding site when DYRK1A induces K465R mutation. Interestingly, the K465R mutation in DYRK1A changes the mode of inhibition of EGCG from a non-competitive to a competitive inhibitor at the ATP site. Although EGCG showed promise as a therapeutic agent (Table [1\)](#page-4-0), its clinical application was limited by its low absorption rate and susceptibility to degradation.

Molecular mechanisms of EGCG activity

This section provides insights into the intricate interactions of EGCG with key cellular targets, shedding light on its multifaceted therapeutic potential. Antioxidant properties of EGCG are central to its role in reducing oxidative stress and mitigating neuro-traumatization, particularly in neurodegenerative diseases. By scavenging ROS, EGCG helps maintain cellular homeostasis and protects neurons from oxidative damage. Additionally, EGCG induces apoptosis in cancer cells by modulating several pro-apoptotic and anti-apoptotic proteins, efectively promoting programmed cell death in abnormal cells while preserving healthy tissue.

Table 1 Target proteins of EGCG

EGCG reduces ROS oxidative stress

EGCG inhibits cellular mechanisms and tissue oxidative injury by preventing pro-oxidant enzymes. EGCG, having high lipophilic features, illustrated a better affinity for free radicals than the unchanged EGCG [\[70](#page-16-10)]. EGCG has been reported as a potent antioxidant and possesses ben-eficial effects in oxidative stress-related diseases [\[71\]](#page-16-11).

A study [[72](#page-16-12)] demonstrated the anti-oxidative capabilities of EGCG in H_2O_2 -mediated oxidative stress of myocardial ischemia injury $[73]$ $[73]$ $[73]$. The levels of superoxide dismutase (SOD) and glutathione peroxidize (GPx) activity ameliorates with the treatment of H2O2-induced control biopsies with EGCG at the concentration of (50 μ g/ml) for 24 h. It has also been observed that when H2O2-induced cervical cancer biopsies treated with EGCG (50 µg/ml) suppress the activity of SOD and GPx by 38.54% and 57.04%, respectively. Similar results were obtained with HeLa cells. Improvement in SOD and GPx activity by co-culture of EGCG indicates it to be an efective natural antioxidant combating ROS generation [[74\]](#page-16-14). Thus, it can be rightfully said that EGCG imparts an inhibitory efect and cancer chemo-preventative agent on the activity of many enzymes and metabolic pathways.

EGG modulates the pathway, which directly inhibits cytokines infammation and formation of ROS in uric acid-induced human umbilical vein endothelial cells [\[75](#page-16-15), [76\]](#page-16-16). EGCG reduces the mRNA levels of Notch1, Hey1, and Hes1 in a dose-dependent manner, leading to the inhibition of cancer growth. Additionally, EGCG suppresses Notch promoter activity, further contributing to its anticancer efects. In vivo studies demonstrated a signifcant reduction in both tumor growth and Notch1 expression in mice injected with EGCG. Similar fndings were observed in EGCG-treated colorectal cancer cells, where decreased levels of Hes1 and Notch2 were reported. Notably, EGCG was also found to cleave Notch1 in 5-fuorouracil-resistant colorectal cancer cells, indicating its potential role in overcoming drug resistance [[77\]](#page-16-17).

 EGCG signifcantly improves the survival rate of human umbilical vein endothelial cells exposed to H_2O_2 -induced oxidative stress [[61](#page-16-18)]. EGCG was found to upregulate key autophagy-related proteins, including Atg5, Atg7, LC3 II/I, and the Atg5–Atg12 complex. It also inhibits the mTOR signaling pathway, further promoting EGCG-induced autophagy **(**Fig. [3](#page-5-0)**).** In addition, EGCG disrupts the Janus kinase (JAK)/signal transducer and activator of transcription (STAT) signaling pathway at multiple stages by inhibiting STAT phosphorylation, thereby preventing its translocation to the nucleus [[78\]](#page-16-19). Specifcally, EGCG inhibits the phosphorylation of the 705th tyrosine residue of STAT, and protein kinase C delta (PKC-delta), efectively blocking STAT1 activity. Moreover, EGCG suppresses the activity of interferongamma (IFNγ), JAK1, JAK2, and STAT1/STAT3. Interestingly, EGCG-mono-palmitate, a derivative of EGCG, activate the Src homology 2 domain-containing tyrosine phosphatase-1 (SHP-1) enzyme, which reduced the phosphorylated levels of BCR-ABL and STAT3 in human chronic myeloid leukemia cells [[79](#page-17-0)].

EGCG has shown protective efects when administrated to patients with contrast-induced nephropathy. It normalized kidney function markers, such as serum creatinine and blood urea nitrogen. It is responsible for improving renal tissue health, as indicated by histological analysis. Metabolic physiology showed reduced cell death and oxidative stress within the kidneys and alleviated infammation in the renal tissue. EGCG-loaded chitosancasino phosphopeptide nanoparticles show free radical scavenging activity [\[80](#page-17-1)].

Furthermore, EGCG decreased both the expression and secretion levels of pro-infammatory cytokine genes, including TNF-α, IL-1β, and IL-6. EGCG also downregulated the expressions of α-SMA, fbronectin, mast cell

Fig. 3 EGCG-Mediated Pathways Targeting Oxidative Stress. Showing the diverse molecular pathways through which EGCG mitigates oxidative stress. EGCG inhibits the phosphorylation of STAT3, thereby disrupting the JAK/STAT signaling pathway, which is crucial for cell proliferation and survival. Additionally, EGCG targets key proteins in the Delta-Notch pathway, including Notch, HEY, and HES1, further infuencing cellular diferentiation and survival mechanisms

trypsin, and chymotrypsin, as well as TGF-β1, CTGF, and PAI-1 [[81\]](#page-17-2) as an emerging potential phytocompound EGCG has come upon with its recently discovered role in fbrosis. Due to its anti-infammatory and anti-proliferative role, EGCG promotes wound healing and prevents scar formation. This is achieved by blocking the pathways of VEGF, CTGF, and TGF-β1. It downregulates the formation of skin scars by reducing the production of COL-1 [\[82\]](#page-17-3). EGCG may be efective in treating skin scars by decreasing blood flow and skin thickness, reducing mast cell activity and angiogenesis while also enhancing heme oxygenase-1 levels, increasing M2 macrophage counts, and boosting elastin, antioxidant activity, and hydration [[81\]](#page-17-2).

EGCG reduces traumatic brain injury

EGCG prevents traumatic brain injury (TBI)-mediated IL-1β and TNF- α mRNA in mice brains [\[83\]](#page-17-4). EGCG considerably diminished the levels of malondialdehyde (MDA), superoxide dismutase (SOD), and glutathione peroxidase (GSH-PX) after TBI. EGCG diminished TBImediated NADPH oxidase activation by inhibiting p47 translocation to the plasma membrane [\[55,](#page-16-29) [84](#page-17-5)]. Preclinical trials refect the therapeutic potential of EGCG in suppressing the occurrence of secondary brain damage after TBI. The intake of EGCG observes a reduction of brain infarction and oedema in rodent models. EGCG preserves blood–brain barrier integrity through the modulation of tight junction proteins, limiting the entry of blood-derived substances into brain parenchyma. It inhibits the activation of microglial, reducing TNF-α, IL-1β, and IL-6 and alters the NF-κB pathway in the injured rat brain. Additionally, it remarkably reduces neutrophil infltration and cytokine pro-infammation [[85\]](#page-17-6).

Therapeutic benefits of green tea consumption were reported in infammatory and NDs [\[86](#page-17-7)]. EGCG efectively inhibited IL-8 in epithelial cells. IL-8 is responsible for the recruitment of neutrophils and supports the formation of ROS. EGCG acts as an antioxidant by inhibiting the phosphorylation of IκB to prevent IL-1β-induced NF-κB activation [\[87](#page-17-8)]. EGCG inhibited IL-1β-mediated expression of iNOS and COX-2 by inhibiting proteasomal degradation of I κ B α [[88\]](#page-17-9). EGCG prevents the binding of AP1 to DNA via IL-1β-mediated phosphorylation of c-Jun, thereby downregulating IL-8 and TNF-α progression. Evidence emerged that supports the inhibition of CD4+T cells, promotes IκB-α, and reduces ROS formation in neurons by EGCG. EGCG prevents the binding of AP1 to DNA via IL-1β-mediated phosphorylation of c-Jun, thereby downregulating IL-8 and TNF-α progression [\[89\]](#page-17-10).

EGCG acts as a neuroprotective agent against Aβ-induced oxidative stress by increasing the antioxidants in BV2 microglial cells [[90\]](#page-17-11). EGCG reduces the NF-κB signaling activation, obstructing the formation of cytokines and VEGF in U373MG cells [[91](#page-17-12)]. EGCG represses Aβ-induced neuroinfammatory reactions of microglia, including TNFα, IL-1β, and iNOS, and enhances the levels of antioxidants [[92\]](#page-17-13). It repressed Aβ-induced NF-κB and MAPK pathways, including JNK and p38 [\[93](#page-17-14)[–96\]](#page-17-15). EGCG blocks infammatory and

oxidative stress markers via the regulation of nuclear receptor PPARγ in N2a/APP695 cells [[97](#page-17-16)].

EGCG induces apoptosis: anti‑proliferation

EGCG, combined with chemotherapeutic drugs, experimentally showed higher sensitivity against tumor cells. At the same time, the anti-infammatory and antioxidant efects of EGCG reduce the negative impact caused by the chemotherapy. Clinically, the nanomodifcation of EGCG signifcantly increases anti-tumor activity. One such pathway illustrated with doxorubicin (DOX). EGCG enhances anti-tumor activity by inhibiting the activity of carbonyl reductase 1 (CBR1) and P-gp in the liver cells. Here, EGCG binds to CBR1 and inhibits the conversion of daunorubicinol (DNROL) to DOX, enhancing antitumor activity and inducing cardiotoxicity [[98\]](#page-17-17).

Combining EGCG and sulforaphane with cisplatin signifcantly increases its efectiveness against both sensitive and resistant ovarian cancer cells. This combination treatment reduces cell survival, accelerates cell death (apoptosis), and halts cell division at the G2/M phase in a time- and dose-dependent manner [[99](#page-17-18)]. EGCG reduced EGFR phosphorylation, suppressed AKT signaling in breast cancer cells, and amplifed raloxifene-induced apoptosis [\[100](#page-17-19)]. In mice, combined EGCG with tamoxifen leads to a decrease in tumor volume by 71% and tumor weight by 80% by inhibiting mTOR and EGFR expression $[101]$ $[101]$ $[101]$ (Fig. [4](#page-6-0)).

EGCG administration not only extended the lifespan of lethally irradiated mice but also mitigated radiationinduced damage to the intestinal mucosa. Treatment with EGCG signifcantly increased the population of Lgr5+intestinal stem cells (ISCs) and their proliferating Ki67+progeny while simultaneously reducing radiation-induced DNA damage and apoptosis. Moreover, EGCG efectively lowered ROS levels and activated the Nrf2 transcription factor, which in turn upregulated antioxidant proteins such as Slc7A11, HO-1, and GPX4. Consequently, EGCG acts as a protective agent against radiation-induced injury by scavenging ROS and inhibiting both apoptosis and ferroptosis via the Nrf2 signaling pathway [\[102\]](#page-17-21).

EGCG induces cellular signaling and epigenetic modifcation

EGCG regulates the cell cycle by modulating cyclindependent kinases (CDKs). It inhibits the activity of CDK1 and CDK2 and activates cyclin-dependent protein kinase inhibitors such as p17, p21, and p16 [\[103](#page-17-22)]. EGCG exerts growth inhibitor effort on human cancer cell lines without affecting normal cells of the body. The efect is dose-dependent, inhibiting cell growth, G0/ G1 phase arrest, and DNA damage. Such physiological

pliferat Fig. 4 Impact of EGCG on tumor-mediated signaling cascades through apoptosis. EGCG blocks signaling cascade activation and promotes apoptosis. Outline the promising gene targets engaged in anti- and pro-apoptotic activities of low and high EGCG concentration. This efect could be attained via the increased regulation of p53 expression. EGCG enhances the ratio of Bax/Bcl-2

and activates apoptosis

role indices apoptosis in many cancer cell lines. Molecular docking studies have revealed CDK-6 as the better ligand for EGCG with binding energy−12.70 and docking energy−11.40 kcal/mol, as compared to other CDK-6 inhibitors. This study was confirmed with experimental immunoblot assay analysis, which proved that EGCG is an inhibitor of CDK-6 [[104,](#page-17-23) [105](#page-17-24)].

The role of EGCG in epigenetic modification has always been appreciated. Sjögren's syndrome (SS) is an autoimmune disease particularly afecting the exocrine glands. There has been a correlation between the syndrome and EGCG, which is responsible for altering the gene expression of related molecules. Such alterations lead to ameliorated salivary gland damage. The mechanism remains the same, including the reduction in ROS activity and accumulation, thereby inhibiting ROS mediating water aqua channel 5. Eventually, EGCG is responsible for increasing the saliva flow $[106]$ $[106]$.

EGCG can inhibit DNA methyltransferases, the enzymes responsible for adding methyl groups to DNA [107]. This inhibition can lead to the demethylation of certain gene promoters, potentially activating genes that were previously silenced. In addition, EGCG has been observed to afect histone acetylation by inhibiting histone deacetylases. Increased histone acetylation typically leads to a more relaxed chromatin structure, which enhances gene transcription. Furthermore, EGCG can infuence the expression of microRNAs, which are small non-coding RNAs that regulate gene expression posttranscriptionally. Changes in miRNA levels can afect various cellular processes and contribute to therapeutic efects of EGCG [\[108](#page-17-27)].

EGCG Therapeutic potential in specifc human diseases

EGCG is a promising therapeutic agent reported for its diverse biological activities, including antioxidant, antiinfammatory, and anticancer properties. Many of these interactions require EGCG concentrations far higher than those obtainable through regular green tea consumption or standard green tea extract supplements. Despite this limitation, recent well-conducted clinical trials have demonstrated the efficacy of green tea extracts and purifed EGCG products in treating specifc conditions. This section focuses on clinically relevant studies, highlighting recent advancements in EGCG-based therapies for diseases. Additionally, this review explores EGCG's mechanisms of action, examining the existing evidence supporting its therapeutic potential in addressing human diseases.

Oral health and dentistry

Gingivitis and periodontitis are infammatory mouth infections in which gums turn red and swollen and sometimes bleed. EGCG helps reduce periodontal disease development by inhibiting the growth of *Porphyromonas gingivalis, Prevotellani crescent*, and *Prevotella* intermedia. These bacteria are engaged in periodontal tissues, causing periodontitis. The efficiency of catechins defends the *gingival* epithelium against the invasion of *Porphyromonas gingivalis,* a potent cause of periodontal disease. Treatment with EGCG blocks the functioning of the Matrix Metalloproteinase-9 (MMP-9) factor and supports osteoclasts in periodontal illness. MMP-2, 8, and 9 present in the dentine region of the oral cavity are responsible for the degradation of tooth decay. EGCG was reported as an MMP inhibitor [[109,](#page-17-28) [110\]](#page-17-29).

