Towards simplified graph neural networks for identifying cancer driver genes in heterophilic networks

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Abstract

The identification of cancer driver genes is crucial for understanding the complex processes involved in cancer development, progression, and therapeutic strategies. Multi-omics data and biological networks provided by numerous databases enable the application of graph deep learning techniques that incorporate network structures into the deep learning framework. However, most existing methods do not account for the heterophily in the biological networks, which hinders the improvement of model performance. Meanwhile, feature confusion often arises in models based on graph neural networks in such graphs. To address this, we propose a Simplified Graph neural network for identifying Cancer Driver genes in heterophilic networks (SGCD), which comprises primarily two components: a graph convolutional neural network with representation separation and a bimodal feature extractor. The results demonstrate that SGCD not only performs exceptionally well but also exhibits robust discriminative capabilities compared to state-ofthe-art methods across all benchmark datasets. Moreover, subsequent interpretability experiments on both the model and biological aspects provide compelling evidence supporting the reliability of SGCD. Additionally, the model can dissect gene modules, revealing clearer connections between driver genes in cancers. We are confident that SGCD holds potential in the field of precision oncology and may be applied to prognosticate biomarkers for a wide range of complex diseases.

Keywords: cancer driver genes; multi-omics data; graph neural networks; heterophilic networks; precision oncology

Introduction

In biomedical research, it is generally accepted that the onset of cancer is associated with the accumulation of mutations in driver genes. Therefore, accurately identifying cancer driver genes is paramount for clarifying the biological processes that underlie carcinogenesis and for the advancement of personalized oncology pharmaceuticals [[1](#page-8-0), [2\]](#page-8-1).

Over the past decade, many statistics methods have emerged to detect cancer driver genes. The approaches based on frequency such as MuSic [\[3](#page-8-2)], MutSigCV [\[4\]](#page-8-3), and OncodriveCLUST [[5](#page-8-4)] generally operate under the assumption that mutations in driver genes tend to recur more often across samples compared to those in nondriver genes, thereby facilitating the recognition of highly mutated genes as cancer driver genes. Nonetheless, these techniques often struggle to identify driver genes with rare mutations.

As machine learning (ML) advances rapidly and large-scale cancer genomics projects involving thousands of patients continue to release multi-omics data [\[6–](#page-8-5)[8\]](#page-8-6), numerous ML-based approaches have also shown noteworthy success in identifying cancer driver genes. Essentially, these methods concentrate on deriving low-dimensional representations of genes from a variety of biological attributes to effectively distinguish driver genes from non-driver genes. For instance, LOTUS [[9\]](#page-8-7) employs

a support vector machine to detect pan-cancer driver genes, while TUSON [[10\]](#page-8-8) utilizes a LASSO regression for the same purpose. Furthermore, there are various methods designed to detect personalized cancer driver genes, such as sysSVM [[11\]](#page-8-9) and IMCDriver [[12](#page-8-10)]. Nonetheless, most existing ML-based methods focus exclusively on multi-omics data to construct gene embeddings for the identification of cancer driver genes, failing to take into account the topological features provided by biological networks.

Some methods focusing on networks are conducted under the assumption that cancers are driven by alterations in numerous genes that interact intimately within biological networks. Therefore, these methods identify cancer driver genes by targeting genes that take on critical topological positions in biological networks using network propagation strategies, such as pgWalk [[13](#page-8-11)], RWRH [[14](#page-8-12)], and BiRW [\[15](#page-8-13)]. However, the lack of omics data has weakened their performance.

With the emergence of graph neural networks (GNNs), the fusion of multi-omics data and networks for identifying cancer driver genes is becoming promising [\[16–](#page-8-14)[19](#page-8-15)]. For example, EMOGI [[20\]](#page-9-0) is a method built on graph convolutional neural network (GCN) [[21\]](#page-9-1) that leverages multi-omics data as gene features along with the protein–protein interaction (PPI) network to identify

Received: November 9, 2024. **Revised:** November 26, 2024. **Accepted:** December 16, 2024

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cancer driver genes. MTGCN [[22](#page-9-2)] merges biological and structural features to develop enhanced representations, utilizing a multitask framework designed to improve the tasks of node and link prediction. SMG [[23\]](#page-9-3) performs a node reconstruction task to obtain a pre-trained encoder, which is then used for downstream tasks including gene identification and disease subnetwork identification. Although the aforementioned methods have yielded some success in cancer driver gene identification, they all rely on the homophily assumption. Recently, many researches have shown that most biological networks tend to exhibit heterophily property [\[24,](#page-9-4) [25](#page-9-5)]. For instance, PPIs encompass both physical interactions and functional associations between various biomolecules, exhibiting a low homophily ratio, as elaborated in Supplementary Materials, [Section 2](https://academic.oup.com/bib/article-lookup/doi/10.1093/bib/bbae691#supplementary-data). Additionally, the quantity of cancer driver genes is markedly fewer than the total number of genes within the biomolecular networks [\[26,](#page-9-6) [27\]](#page-9-7).

