Evidence for NF-κB- and CBP-Independent Repression of p53's Transcriptional Activity by Human T-Cell Leukemia Virus Type 1 Tax in Mouse Embryo and Primary Human Fibroblasts

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The human T-cell leukemia virus type 1 (HTLV-1) Tax oncoprotein can repress the transcriptional activity of the tumor suppressor protein p53. However, it remains controversial whether Tax requires NF- κ B factors/ activity and/or p300/CBP in order to inactivate p53 function. To address this issue, we have investigated Tax's effect on p53's transcriptional activation in I κ B-kinase-deficient mouse embryonic fibroblasts (MEFs); some of which are entirely silent for Tax-induced NF- κ B activity. We found that, in IKK $\alpha^{-/-}$, IKK $\beta^{-/-}$, and IKK $\gamma^{-/-}$ MEFs, p53 activation of a prototypic responsive plasmid (pG13-luciferase) was repressed by wild-type Tax. Curiously, p53's activity in MEFs was also repressed by a p300/CBP-binding deficient Tax protein. Our results highlight the complex nature of Tax-mediated repression of p53- activity, which requires further investigation.

Human T-cell leukemia virus type 1 (HTLV-1) is the etiological agent for adult T-cell leukemia (ATL) (22, 33). While the mechanism of tumorigenesis by HTLV-1 is not fully understood, the viral Tax protein is strongly implicated in the transformation of cells (7, 8, 16, 23, 28). Tax has pleiotropic effects on host cell gene expression and activates both CREB/ ATF (reviewed in reference 3) and NF- κ B (26) pathways.

Tumor suppressor p53 is a DNA-binding transcription factor which plays a key role in the response of cells to DNA damage and transformation (31). Approximately 50% of all cancers have mutations in the TP53 gene (which encodes p53). Relevant to HTLV-1, it was first observed that p53 mutation is present in only a small fraction of patients with ATL and that within ATL cells the p53 protein appears to be stabilized (27). Subsequently, many investigators have independently demonstrated that p53's transcriptional activity is suppressed by the HTLV-1 Tax oncoprotein (1, 2, 15, 18, 24, 30).

Relevant to the mechanism of p53 inactivation by Tax, there are two schools of thought. One subscribes to the concept that Tax competes with p53 for the binding of the p300/CBP transcriptional coactivator (2, 29). The second suggests that Tax acts through an NF- κ B/RelA(p65) activation pathway to perturb p53 function (19, 20). In this study, we have interrogated mouse embryonic fibroblasts (MEFs) genotypically deficient in IKK α (IKK $\alpha^{-/-}$), IKK β (IKK $\beta^{-/-}$), and IKK γ (IKK $\gamma^{-/-}$) for Tax-p53 functional interplay. We also asked whether in the same three MEFs, a Tax mutant (K88A), previously shown to be incapable of binding CBP, remains proficient for suppressing p53's transcriptional activity.

Tax repressed p53's transcriptional activity in IKK $\alpha^{-/-}$,

IKK $\beta^{-/-}$, and IKK $\gamma^{-/-}$ MEFs. Previous findings showed that Tax can repress the transcriptional activity of p53 in various transformed cell lines. We began our present work by reconfirming Tax's repressive activity in HeLa cells. Accordingly, we transfected HeLa cells with a luciferase reporter plasmid containing 13 copies of a p53 consensus binding site (pG13-Luc) with a p53 expression plasmid, alone or with p53 plus a second expression plasmid for Tax (WT; Fig. 1A, left). As expected, p53 robustly activated the expression of pG13-Luc, while the further addition of Tax repressed activation by approximately 70% (Fig. 1A). Indeed, p53's activity was repressed by Tax in a dose-dependent manner (Fig. 1B). As controls, we verified the expression and activity of three Tax mutants (to be used in later transfections) by Western blotting and by assays which employed HTLV-long terminal repeat (LTR)-Luc (Fig. 1A). A dose-dependent expression of transfected p53 plasmid in HeLa cells was also verified (Fig. 1C).