Acrolein was found to prevent *gingival* fbroblasts, usually because of cell disruptions and growth. Hence, this might increase several infammatory situations in the oral cavity, which could cause gingival and periodontal disease. EGCG may decrease the toxicity of acrolein. Therefore, green tea/EGCG has been confrmed as a wonder drug for oral health [\[111\]](#page-17-30). Due to inadequate diets among the youth, the uptake of large amounts of carbohydrates and fermented sugars leads to the accumulation of acidproducing microbiota. Among all microbiota, *S. mutans, Lactobacillus,* and *Actinomyces* viscous are dominant and highly implicated in the development of dental caries.

 A high dose of EGCG kills bacterial structure whereas a low dose is effective in anti-bacterial nature by inhibiting virulence factors. In either dose, EGCG leads to destruction in the biofilm growth. Such efficacy is prominent against oral *Candida*. A high dose induces damage to mitochondrial membranes and uncoupling of oxidative phosphorylation. EGCG when combined with other drugs, shows a higher potency towards drug-resistant strains. EGCG is directly involved in the inactivation of oral viruses such as HPV, HSV, and other viral proteins [[112](#page-17-31)].

Tea leaves are rich in fuoride, which improves dental health. Dental caries are induced by oral microfora. Microbial dysbiosis containing Gram-positive and Gram-negative aerobic and anaerobic bacteria afects the progression of cariogenic dental plaque [[112–](#page-17-31)[115](#page-18-0)]. EGCG inhibits sugar transport and acid formation via lactate dehydrogenase [\[116\]](#page-18-1). EGCG is known as a competitive inhibitor of NADPH. In addition, EGCG inhibits the NADPH oxidase translocation and ROS production $[117]$ $[117]$ $[117]$. Consuming catechins daily efficiently decreases dental caries, and using EGCG/green tea mouthwash diminishes the acidity of saliva and prevents bacterial colonization. The study enhanced salivary pH with EGCG/green tea and decreased dental caries [[118](#page-18-3), [119](#page-18-4)].

EGCG acts as the main compound in green tea that was reported to inhibit the growth and virulence factor of microbes of oral pathogens. Though the mechanism is unclear, it has been validated that EGCG is responsible for reducing volatile sulfur compounds by suppressing the *mgl* gene. *Mgl* gene is encoded for enzymes L-methionine-α-deamino-γ-mercaptomethane-lyase, responsible for methyl mercaptan (CH3SH) production by oral anaerobes. EGCG inhibits the growth of *P. gingivalis* and reduces CH3SH production and *mgl* gene expression $[112]$ $[112]$ $[112]$. The reaction has been carried out by introducing a methyl sulfonyl group into the B-ring of EGCG. A methylation reaction is set to attach the orthoquinone site via oxidation, which facilitates reduction in halitosis. Hence, green tea helps to lower the oral odor [\[114](#page-18-5)].

Solobacterium moorei is an anaerobic bacterium grouped under a volatile sulfde compound (VSC) associated with halitosis. Upon performing a microplate dilution assay, EGCG was found to show anti-bacterial activity against *S. moorei*. EGCG inhibits the growth of *S. moorei*, with MIC values of 250 μg/ml [\[120\]](#page-18-6). Through transmission electron microscopy and a permeabilization assay, it was found that EGCG is responsible for targeting bacterial cell membranes and attenuating their integrity. Similar results were obtained when an analysis was performed on colonization properties. It was found that EGCG signifcantly reduces colonization and adherence to oral epithelial cells. In addition, EGCG at ½ MIC signifcantly decreased the $β$ -galactosidase gene expression $[120]$.

Cancer

Oral cancer

EGCG inhibits cell cycle progression and modulates signalling pathways that cause cancer. Its therapeutic potential has been efective against oral cancer. EGCG imparts the G1 arrest of tumor cells, and its treatment signifcantly enhances caspase-7, 9, and −3 activity along with increased expression of Bax [[121\]](#page-18-7). Indirectly, EGCG is involved in caspases-induced apoptosis.

EGCG causes cell death in cancerous cells by modulation of several signalling cascades**.** It diminishes cell proliferation via suppressed Akt, NF-κB, EGFR, and MAPK/ERK1/2 pathways [[47\]](#page-16-0). EGCG acts as a potent drug for tumor chemoprevention capability for cancer transformation of oral premalignant lesions, tumor proliferation blockage, and cell death initiation (Fig. [5](#page-8-0)).

MMP-9 is associated with oral tumour development [[122](#page-18-8), [123\]](#page-18-9). EGCG blocks the invasion and migration of human oral tumour cells by downregulating the MMP 2 and 9 [[124](#page-18-10)] (Table [2](#page-9-0)). By regulating MMP, metastasis is also prevented. A partial decrease in angiogenesis was observed in VEGF receptor phosphorylation in oral tumours $[5]$ $[5]$. The effects of EGCG in hindering HGF-mediated proliferation of oral cancers have been reported. EGCG blocks HGF-induced Met phosphorylation and the expression of MMP-9 and -2 [\[125](#page-18-11), [126\]](#page-18-12). It causes an inhibitory efect on cell motility, migration, and adhesion $[127]$ $[127]$. The outcomes of green tea/EGCG hinder the development of premalignant lesions for the tumor [[128\]](#page-18-14).

Breast cancer

EGCG has demonstrated a signifcant potential in the prevention of various types of cancer. Studies revealed that in a concentration-dependent manner, EGCG shows antioxidant or pro-oxidant properties. Such characteristics endure EGCG to block cell cycle progression and modulate signaling pathways during cancer prognosis. EGCG inhibits the transpiration of VEGF by inducing apoptosis and negatively modulates metastasis . Similar results were obtained from in vivo studies on xenograft animal models [[143\]](#page-18-15).

EGCG interferes with estrogen receptor signaling, which is critical in estrogen receptor-positive breast cancers, potentially reducing the proliferation of these cancer cells. It inhibits the formation of new blood vessels (angiogenesis), which is essential for tumour growth and metastasis in breast cancer. EGCG enhances the efficacy of common breast cancer treatments, such as tamoxifen, by sensitizing cancer cells to these therapies.

Even the overall expression of cyclins (Cyclin D, Cyclin E, CDK 4, CDK 1, and PCNA) was down-regulated in time-dependent increasing concertation of the EGCGtreated group as compared to the untreated control group by Western blot analysis that blocked the G1 phase of the cell cycle [[47\]](#page-16-0). Experiments on nude mice induced with human MDA-MB-231 breast cancer cells

Fig. 5 Role of EGCG in inhibition of cancer growth**.** EGCG acts as an anti-cancer agent either by responding through the RTK pathway or by binding to its 67 -ligand-receptor (67-LR). RTK method mediates via RAS pathway regulating nuclear and cytosolic genes. RTK also alter PI3K-AKT pathway thereby inhibiting angiogenesis. EGCG regulates the process of apoptosis via FLA and TNFα

and treated with EGCG show an efective reduction in tumour incidence [[144](#page-18-16)].

EGCG has demonstrated prominent inhibition in the growth of cancer stem cells (CSCs) when used alone and when used in combination with other chemotherapeutic drugs. It reduces the resistance of CSCs and intracellular efflux. Moreover, EGCG increases the drug concentration in CSCs by inhibiting the activity of the intracellular transporter gene family, ABC transporters, at minimal concentration [\[145](#page-18-17)]. EGCG does not afect the ABC transporter in glioma CSCs but is able to downregulate P-glycoprotein, thus reducing colony formation and the migration of tumour cells $[146]$ $[146]$ $[146]$. EGCG at low concentration inhibits colony formation in pancreatic ductal carcinoma, but when used in combination with PDE3A inhibitor trequinsin, it lowers the expression of proteins FOXO3 and CD44 [[147](#page-18-19)]. Upregulation of cGMP expression is observed, which showed a signifcant reduction in colony formation [\[148](#page-18-20)].

Salivary gland tumors

EGCG inhibits β1 integrin, thereby reducing the expression of MMP-2 and MMP-9, thus providing molecular evidence for the inhibitory efect of EGCG on salivary gland cancer metastasis [\[149\]](#page-18-21). Studies have reported that EGCG inhibits cell proliferation and expression of EGFR, downregulates Bcl-2, and upregulates Bax;, therefore, inducing apoptosis of adenoid cystic carcinoma [150]. EGCG mitigates inflammatory damage to the salivary glands by suppressing infammatory cytokines and reducing oxidative stress. Moreover, it promotes the growth and repair of salivary gland tissue. Additionally, EGCG inhibits the metastasis of salivary gland cancer by downregulating MMP proteins [[151](#page-19-1), [152\]](#page-19-2).

Prostate cancer

EGCG administration leads to the downregulation of cyclin D and cyclin E, which are involved in G1/S progression. This indicates that EGCG induces G1 phase arrest in prostate carcinoma cells. Similar role was observed in breast cancers suggesting chemo-preventiverole of EGCG [[153](#page-19-3)]. Treatment with ECGC on LNCaP prostate cancer cell lines reduces cell proliferation. There was increased expression of androgen receptor and prostate-specifc antigens on the cancer cell lines [[154\]](#page-19-4). EGCG induces apoptosis in prostate cancer cells. It activates pro-apoptotic proteins and inhibits anti-apoptotic proteins, leading to the selective killing of cancer cells while sparing normal cells [\[155](#page-19-5)]. EGCG can cause cell cycle arrest in prostate cancer cells, particularly at the G1 phase, preventing cells from progressing to the S phase and thereby inhibiting tumor growth. Prostate cancer growth is often driven by androgen receptor signalling. EGCG downregulates androgen receptor expression and function, reducing the proliferation of androgendependent prostate cancer cells. In addition, EGCGinhibits the expression of MMP enzymes that degrade the extracellular matrix and facilitate cancer cell invasion and metastasis. By inhibiting MMPs, EGCG reduces the metastatic potential of prostate cancer cells [[149](#page-18-21)].

Others

EGCG exerts anti-cancer efects across various cancer types through multiple mechanisms, including the induction of apoptosis, inhibition of cell proliferation, suppression of metastasis, and modulation of epigenetic changes [[58](#page-16-8), [156](#page-19-6)]. Its ability to target specifc signaling pathways and enhance the efectiveness of existing treatments for cancer prevention and therapy [\[157](#page-19-7)]. EGCG diminished the geftinib-induced overexpression of CYP1A1, CYP1B1, EGFR, cyclin D1, p-Akt (Ser473), and survivin at both the transcriptional and translational levels. EGCG upregulated phosphorylation p53 at Ser15, thereby enhancing gefitinib's therapeutic efficacy against benzo[a]pyrene-induced lung cancer [[158\]](#page-19-8).

EGCG inhibit lung cancer cell growth by causing cell cycle arrest and promoting apoptosis. It reduces the metastatic potential of lung cancer cells by downregulating the expression of MMPs and other factors involved in cell migration and invasion [\[33\]](#page-15-21). By reducing oxidative stress, EGCG decreases the risk of lung cancer initiation and progression, especially in smokers and individuals exposed to environmental carcinogens [\[159](#page-19-9)].

Due to its numerous health benefts, EGCG has been increasingly used in the treatment of acute and chronic respiratory diseases. EGCG afects tumour initiation and progression by downregulating angiogenesis and metastasis. Its involvement is remarkable in increasing tumor suppressor genes, apoptosis, neoplastic cells, and cell cycle arrest. EGCG downregulates (Intercellular adhesion molecule) ICAM-1 expression and the counts of neutrophils and eosinophils in the bronchoalveolar lavage fuid (BALF) in human pulmonary alveolar epithelial cells [[160\]](#page-19-10).

An evident correlation was observed between EGCG and Nonalcoholic fatty liver disease (NAFLD). EGCG efectively ameliorated NAFLD disorder and its phenotypic [\[161](#page-19-11)]. It inhibits intestinal barrier and infammations. This is achieved by increasing the concentration of gut microbes, including short-chain fatty acid (SCFA) producers such as gram-negative *Lactobacillus*. An increase in SCFA level inhibits the TLR4/NF-κB pathway, thereby alleviating liver inflammation. Thus, it suggests that the role of EGCG on gut microbiota is to reduce the infammation in hepatocytes [\[141\]](#page-18-34).

EGCG targets survival pathways such as PI3K/Akt and NF-κB, which are often upregulated in pancreatic cancer $[162]$ $[162]$. This leads to reduced cell proliferation and increased apoptosis. EGCG enhances the sensitivity of pancreatic cancer cells to chemotherapy, potentially overcoming resistance to conventional treatments [\[163](#page-19-13)]. EGCG inhibits the metastatic spread of pancreatic cancer cells by downregulating enzymes and signalling molecules involved in invasion and migration (Fig. [5](#page-8-0)).

EGCG was reported as a noncompetitive inhibitor of NAD kinase with a *K*i of 3.28±0.32 μM. SPR analysis revealed a K_d of 1.78 ± 1.15 μ M, while HDX-MS indicated binding at non-substrate sites on NADK [\[63\]](#page-16-22). EGCG selectively inhibited the growth of KRAS-mutant lung cancer cells without afecting KRAS wild-type cells [\[164\]](#page-19-14).

Cardiovascular diseases *Atherosclerosis*

Atherosclerosis is a disease of the arteries caused by endothelial dysfunction, infammatory vascular cells, and lipid accumulation. EGCG-treated mice showed a 23% decline in aortic weights and cholesterol as well as TG [\[165](#page-19-15)]. Studies exhibited a considerable decrease in LDL-C, which was examined in EGCG-treated subjects. Therefore, these outcomes recommend that EGCG has the potential to block the progression of CVD and hypertension via an important decrease in LDL-C [[166](#page-19-16), [167](#page-19-17)]. Studies revealed that EGCG acts as an inhibitor of HDAC1, leading to better functioning of the heart. It decreases heart/body weight and the ratio of mtDNA/ nDNA. In addition to inhibiting HDAC1, EGCG increases the binding of acetylated H3K9 or H3K14 in the promotor regions of peroxisome proliferator-activated receptor-1α and nuclear respiratory factors $[168]$.

Cardiac hypertrophy and heart failure

The protection of cardiac homeostasis comprises hypertrophy and enlargement of heart size, as well as increased protein synthesis, which are the major symptoms of cardiac hypertrophy that generally cause a decline in the heart's capability to pump blood to the organs. The therapeutic potential effect of EGCG in treating hypertension-mediated learning as well as memory impairment is possibly due to its infuential antioxidative roles [[169\]](#page-19-19). EGCG is helpful in fghting against aging-mediated cardiac hypertrophy, fbrosis, and cell death [[170\]](#page-19-20). EGCG prevents the development of heart failure in mice via inhibition of myocardial fbrosis and decline of ventricular collagen remodelling [[171\]](#page-19-21). EGCG inhibits NF-κB activation and subsequent TGF overexpression. However, it diminished the expression of fbronectin and the proliferation of rat cardiac fibroblasts $[165, 172]$ $[165, 172]$ $[165, 172]$ $[165, 172]$. The treatment of EGCG drastically recovered cardiac diastolic role via up-regulating cTnI via preventing histone deacetylase 1/3 expression. EGCG can contribute to the hindrance of cardiac diastolic dysfunction [\[173\]](#page-19-23).