A few methods have tried to enhance the accuracy of identifying cancer driver genes on heterophilic graphs. For example, HGDC [\[27](#page-9-7)] is designed to tackle the heterophilic nature of biological networks to identify cancer driver genes, integrating graph diffusion technique with hierarchical attention mechanisms. However, the graph diffusion convolution (GDC) employed by HGDC necessitates the calculation and storage of diffusion matrices such as the heat diffusion kernel or personalized PageRank, which can be computationally burdensome for large-scale graphs. Moreover, based on GCN, HGDC also encounters the issue of feature confusion, a common challenge for traditional GNNs in heterophilic graphs.

In this study, we propose an efficient approach named SGCD (Simplified Graph neural network for identifying Cancer Driver genes in heterophilic networks), which utilizes simplified GNNs for identifying cancer driver genes in heterophilic networks. We innovatively introduce representation separation (RS) to replace the traditional message-passing mechanism of GCN, effectively mitigating the issue of feature confusion in GNNs when dealing with these graphs. The experimental findings consistently indicate that SGCD surpasses the stateof-the-art approaches, highlighting its excellence. Furthermore, subsequent model interpretability experiments and biological interpretability experiments provide compelling evidence for the powerful interpretability of SGCD. In addtion, SGCD can dissect gene modules, enabling a comprehensive analysis of the association between gene modules and cancer mechanisms.

Material and methods Data collection and preprocessing

From the Cancer Genome Atlas ([https://portal.gdc.cancer.gov/\)](https://portal.gdc.cancer.gov/), we collect oncogenomics (mutations and copy number variations), epigenomics (DNA methylation), and transcriptomics (gene expression) data, comprising over 29 446 samples across 16 distinct malignancies.

For each gene, we calculate gene mutation rate, copy number aberrations, differential DNA methylation rate, and differential gene expression rate across the 16 cancer types (see Supplementary Materials [Section 3](https://academic.oup.com/bib/article-lookup/doi/10.1093/bib/bbae691#supplementary-data) for details). By integrating the feature vectors from all cancer types, we construct a 64 dimensional feature vector for every gene. Subsequently, minmax normalization is conducted on the features of each gene.

The lists of known driver genes are sourced from the Network of Cancer Genes (NCG) v6.0 [\[28\]](#page-9-8), COSMIC Cancer Gene Census

(CGC v91) [\[26\]](#page-9-6), and DigSEE [\[29\]](#page-9-9), which serve as positive samples. In contrast, negative samples are obtained by excluding gene lists from NCG, COSMIC, OMIM [\[30\]](#page-9-10), as well as pathways from KEGG [[31](#page-9-11)].

The PPI data is from CPDB [[32](#page-9-12)], MULTINET [[33](#page-9-13)], PCNet [[34](#page-9-14)], STRINGdb [\[35\]](#page-9-15), and IRefIndex [[36](#page-9-16)]. Particularly, we exclusively consider interactions with high confidence. For the CPDB network, only interactions with a confidence score above 0.5 are included, while for STRINGdb, a threshold of 0.85 is applied. MULTINET and the 2015 IRefIndex version are directly retrieved from the Hotnet2 github repository. For the updated IRefIndex, our primary focus is on binary interactions between two proteins as well as human interactions. The process of PCNet is the same as EMOGI [\[20\]](#page-9-0). To integrate different PPI data into the consistent format, we first convert gene names from different formats into uniform symbol names. Each gene is then characterized as a node in the graph, with edges constructed between nodes to reflect the corresponding protein-protein interactions. Consequently, we obtain a total of six PPIs in a unified format, and the overview of PPIs can be found in [Table S2](https://academic.oup.com/bib/article-lookup/doi/10.1093/bib/bbae691#supplementary-data) in the Supplementary Materials.

Overview of SGCD

SGCD is a straightforward and effective method comprising primarily two components: a GCN with RS and a bimodal feature extractor. The overview of SGCD is shown in [Fig. 1.](#page-2-0) Firstly, SGCD leverages a GCN with RS to learn node embeddings from multiomics and PPIs. Secondly, SGCD incorporates a bimodal feature extractor to preserve the topological information from PPIs and the multi-omics information. Finally, SGCD aggregates the representations obtained from the above two modules to calculate the likelihood of a gene functioning as a driver gene. Additionally, SGCD employs GNNExplainer to identify cancer gene modules by detecting compact subgraph structures through a masking method.

GCN with RS

Recently, many studies have theoretically demonstrated that RS can enhance the generalization capability of GNNs on heterophilic graphs [[37](#page-9-17), [38\]](#page-9-18). Therefore, SGCD introduces RS to replace the traditional message-passing mechanism of the GCN. As far as we know, we are the first to utilize RS within the framework for identifying cancer driver genes. Considering there is an attributed biological network denoted as $G = \{A, X\}$. A represents the adjacency matrix and $X \in \mathbb{R}^{n \times f}$ is the features matrix, where *n* indicates the overall number of genes and *f* is the dimension of multi-omics data. Firstly, to prevent significant differences in features distribution between high-degree and low-degree nodes during information passing, SGCD applies symmetric normalization as follows:

$$
\tilde{A} = \tilde{D}^{-\frac{1}{2}} (A + I) \tilde{D}^{-\frac{1}{2}}
$$
\n(1)

where $\tilde{D} = D + I$, *D* is the diagonal degree matrix of *A*, and *I* is the identity matrix.