We next proceeded to investigate Tax-p53 interaction in MEFs which are individually knocked out for IKK α , IKK β , or IKK γ . We chose this approach because the IKK $\alpha/\beta/\gamma$ complex is critically important for downstream nuclear NF- κ B activation (34) and because others have suggested a role for Tax-induced NF- κ B activation in repressing p53 function (19, 20). Currently, there is wide agreement that Tax directly binds IKK γ (4, 9, 13) and that in IKK $\gamma^{-/-}$ settings NF- κ B cannot be activated by tumor necrosis factor alpha (TNF- α), interleukin-1 β (IL-1 β), or Tax (11, 32).

We cotransfected pG13-Luc, p53, and Tax in the indicated combinations into IKK $\alpha^{-/-}$, IKK $\beta^{-/-}$, and IKK $\gamma^{-/-}$ MEFs (Fig. 1D). Since all of our assays in the MEFs were done side by side under identical conditions, we could compare and observe that the basal activity of MEF-endogenous p53 varied, in fact, quite a bit between WT, IKK $\alpha^{-/-}$, IKK $\beta^{-/-}$, and IKK $\gamma^{-/-}$ cells (Fig. 1D, first bar of each graph). Basal expression of pG13-Luc was lower (approximately 5- to 10-fold; data not shown) in IKK $\alpha^{-/-}$ and IKK $\beta^{-/-}$ cells than in either WT

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FIG. 1. Repression of p53's transcriptional activity in IKK-deficient MEFs by Tax. (A) Cells were transfected with 1 μ g of Tax or indicated Tax mutants, and cell lysates were subjected to immunoblotting using polyclonal anti-Tax antibody (upper panel) or luciferase assay (bottom panel). For the luciferase assay, HeLa cells were cotransfected with 0.5 μ g of HTLV-LTR-Luc and 0.5 μ g of either Tax (WT) or a Tax mutant (H43Q, S274A, or L320G), respectively. (B) HeLa cells were cotransfected with 0.1 μ g of reporter plasmid pG13-Luc, 0.5 μ g of p53, and progressively increased amounts of wild-type Tax plasmid as indicated. (C) Expression of p53 in HeLa cells from transfected plasmids. p53 expression vectors, at the indicated amounts, were transfected into HeLa cells, and expression of protein was monitored by Western blotting. (D) Wild-type MEFs (WT) and IKK $\alpha^{-/-}$, IKK $\beta^{-/-}$, and IKK $\gamma^{-/-}$ MEFs were cotransfected in parallel with combinations of 0.1 μ g of pG13-Luc, 0.5 μ g of p53 expression plasmid, and 0.5 μ g of Tax. Total transfected DNA was adjusted to the same amount with empty carrier plasmid. Luciferase assays were performed 24 h after transfection (upper panel). Cell lysates from a representative experiment were subjected to immunoblotting using anti-p53 (DO-1) and anti- β -actin (lower panel). All luciferase activities were normalized to β -galactosidase (β -Gal) readouts from a cotransfected basal luciferase value. Results represent averages \pm standard deviations from at least three independent experiments. *P* values are indicated.

or IKK $\gamma^{-/-}$ cells, and correspondingly activation by p53 was commensurately much higher (Fig. 1D). Nonetheless, expression of exogenous p53 consistently elevated pG13-Luc (Fig. 1C, second bar of each graph) expression to decent levels in all cells. In all cases, we further observed that this activation was repressed by Tax (Fig. 1D, third bar of each graph). Consistent with previous reports which correlated stabilization of p53 by Tax with inactivation of p53 activity (24, 27), immunoblotting



FIG. 2. Repression of p53's transcriptional activity in IKK $\gamma^{-/-}$ MEFs by Tax mutants. IKK $\gamma^{-/-}$ cells were cotransfected with the indicated combinations of 0.1 µg of pG13-Luc, 0.5 µg of p53, 0.5 µg of Tax S258A (left), L320G (middle), and H43Q (right). Luciferase assays were performed 24 h later. Results are average values ± standard deviations from at least three independent experiments.

analyses of the transfected cells showed slightly higher steadystate amounts of p53 in the presence of Tax (Fig. 1D; middle panel, p53). These analyses indicate that the decreases seen in p53's transcriptional activity cannot be trivially explained by reduced p53 protein expression.