Neurodegenerative disorders

An efective and well-mentioned role of EGCG is to reduce the formation and accumulation of ROS production and increase pro-apoptotic markers in various tissues. EGCG imparts neuroprotective efects and widens neuro-rescue actions; thus, EGCG shows neuroprotective efects. Some human studies have revealed a dose-dependent relationship between EGCG intake and neuro-related disorders such as AD and PD [\[10](#page-15-1), [174](#page-19-24)].

PD is a neurodegenerative disorder associated with the accumulation of Fe, oxidative stress, and infammation. It is distinguished by nigrostriatal degeneration, which may involve $α$ -synuclein $(αS)$ aggregation, dysregulation of redox metal homeostasis, and neurotoxicity. However, tea polyphenols play a vital function, halting the progression of PD development. Tea polyphenols can directly obstruct the aggregation of the αS protein and alter signaling cascades in animal models [\[175\]](#page-19-25).

EGCG prevents αS aggregation and interferes with PD development. The aggregation has reduced via EGCG in a concentration-dependent way, as revealed through a lack of thioflavin T binding $[176, 177]$ $[176, 177]$ $[176, 177]$ $[176, 177]$ $[176, 177]$. αS aggregates have been identifed via incubation with A-Syn-HiLyte488. This binding has been inhibited via EGCG [176]. The α S amino acid positions that are interrelated with EGCG have been identifed on peptide membranes. EGCG attaches to αS through unstable hydrophobic interfaces. EGCG might be a promising altering drug of αS aggregates for the management of PD and other α-synucleinopathies [\[176](#page-19-26)].

EGCG post-treatment signifcantly rescued 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP)-induced neurotoxicity by increasing the rotational latency and increasing dopamine [[178](#page-19-28)[–181\]](#page-19-29). EGCG has antioxidant and iron-chelating properties. EGCG blocked MPTP-mediated neurotoxicity by raising the locomotor

activity, positive neurons, and striatal dopamine (DA) concentrations. A decline in TH action with iron, EGCG inhibited MPTP-induced neurodegeneration and rescued dopaminergic neurons after MPTP treatment in iron excess condition [\[182](#page-19-30)[–184\]](#page-19-31).

The neuroprotective functions of EGCG were reported in PD models. Because of its multi-targeted activities, EGCG might have the potential to be a new drug for the treatment of PD and for preventing neurodegeneration. The molecular mechanism by which EGCG exerts neuroprotective advantages, such as inducing apoptosis and inhibition of infammation, oxidative stress, modulation of dopamine making, and the aggregation of αS , is described in Fig. [6](#page-11-0).

The main pathological characteristics of AD is amy-loid β peptide (Aβ) accumulation in the brain [[185](#page-19-32)[–187](#page-20-0)]. Aβ is believed to be a vital neuroinfammatory stimulus to microglia. Treatment with EGCG has been found to reduce the aggregation of amyloid $[16]$ $[16]$. Aβ aggregates have been diminished after EGCG treatment in APP transgenic animal models. EGCG controlled (amyloid precursor protein) APP processing and subsequently reduced $\text{A}\beta$ deposition [\[16](#page-15-7)].

EGCG is involved in the non-amyloidogenic procedure via stimulating α-secretase cleavage in SweAPP N2a cells.

Fig. 6 The potential neuroprotective efects of EGCG in PD**.** The mechanisms of EGCG exert neuroprotective advantages. EGCG can prevent protein misfolding, neuronal apoptosis, oxidative stress, and neuroinfammatory responses

EGCG enhances sAPP-α and diminished Aβ formation in MC65 cells, hence showing the defensive nature of EGCG on brain edema and neuronal injury $[16]$ $[16]$. EGCG bypassed the blood–brain barrier (BBB) to accomplish the efficient parts of the brain. EGCG is being considered a promising agent for managing NDs [\[188–](#page-20-1)[190\]](#page-20-2). The function of EGCG in treating AD has been studied, demonstrating that EGCG participates in a neuroprotective function and is promising to be utilized as a therapeutic drug for treating AD (Fig. [7](#page-12-0)).

EGCG enhances the production of soluble proteins of AβPP, sAβPPα through PKC-dependent α-secretase activation. Cleavage of AβPP to sAβPPα leads to a non-amyloidogenic secretory pathway accomplished via a putative α-secretase, therefore precluding the formation of Aβ, the last is controlled through the sequential activity of $β$ and γ -secretases [[191\]](#page-20-3). The neuro-protecting results of EGCG against Aβ-mediated neuronal loss and tau toxicity in AD models have been identifed in multiple studies. EGCG, with its APP processing ability, provides a hopeful and diferent strategy for AD prevention [[16](#page-15-7)].

Metabolic disorders

Diabetes mellitus

Metabolic diseases (MDs), diabetes, and obesity are the most common disorders [\[192\]](#page-20-4). Diabetes mellitus (DM), one of the most frequent metabolic disorders worldwide, is attributed to hyperglycemia caused by either reduced insulin emission or insulin resistance. Multiple studies

Fig. 7 The promising efects of EGCG in AD pathogenesis**.** EGCG participates in a neuroprotective function and is prospectively utilized as a therapeutic drug for managing AD

have revealed that type 2 diabetes mellitus (T2DM) might stimulate various complications, including diabetic cardiovascular complications, diabetic nephropathy, and neuropathy, which were the main reasons for its mortal-ity and morbidity [\[193](#page-20-5)]. The chief pathophysiologic factors that cause T2DM are peripheral insulin resistance and the last devastation of insulin creator pancreatic cells [[194,](#page-20-6) [195\]](#page-20-7).

The antioxidant properties of EGCG have been studied in multiple diseases. However, improving the bioavailability of EGCG via nano-formulation might contribute to a more productive treatment of DM metabolic consequences and vascular complications [\[196\]](#page-20-8). Although the mechanisms by which EGCG is related to the onset of DM are still unknown, the possibility might explain the hyperglycemia-induced infammation (Fig. [8\)](#page-13-0). A report has discussed a case of popular interstitial EGCG composed of MMP-9-bearing cells in a type II DM patient. EGCG improves insulin sensitivity and glycaemic control and drastically diminish serum triglycerides and total cholesterol levels following long-term supplementation [\[142](#page-18-35)]. Additionally, EGCG declined triglycerides and considerably enhanced HDL and glucagon-like peptide 1 levels in a randomized, double-blinded, placebocontrolled clinical trial linking patients with T2DMs and lipid abnormalities [[197\]](#page-20-9).

Obesity

Obesity is predominantly the outcome of a positive energy equilibrium determined by enhanced calorieenrich food utilization and minimal exercise. It is considered a low-grade systematic infammation disease. It could react quickly and animatedly to modifcations in nutrient excess via adipocyte hypertrophy as well as hyperplasia. Obesity is principally motivated by an inequity between energy intake and expenditure; this modifcation leads to an improvement of the adipose tissue due to the gathering of lipids that happens in peripheralrelated organs [\[198\]](#page-20-10). EGCG is shown to have anti-obesity as well as anti-diabetic effects $[199]$ $[199]$ $[199]$. The promising role of ECGC as an antioxidant and anti-infammatory in the formation of gut microbiota and favours the formation of bacteria such as *Alloprevotella* and *Muribaculaceae* [[200\]](#page-20-12). EGCG enables the reduction of negative regulating bacteria*, Blautia,* hence mining the functions of probiotics to improve intestinal infammation and treat obesity.

Apart from gut improvement, EGCG shows its prominent role in balancing the level and proportion of gut SCFAs, the expression levels of infammatory and transcription factors, and alterations in hypothalamic neurotransmitters. This suggests the effect of EGCG on the overall body axis, thereby improving obesity and its induced infammatory response [[201\]](#page-20-13).

Fig. 8 The Role of EGCG in Diabetes Management. Showing the pivotal role of EGCG in diabetes, particularly through its interaction with the KEAP1-Nrf2 signaling pathway. EGCG forms a hybrid complex with glutathione (GSH), which binds to KEAP1, resulting in the dissociation of KEAP1 from Nrf2. This dissociation allows Nrf2 to translocate into the nucleus, where it initiates the transcription of crucial antioxidant proteins, including heme oxygenase-1 (HO-1) and NAD(P)H quinone oxidoreductase 1 (NQO1). These antioxidant proteins play an important role in reducing oxidative stress and infammation, key contributors to diabetic complications

Although epidemiological and clinical studies explain the health advantages of EGCG on diabetes and obesity, the mechanisms of its actions are promising based on several laboratory data. EGCG had a considerable outcome on the diminished obesity in body weight gain and decline in epididymal adipose tissue weight, which infuenced serum lipid attributes, such as triglyceride and cholesterol [\[202\]](#page-20-14).

Conclusions and future prospects

EGCG, a primary polyphenol present as a major composition in green tea, has achieved a signifcant attantion due to its proven importance in maintaining human health. It has a wide range of biological activities in preventing various disease. EGCG has antioxidant, antiinflammatory, and anti-microbial properties. These activities are connected to its ability to inhibit the growth of cancer cells, prevent tooth decay, reduce infammation, and protect neurons from damage. EGCG improves overall health in oral dentistry, including bad breath, oral cancer, tooth decay, and oral cavity infammation.

While preclinical studies highlight the potential of EGCG in treating various diseases, rigorous clinical trials are needed to establish its efficacy and safety in humans. Future research should focus on determining

the optimal dosage, treatment duration, and delivery methods to maximize therapeutic benefts while minimizing potential side effects. Efficient delivery of EGCG to target tissues remains a challenge due to its poor bioavailability and rapid metabolism. Future research could explore advanced delivery systems such as nanoparticles, liposomes, or other innovative drug delivery technologies that enhance stability and bioavailability of EGCG, allowing it to reach specifc tissues more efectively.

EGCG modulates various signaling pathways and opens avenue for its use in combination with other therapeutic agents. EGCG could be tailored to target specifc signaling pathways unique to certain cancer types. This precision approach may lead to more personalized cancer therapies with higher efficacy. Future studies could investigate the synergistic efects of EGCG when combined with other drugs or natural compounds, potentially enhancing treatment outcomes for conditions like cancer, neurodegenerative diseases, and metabolic disorders.

In addition to advancements in genomics and personalized medicine, there is potential to tailor EGCGbased therapies to individual genetic profles. Although EGCG is generally considered safe, long-term studies are needed to assess its safety profle, particularly at higher doses or with prolonged use. Investigating

potential interactions with other medications and understanding the long-term efects on diferent organ systems will be crucial for its safe therapeutic application.

EGCG has signifcant potential as a chemopreventive agent, particularly in individuals at high risk of developing certain cancers. Future studies could explore the role of EGCG in preventing cancer initiation and progression, potentially leading to its use in preventive strategies for at-risk populations. The role of EGCG in neuroprotection, cancer prevention, and oral health, future studies could explore its potential in other areas such as cardiovascular health, diabetes management, and immune system modulation. Expanding the scope of research could uncover new therapeutic applications for EGCG. The future of EGCG in disease therapy lies in its potential to serve as both a preventive agent and a therapeutic adjunct. By developing advanced delivery systems, combining EGCG with existing therapies, and conducting rigorous clinical trials, EGCG could become an integral part of newer treatments.

For EGCG to transition from experimental studies to clinical practice, it must undergo rigorous evaluation . EGCG holds signifcant promise as a therapeutic agent for various diseases, particularly in neuroprotection, cancer prevention, and oral health. However, realizing its full potential requires continued research, clinical validation, and the development of innovative delivery methods. Overall, the available evidence suggests that EGCG has the potential to be a safe and efective treatment for several oral diseases and neurodegenerative diseases. Hence, more investigation is needed to validate these fndings and to develop more efective delivery methods for EGCG.

Abbreviations

Acknowledgements

MA thanks the Indian Council of Medical Research for fnancial support (Grant No. 45/6/2020-DDI/BMS). MIH acknowledges the Indian Council of Medical Research for fnancial support (Grant No. ISRM/12(22)/2020). AS thanks to the Ajman University for the payment of APC.

Authors' contributions

All authors have read and agreed to publish the current version of the manuscript. Manzar Alam: Conceptualization, Writing- Original draft preparation, Data curation, Investigation, Methodology; Mehak Gulzar: Writing- Original draft preparation, Investigation, Methodology; Mohammad Salman Akhtar: Writing- Original draft preparation, Investigation, Methodology; Summya Rashid: Writing- Original draft preparation, Data curation, Methodology, Zulfareen: Data curation, Investigation, Methodology; Tanuja: Writing- Original draft preparation, Data curation, Investigation, Methodology; Anas Shamsi: Writing-Original draft preparation, Data curation, Md. Imtaiyaz Hassan: Conceptualization, Writing- Original draft preparation review and editing, Investigation, Supervision, project administration. All authors have read and approved the fnal version of the manuscript.

Funding

This work is supported by the Central Council for Research in Unani Medicine (CCRUM), Ministry of AYUSH, Government of India (Grant No. 3–69/2020- CCRUM/Tech).

Data availability

All data generated or analyzed during this study are included in this manuscript and are attached to this article.

Declarations

Ethics approval and consent to participate Not applicable.

Consent for publication

Not applicable.

Competing interests

The authors declare no competing interests.