To fully leverage node representations, SGCD extracts both the representations of first-order neighboring nodes and secondorder neighboring nodes. Besides, since potentially large differences in features between connected nodes in heterophilic graph, aggregating neighbor information through layer-by-layer stacking (i.e., sum operation) can cause node representations to become

Figure 1. Overview of SGCD. (a) Collection of PPIs and 64-dimensional multi-omics data. (b) GCN with RS Module.We leverage a GCN with RS to learn node embeddings from multi-omics and PPI networks. (c) Bimodal feature extractor. It incorporates two Multilayer Perceptrons (MLP) to separately embed the topological information and the multi-omics. (d) Prediction module. We combine the convolution-derived representation with the bimodal MLP-derived representation through a linear layer to estimate whether a gene serves as a driver gene. (e) GNNExplainer is adopted to identify cancer gene modules.

similar, which hinders the performance of the model from effectively distinguishing between different classes of nodes [[39](#page-9-19)]. SGCD alleviates the confusion in representing central nodes in dissimilar node pairs through concatenation. The node representation h_G can be defined as follows:

$$
r_1^{(l)} = \tilde{A}r^{(l)} \tag{2}
$$

$$
r_2^{(l)} = \tilde{A}^2 r^{(l)} \tag{3}
$$

 $r^{(l+1)}$ = ReLU(concat($[r_1^{(l)}, r_2^{(l)}]$ $\binom{10}{2}$)) (4)

$$
h_G = \log \sigma \left(\text{concat}([r^{(1)}W^{(1)}, ..., r^{(l)}W^{(l)}, ..., r^{(L)}W^{(L)}]) \right) \tag{5}
$$

 w here $r^{(0)} = XW^{(0)}, l ∈ {0, 1, ..., L}, L$ is the layer number of convolutions, {*W(*0*)* , ...,*W(l)* , ...,*W(L)* } are a series of trainable matrices, ReLU is the rectified linear unit, σ is softmax function.

Bimodal feature extractor

Due to the complex relationships between graph topology and label distributions in heterophilic graphs, many conventional GNNs may fail to fully leverage the graph topology in such contexts [[40](#page-9-20), [41\]](#page-9-21). In contrast, MLPs are essentially fully connected neural networks, where each layer performs a linear transformation on the feature vectors followed by a nonlinear activation function. This design makes MLPs rely solely on the features of the nodes without considering adjacency relationships, thereby maintaining local independence of nodes and effectively avoiding interference from neighboring nodes. In our study, regarding G as a combination of two modalities: topology and multi-omics, SGCD designs a bimodal feature extractor based on MLP to separately embed the adjacency matrix into h_A and the multi-omics into h_X. This design enables SGCD to acquire distinct topological and omics information, avoiding a conflation of them. Then,

SGCD integrates information from two modalities to generate the representations of nodes. The details are as follows:

$$
h_A = \text{MLP}_A(A) \tag{6}
$$

$$
h_X = \text{MLP}_X(X) \tag{7}
$$

$$
h_{\rm BI} = \alpha h_{\rm A} + \beta h_{\rm X} \tag{8}
$$

where α and β are hyperparameters.

Model prediction

Ultimately, we combine the convolution-derived representation *h_G* with the bimodal MLP-derived representation *h_{BI}* through a linear layer to calculate the likelihood of a gene functioning as a driver gene as follows:

$$
p = \log \sigma [(h_G + h_{BI})W'] \tag{9}
$$

where *W* is a trainable matrix.

In our study, we utilize the binary cross-entropy loss to train the model:

$$
L = -\sum_{i=1}^{n} (y_i \log (p_i)) + (1 - y_i) \log (1 - p_i)
$$
 (10)

where *yi* is the true label of gene *i*, *pi* is the prediction score of gene *i*, and *n* is the number of nodes.

Our model is built using Python 3.8, PyTorch Geometric 2.0.1 [[42\]](#page-9-22) and PyTorch 1.10.1. To identify the optimal hyperparameters for SGCD, we apply Optuna [[43\]](#page-9-23) to automate hyperparameter search. This optimization is obtained using stratified five-fold cross-validation on the training set, ensuring consistent proportions of known cancer genes and non-cancer genes across all sets. In our experiments, the hidden layer dimension of GCN with RS is set to 64. To regularize the model and mitigate overfitting, a weight decay of 5 × 10[−]⁵ is applied. The learning rate is set to 0.0149. The coefficient for MLP_A is 0.0204, and for MLP_X it is 0.001. The model is trained for 30 epochs, and the number of hops is 2.

GNNExplainer

We apply GNNExplainer [\[44](#page-9-24)] to interpret key interactions for genes. For a given node *i*, GNNExplainer identifies a connected subgraph $G_{\text{Si}} \subseteq G$. It is described below:

$$
\min_{M_i, f_i} -\log P_{\Phi} \left(Y = \hat{y}_i \mid G = A_i \odot \sigma \left(M_i \right), X = \mathbf{X}_i \odot \sigma \left(f_i \right) \right) \tag{11}
$$

where *Ai* represents the adjacency matrix and *Xi* represents multiomics features. $M_i \in \mathbb{R}^{n \times n}$ denotes the mask matrix for the adjacency matrix and *fi* ∈ R*^m* denotes multi-omics features, which need to be learned for node *i*. *n* is the total number of genes, and *m* denotes the dimension of the multi-omics features. The trained GNN model is denoted by Φ . \odot stands for Hadamard product, and *σ* refers to the activation function (sigmoid).