Elsewhere, we noted that Tax has residual NF-KB-activating activities in both IKK $\alpha^{-/-}$ and IKK $\beta^{-/-}$ MEFs; by comparison, its activation of NF- κ B is completely silent in IKK $\gamma^{-/-}$ cells (unpublished observations). We, thus, focused our analyses of NF- κ B requirement for p53 repression using IKK $\gamma^{-/-}$ cells. To provide an additional degree of stringency, we tested the TaxS258A mutant, a form of Tax previously characterized to be CREB active but NF-KB defective (25). We found that in IKK $\gamma^{-/-}$ MEFs, TaxS258A did, in fact, repress p53-activation of pG13-Luc (Fig. 2, left). Hence, although NF-KB activation was thwarted through two independent constraints (i.e., a cellular defect enforced through $IKK\gamma^{-/-}$ and a viral defect stipulated by using an NF-kB-incompetent Tax mutant), p53 activation of pG13-Luc was still suppressed by Tax. This result (Fig. 2, left) posits that NF-kB activation is dispensable for Tax's repression of p53-activity.

In parallel, we used another mutant, TaxL320G (25), as a control for TaxS258A. Whereas TaxS258A is NF-κB⁻ CREB⁺, TaxL320G is NF- κ B⁺ CREB⁻. One possible interpretation of the above TaxS258A result is that suppression of p53 activity requires CREB activation by Tax. However, when we analyzed the TaxL320G results in IKK $\gamma^{-/-}$ MEFs, we were surprised to see comparably strong suppression of p53 activation (Fig. 2, middle). We also analyzed a third mutant, TaxH43Q. Previously, Fu and colleagues have delineated that TaxH43Q cannot bind protein phosphatase 2A (PP2A) and failed to activate NF-KB (5). Here, we found that TaxH43Q, nevertheless, remained competent for suppressing p53's transcriptional activity (Fig. 2, right). Taken together, our results argue that in IKK $\gamma^{-/-}$ cells neither NF- κ B (Fig. 2, left and right panels) nor CREB (Fig. 2, middle panel) activity appears to be needed for Tax's repression of p53-pG13-Luc activation.

TaxK88A, a p300/CBP-binding defective mutant, repressed



FIG. 3. A TaxK88A mutant deficient for CBP binding still repressed p53's transcriptional activity. (A) Cells were cotransfected with 0.5 µg of NF- κ B-Luc or HTLV-LTR-Luc and either 0.5 µg of Tax or Tax K88A, as indicated. (B) Wild-type MEF (WT), IKK $\alpha^{-/-}$, IKK $\beta^{-/-}$, and IKK $\gamma^{-/-}$ cells were cotransfected with 0.1 µg of pG13-Luc, 0.5 µg of p53, and 0.5 µg of TaxK88A. Luciferase assays were performed 24 h later. Results represent averages +/- standard deviations from at least three independent experiments.

p53 function. Although TaxL320G cannot functionally activate CREB, it remains competent for physical association with CREB and p300/CBP (6). To test directly the hypothesis that p300/CBP sequestration by Tax is a mechanism for suppressing p53's transcriptional activity, we next checked, in MEFs, the TaxK88A mutant previously shown by Harrod and colleagues to be defective in p300/CBP-binding (10). We confirmed that TaxK88A is NF-κB active (Fig. 3A, left); and, consistent with its p300/CBP binding defect, TaxK88A could not activate the HTLV-1 LTR (Fig. 3A, right). Provocatively, in WT, IKKα^{-/-}, IKKβ^{-/-}, and IKKγ^{-/-} MEFs, we found that TaxK88A repressed p53's activation of pG13-Luc (Fig. 3B).

It is possible that the TaxK88A results are idiosyncratic to mouse embryo fibroblasts and harbor no general implication for the behavior of Tax in primary nontransformed human cells. To address this question, we obtained primary human fibroblasts from the American Type Culture Collection and



FIG. 4. Tax-p300/CBP interaction appears not to be needed to repress p53 activity in human primary fibroblasts. Human primary fibroblast cells were cotransfected with indicated combinations of 0.25 μ g of pG13-Luc, 0.25 μ g of p53, and 0.5 μ g of either Tax (WT) or the TaxK88A mutant. In the right panel 3 μ g of p300 Δ HAT shown previously to be dominant negative for Tax-p300/CBP interaction (17) was employed. Luciferase assays were performed 24 h later. Results represent averages \pm standard deviations from at least three independent transfections.