Received: 16 May 2024 Revised: 27 September 2024 Accepted: 29 October 2024
Published online: 27 December 2024

References

- 1. Fraga CG, Croft KD, Kennedy DO, Tomás-Barberán FA. The efects of polyphenols and other bioactives on human health. Food Funct. 2019;10(2):514–28. [https://doi.org/10.1039/C8FO01997E.](https://doi.org/10.1039/C8FO01997E)
- 2. Zhang S, Mao B, Cui S, Zhang Q, Zhao J, Tang X, et al. Absorption, metabolism, bioactivity, and biotransformation of epigallocatechin gallate. Crit Rev Food Sci Nutr. 2024;64(19):6546–66. [https://doi.org/10.](https://doi.org/10.1080/10408398.2023.2170972) [1080/10408398.2023.2170972.](https://doi.org/10.1080/10408398.2023.2170972)
- 3. Suzuki T, Miyoshi N, Hayakawa S, Imai S, Isemura M, Nakamura Y. Health benefts of tea consumption. Beverage impacts on health and nutrition. Springer; 2016. p. 49–67. https://doi.org/10.1007/978-3-319-23672-8_4
- 4. Hayakawa S, Oishi Y, Tanabe H, Isemura M, Suzuki Y. Tea, Coffee and Health Benefts. Bioactive Molecules in Food; Mérillon, J-M, Ramawat, KG, Eds. 2018:1–58.<https://doi.org/10.3390/molecules25194553>
- 5. Kciuk M, Alam M, Ali N, Rashid S, Głowacka P, Sundaraj R, et al. Epigallocatechin-3-gallate therapeutic potential in cancer: mechanism of action and clinical implications. Molecules. 2023;28(13):5246. [https://](https://doi.org/10.3390/molecules28135246) [doi.org/10.3390/molecules28135246.](https://doi.org/10.3390/molecules28135246)
- 6. Reygaert WC. Green tea catechins: Their use in treating and preventing infectious diseases. BioMed Res Int. 2018;2018:9105261. [https://doi.org/](https://doi.org/10.1155/2018/9105261) [10.1155/2018/9105261](https://doi.org/10.1155/2018/9105261).
- 7. Chakrawarti L, Agrawal R, Dang S, Gupta S, Gabrani R. Therapeutic efects of EGCG: a patent review. Expert Opin Ther Pat. 2016;26(8):907– 16.<https://doi.org/10.1080/13543776.2016.1203419>.
- 8. Talib WH, Awajan D, Alqudah A, Alsawwaf R, Althunibat R, Abu AlRoos M, et al. Targeting cancer hallmarks with Epigallocatechin Gallate (EGCG): mechanistic basis and therapeutic targets. Molecules. 2024;29(6):1373. [https://doi.org/10.3390/molecules29061373.](https://doi.org/10.3390/molecules29061373)
- 9. Siriphap A, Kiddee A, Duangjai A, Yosboonruang A, Pook-In G, Saokaew S, et al. Antimicrobial activity of the green tea polyphenol (-)-epigallocatechin-3-gallate (EGCG) against clinical isolates of multidrug-resistant Vibrio cholerae. Antibiotics. 2022;11(4):518. [https://doi.org/10.3390/](https://doi.org/10.3390/antibiotics11040518) [antibiotics11040518.](https://doi.org/10.3390/antibiotics11040518)
- 10. Singh NA, Mandal AKA, Khan ZA. Potential neuroprotective properties of epigallocatechin-3-gallate (EGCG). Nutr J. 2015;15:1–17. [https://doi.](https://doi.org/10.1186/s12937-016-0179-4) [org/10.1186/s12937-016-0179-4](https://doi.org/10.1186/s12937-016-0179-4).
- 11. Cascella M, Bimonte S, Muzio MR, Schiavone V, Cuomo A. The efficacy of Epigallocatechin-3-gallate (green tea) in the treatment of Alzheimer's disease: An overview of pre-clinical studies and translational perspectives in clinical practice. Infect Agents Cancer. 2017;12(1):1–7. [https://](https://doi.org/10.1186/s13027-017-0145-6) [doi.org/10.1186/s13027-017-0145-6.](https://doi.org/10.1186/s13027-017-0145-6)
- 12. Chowdhury A, Sarkar J, Chakraborti T, Pramanik PK, Chakraborti S. Protective role of epigallocatechin-3-gallate in health and disease: a perspective. Biomed Pharmacother. 2016;78:50–9. [https://doi.org/10.](https://doi.org/10.1016/j.biopha.2015.12.013) [1016/j.biopha.2015.12.013](https://doi.org/10.1016/j.biopha.2015.12.013).
- 13. Zhang S, Zhu Q, Chen J-Y, OuYang D, Lu J-H. The pharmacological activity of epigallocatechin-3-gallate (EGCG) on Alzheimer's disease animal model: a systematic review. Phytomedicine. 2020;79:153316. [https://](https://doi.org/10.1016/j.phymed.2020.153316) doi.org/10.1016/j.phymed.2020.153316.
- 14. Singh R, Akhtar N, Haqqi TM. Green tea polyphenol epigallocatechi3 gallate: Infammation and arthritis. Life Sci. 2010;86(25–26):907–18. [https://doi.org/10.1016/j.lfs.2010.04.013.](https://doi.org/10.1016/j.lfs.2010.04.013)
- 15. Khalatbary AR, Khademi E. The green tea polyphenolic catechin epigallocatechin gallate and neuroprotection. Nutr Neurosci. 2020;23(4):281– 94. [https://doi.org/10.1080/1028415X.2018.1500124.](https://doi.org/10.1080/1028415X.2018.1500124)
- 16. Youn K, Ho C-T, Jun M. Multifaceted neuroprotective effects of (-)-epigallocatechin-3-gallate (EGCG) in Alzheimer's disease: An overview of pre-clinical studies focused on β-amyloid peptide. Food Sci Human Wellness. 2022;11(3):483–93. [https://doi.org/10.1016/j.fshw.2021.12.006.](https://doi.org/10.1016/j.fshw.2021.12.006)
- 17. Nasb M, Li F, Dayoub L, Wu T, Wei M, Chen N. Bridging the gap: Integrating exercise mimicry into chronic disease management through suppressing chronic infammation. J Adv Res. 2024;3(24):00176. [https://](https://doi.org/10.1016/j.jare.2024.04.034) doi.org/10.1016/j.jare.2024.04.034.
- 18. Wang Y, Ge S, Ahammed GJ, Gao H, Shen K, Wang Q, et al. Epigallocatechin-3-gallate-induced tolerance to cadmium stress involves increased flavonoid synthesis and nutrient homeostasis in tomato roots. Plant Physiol Biochem. 2024;208:108468. [https://doi.org/10.1016/j.plaphy.](https://doi.org/10.1016/j.plaphy.2024.108468) [2024.108468](https://doi.org/10.1016/j.plaphy.2024.108468). S0981-9428(24)00136-0.
- 19. Xu J, Xu S, Luo J, Zhang S, Wu D, Yang Q, et al. Epigallocatechin-3-gallate alleviates ethanol-induced endothelia cells injury partly through alteration of NF-kappaB translocation and activation of the Nrf2 signaling pathway. Biol Pharm Bull. 2024;47(7):1248–54. [https://doi.org/10.1248/](https://doi.org/10.1248/bpb.b23-00773) [bpb.b23-00773.](https://doi.org/10.1248/bpb.b23-00773)
- 20. Zhang Q, Fei X, Li Y, Zhang H, Chen L, Ruan J, et al. Epigallocatechin-3-gallate attenuates fuoride induced apoptosis via PI3K/FoxO1 pathway in ameloblast-like cells. Toxicon. 2024;247:107857. [https://doi.](https://doi.org/10.1016/j.toxicon.2024.107857) [org/10.1016/j.toxicon.2024.107857](https://doi.org/10.1016/j.toxicon.2024.107857). S0041-0101(24)00429-X.
- 21. Zhang Y, Wang Q, Zhu F. Epigallocatechin-3-gallate attenuates the sulfamethoxazole-induced immunotoxicity and reduces SMZ residues in Procambarus clarkia. J Hazard Mater 2024 472;134602.[https://doi.org/](https://doi.org/10.1016/j.jhazmat.2024.134602) [10.1016/j.jhazmat.2024.134602](https://doi.org/10.1016/j.jhazmat.2024.134602) S0304-3894(24)01181-6
- 22. Kim E, Hwang K, Lee J, Han SY, Kim E-M, Park J, et al. Skin protective efect of epigallocatechin gallate. Int J Mol Sci. 2018;19(1):173. [https://](https://doi.org/10.3390/ijms19010173) doi.org/10.3390/ijms19010173.
- 23. Wolfe KL, Liu RH. Cellular antioxidant activity (CAA) assay for assessing antioxidants, foods, and dietary supplements. J Agric Food Chem. 2007;55(22):8896–907.<https://doi.org/10.1021/jf0715166>.
- 24. Vishnoi H, Bodla RB, Kant R, Bodla R. Green Tea (Camellia sinensis) and its antioxidant property: A review. International Journal of Pharmaceutical Sciences and Research. 2018 9(5);1723–36. [https://doi.org/10.13040/](https://doi.org/10.13040/IJPSR.0975-8232) [IJPSR.0975-8232](https://doi.org/10.13040/IJPSR.0975-8232)
- 25. Nobari H, Saedmocheshi S, Chung LH, Suzuki K, Maynar-Mariño M, Pérez-Gómez J. An overview on how exercise with green tea consumption can prevent the production of reactive oxygen species and improve sports performance. Int J Environ Res Public Health. 2021;19(1):218. [https://doi.org/10.3390/ijerph19010218.](https://doi.org/10.3390/ijerph19010218)
- 26. Alam M, Shamsi A, Hassan MI. 11 Biological roles and mechanism of phytochemicals in disease prevention and treatment. 2023. [https://doi.](https://doi.org/10.1201/b22842-11) [org/10.1201/b22842-11](https://doi.org/10.1201/b22842-11)
- 27. Dube A, Ng K, Nicolazzo JA, Larson I. Efective use of reducing agents and nanoparticle encapsulation in stabilizing catechins in alkaline solution. Food Chem. 2010;122(3):662–7. [https://doi.org/10.1016/j.foodc](https://doi.org/10.1016/j.foodchem.2010.03.027) [hem.2010.03.027](https://doi.org/10.1016/j.foodchem.2010.03.027).
- 28. Nagle DG, Ferreira D, Zhou Y-D. Epigallocatechin-3-gallate (EGCG): chemical and biomedical perspectives. Phytochemistry. 2006;67(17):1849–55. <https://doi.org/10.1016/j.phytochem.2006.06.020>.
- 29. Kochman J, Jakubczyk K, Antoniewicz J, Mruk H, Janda K. Health benefts and chemical composition of matcha green tea: a review. Molecules. 2020;26(1):85.<https://doi.org/10.3390/molecules26010085>.
- 30. Yuskavage JK. Epigallocatechin gallate in the regulation of insulin secretion: Virginia Tech; 2008. <http://hdl.handle.net/10919/32761>
- 31. Mokra D, Joskova M, Mokry J. Therapeutic effects of green tea polyphenol (-)-Epigallocatechin-3-Gallate (EGCG) in relation to molecular pathways controlling infammation, oxidative stress, and apoptosis. Int J Mol Sci. 2022;24(1):340.<https://doi.org/10.1039/C3RA45933K>.
- 32. Makarewicz M, Drożdż I, Tarko T, Duda-Chodak A. The interactions between polyphenols and microorganisms, especially gut microbiota. Antioxidants. 2021;10(2):188.<https://doi.org/10.3390/antiox10020188>.
- 33. Mokra D, Adamcakova J, Mokry J. Green tea polyphenol (-)-epigallocatechin-3-gallate (EGCG): a time for a new player in the treatment of respiratory diseases? Antioxidants. 2022;11(8):1566. [https://doi.org/10.](https://doi.org/10.3390/antiox11081566) [3390/antiox11081566](https://doi.org/10.3390/antiox11081566).
- 34. Kitichalermkiat A, Katsuki M, Sato J, Sonoda T, Masuda Y, Honjoh K-i, et al. Efect of epigallocatechin gallate on gene expression of Staphylococcus aureus. J Glob Antimicrob Resist. 2020;22:854–9. [https://doi.org/](https://doi.org/10.1016/j.jgar.2020.06.006) [10.1016/j.jgar.2020.06.006.](https://doi.org/10.1016/j.jgar.2020.06.006)
- 35. Steinmann J, Buer J, Pietschmann T, Steinmann E. Anti-infective properties of epigallocatechin-3-gallate (EGCG), a component of green tea. Br J Pharmacol. 2013;168(5):1059–73. <https://doi.org/10.1111/bph.12009>.
- 36. Ali A, Parisi A, Normanno G. Polyphenols as emerging antimicrobial agents. Emerging Modalities in Mitigation of Antimicrobial Resistance. Springer; 2022. p. 219–59. [https://doi.org/10.1007/978-3-030-84126-3_](https://doi.org/10.1007/978-3-030-84126-3_10) [10.](https://doi.org/10.1007/978-3-030-84126-3_10)
- 37. Gemma S, Brogi S, Patil PR, Giovani S, Lamponi S, Cappelli A, et al. From (+)-epigallocatechin gallate to a simplifed synthetic analogue as a cytoadherence inhibitor for P. falciparum. Rsc Adv. 2014;4(9):4769–81. <https://doi.org/10.1039/C3RA45933K>.
- 38. Serra DO, Mika F, Richter AM, Hengge R. The green tea polyphenol EGCG inhibits E. coli bioflm formation by impairing amyloid curli fbre assembly and downregulating the bioflm regulator CsgD via the σE-dependent sRNA RybB. Mol Microbiol. 2016;101(1):136–51. [https://](https://doi.org/10.1111/mmi.13379) doi.org/10.1111/mmi.13379.
- 39. Gao T, Ye F, Tan Y, Peng M, Yuan F, Liu Z, et al. Metabolomics and proteomics analyses revealed mechanistic insights on the antimicrobial activity of epigallocatechin gallate against Streptococcus suis. Front Cell Infect Microbiol. 2022;12:973282. [https://doi.org/10.3389/fcimb.](https://doi.org/10.3389/fcimb.2022.973282) [2022.973282](https://doi.org/10.3389/fcimb.2022.973282).
- 40. Lee S-B, Choi E-H, Jeong K-H, Kim K-S, Shim S-M, Kim G-H. Efect of catechins and high-temperature-processed green tea extract on scavenging reactive oxygen species and preventing Aβ1–42 fbrils' formation in brain microvascular endothelium. Nutr Neurosci. 2020;23(5):363–73. [https://doi.org/10.1080/1028415X.2018.1507618.](https://doi.org/10.1080/1028415X.2018.1507618)
- 41. Cano A, Ettcheto M, Chang J-H, Barroso E, Espina M, Kühne BA, et al. Dual-drug loaded nanoparticles of Epigallocatechin-3-gallate (EGCG)/ Ascorbic acid enhance therapeutic efficacy of EGCG in a APPswe/ PS1dE9 Alzheimer's disease mice model. J Contr Rel. 2019;301:62–75. <https://doi.org/10.1016/j.jconrel.2019.03.010>.
- 42. Singh AP, Biswas A, Shukla A, Maiti P. Targeted therapy in chronic diseases using nanomaterial-based drug delivery vehicles. Signal Transduct Target Ther. 2019;4(1):33. [https://doi.org/10.1038/](https://doi.org/10.1038/s41392-019-0068-3) [s41392-019-0068-3](https://doi.org/10.1038/s41392-019-0068-3)
- 43. Cano A, Ettcheto M, Chang JH, Barroso E, Espina M, Kuhne BA et al. Dual-drug loaded nanoparticles of Epigallocatechin-3-gallate (EGCG)/Ascorbic acid enhance therapeutic efficacy of EGCG in a APPswe/PS1dE9 Alzheimer's disease mice model. J Control Release. 2019;301:62–75<https://doi.org/10.1016/j.jconrel.2019.03.010> S0168-3659(19)30157-9
- Zheng M, Pan M, Zhang W, Lin H, Wu S, Lu C, et al. Poly (α-l-lysine)based nanomaterials for versatile biomedical applications: Current

advances and perspectives. Bioact Mat. 2021;6(7):1878–909. [https://doi.](https://doi.org/10.1016/j.bioactmat.2020.12.001) [org/10.1016/j.bioactmat.2020.12.001](https://doi.org/10.1016/j.bioactmat.2020.12.001).