For undirected graphs like PPI, *Mi* is preserved while optimizing. The values of *M_i* reflect the significance of the associated edges. The explanatory subgraph G_{Si} used for predicting \hat{y}_i at node *i* is determined as follows:

$$
G_{\text{Si}} = A_i \odot \mathbf{1} \{ M_i \ge \theta \} \tag{12}
$$

where *θ* stands for edge threshold.

Results

Performance assessment of SGCD

To assess the effectiveness of SGCD in cancer driver gene identification, we compare it with five methods, including the standard GNN method GCN [\[21\]](#page-9-1) and four advanced GCN-based approaches specifically designed for cancer driver gene identification, including EMOGI [[20\]](#page-9-0), MTGCN [\[22\]](#page-9-2), SMG [\[23\]](#page-9-3), and HGDC [\[27](#page-9-7)].

- GCN [\[21\]](#page-9-1) is a quintessential type of GNN that aggregates features from itself along with features from its direct neighbors. This mechanism captures the local information in the graph, allowing for richer node representations.
- EMOGI [\[20\]](#page-9-0) is an explainable method built on GCN that utilizes pan-cancer multi-omics data as gene features in combination alongside the PPI networks to generate more meaningful representations.
- • MTGCN [[22](#page-9-2)] merges biological and structural features to develop enhanced representations. It introduces a Chebnetbased multi-task framework [\[45](#page-9-25)], boosting both the main and auxiliary tasks. In addition, it includes a weight learning mechanism that dynamically adjusts the relative contributions of each task.
- SMG [[23](#page-9-3)] adopts the pretrain-finetune strategy. During the pretraining phase, it randomly masks certain nodes, and utilizes an GNN-based encoder to recover these masked nodes by referring to the information from their surrounding neighbors. During the subsequent fine-tuning stage, SMG takes advantage of the pre-trained encoder to represent the PPIs and utilizes a tailored layer to predict results for the specific tasks.

• HGDC [\[27](#page-9-7)] is designed to tackle heterophily within biological networks for the identification of driver genes. HGDC combines graph diffusion methods with hierarchical attention mechanisms. By utilizing graph diffusion to create supplementary views, HGDC significantly improves prediction accuracy across various biological networks.

To guarantee an equitable comparison, all methods utilize identical processed feature vectors and PPI networks, including CPDB [[32\]](#page-9-12), MULTINET [[33\]](#page-9-13), PCNet [[34](#page-9-14)], STRINGdb [\[35\]](#page-9-15), IRefIndex [[36](#page-9-16)], and IRefIndex-2015 [\[36](#page-9-16)]. The parameters for the baseline models are either configured according to their papers or adjusted as needed to maximize their performance. The datasets are divided into a training set (75%) and a testing set (25%), and then we train the SGCD and baseline models separately to attain optimal performance. As shown in [Fig. 2\(a\)](#page-4-0) to [Fig. 2\(f\),](#page-4-0) SGCD achieves the best AUPRC in different PPIs than other baseline models, indicating the advantage of SGCD in detecting potential cancer driver genes.

To validate the robustness of SGCD, we evaluate performance under both feature perturbation and network perturbation. Feature perturbation is implemented by randomly masking node features, where the values are set to 0, with masking rates of 0.25, 0.50, 0.75, and 0.90. Network perturbation is performed by randomly deleting edges in the network, with deletion rates set at 0.25, 0.50, 0.75, and 0.90. Then, we train SGCD and baseline models at each perturbation rate. The results shown in [Fig. 2\(g\)](#page-4-0) and [2\(h\)](#page-4-0) demonstrate that with the increasing rate of feature perturbation or network perturbation, SGCD consistently shows the best performance and experiences a relatively smaller decrease in performance compared to baseline models. These results indicate that SGCD evinces strong robustness, adapts well to real-world conditions, and consistently performs with stability and excellence.

The time overhead of an algorithm is a crucial metric for evaluating its efficiency. In this research, we record the running time of SGCD and baseline models across different PPIs. As shown in the [Fig. 2\(i\),](#page-4-0) SGCD persistently achieves minimal time overhead (log 10-transformed) across all datasets, highlighting its exceptional time efficiency.

Ablation study

To analyze the contribution of every step within SGCD model architecture, we carry out ablation experiments on MULTINET. The results of ablation experiments are shown in [Table 1](#page-4-1). We notice that the performance of merely using classic MLP architecture ($MLP_A + MLP_X$) significantly surpasses that of GCN (as shown in [Fig. 2a](#page-4-0)), which suggests that traditional convolutional methods may lead to feature confusion, resulting in even worse performance than MLP. Additionally, the comparison between GCN with RS and the GCN-based method EMOGI (as shown in [Fig. 2a](#page-4-0)) also demonstrates that feature separation can effectively reduce the confusion in node information aggregation within heterophilic networks, thereby significantly improving the performance of models in such networks. Finally, the complete framework of SGCD model shows superior performance compared to GCN with $RS + MLP_A$ or GCN with $RS + MLP_X$, indicating that SGCD successfully improves cancer driver gene identification by integrating GCN with RS and bimodal feature extractor.