tested for functional interplay between TaxK88A and p53. In human fibroblasts, WT Tax suppressed p53 activation of pG13-Luc (Fig. 4, left), and TaxK88A also did so comparably (Fig. 4, middle). Thus, the ability of TaxK88A to influence p53 activity is conserved from MEFs to human primary cells. To further establish that a functional interplay between Tax and p300/ CBP may not be required for the former to suppress p53's transcriptional activity, we employed the p300 Δ HAT protein. Previously, we found that $p300\Delta HAT$ is dominant negative for Tax's interaction with p300/CBP (17). At concentration of p300AHAT which suppressed the ability of Tax to activate HTLV-1 LTR (data not shown), we found that Tax nevertheless repressed p53 activity in primary human fibroblasts (Fig. 4, right). This result further supports the notion that the p300/ CBP-interaction may be dispensable for Tax's action on p53 in human cells.

Here, using IKK-deficient MEFs plus two NF- κ B-deficient Tax mutants we accrued evidence that repression of p53 by Tax is independent of NF- κ B signaling (Fig. 1 and 2). Astonishingly, we also found that, in IKK-deficient MEFs and in human primary fibroblasts, CREB- and p300/CBP interaction by Tax also seems to be unnecessary for repressing p53 activity (Fig. 3 and 4). Our results here using primary mouse and human fibroblasts add further complexity to the Tax-p53 interaction which cannot be easily explained by currently proposed mechanistic models.

After this work was completed and during the writing of the manuscript, we came across a paper from Jeong and colleagues suggesting that Tax repression of p53 is operative in IKK $\alpha^{-/-}$ MEFs but not in IKK $\beta^{-/-}$ MEFs (12). Perplexingly, these investigators did not test IKK $\gamma^{-/-}$ MEFs, which in our experience is the most stringent cell that restricts Tax's ability to activate NF- κ B. The results of Jeong and coworkers in IKK $\alpha^{-/-}$ agree with our findings, but their results in IKK $\beta^{-/-}$ do not (Fig. 1 and 3). There are several possible reasons for the

difference. First, Jeong et al. used an MDM2-Luc reporter as a p53-responsive surrogate, while we used the arguably more sensitive Vogelstein reporter, pG13-Luc, which contains 13 consecutive p53-binding sites (14). Second, we assayed for suppression by Tax of transfected p53; by contrast, Jeong et al. relied on Tax suppression of presumptively active MEF-endogenous p53. This difference in approach is significant because, as we have noted above, the basal activity of p53 is extremely low in IKK $\alpha^{-/-}$ and IKK $\beta^{-/-}$ MEFs. In fact, p53 protein is undetectable by immunoblotting in MEF cells (Fig. 1D, bottom), and we have found that interpretations of perturbations based on low basal luciferase readouts are frequently unreliable. Moreover, in the absence of data, it was unclear whether the ambient expression of MDM2-Luc in IKK $\beta^{-/-}$ cells reported by Jeong et al. was in fact p53 dependent and not simply driven by TATAA-specific basal transcription factors. Nevertheless, despite these differences in detail, both we and Jeong et al. (12) arrived at the same major conclusion that Tax activation of NF-KB is dispensable for suppression of p53 function. We want to provide a word of caution that our findings stem from overexpression of Tax. Whether the results accurately reflect Tax effects from integrated provirus requires further verification.

If not NF-κB activation or CBP sequestration, how then does Tax suppress p53 function? We profess that we do not know the answer; however, we can offer a preliminary speculation. Previously, Pise-Masison et al. have reported that p53 is phosphorylated at Ser15 and Ser 392 in HTLV-1-transformed cells (21), while Ariumi et al. (2) demonstrated instead Taxdependent phosphorylation of p53 at Ser 6, 9, 20, 33, and 37 and at Ser 15 only after DNA damage. In agreement with these independent findings, we have found a Tax-associated kinase which specifically phosphorylates p53; and on our two-dimensional gel analyses Tax-dependent phosphorylation of p53 correlates with repression of function (data not shown). As yet, we have not identified this Tax-associated kinase, and we do not yet know whether it has a role(s) in either NF-κB or CREB/ CBP signaling.

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