- 45. Alam M, Kashyap T, Mishra P, Panda AK, Nagini S, Mishra R. Role and regulation of proapoptotic Bax in oral squamous cell carcinoma and drug resistance. Head Neck. 2019;41(1):185–97. [https://doi.org/10.1002/](https://doi.org/10.1002/hed.25471) [hed.25471](https://doi.org/10.1002/hed.25471).
- 46. Alam M, Ali S, Mohammad T, Hasan GM, Yadav DK, Hassan MI. B cell lymphoma 2: a potential therapeutic target for cancer therapy. Int J Mol Sci. 2021;22(19):10442. [https://doi.org/10.3390/ijms221910442.](https://doi.org/10.3390/ijms221910442)
- 47. Almatroodi SA, Almatroudi A, Khan AA, Alhumaydhi FA, Alsahli MA, Rahmani AH. Potential therapeutic targets of epigallocatechin gallate (EGCG), the most abundant catechin in green tea, and its role in the therapy of various types of cancer. Molecules. 2020;25(14):3146. [https://](https://doi.org/10.3390/molecules25143146) [doi.org/10.3390/molecules25143146.](https://doi.org/10.3390/molecules25143146)
- 48. Wang Y, Wu S, Li Q, Lang W, Li W, Jiang X, et al. Epigallocatechin-3-gallate: a phytochemical as a promising drug candidate for the treatment of Parkinson's disease. Front Pharmacol. 2022;13:977521. [https://doi.](https://doi.org/10.3389/fphar.2022.977521) [org/10.3389/fphar.2022.977521.](https://doi.org/10.3389/fphar.2022.977521)
- 49. Gonçalves PB, Sodero ACR, Cordeiro Y. Green tea epigallocatechin-3-gallate (EGCG) targeting protein misfolding in drug discovery for neurodegenerative diseases. Biomolecules. 2021;11(5):767. [https://doi.](https://doi.org/10.3390/biom11050767) [org/10.3390/biom11050767](https://doi.org/10.3390/biom11050767).
- 50. Bakun P, Mlynarczyk DT, Koczorowski T, Cerbin-Koczorowska M, Piwowarczyk L, Kolasiński E, et al. Tea-break with epigallocatechin gallate derivatives - powerful polyphenols of great potential for medicine. Eur J Med Chem. 2023;261(115820):14. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.ejmech.2023.115820) [ejmech.2023.115820.](https://doi.org/10.1016/j.ejmech.2023.115820)
- 51. Zhang L, Liu W, You H, Chen Z, Xu L, He H. Assessing the analgesic efficacy of oral epigallocatechin-3-gallate on epidural catheter analgesia in patients after surgical stabilisation of multiple rib fractures: a prospective double-blind, placebo-controlled clinical trial. Pharm Biol. 2020;58(1):741–4. [https://doi.org/10.1080/13880209.2020.1797123.](https://doi.org/10.1080/13880209.2020.1797123)
- 52. Chu C, Deng J, Man Y, Qu Y. Green tea extracts epigallocatechin-3-gallate for diferent treatments. BioMed Res Int. 2017;2017:5615647. [https://doi.org/10.1155/2017/5615647.](https://doi.org/10.1155/2017/5615647)
- Alam M, Naqvi AAT, Hassan MI. Emerging Role of Structural and Systems Biology in Anticancer Therapeutics. Systems Biomedicine Approaches in Cancer Research. Springer; 2022. p. 97–114. [https://doi.org/10.1007/](https://doi.org/10.1007/978-981-19-1953-4_5) [978-981-19-1953-4_5](https://doi.org/10.1007/978-981-19-1953-4_5)
- 54. Lecumberri E, Dupertuis YM, Miralbell R, Pichard C. Green tea polyphenol epigallocatechin-3-gallate (EGCG) as adjuvant in cancer therapy. Clin Nutr. 2013;32(6):894–903. [https://doi.org/10.1016/j.clnu.2013.03.](https://doi.org/10.1016/j.clnu.2013.03.008) [008](https://doi.org/10.1016/j.clnu.2013.03.008).
- 55. Alam M, Ali S, Ahmed S, Elasbali AM, Adnan M, Islam A, et al. Therapeutic potential of ursolic acid in cancer and diabetic neuropathy diseases. Int J Mol Sci. 2021;22(22):12162. [https://doi.org/10.3390/ijms222212](https://doi.org/10.3390/ijms222212162) [162](https://doi.org/10.3390/ijms222212162).
- 56. Ali S, Alam M, Hassan MI. Kinase inhibitors: An overview. Protein Kinase Inhibitors. 2022:1–22. [https://doi.org/10.1016/B978-0-323-91287-7.](https://doi.org/10.1016/B978-0-323-91287-7.00026-0) [00026-0](https://doi.org/10.1016/B978-0-323-91287-7.00026-0)
- 57. Ono K, Yoshiike Y, Takashima A, Hasegawa K, Naiki H, Yamada M. Potent anti-amyloidogenic and fbril-destabilizing efects of polyphenols in vitro: implications for the prevention and therapeutics of Alzheimer's disease. J Neurochem. 2003;87(1):172–81. [https://doi.org/10.1046/j.](https://doi.org/10.1046/j.1471-4159.2003.01976.x) [1471-4159.2003.01976.x.](https://doi.org/10.1046/j.1471-4159.2003.01976.x)
- 58. Huang Y-J, Wang K-L, Chen H-Y, Chiang Y-F, Hsia S-M. Protective efects of epigallocatechin gallate (EGCG) on endometrial, breast, and ovarian cancers. Biomolecules. 2020;10(11):1481. [https://doi.org/10.3390/biom1](https://doi.org/10.3390/biom10111481) [0111481.](https://doi.org/10.3390/biom10111481)
- 59. Delabar JM, Gomes MAG, Fructuoso M, Sarrazin N, George N, Fleary-Roberts N, et al. EGCG-like non-competitive inhibitor of DYRK1A rescues cognitive defect in a down syndrome model. Eur J Med Chem. 2024;265:116098. [https://doi.org/10.1016/j.ejmech.2023.116098.](https://doi.org/10.1016/j.ejmech.2023.116098)
- 60. Sharma SK, Parasuraman P, Kumar G, Surolia N, Surolia A. Green tea catechins potentiate triclosan binding to enoyl-ACP reductase from Plasmodium falciparum (PfENR). J Med Chem. 2007;50(4):765–75. [https://doi.org/10.1021/jm061154d.](https://doi.org/10.1021/jm061154d)
- 61. Meng J, Chen Y, Wang J, Qiu J, Chang C, Bi F, et al. EGCG protects vascular endothelial cells from oxidative stress-induced damage by targeting the autophagy-dependent PI3K-AKT-mTOR pathway. Ann Transl Med. 2020;8(5):200. <https://doi.org/10.21037/atm.2020.01.92>.
- 62. Liu H, Wang L, Li F, Jiang Y, Guan H, Wang D, et al. The synergistic protection of EGCG and quercetin against streptozotocin (STZ)-induced NIT-1 pancreatic β cell damage via upregulation of BCL-2 expression by miR-16-5p. J Nutr Biochem. 2021;96:108748. [https://doi.org/10.](https://doi.org/10.1016/j.jnutbio.2021.108748) [1016/j.jnutbio.2021.108748.](https://doi.org/10.1016/j.jnutbio.2021.108748)
- 63. Liu T, Shi W, Ding Y, Wu Q, Zhang B, Zhang N, et al. (−)-Epigallocatechin gallate is a noncompetitive inhibitor of NAD kinase. ACS Med Chem Lett. 2022;13(11):1699–706. [https://doi.org/10.1021/acsme](https://doi.org/10.1021/acsmedchemlett.2c00163) [dchemlett.2c00163.](https://doi.org/10.1021/acsmedchemlett.2c00163)
- 64. Guo C, Huang Q, Wang Y, Yao Y, Li J, Chen J, et al. Therapeutic application of natural products: NAD+ metabolism as potential target. Phytomedicine. 2023;114:154768. [https://doi.org/10.1016/j.phymed.](https://doi.org/10.1016/j.phymed.2023.154768) [2023.154768](https://doi.org/10.1016/j.phymed.2023.154768).
- 65. Sharma C, Sadek B, Goyal SN, Sinha S, Kamal MA, Ojha S. Small molecules from nature targeting G-protein coupled cannabinoid receptors: potential leads for drug discovery and development. Evid-Based Complement Altern Med. 2015;2015(1):238482. [https://doi.org/10.](https://doi.org/10.1155/2015/238482) [1155/2015/238482](https://doi.org/10.1155/2015/238482).
- 66. Cunningham CW. Plant-based modulators of endocannabinoid signaling. J Nat Prod. 2019;82(3):636–46. [https://doi.org/10.1021/acs.](https://doi.org/10.1021/acs.jnatprod.8b00874) [jnatprod.8b00874](https://doi.org/10.1021/acs.jnatprod.8b00874).
- 67. Abdel-Hamid NM, Abass SA. Matrix metalloproteinase contribution in management of cancer proliferation, metastasis and drug targeting. Mol Biol Rep. 2021;48(9):6525–38. [https://doi.org/10.1007/](https://doi.org/10.1007/s11033-021-06635-z) [s11033-021-06635-z](https://doi.org/10.1007/s11033-021-06635-z).
- 68. Wang W, Zhang Q, Xiong X, Zheng Y, Yang W, Du L. Investigation on the infuence of galloyl moiety to the peptidyl prolyl cis/trans isomerase Pin1: a spectral and computational analysis. J Mol Liq. 2020;316:113870. [https://doi.org/10.1016/j.molliq.2020.113870.](https://doi.org/10.1016/j.molliq.2020.113870)
- 69. Zueva IV, Vasilieva EA, Gaynanova GA, Moiseenko AV, Burtseva AD, Boyko KM, et al. Can activation of acetylcholinesterase by β-Amyloid peptide decrease the efectiveness of cholinesterase inhibitors? Int J Mol Sci. 2023;24(22):16395. [https://doi.org/10.3390/ijms242216395.](https://doi.org/10.3390/ijms242216395)
- 70. Zhu S, Li Y, Li Z, Ma C, Lou Z, Yokoyama W, et al. Lipase-catalyzed synthesis of acetylated EGCG and antioxidant properties of the acetylated derivatives. Food Res Int. 2014;56:279–86. [https://doi.org/](https://doi.org/10.1016/j.foodres.2013.10.026) [10.1016/j.foodres.2013.10.026](https://doi.org/10.1016/j.foodres.2013.10.026).
- 71. El-Mowafy A, Salem H, Al-Gayyar M, El-Mesery M, El-Azab M. Evaluation of renal protective effects of the green-tea (EGCG) and red grape resveratrol: role of oxidative stress and infammatory cytokines. Nat Prod Res. 2011;25(8):850–6. [https://doi.org/10.1080/14786419.2010.](https://doi.org/10.1080/14786419.2010.533669) [533669](https://doi.org/10.1080/14786419.2010.533669).
- 72. Chen W-C, Hsieh S-R, Chiu C-H, Hsu B-D, Liou Y-M. Molecular identifcation for epigallocatechin-3-gallate-mediated antioxidant intervention on the H 2 O 2-induced oxidative stress in H9c2 rat cardiomyoblasts. J Biomed Sci. 2014;21(1):1–12. [https://doi.org/10.](https://doi.org/10.1186/1423-0127-21-56) [1186/1423-0127-21-56](https://doi.org/10.1186/1423-0127-21-56).
- 73. Gao Z, Han Y, Hu Y, Wu X, Wang Y, Zhang X, et al. Targeting HO-1 by epigallocatechin-3-gallate reduces contrast-induced renal injury via anti-oxidative stress and anti-infammation pathways. PLoS ONE. 2016;11(2):e0149032. <https://doi.org/10.1371/journal.pone.0149032>.
- 74. Simos YV, Verginadis II, Toliopoulos IK, Velalopoulou AP, Karagounis IV, Karkabounas SC, et al. Effects of catechin and epicatechin on superoxide dismutase and glutathione peroxidase activity, in vivo. Redox Rep. 2012;17(5):181–6. <https://doi.org/10.1179/1351000212Y.0000000020>.
- 75. Huang W, Zhang M, Qiu Q, Zhang J, Hua C, Chen G, et al. Metabolomics of human umbilical vein endothelial cell-based analysis of the relationship between hyperuricemia and dyslipidemia. Nutr Metab Cardiovasc Dis. 2024;34(6):1528–37.<https://doi.org/10.1016/j.numecd.2024.02.001>.
- 76. Xie H, Sun J, Chen Y, Zong M, Li S, Wang Y. EGCG attenuates uric acidinduced infammatory and oxidative stress responses by medicating the NOTCH pathway. Oxid Med Cell Longev. 2015;2015(1):214836. <https://doi.org/10.1155/2015/214836>.
- 77. Cai J, Qiao Y, Chen L, Lu Y, Zheng D. Regulation of the Notch signaling pathway by natural products for cancer therapy. J Nutr Biochem. 2023;123:109483. [https://doi.org/10.1016/j.jnutbio.2023.109483.](https://doi.org/10.1016/j.jnutbio.2023.109483)
- 78. Farooqi AA, Pinheiro M, Granja A, Farabegoli F, Reis S, Attar R, et al. EGCG mediated targeting of deregulated signaling pathways and non-coding RNAs in diferent cancers: focus on JAK/STAT, Wnt/β-Catenin, TGF/ SMAD, NOTCH, SHH/GLI, and TRAIL mediated signaling pathways. Cancers. 2020;12(4):951.<https://doi.org/10.3390/cancers12040951>.
- 79. Kiesel VA, Stan SD. Modulation of notch signaling pathway by bioactive dietary agents. Int J Mol Sci. 2022;23(7):3532. [https://doi.org/10.3390/](https://doi.org/10.3390/ijms23073532) ijms23073532
- 80. Gadapa S, Battula SN, Mor S, Pushpadass HA, Naik LN, Emerald ME. Green tea catechin loaded niosomes: Formulation and their characterization for food fortifcation. J Food Sci Technol. 2022;59(9):3669–82. [https://doi.org/10.1007/s13197-022-05384-6.](https://doi.org/10.1007/s13197-022-05384-6)
- 81. Zhang Y, Liu E, Gao H, He Q, Chen A, Pang Y, et al. Natural products for the treatment of hypertrophic scars: preclinical and clinical studies. Heliyon. 2024;10(17):e37059. [https://doi.org/10.1016/j.heliyon.2024.](https://doi.org/10.1016/j.heliyon.2024.e37059) [e37059](https://doi.org/10.1016/j.heliyon.2024.e37059).
- 82. Garcia Garcia JM, Vannuzzi V, Donati C, Bernacchioni C, Bruni P, Petraglia F. Endometriosis: cellular and molecular mechanisms leading to fbrosis. Reprod Sci. 2023;30(5):1453–61. [https://doi.org/10.1007/](https://doi.org/10.1007/s43032-022-01083-x) [s43032-022-01083-x](https://doi.org/10.1007/s43032-022-01083-x).
- 83. Zhang B, Wang B, Cao S, Wang Y. Epigallocatechin-3-gallate (EGCG) attenuates traumatic brain injury by inhibition of edema formation and oxidative stress. Kor J Physiol Pharmacol. 2015;19(6):491–7. [https://doi.](https://doi.org/10.1201/9781003402374-70) [org/10.1201/9781003402374-70](https://doi.org/10.1201/9781003402374-70).
- 84. Abo-Salem OM, Ali TM, Harisa GI, Mehanna OM, Younos IH, Almalki WH. Beneficial effects of (-)-epigallocatechin-3-O-gallate on diabetic peripheral neuropathy in the rat model. J Biochem Mol Toxicol. 2020;34(8):e22508. [https://doi.org/10.1002/jbt.22508.](https://doi.org/10.1002/jbt.22508)
- 85. Arafa MH, Atteia HH. Protective role of epigallocatechin gallate in a rat model of cisplatin-induced cerebral infammation and oxidative damage: impact of modulating NF-κB and Nrf2. Neurotox Res. 2020;37(2):380–96. [https://doi.org/10.1007/s12640-019-00095-x.](https://doi.org/10.1007/s12640-019-00095-x)
- 86. Payne A, Nahashon S, Taka E, Adinew GM, Soliman KF. Epigallocatechin-3-Gallate (EGCG): New therapeutic perspectives for neuroprotection, aging, and neuroinfammation for the modern age. Biomolecules. 2022;12(3):371.<https://doi.org/10.3390/biom12030371>.
- 87. Sang S, Lambert JD, Ho C-T, Yang CS. The chemistry and biotransformation of tea constituents. Pharmacol Res. 2011;64(2):87–99. [https://doi.](https://doi.org/10.1016/j.phrs.2011.02.007) [org/10.1016/j.phrs.2011.02.007.](https://doi.org/10.1016/j.phrs.2011.02.007)
- 88. Jude S, Gopi S. Multitarget approach for natural products in infammation. Infammation and natural products. Elsevier; 2021. p. 39–67. <https://doi.org/10.1016/j.drudis.2014.08.006>