Overall, the results of the ablation study confirm the effectiveness of each step of SGCD. Specifically, the results demonstrate that substituting RS for the traditional message-passing mechanism of GCN significantly reduces feature confusion, ultimately

Figure 2. Performance assessment of SGCD. (a) AUPRC performance comparison on CPDB. (b) AUPRC performance comparison on MULTINET. (c) AUPRC performance comparison on IRedfIndex. (d) AUPRC performance comparison on PCNet. (e) AUPRC performance comparison on STRINGdb. (f) AUPRC performance comparison on IRedfIndex_2015. (g) Feature robustness analysis of SGCD and baseline models. (h) Network robustness analysis of SGCD and baseline models. (i) The time overhead analysis of SGCD and baseline models.

Table 1. The ablation results of SGCD

Model	AUPRC	AUROC	F1	ACC
GCN with $RS + MLP_A$	0.7881	0.9320	0.6917	0.9015
GCN with $RS + MLP_X$	0.7560	0.9242	0.6648	0.8948
GCN with RS	0.7557	0.9242	0.6634	0.8946
$MLP_A + MLP_X$ (bimodal feature extractor)	0.6021	0.7791	0.5288	0.8755
GCN with $RS + MLP_A + MLP_X$ (SGCD)	0.7900	0.9324	0.6939	0.9021

improving the generalization ability of GNNs in heterophilic graphs. Furthermore, given the complex information between graph topology and label distribution in heterophilic graphs, the use of bimodal feature extractor can effectively extract distinct topological and omics information from these graphs, thus enabling a more optimized utilization of graph information.

Performance on independent test sets

To verify if the performance of SGCD and baseline models are biased towards any cancer-related datasets, we evaluate them on two independent datasets. We train SGCD the baseline methods using labeled samples, including both positive and negative instances. The trained models are next used to predict cancerrelated genes in two independent test sets: one consists of 320 genes from the OncoKB [\[46](#page-9-26)] and another with 388 genes from the ONGene [\[47\]](#page-9-27). After excluding genes that overlap with the training samples, we regard the predicted cancer driver genes in the test set as true positives, while genes absent from the test set are classified as false positives. Although all methods perform relatively poorly due to the insufficient number of true positives in the independent test sets, [Fig. 3](#page-5-0) shows that SGCD consistently surpasses the baseline models on both OncoKB and ONGene.

Prediction of potential cancer driver genes

We use SGCD to train and predict on six PPIs. Then, by aggregating the top 100 predicted driver genes from each PPIs, a list of 315 potential cancer driver genes is obtained, as demonstrated in [Table S1](https://academic.oup.com/bib/article-lookup/doi/10.1093/bib/bbae691#supplementary-data) in the Supplementary Materials. Comparing SGCD with

Figure 3. Performance of SGCD and baseline models across two independent sets based on OncoKB and ONGene.

Figure 4. Venn diagram of the overlap between the SGCD and other driver genes identification methods.

several other identification methods, we observe that SGCD can predict unique driver genes not observed by the other methods, as shown in [Fig. 4.](#page-5-1) This underscores the capability of SGCD to uncover overlooked driver genes, highlighting its valuable genetic insights. Among these unique genes, most of them are linked to cancer onset and development. For example, many studies have demonstrated that GNB1 is involved in the progression and drug resistance of multiple cancer types [\[48\]](#page-9-28). Extensive research has demonstrated that NR2C2 potentially function as either a tumorigenic gene or a tumor-suppressive gene, depending on the type of tumors [\[49](#page-9-29), [50\]](#page-9-30). A previous study has indicated that PPP2CA has potential to serve as a tumor suppressor gene across various cancers, with its expression potentially modulated by rs13187105 or other SNPs that exhibit strong linkage disequilibrium [[51](#page-9-31)].

To further analyze these potential cancer genes, we compare them with two sets of candidate cancer driver genes derived from sources based on published literature. The first source is the CancerMine [\[52\]](#page-9-32), a text-mined and regularly updated resource that catalogs drivers, oncogenes and tumor suppressor genes across various cancer types. The second source consists of a highly reliable gene set gathered from the Candidate Cancer Gene Database (CCGD) [\[53](#page-9-33)], encompassing all available data from transposon-based forward genetic screens related to cancer. Overall, approximately 91% (287/315) of the potential driver genes have evidences supporting their association with cancer. Furthermore, among these evidence-supported genes, over 88% (253/287) are supported by CancerMine, over 76% (220/287) are supported by CCGD and over 64% (186/287) are supported by both CancerMine and CCGD. These experimental results further substantiate the strong reliability of the cancer driver genes identified by SGCD.

Enrichment analysis

We perform enrichment analysis of Gene Ontology (GO) and KEGG pathways on the predicted cancer driver genes, and the results show that these predicted genes exhibit notable enrichment in numerous cancer-related pathways. For instance, as shown in [Fig. 5\(a\),](#page-6-0) in Go biological process enrichment, cell–cell adhesion is essential for enabling extravasation from the primary tumor and subsequent metastasis, while the loss of cell adhesion molecules is closely linked to tumor progression [[54\]](#page-9-34). As shown in [Fig. 5\(b\),](#page-6-0) in cellular component enrichment, membrane rafts, as targets for cancer treatment, significantly contribute to cell survival regulation by enhancing Akt activation. They are closely tied to their pivotal role in regulating multiple stages of malignant cell transformation, including growth, apoptosis susceptibility, invasiveness, and metastatic capacity [[55](#page-9-35)]. As shown in [Fig. 5\(c\),](#page-6-0) in GO molecular function enrichment, phosphatases act as molecular switches capable of activating or deactivating various signaling pathways, leading to abnormal cellular activities such as unchecked proliferation, differentiation, angiogenesis, and metastasis. Numerous phosphatases have been associated with the initiation and pathogenesis of various types of cancer [\[56](#page-10-0)]. As shown in [Fig. 5\(d\)](#page-6-0), in KEGG pathway enrichment, the PI3K-Akt signaling pathway, a pivotal regulator of diverse cellular functions, is frequently dysregulated in cancer, fostering tumor initiation and progression. Targeting this pathway, either as a standalone approach or in conjunction with other therapeutic modalities, has emerged as a highly efficacious strategy in the battle against cancer [[57\]](#page-10-1).

Drug sensitivity analysis

Given that drug sensitivity reveals how cancer driver genes influence cancer cell responses to specific drugs, we select the top 10 predicted cancer driver genes in each PPIs for Cancer Therapeutics Response Portal drug sensitivity analysis using Gene Set Cancer Analysis [\(http://bioinfo.life.hust.edu.cn/GSCA\)](http://bioinfo.life.hust.edu.cn/GSCA) [\[58,](#page-10-2) [59](#page-10-3)]. [Figure 6](#page-7-0) illustrates the drug sensitivity analysis results for MULTI-NET, while the results of other PPIs are present in [Fig. S1](https://academic.oup.com/bib/article-lookup/doi/10.1093/bib/bbae691#supplementary-data) in the Supplementary Materials. The results of the drug sensitivity analysis demonstrate that cancer driver genes identified by SGCD has the potential to provide important perspectives on drug targets, thereby enhancing both the effectiveness and precision of cancer treatment. As exemplified by MULTINET in [Fig. 6](#page-7-0), the majority of these genes exhibit significant correlations with drug sensitivity, highlighting their potential involvement in affecting responses to particular cancer therapies. For instance, AR-42 is an innovative histone deacetylase inhibitor, and it demonstrates antitumor effects in pancreatic cancer cells by impacting various biochemical pathways [\[60\]](#page-10-4). PIK-93 promotes a treatment-friendly tumor microenvironment when used in conjunction with anti– PD-L1 antibodies, thus improving the effectiveness of PD-1/PD-L1 blockade cancer immunotherapy [[61](#page-10-5)]. FK866, by blocking NAMPTdriven NAD+ production, can reduce the activation and stemness of CAFs, diminish the release of inflammatory cytokines and

Figure 5. Enrichment analysis. (a) Go biological process enrichment. (b) GO cellular component enrichment. (c) Go molecular function enrichment. (d) KEGG pathway enrichment.

chemokines by suppressing PITX3 expression, and thereby inhibit colorectal cancer metastasis [[62](#page-10-6)].

Gene module dissection in pan-cancer

We utilize the GNNExplainer [[44](#page-9-24)] to elucidate the influential factors associated with cancer driver genes across multi-omics data, subsequently detecting the cancer gene modules. GNNExplainer [[44](#page-9-24)] utilizes a masking strategy to maximize the mutual information between the predictions and the distribution of potential subgraph structures, thereby identifying the most compact gene module.

Firstly, we compare the topological characteristics of cancer gene modules with non-cancer gene modules using graphical metrics, such as PageRank, clustering coefficient, degree centrality, and betweenness centrality (see Supplementary Materials [Section 5](https://academic.oup.com/bib/article-lookup/doi/10.1093/bib/bbae691#supplementary-data) for details). These modules are obtained by separately applying GNNExplainer to the predicted cancer driver genes and the non-cancer driver genes. The results of MULTINET are shown in [Fig. 7](#page-7-1), which demonstrate that the topological structures of cancer gene modules exhibit significantly greater consistency compared to those of non-cancer gene modules (*P <* 9.52e-25, *t*-test). The results of other PPIs can be found in the [Fig. S2](https://academic.oup.com/bib/article-lookup/doi/10.1093/bib/bbae691#supplementary-data) in the Supplementary Materials.

Furthermore, we analyze the relationship between different gene modules. In lung cancer, MET and ALK are the most commonly encountered driver genes [\[63\]](#page-10-7), but they have different carcinogenic mechanisms. MET gene abnormalities primarily include MET exon 14 mutations, MET amplification, MET gene fusions, and MET protein overexpression. These alterations can result in abnormal activation of the MET signaling pathway, which, in turn, can drive tumor development and progression [[64\]](#page-10-8). By contrast, the majority of mutations in the ALK gene occur as translocations with a partner gene, resulting in a fusion oncogene, which is subsequently overexpressed in cancers [[65\]](#page-10-9). The gene modules of MET and ALK identified by our method are shown in [Fig. 7\(b\)](#page-7-1) and [7\(c\).](#page-7-1) Moreover, we notice that the MET gene module and the ALK gene module share three common genes: STAT3, SHC1, and GRB2, which combine the two gene modules into a high-order gene module as shown in [Fig. 7\(d\).](#page-7-1) It is worth noting that among these genes, STAT3 is the known cancer driver gene, while SHC1 and GRB2 are cancer driver genes predicted by SGCD (ranked in the top 50 predicted cancer driver genes of MULTINET). The research has shown that cancer driver genes are often found together in the same modules instead of being randomly scattered [[66\]](#page-10-10), and this suggests that the identified cancer genes SHC1 and GRB2 are promising candidates for cancer driver genes.