- 89. Sah DK, Khoi PN, Li S, Arjunan A, Jeong J-U, Jung YD. (-)-Epigallocatechin-3-gallate prevents IL-1β-Induced uPAR expression and invasiveness via the suppression of NF-κB and AP-1 in human bladder cancer cells. Int J Mol Sci. 2022;23(22):14008. [https://doi.org/10.3390/ijms2](https://doi.org/10.3390/ijms232214008) [32214008](https://doi.org/10.3390/ijms232214008).
- 90. Pesapane A, Di Giovanni C, Rossi FW, Alfano D, Formisano L, Ragno P, et al. Discovery of new small molecules inhibiting 67 kDa laminin receptor interaction with laminin and cancer cell invasion. Oncotarget. 2015;6(20):18116. <https://doi.org/10.18632/oncotarget.4016>.
- 91. Li Y, Li D, Chen J, Wang S. A polysaccharide from Pinellia ternata inhibits cell proliferation and metastasis in human cholangiocarcinoma cells by targeting of Cdc42 and 67 kDa Laminin Receptor (LR). Int J Biol Macromol. 2016;93:520–5. [https://doi.org/10.1016/j.ijbiomac.2016.08.069.](https://doi.org/10.1016/j.ijbiomac.2016.08.069)
- 92. Farkhondeh T, Pourbagher-Shahri AM, Ashrafzadeh M, Folgado SL, Rajabpour-Sanati A, Khazdair MR, et al. Green tea catechins inhibit microglial activation which prevents the development of neurological disorders. Neural Regen Res. 2020;15(10):1792–8. [https://doi.org/10.](https://doi.org/10.4103/1673-5374.280300) [4103/1673-5374.280300](https://doi.org/10.4103/1673-5374.280300).
- 93. Fujimura Y, Sumida M, Sugihara K, Tsukamoto S, Yamada K, Tachibana H. Green tea polyphenol EGCG sensing motif on the 67-kDa laminin receptor. PLoS ONE. 2012;7(5):e37942. [https://doi.org/10.1371/journal.](https://doi.org/10.1371/journal.pone.0037942) [pone.0037942.](https://doi.org/10.1371/journal.pone.0037942)
- Yu H-N, Zhang L-C, Yang J-G, Das UN, Shen S-R. Effect of laminin tyrosine–isoleucine–glycine–serine–arginine peptide on the growth of human prostate cancer (PC-3) cells in vitro. Eur J Pharmacol. 2009;616(1):251–5.<https://doi.org/10.1371/journal.pone.0037942>.
- 95. Fujimura Y, Yamada K, Tachibana H. A lipid raft-associated 67 kDa laminin receptor mediates suppressive effect of epigallocatechin-3-Ogallate on FcεRI expression. Biochem Biophys Res Commun. 2005;336(2):674–81.<https://doi.org/10.1016/j.bbrc.2005.08.146>.
- 96. Jha A, Alam M, Kashyap T, Nath N, Kumari A, Pramanik KK, et al. Crosstalk between PD-L1 and Jak2-Stat3/MAPK-AP1 signaling promotes oral cancer progression, invasion and therapy resistance. Int Immunopharmacol. 2023;124:110894. <https://doi.org/10.1016/j.intimp.2023.110894>.
- 97. Zhang Z-X, Li Y-B, Zhao R-P. Epigallocatechin gallate attenuates β-amyloid generation and oxidative stress involvement of PPARγ in N2a/APP695 cells. Neurochem Res. 2017;42:468–80. [https://doi.org/](https://doi.org/10.1007/s11064-016-2093-8) [10.1007/s11064-016-2093-8.](https://doi.org/10.1007/s11064-016-2093-8)
- 98. Zhou H, Fu L-X, Li L, Chen Y-Y, Zhu H-Q, Zhou J-L, et al. The epigallocatechin gallate derivative Y6 reduces the cardiotoxicity and enhances the efficacy of daunorubicin against human hepatocellular carcinoma by inhibiting carbonyl reductase 1 expression. J Ethnopharmacol. 2020;261:113118. [https://doi.org/10.1016/j.jep.2020.](https://doi.org/10.1016/j.jep.2020.113118) [113118.](https://doi.org/10.1016/j.jep.2020.113118)
- 99. Zan L, Chen Q, Zhang L, Li X. Epigallocatechin gallate (EGCG) suppresses growth and tumorigenicity in breast cancer cells by downregulation of miR-25. Bioengineered. 2019;10(1):374–82. [https://doi.](https://doi.org/10.1080/21655979.2019.1657327) [org/10.1080/21655979.2019.1657327.](https://doi.org/10.1080/21655979.2019.1657327)
- 100. Wang L, Li P, Feng K. EGCG adjuvant chemotherapy: Current status and future perspectives. Eur J Med Chem. 2023;250:115197. [https://](https://doi.org/10.1016/j.ejmech.2023.115197) doi.org/10.1016/j.ejmech.2023.115197.
- 101. Yen C, Zhao F, Yu Z, Zhu X, Li CG. Interactions between natural products and tamoxifen in breast cancer: a comprehensive literature review. Front Pharmacol. 2022;13:847113. [https://doi.org/10.3389/](https://doi.org/10.3389/fphar.2022.847113) [fphar.2022.847113.](https://doi.org/10.3389/fphar.2022.847113)
- 102. Xie L-W, Cai S, Zhao T-S, Li M, Tian Y. Green tea derivative (−)-epigallocatechin-3-gallate (EGCG) confers protection against ionizing radiation-induced intestinal epithelial cell death both in vitro and in vivo. Free Radical Biol Med. 2020;161:175–86. [https://doi.org/10.](https://doi.org/10.1016/j.freeradbiomed.2020.10.012) [1016/j.freeradbiomed.2020.10.012](https://doi.org/10.1016/j.freeradbiomed.2020.10.012).
- 103. Zhao Y, Hu X, Zuo X, Wang M. Chemopreventive efects of some popular phytochemicals on human colon cancer: a review. Food Funct. 2018;9(9):4548–68.<https://doi.org/10.1039/C8FO00850G>.
- 104. Singla RK, Behzad S, Khan J, Tsagkaris C, Gautam RK, Goyal R, et al. Natural kinase inhibitors for the treatment and management of endometrial/uterine cancer: preclinical to clinical studies. Front Pharmacol. 2022;13:801733.<https://doi.org/10.3389/fphar.2022.801733>.
- 105. Phosrithong N, Ungwitayatorn J. Molecular docking study on anticancer activity of plant-derived natural products. Med Chem Res. 2010;19:817–35. [https://doi.org/10.1007/s00044-009-9233-5.](https://doi.org/10.1007/s00044-009-9233-5)
- 106. Errachid A, Nohawica M, Wyganowska-Swiatkowska M. A comprehensive review of the infuence of Epigallocatechin gallate on Sjögren's syndrome associated molecular regulators of exocytosis. Biomed Rep. 2021;15(5):95. <https://doi.org/10.3892/br.2021.1471>.
- 107. Remely M, Ferk F, Sterneder S, Setayesh T, Roth S, Kepcija T, et al. EGCG prevents high fat diet-induced changes in gut microbiota, decreases of DNA strand breaks, and changes in expression and DNA methylation of Dnmt1 and MLH1 in C57BL/6J male mice. Oxid Med Cell Longev. 2017;2017(1):3079148. [https://doi.org/10.1155/2017/](https://doi.org/10.1155/2017/3079148) [3079148.](https://doi.org/10.1155/2017/3079148)
- 108. Dharshini LCP, Mandal AKA. Regulation of gene expression by modulating microRNAs through Epigallocatechin-3-gallate in cancer. Mol Biol Rep. 2024;51(1):023–09145. [https://doi.org/10.1007/](https://doi.org/10.1007/s11033-023-09145-2) [s11033-023-09145-2](https://doi.org/10.1007/s11033-023-09145-2).
- 109. Li Y, Zhao Y, Han J, Wang Y, Lei S. Efects of epigallocatechin gallate (EGCG) on the biological properties of human dental pulp stem cells and infammatory pulp tissue. Arch Oral Biol. 2021;123:105034. [https://](https://doi.org/10.1016/j.archoralbio.2020.105034) [doi.org/10.1016/j.archoralbio.2020.105034.](https://doi.org/10.1016/j.archoralbio.2020.105034)
- 110. Demeule M, Brossard M, Pagé M, Gingras D, Béliveau R. Matrix metalloproteinase inhibition by green tea catechins. Biochim et Biophys Acta (BBA)-Protein Struct Mol Enzymol. 2000;1478(1):51–60. [https://doi.org/](https://doi.org/10.1016/S0167-4838(00)00009-1) [10.1016/S0167-4838\(00\)00009-1.](https://doi.org/10.1016/S0167-4838(00)00009-1)
- 111. Huang Q, Zhu Y, Lv L, Sang S. Translating in vitro acrolein-trapping capacities of tea polyphenol and soy genistein to in vivo situation is mediated by the bioavailability and biotransformation of individual polyphenols. Mol Nutr Food Res. 2020;64(1):1900274. [https://doi.org/](https://doi.org/10.1002/mnfr.201900274) [10.1002/mnfr.201900274](https://doi.org/10.1002/mnfr.201900274).
- 112. Kong C, Zhang H, Li L, Liu Z. Efects of green tea extract epigallocatechin-3-gallate (EGCG) on oral disease-associated microbes: a review. J Oral Microbiol. 2022;14(1):2131117. [https://doi.org/10.1080/20002297.](https://doi.org/10.1080/20002297.2022.2131117) [2022.2131117](https://doi.org/10.1080/20002297.2022.2131117).
- 113. Mosaddad SA, Tahmasebi E, Yazdanian A, Rezvani MB, Seifalian A, Yazdanian M, et al. Oral microbial bioflms: an update. Eur J Clin Microbiol Infect Dis. 2019;38:2005–19. [https://doi.org/10.1007/](https://doi.org/10.1007/s10096-019-03641-9) [s10096-019-03641-9](https://doi.org/10.1007/s10096-019-03641-9).
- 114. Vyas T, Nagi R, Bhatia A, Bains SK. Therapeutic effects of green tea as an antioxidant on oral health-a review. J Fam Med Prim Care. 2021;10(11):3998. [https://doi.org/10.4103/jfmpc.jfmpc_943_21.](https://doi.org/10.4103/jfmpc.jfmpc_943_21)
- 115. Taylor PW, Hamilton-Miller J, Stapleton PD. Antimicrobial properties of green tea catechins. Food Sci Technol Bull. 2006;2:71–8. [https://doi.org/](https://doi.org/10.1616/1476-2137.14184) [10.1616/1476-2137.14184](https://doi.org/10.1616/1476-2137.14184).
- 116. Ni D, Ai Z, Munoz-Sandoval D, Suresh R, Ellis PR, Yuqiong C, et al. Inhibition of the facilitative sugar transporters (GLUTs) by tea extracts and catechins. FASEB J. 2020;34(8):9995–10010. [https://doi.org/10.1096/f.](https://doi.org/10.1096/fj.202000057RR) [202000057RR.](https://doi.org/10.1096/fj.202000057RR)
- 117. Han JH, Kim M, Kim HJ, Jang SB, Bae S-J, Lee I-K, et al. Targeting lactate dehydrogenase a with catechin resensitizes SNU620/5FU gastric cancer cells to 5-Fluorouracil. Int J Mol Sci. 2021;22(10):5406. [https://doi.org/10.](https://doi.org/10.3390/ijms22105406) [3390/ijms22105406.](https://doi.org/10.3390/ijms22105406)
- 118. Manikandan S, Behera S, Karthikeyan R, Niranjana A, Bharathan R, Mohammed OFB. Efect of green tea extract mouthrinse and probiotic mouthrinse on salivary pH in a group of schoolchildren: an in vivo study. J Pharm Bioallied Sci. 2020;12(Suppl 1):S404. [https://doi.org/10.](https://doi.org/10.4103/jpbs.jpbs_119_20) [4103/jpbs.jpbs_119_20.](https://doi.org/10.4103/jpbs.jpbs_119_20)
- 119. Dakshinamoorthy M, Subramanian M, Padmavathi K, Mahalakshmi K, Arumugam K, Paramasivam V. Efect of probiotic chocolate in the reduction of Streptococcus Mutans count. Biomed Pharmacol J. 2016;9(3).<https://doi.org/10.13005/bpj/1051>.
- 120. Morin M-P, Bedran TBL, Fournier-Larente J, Haas B, Azelmat J, Grenier D. Green tea extract and its major constituent epigallocatechin-3-gallate inhibit growth and halitosis-related properties of Solobacterium moorei. BMC Complement Altern Med. 2015;15:1–11. [https://doi.org/](https://doi.org/10.1186/s12906-015-0557-z) [10.1186/s12906-015-0557-z.](https://doi.org/10.1186/s12906-015-0557-z)
- 121. Kwon OS, Jung JH, Shin EA, Park JE, Park WY, Kim S-H. Epigallocatechin-3-gallate induces apoptosis as a TRAIL sensitizer via activation of caspase 8 and death receptor 5 in human colon cancer cells. Biomedicines. 2020;8(4):84.<https://doi.org/10.3390/biomedicines8040084>.
- 122. Pramanik KK, Nagini S, Singh AK, Mishra P, Kashyap T, Nath N, et al. Glycogen synthase kinase-3β mediated regulation of matrix metalloproteinase-9 and its involvement in oral squamous cell carcinoma progression and invasion. Cell Oncol. 2018;41(1):47–60. [https://doi.org/](https://doi.org/10.1007/s13402-017-0358-0) [10.1007/s13402-017-0358-0](https://doi.org/10.1007/s13402-017-0358-0).
- 123. Pramanik KK, Singh AK, Alam M, Kashyap T, Mishra P, Panda AK, et al. Reversion-inducing cysteine-rich protein with Kazal motifs and its regulation by glycogen synthase kinase 3 signaling in oral cancer. Tumor Biol. 2016;37(11):15253–64. [https://doi.org/10.1007/s13277-016-5362-x.](https://doi.org/10.1007/s13277-016-5362-x)
- 124. Ho YC, Yang SF, Peng CY, Chou MY, Chang YC. Epigallocatechin-3-gallate inhibits the invasion of human oral cancer cells and decreases the productions of matrix metalloproteinases and urokinase-plasminogen activator. J Oral Pathol Med. 2007;36(10):588–93. [https://doi.org/10.](https://doi.org/10.1111/j.1600-0714.2007.00588.x) [1111/j.1600-0714.2007.00588.x.](https://doi.org/10.1111/j.1600-0714.2007.00588.x)
- 125. Koh YW, Choi EC, Kang SU, Hwang HS, Lee MH, Pyun J, et al. Green tea (−)-epigallocatechin-3-gallate inhibits HGF-induced progression in oral cavity cancer through suppression of HGF/c-Met. J Nutr Biochem. 2011;22(11):1074–83. [https://doi.org/10.1016/j.jnutbio.2010.09.005.](https://doi.org/10.1016/j.jnutbio.2010.09.005)
- 126. Xie L, Yi J, Song Y, Zhao M, Fan L, Zhao L. Suppression of GOLM1 by EGCG through HGF/HGFR/AKT/GSK-3β/β-catenin/c-Myc signaling pathway inhibits cell migration of MDA-MB-231. Food Chem Toxicol. 2021;157:112574. [https://doi.org/10.1016/j.fct.2021.112574.](https://doi.org/10.1016/j.fct.2021.112574)
- 127. Chen P-N, Chu S-C, Kuo W-H, Chou M-Y, Lin J-K, Hsieh Y-S. Epigallocatechin-3 gallate inhibits invasion, epithelial− mesenchymal transition, and tumor growth in oral cancer cells. J Agri Food Chem. 2011;59(8):3836–44.<https://doi.org/10.1021/jf1049408>.
- 128. Yuan J-M. Cancer prevention by green tea: evidence from epidemiologic studies. Am J Clin Nutr. 2013;98(6):1676S-S1681. [https://doi.org/](https://doi.org/10.3945/ajcn.113.058271) [10.3945/ajcn.113.058271](https://doi.org/10.3945/ajcn.113.058271).
- 129. Xu X, Dai Z, Zhang Z, Kou X, You X, Sun H, et al. Fabrication of oral nanovesicle in-situ gel based on Epigallocatechin gallate phospholipid complex: Application in dental anti-caries. Eur J Pharmacol. 2021;897:173951.<https://doi.org/10.1016/j.ejphar.2021.173951>.
- 130. Shinde S, Lee LH, Chu T. Inhibition of bioflm formation by the synergistic action of EGCG-S and antibiotics. Antibiotics. 2021;10(2):102. [https://](https://doi.org/10.3390/antibiotics10020102) doi.org/10.3390/antibiotics10020102.
- 131. Schneider-Rayman M, Steinberg D, Sionov RV, Friedman M, Shalish M. Efect of epigallocatechin gallate on dental bioflm of Streptococcus