Figure 6. Correlation between drug sensitivity and mRNA expression for the top 10 predicted cancer driver genes.

Figure 7. Cancer gene module analysis on SGCD. (a) Graphical metrics of gene modules. (b) The structure of MET gene module. (c) The structure of ALK gene module. (d) high-order gene module of MET and ALK.

Conclusion

Nowadays, cancer is one of the major threats to human health, and its underlying mechanisms are complex. It is widely accepted in the biomedical field that cancer emerges due to the accumulation of mutations across various genes. Therefore, cancer driver gene identification is important for uncovering the processes of cancer initiation and progression.

In this research, we present an innovative model, SGCD, which employs simplified GNNs to identify cancer driver genes in heterophilic networks. A key innovation of SGCD is the introduction of the GCN with RS module, where RS replaces the traditional message-passing mechanism, effectively mitigating potential feature confusion issues inherent in conventional GNNs. Additionally, SGCD utilizes a bimodal feature extractor to capture both

topological and omics information, thereby enhancing identification performance. The experimental findings indicate that SGCD surpasses the state-of-the-art approaches, strengthening the predictive accuracy and robustness of the model. Furthermore, subsequent model interpretability experiments and biological interpretability experiments reveal that the potential cancer driver genes identified by SGCD are closely associated with cancer, validating the strong interpretability of SGCD. In addition, the model is capable of dissecting gene modules, providing deeper insights into the relationships between genes and their roles in cancer. We believe that SGCD is a general method, offering novel perspectives on the identification of cancer driver genes and allowing its application beyond the field of cancer genomics to other complex diseases.

Key Points

- An efficient method SGCD is developed for identifying cancer driver genes in heterophilic network by utilizing simplified graph neural networks.
- SGCD introduces representation separation to replace the traditional message-passing mechanism in GCN, significantly mitigating potential feature confusion.
- The computation experiments show the superiority of SGCD compared to other baseline models.
- The biological interpretability experiments provide compelling evidence supporting the reliability of SGCD.
- SGCD can dissect gene modules, revealing clearer connections between driver genes in cancers.

Supplementary data

[Supplementary data](https://academic.oup.com/bib/article-lookup/doi/10.1093/bib/bbae691#supplementary-data) is available at *Briefings in Bioinformatics* online.

Conflict of interest: None declared.

Funding

This work is supported in part by the National Natural Science Foundation of China [62202383, 62433016], Guangdong Basic and Applied Basic Research Foundation [2024A1515012602], the National Key Research and Development Program of China [2022YFD1801200], the State Key Laboratory for Animal Disease Control and Prevention Foundation [SKLADCPKFKT202407], the Macau Young Scholars Program [AM2024027], and the Science and Technology Development Fund of Macao [0002/2024/RIA1].

Data available

All data is publicly available and the code of SGCD can be freely downloaded from [https://github.com/xingyili/SGCD.](https://github.com/xingyili/SGCD)