mutans: an in vitro study. BMC Oral Health. 2021;21:1–11. [https://doi.](https://doi.org/10.1186/s12903-021-01798-4) [org/10.1186/s12903-021-01798-4.](https://doi.org/10.1186/s12903-021-01798-4)

- 132. Aragão MGB, He X, Aires CP, Corona SAM. Epigallocatechin gallate reduces the virulence of cariogenic Streptococcus mutans bioflm by afecting the synthesis of bioflm matrix components. Arch Oral Biol. 2024;164:105990.<https://doi.org/10.1016/j.archoralbio.2024.105990>.
- 133. Lagha AB, Grenier D. Tea polyphenols protect gingival keratinocytes against TNF-α-induced tight junction barrier dysfunction and attenuate the infammatory response of monocytes/macrophages. Cytokine. 2019;115:64–75. <https://doi.org/10.1016/j.cyto.2018.12.009>.
- 134. Messire G, Serreau R, Berteina-Raboin S. Antioxidant effects of catechins (EGCG), andrographolide, and curcuminoids compounds for skin protection, cosmetics, and dermatological uses: an update. Antioxidants. 2023;12(7):1317. [https://doi.org/10.3390/antiox12071317.](https://doi.org/10.3390/antiox12071317)
- 135. Ben Lagha A, Haas B, Grenier D. Tea polyphenols inhibit the growth and virulence properties of Fusobacterium nucleatum. Sci Rep. 2017;7(1):44815.<https://doi.org/10.1038/srep44815>.
- 136. Wang X, Liu Y, Wu Z, Zhang P, Zhang X. Tea polyphenols: a natural antioxidant regulates gut flora to protect the intestinal mucosa and prevent chronic diseases. Antioxidants. 2022;11(2):253. [https://doi.org/](https://doi.org/10.3390/antiox11020253) [10.3390/antiox11020253](https://doi.org/10.3390/antiox11020253).
- 137. Moghadam ET, Yazdanian M, Tahmasebi E, Tebyanian H, Ranjbar R, Yazdanian A, et al. Current herbal medicine as an alternative treatment in dentistry: In vitro, in vivo and clinical studies. Eur J Pharmacol. 2020;889:173665.<https://doi.org/10.1016/j.ejphar.2020.173665>.
- 138. Guo Y, Li Z, Chen F, Chai Y. Polyphenols in oral health: homeostasis maintenance, disease prevention, and therapeutic applications. Nutrients. 2023;15(20):4384.<https://doi.org/10.1016/j.ejphar.2020.173665>.
- 139. Aggarwal V, Tuli HS, Tania M, Srivastava S, Ritzer EE, Pandey A et al., editors. Molecular mechanisms of action of epigallocatechin gallate in cancer: Recent trends and advancement. Seminars in cancer biology; 2022: Elsevier. [https://doi.org/10.1016/j.semcancer.2020.05.011.](https://doi.org/10.1016/j.semcancer.2020.05.011)
- 140. da Silva-Júnior EF, Silva LR. Multi-target approaches of Epigallocatechin-3-O-gallate (EGCG) and its derivatives against Infuenza Viruses. Curr Top Med Chem. 2022;22(18):1485–500. [https://doi.org/10.2174/](https://doi.org/10.2174/1568026622666220127112056) [1568026622666220127112056.](https://doi.org/10.2174/1568026622666220127112056)
- 141. Tang G, Xu Y, Zhang C, Wang N, Li H, Feng Y. Green Tea and Epigallocatechin Gallate (EGCG) for the management of Nonalcoholic Fatty Liver Diseases (NAFLD): insights into the role of oxidative stress and antioxidant mechanism. Antioxidants. 2021;10(7):1076. [https://doi.org/](https://doi.org/10.3390/antiox10071076) [10.3390/antiox10071076](https://doi.org/10.3390/antiox10071076).
- 142. Zhu W, Tang H, Cao L, Zhang J, Li J, Ma D, et al. Epigallocatechin-3-Ogallate ameliorates oxidative stress-induced chondrocyte dysfunction and exerts chondroprotective effects via the Keap1/Nrf2/ARE signaling pathway. Chem Biol Drug Des. 2022;100(1):108–20. [https://doi.org/10.](https://doi.org/10.1111/cbdd.14056) [1111/cbdd.14056.](https://doi.org/10.1111/cbdd.14056)
- 143. Wang Y, Huang M, Zhou X, Li H, Ma X, Sun C. Potential of natural favonoids to target breast cancer angiogenesis. Br J Pharmacol. 2023. <https://doi.org/10.1111/bph.16275>.
- 144. Marín V, Burgos V, Pérez R, Maria DA, Pardi P, Paz C. The potential role of Epigallocatechin-3-Gallate (EGCG) in breast cancer treatment. Int J Mol Sci. 2023;24(13):10737. [https://doi.org/10.3390/ijms241310737.](https://doi.org/10.3390/ijms241310737)
- 145. Chu M, Zheng C, Chen C, Song G, Hu X, Wang Z-w, editors. Targeting cancer stem cells by nutraceuticals for cancer therapy. Seminars in cancer biology; 2022: Elsevier. [https://doi.org/10.1016/j.semcancer.2021.07.](https://doi.org/10.1016/j.semcancer.2021.07.008) [008](https://doi.org/10.1016/j.semcancer.2021.07.008).
- 146. Park IS, Cho JH, Han Y, Lee KW, Song YS. Targeting cancer stem cells with phytoceuticals for cancer therapy. Functional Foods in Cancer Prevention and Therapy. Elsevier; 2020. p. 329–57. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.jnutbio.2021.108843) [jnutbio.2021.108843](https://doi.org/10.1016/j.jnutbio.2021.108843).
- 147. Wei H, Ge Q, Zhang L-Y, Xie J, Gan R-H, Lu Y-G, et al. EGCG inhibits growth of tumoral lesions on lip and tongue of K-Ras transgenic mice through the Notch pathway. J Nutr Biochem. 2022;99:108843. [https://](https://doi.org/10.1016/j.jnutbio.2021.108843) doi.org/10.1016/j.jnutbio.2021.108843.
- 148. Lu X, Friedrich LJ, Eferth T. Natural products targeting tumour angiogenesis. Br J Pharmacol. 2023. [https://doi.org/10.1111/bph.16232.](https://doi.org/10.1111/bph.16232)
- 149. Tanabe H, Suzuki T, Ohishi T, Isemura M, Nakamura Y, Unno K. Efects of epigallocatechin-3-gallate on matrix metalloproteinases in terms of its anticancer activity. Molecules. 2023;28(2):525. [https://doi.org/10.3390/](https://doi.org/10.3390/molecules28020525) [molecules28020525](https://doi.org/10.3390/molecules28020525).
- 150. Alam M, Alam S, Shamsi A, Adnan M, Elasbali AM, Al-Soud WA, et al. Bax/Bcl-2 cascade is regulated by the EGFR pathway: therapeutic targeting of non-small cell lung cancer. Front Oncol. 2022;12:869672. <https://doi.org/10.3389/fonc.2022.869672>.
- 151. Pan Y, Lv H, Feng X, Zhou S, Hu H, Chen S, et al. Epigallocatechin gallate (EGCG) alleviates the infammatory response and recovers oral microbiota in acetic acid-induced oral infammation mice. Food Funct. 2023;14(22):10069–82. [https://doi.org/10.1039/D3FO03107A.](https://doi.org/10.1039/D3FO03107A)
- 152. Bag N, Bag A. Antimetastatic properties of tea polyphenols. Nutr Cancer. 2020;72(3):365–76. [https://doi.org/10.1080/01635581.2019.16384](https://doi.org/10.1080/01635581.2019.1638426) [26.](https://doi.org/10.1080/01635581.2019.1638426)
- 153. Yang L, Zhang W, Chopra S, Kaur D, Wang H, Li M, et al. The epigenetic modifcation of epigallocatechin gallate (EGCG) on cancer. Curr Drug Targets. 2020;21(11):1099–104. [https://doi.org/10.2174/1389450121](https://doi.org/10.2174/1389450121666200504080112) [666200504080112](https://doi.org/10.2174/1389450121666200504080112).
- 154. Bakhshandeh N, Mohammadi M, Mohammadi P, Nazari E, Damchi M, Khodabandelu S, et al. Increased expression of androgen receptor and PSA genes in LNCaP (prostate cancer) cell line due to high concentrations of EGCG, an active ingredient in green tea. Horm Mol Biol Clin Invest. 2023;44(2):181–6.<https://doi.org/10.1515/hmbci-2022-0054>.
- 155. Thomas P, Dong J. (-)-Epicatechin acts as a potent agonist of the membrane androgen receptor, ZIP9 (SLC39A9), to promote apoptosis of breast and prostate cancer cells. J Steroid Biochem Mol Biol. 2021;211:105906. [https://doi.org/10.1016/j.jsbmb.2021.105906.](https://doi.org/10.1016/j.jsbmb.2021.105906)
- 156. Ferrari E, Bettuzzi S, Naponelli V. The potential of epigallocatechin gallate (EGCG) in targeting autophagy for cancer treatment: a narrative review. Int J Mol Sci. 2022;23(11):6075. [https://doi.org/10.3390/ijms2](https://doi.org/10.3390/ijms23116075) [3116075.](https://doi.org/10.3390/ijms23116075)
- 157. Man GCW, Wang J, Song Y, Wong JH, Zhao Y, Lau TS, et al. Therapeutic potential of a novel prodrug of green tea extract in induction of apoptosis via ERK/JNK and Akt signaling pathway in human endometrial cancer. BMC Cancer. 2020;20:1–14. [https://doi.org/10.1186/](https://doi.org/10.1186/s12885-020-07455-3) [s12885-020-07455-3.](https://doi.org/10.1186/s12885-020-07455-3)
- 158. Dev SS, Farghadani R, Abidin SAZ, Othman I, Naidu R. Flavonoids as receptor tyrosine kinase inhibitors in lung cancer. J Funct Foods. 2023;110:105845. [https://doi.org/10.1016/j.jf.2023.105845.](https://doi.org/10.1016/j.jff.2023.105845)
- 159. Li F, Hao S, Gao J, Jiang P. EGCG alleviates obesity-exacerbated lung cancer progression by STAT1/SLC7A11 pathway and gut microbiota. J Nutr Biochem. 2023;120:109416. [https://doi.org/10.1016/j.jnutbio.2023.](https://doi.org/10.1016/j.jnutbio.2023.109416) [109416](https://doi.org/10.1016/j.jnutbio.2023.109416).
- 160. Bernini R, Velotti F. Natural polyphenols as immunomodulators to rescue immune response homeostasis: Quercetin as a research model against severe COVID-19. Molecules. 2021;26(19):5803. [https://doi.org/](https://doi.org/10.3390/molecules26195803) [10.3390/molecules26195803](https://doi.org/10.3390/molecules26195803).
- 161. Dey P, Olmstead BD, Sasaki GY, Vodovotz Y, Yu Z, Bruno RS. Epigallocatechin gallate but not catechin prevents nonalcoholic steatohepatitis in mice similar to green tea extract while diferentially afecting the gut microbiota. J Nutr Biochem. 2020;84:108455. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.jnutbio.2020.108455) [jnutbio.2020.108455.](https://doi.org/10.1016/j.jnutbio.2020.108455)
- 162. Suhail M, Rehan M, Tarique M, Tabrez S, Husain A, Zughaibi TA. Targeting a transcription factor NF-κB by green tea catechins using in silico and in vitro studies in pancreatic cancer. Front Nutr. 2023;9:1078642. [https://](https://doi.org/10.3389/fnut.2022.1078642) doi.org/10.3389/fnut.2022.1078642.
- 163. Khan H, Ullah H, Castilho PCMF, Gomila AS, D'Onofrio G, Filosa R, et al. Targeting NF-KB signaling pathway in cancer by dietary polyphenols. Crit Rev Food Sci Nutr. 2020;60(16):2790–800. [https://doi.org/10.1080/](https://doi.org/10.1080/10408398.2019.1661827) [10408398.2019.1661827](https://doi.org/10.1080/10408398.2019.1661827).
- 164. Niloy MS, Shakil MS, Alif MM, Rosengren RJ. Using natural compounds to target KRAS mutated non-small cell lung cancer. Curr Med Chem. 2021;28(39):8098–115. [https://doi.org/10.2174/09298673286662103011](https://doi.org/10.2174/0929867328666210301105856) [05856](https://doi.org/10.2174/0929867328666210301105856).
- 165. Eng QY, Thanikachalam PV, Ramamurthy S. Molecular understanding of Epigallocatechin gallate (EGCG) in cardiovascular and metabolic diseases. J Ethnopharmacol. 2018;210:296–310. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.jep.2017.08.035) [jep.2017.08.035](https://doi.org/10.1016/j.jep.2017.08.035).
- 166. Momose Y, Maeda-Yamamoto M, Nabetani H. Systematic review of green tea epigallocatechin gallate in reducing low-density lipoprotein cholesterol levels of humans. Int J Food Sci Nutr. 2016;67(6):606–13. <https://doi.org/10.1080/09637486.2016.1196655>.
- 167. Chen I-J, Liu C-Y, Chiu J-P, Hsu C-H. Therapeutic effect of high-dose green tea extract on weight reduction: a randomized, double-blind,

placebo-controlled clinical trial. Clin Nutr. 2016;35(3):592–9. [https://doi.](https://doi.org/10.1016/j.clnu.2015.05.003) [org/10.1016/j.clnu.2015.05.003.](https://doi.org/10.1016/j.clnu.2015.05.003)

- 168. Li G, Pan B, Liu L, Xu X, Zhao W, Mou Q, et al. Epigallocatechin-3-gallate restores mitochondrial homeostasis impairment by inhibiting HDAC1-mediated NRF1 histone deacetylation in cardiac hypertrophy. Mol Cell Biochem. 2024;479(4):963–73. [https://doi.org/10.1007/](https://doi.org/10.1007/s11010-023-04768-2) [s11010-023-04768-2](https://doi.org/10.1007/s11010-023-04768-2).
- 169. Wang M-H, Chang W-J, Soung H-S, Chang K-C. (−)-Epigallocatechin-3-gallate decreases the impairment in learning and memory in spontaneous hypertension rats. Behav Pharmacol. 2012;23(8):771–80. [https://](https://doi.org/10.1097/fbp.0b013e32835a3bc8) doi.org/10.1097/fbp.0b013e32835a3bc8.
- 170. Muhammed I, Sankar S, Govindaraj S. Ameliorative efect of epigallocatechin gallate on cardiac hypertrophy and fbrosis in aged rats. J Cardiovasc Pharmacol. 2018;71(2):65–75. [https://doi.org/10.1097/fc.](https://doi.org/10.1097/fjc.0000000000000545) [0000000000000545.](https://doi.org/10.1097/fjc.0000000000000545)
- 171. Chen K, Chen W, Liu SL, Wu TS, Yu KF, Qi J, et al. Epigallocatechingallate attenuates myocardial injury in a mouse model of heart failure through TGF-β1/Smad3 signaling pathway. Mol Med Rep. 2018;17(6):7652–60. <https://doi.org/10.3892/mmr.2018.8825>.
- 172. Cai Y, Yu S-S, Chen T-T, Gao S, Geng B, Yu Y, et al. EGCG inhibits CTGF expression via blocking NF-κB activation in cardiac fbroblast. Phytomedicine. 2013;20(2):106–13. [https://doi.org/10.1016/j.phymed.2012.](https://doi.org/10.1016/j.phymed.2012.10.002) [10.002](https://doi.org/10.1016/j.phymed.2012.10.002).
- 173. Pan B, Quan J, Liu L, Xu Z, Zhu J, Huang X, et al. Epigallocatechin gallate reverses cTnI-low expression-induced age-related heart diastolic dysfunction through histone acetylation modifcation. J Cell Mol Med. 2017;21(10):2481–90. <https://doi.org/10.1111/jcmm.13169>.
- 174. Chan D, Woo J, Ho S, Pang C, Law L, Ng P, et al. Genetic and environmental risk factors for Parkinson's disease in a Chinese population. J Neurol Neurosurg Psychiatry. 1998;65(5):781–4. [https://doi.org/10.1136/](https://doi.org/10.1136/jnnp.65.5.781) [jnnp.65.5.781.](https://doi.org/10.1136/jnnp.65.5.781)
- 175. Ono K, Tsuji M, Yamasaki TR, Pasinetti GM. Anti-aggregation efects of phenolic compounds on α-synuclein. Molecules. 2020;25(10):2444. <https://doi.org/10.3390/molecules25102444>.
- 176. Xu Y, Zhang Y, Quan Z, Wong W, Guo J, Zhang R, et al. Epigallocatechin gallate (EGCG) inhibits alpha-synuclein aggregation: a potential agent for Parkinson's disease. Neurochem Res. 2016;41:2788–96. [https://doi.](https://doi.org/10.1007/s11064-016-1995-9) [org/10.1007/s11064-016-1995-9](https://doi.org/10.1007/s11064-016-1995-9).
- 177. Kumar S, Kumar R, Kumari M, Kumari R, Saha S, Bhavesh NS, et al. Ellagic acid inhibits α-synuclein aggregation at multiple stages and reduces its cytotoxicity. ACS Chem Neurosci. 2021;12(11):1919–30. [https://doi.org/](https://doi.org/10.1021/acschemneuro.1c00001) [10.1021/acschemneuro.1c00001](https://doi.org/10.1021/acschemneuro.1c00001).
- 178. Xu Q, Langley M, Kanthasamy A, Reddy M. Neurorescue effect of EGCG in an animal model of Parkinson's disease. The FASEB Journal. 2016 30;1174.11-11. [https://doi.org/10.1007/s11064-016-1995-9.](https://doi.org/10.1007/s11064-016-1995-9)
- 179. Kujawska M, Jodynis-Liebert J. Polyphenols in Parkinson's disease: a systematic review of in vivo studies. Nutrients. 2018;10(5):642. [https://](https://doi.org/10.3390/nu10050642) doi.org/10.3390/nu10050642.
- 180. Chen P, Totten M, Zhang Z, Bucinca H, Erikson K, Santamaría A, et al. Iron and manganese-related CNS toxicity: mechanisms, diagnosis and treatment. Expert Rev Neurother. 2019;19(3):243–60. [https://doi.org/10.](https://doi.org/10.3390/nu10050642) [3390/nu10050642](https://doi.org/10.3390/nu10050642).
- 181. Chu AJ. Quarter-century explorations of bioactive polyphenols: Diverse health benefts. Front Biosci-Landmark. 2022;27(4):134. [https://doi.org/](https://doi.org/10.31083/j.fbl2704134) [10.31083/j.fbl2704134](https://doi.org/10.31083/j.fbl2704134).
- 182. Mandel S, Youdim MB. Catechin polyphenols: neurodegeneration and neuroprotection in neurodegenerative diseases. Free Radical Biol Med. 2004;37(3):304–17. [https://doi.org/10.1016/j.freeradbiomed.2004.04.](https://doi.org/10.1016/j.freeradbiomed.2004.04.012) [012](https://doi.org/10.1016/j.freeradbiomed.2004.04.012).
- 183. Mandel SA, Amit T, Weinreb O, Youdim MB. Understanding the broad-spectrum neuroprotective action profle of green tea polyphenols in aging and neurodegenerative diseases. J Alzheimers Dis. 2011;25(2):187–208. [https://doi.org/10.1016/j.freeradbiomed.2004.04.](https://doi.org/10.1016/j.freeradbiomed.2004.04.012) [012](https://doi.org/10.1016/j.freeradbiomed.2004.04.012).
- 184. Alam M, Ahmed S, Abid M, Hasan GM, Islam A, Hassan MI. Therapeutic targeting of microtubule affinity-regulating kinase 4 in cancer and neurodegenerative diseases. J Cell Biochem. 2023;124(9):1223–40. [https://](https://doi.org/10.1002/jcb.30468) doi.org/10.1002/jcb.30468.
- 185. Iqbal K, Grundke-Iqbal I. Alzheimer neurofbrillary degeneration: signifcance, etiopathogenesis, therapeutics and prevention. J Cell Mol Med. 2008;12(1):38–55.<https://doi.org/10.1111/j.1582-4934.2008.00225.x>.
- 186. Xue B, DasGupta D, Alam M, Khan MS, Wang S, Shamsi A, et al. Investi gating binding mechanism of thymoquinone to human transferrin, tar geting Alzheimer's disease therapy. J Cell Biochem. 2022;123(8):1381– 93.<https://doi.org/10.1002/jcb.30299> .
- 187. Hasan GM, Shamsi A, Sohal SS, Alam M, Hassan MI. Structure-Based Identifcation of Natural Compounds as Potential RET-Kinase Inhibitors for Therapeutic Targeting of Neurodegenerative Diseases. Journal of Alzheimer's disease. 2023(Preprint):1–15. [https://doi.org/10.3233/](https://doi.org/10.3233/jad-230698) [jad-230698](https://doi.org/10.3233/jad-230698) .
- 188. Cascella M, Muzio MR. Potential application of the Kampo medicine goshajinkigan for prevention of chemotherapy-induced peripheral neuropathy. J Integr Med. 2017;15(2):77–87. [https://doi.org/10.1016/](https://doi.org/10.1016/S2095-4964(17)60313-3) [S2095-4964\(17\)60313-3](https://doi.org/10.1016/S2095-4964(17)60313-3) .
- 189. Soeda S, Aritake K, Urade Y, Sato H, Shoyama Y. Neuroprotective activities of safron and crocin. The benefts of natural products for neurodegenerative diseases. 2016:275–92. [https://doi.org/10.1007/](https://doi.org/10.1007/978-3-319-28383-8_14) [978-3-319-28383-8_14](https://doi.org/10.1007/978-3-319-28383-8_14)
- 190. Grabska-Kobyłecka I, Szpakowski P, Król A, Książek-Winiarek D, Kobyłecki A, Głąbiński A, et al. Polyphenols and their impact on the preven tion of neurodegenerative diseases and development. Nutrients. 2023;15(15):3454. <https://doi.org/10.3390/nu15153454> .
- 191. Manzine PR, Ettcheto M, Cano A, Busquets O, Marcello E, Pelucchi S, et al. ADAM10 in Alzheimer's disease: Pharmacological modulation by natural compounds and its role as a peripheral marker. Biomed Pharmacother. 2019;113:108661. [https://doi.org/10.1016/j.biopha.2019.](https://doi.org/10.1016/j.biopha.2019.108661) [108661](https://doi.org/10.1016/j.biopha.2019.108661) .
- 192. Saklayen MG. The global epidemic of the metabolic syndrome. Curr Hypertens Rep. 2018;20(2):1–8. [https://doi.org/10.1007/](https://doi.org/10.1007/s11906-018-0812-z) [s11906-018-0812-z](https://doi.org/10.1007/s11906-018-0812-z) .
- 193. Zheng Y, Ley SH, Hu FB. Global aetiology and epidemiology of type 2 diabetes mellitus and its complications. Nat Rev Endocrinol. 2018;14(2):88–98. <https://doi.org/10.1038/nrendo.2017.151> .
- 194. Martin MA, Goya L, Ramos S. Protective effects of tea, red wine and cocoa in diabetes. Evidences from human studies. Food Chem Toxicol. 2017;109:302–14.<https://doi.org/10.1016/j.fct.2017.09.015> .
- 195. Forbes JM, Cooper ME. Mechanisms of diabetic complications. Physiol Rev. 2013;93(1):137–88.<https://doi.org/10.1152/physrev.00045.2011> .
- 196. Bulboaca AE, Boarescu PM, Porfre AS, Dogaru G, Barbalata C, Valeanu M, et al. The efect of nano-epigallocatechin-gallate on oxidative stress and matrix metalloproteinases in experimental diabetes mellitus. Anti oxidants (Basel). 2020;9(2):172. <https://doi.org/10.3390/antiox9020172> .
- 197. Liu C-Y, Huang C-J, Huang L-H, Chen I-J, Chiu J-P, Hsu C-H. Efects of green tea extract on insulin resistance and glucagon-like peptide 1 in patients with type 2 diabetes and lipid abnormalities: a rand omized, double-blinded, and placebo-controlled trial. PLoS ONE. 2014;9(3):e91163. <https://doi.org/10.1371/journal.pone.0091163> .
- 198. Ali F, Ismail A, Kersten S. Molecular mechanisms underlying the poten tial antiobesity-related diseases efect of cocoa polyphenols. Mol Nutr Food Res. 2014;58(1):33–48. <https://doi.org/10.1002/mnfr.201300277> .
- 199. Chen L, Pu Y, Xu Y, He X, Cao J, Ma Y, et al. Anti-diabetic and anti-obesity: Efficacy evaluation and exploitation of polyphenols in fruits and vegetables. Food Res Int. 2022;157:111202. [https://doi.org/10.1016/j.foodr](https://doi.org/10.1016/j.foodres.2022.111202) [es.2022.111202](https://doi.org/10.1016/j.foodres.2022.111202) .
- 200. Rodríguez-Daza MC, de Vos WM. Polyphenols as drivers of a homeo static gut microecology and immuno-metabolic traits of Akkermansia muciniphila: from mouse to man. Int J Mol Sci. 2022;24(1):45. [https://](https://doi.org/10.3390/ijms24010045) doi.org/10.3390/ijms24010045 .
- 201. Wu Z, Huang S, Li T, Li N, Han D, Zhang B, et al. Gut microbiota from green tea polyphenol-dosed mice improves intestinal epithelial home ostasis and ameliorates experimental colitis. Microbiome. 2021;9:1–22. <https://doi.org/10.1186/s40168-021-01115-9> .
- 202. Legeay S, Rodier M, Fillon L, Faure S, Clere N. Epigallocatechin gallate: a review of its benefcial properties to prevent metabolic syndrome. Nutrients. 2015;7(7):5443–68.<https://doi.org/10.3390/nu7075230> .

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in pub lished maps and institutional affiliations.