References

- [1.](#page-0-9) Alexandrov LB, Nik-Zainal S, Wedge DC. *et al*. Signatures of mutational processes in human cancer. *Nature* 2013;**500**:415–21. <https://doi.org/10.1038/nature12477>.
- [2.](#page-0-10) Martincorena I, Campbell PJ. Somatic mutation in cancer and normal cells. *Science* 2015;**349**:1483–9. [https://doi.org/10.1126/](https://doi.org/10.1126/science.aab4082) [science.aab4082.](https://doi.org/10.1126/science.aab4082)
- [3.](#page-0-11) Dees ND, Zhang Q, Kandoth C. *et al*. MuSic: identifying mutational significance in cancer genomes. *Genome Res* 2012;**22**: 1589–98. <https://doi.org/10.1101/gr.134635.111>.
- [4.](#page-0-12) Lawrence MS, Stojanov P, Polak P. *et al*. Mutational heterogeneity in cancer and the search for new cancer-associated genes. *Nature* 2013;**499**:214–8. [https://doi.org/10.1038/nature12213.](https://doi.org/10.1038/nature12213)
- [5.](#page-0-13) Tamborero D, Gonzalez-Perez A, Lopez-Bigas N. Oncodrive CLUST: exploiting the positional clustering of somatic mutations to identify cancer genes. *Bioinformatics* 2013;**29**:2238–44. [https://doi.org/10.1093/bioinformatics/btt395.](https://doi.org/10.1093/bioinformatics/btt395)
- [6.](#page-0-14) Weinstein JN, Collisson EA, Mills GB. *et al*. The cancer genome atlas pan-cancer analysis project. *Nat Genet* 2013;**45**:1113–20. <https://doi.org/10.1038/ng.2764>.
- 7. Zhang J, Bajari R, Andric D. *et al*. The International Cancer Genome Consortium Data Portal. *Nat Biotechnol* 2019;**37**:367–9. [https://doi.org/10.1038/s41587-019-0055-9.](https://doi.org/10.1038/s41587-019-0055-9)
- [8.](#page-0-15) Tate JG, Bamford S, Jubb HC. *et al*. COSMIC: the catalogue of somatic mutations in cancer. *Nucleic Acids Res* 2019;**47**:D941–7. [https://doi.org/10.1093/nar/gky1015.](https://doi.org/10.1093/nar/gky1015)
- [9.](#page-0-16) Collier O, Stoven V, Vert J-P. Lotus: a single-and multitask machine learning algorithm for the prediction of cancer driver genes. *PLoS Comput Biol* 2019;**15**:e1007381. [https://doi.](https://doi.org/10.1371/journal.pcbi.1007381) [org/10.1371/journal.pcbi.1007381](https://doi.org/10.1371/journal.pcbi.1007381).
- [10.](#page-0-17) Davoli T, Andrew Wei X, Mengwasser KE. *et al*. Cumulative haploinsufficiency and triplosensitivity drive aneuploidy patterns and shape the cancer genome. *Cell* 2013;**155**:948–62. [https://doi.](https://doi.org/10.1016/j.cell.2013.10.011) [org/10.1016/j.cell.2013.10.011](https://doi.org/10.1016/j.cell.2013.10.011).
- [11.](#page-0-18) Mourikis TP, Benedetti L, Foxall E. *et al*. Patient-specific cancer genes contribute to recurrently perturbed pathways and establish therapeutic vulnerabilities in esophageal adenocarcinoma. *Nat Commun* 2019;**10**:3101. [https://doi.org/10.1038/](https://doi.org/10.1038/s41467-019-10898-3) [s41467-019-10898-3](https://doi.org/10.1038/s41467-019-10898-3).
- [12.](#page-0-19) Zhang T, Zhang S-W, Li Y. Identifying driver genes for individual patients through inductive matrix completion. *Bioinformatics* 2021;**37**:4477–84. [https://doi.org/10.1093/bioinformatics/](https://doi.org/10.1093/bioinformatics/btab477) [btab477.](https://doi.org/10.1093/bioinformatics/btab477)
- [13.](#page-0-20) Jiang R. Walking on multiple disease-gene networks to prioritize candidate genes. *J Mol Cell Biol* 2015;**7**:214–30. [https://doi.](https://doi.org/10.1093/jmcb/mjv008) [org/10.1093/jmcb/mjv008.](https://doi.org/10.1093/jmcb/mjv008)
- [14.](#page-0-21) Li Y, Patra JC. Genome-wide inferring gene–phenotype relationship by walking on the heterogeneous network. *Bioinformatics* 2010;**26**:1219–24. [https://doi.org/10.1093/bioinformatics/](https://doi.org/10.1093/bioinformatics/btq108) [btq108](https://doi.org/10.1093/bioinformatics/btq108).
- [15.](#page-0-22) Xie M, Hwang T, Kuang R. Prioritizing disease genes by birandom walk. In: *Advances in Knowledge Discovery and Data Mining: 16th Pacific-Asia Conference, PAKDD 2012, Kuala Lumpur, Malaysia, May 29–June 1, 2012, Proceedings, Part II 16*, pp. 292–303. Berlin, Heidelberg: Springer Berlin Heidelberg, 2012.
- [16.](#page-0-23) Zhao W, Xun G, Chen S. *et al*. MODIG: integrating multi-omics and multi-dimensional gene network for cancer driver gene identification based on graph attention network model. *Bioinformatics* 2022;**38**:4901–7. [https://doi.org/10.1093/bioinformatics/](https://doi.org/10.1093/bioinformatics/btac622) [btac622](https://doi.org/10.1093/bioinformatics/btac622).
- 17. Li X, Li J, Hao J. *et al*. Multiplex networks and pan-cancer multiomics-based driver gene identification using graph neural networks. *Big Data Min Anal* 2024;**7**:1262–72. [https://doi.](https://doi.org/10.26599/BDMA.2024.9020043) [org/10.26599/BDMA.2024.9020043](https://doi.org/10.26599/BDMA.2024.9020043).
- 18. Yang J, Haitao F, Xue F. *et al*. Multiview representation learning for identification of novel cancer genes and their causative biological mechanisms. *Brief Bioinform* 2024;**25**:bbae418. [https://](https://doi.org/10.1093/bib/bbae418) doi.org/10.1093/bib/bbae418.
- [19.](#page-0-24) Peng W, Zhou Z, Dai W. *et al*. Multi-network graph contrastive learning for cancer driver gene identification. *IEEE*

Trans Netw Sci Eng 2024;**11**:3430–40. [https://doi.org/10.1109/](https://doi.org/10.1109/TNSE.2024.3373652) [TNSE.2024.3373652](https://doi.org/10.1109/TNSE.2024.3373652).

- [20.](#page-0-25) Schulte-Sasse R, Budach S, Hnisz D. *et al*. Integration of multiomics data with graph convolutional networks to identify new cancer genes and their associated molecular mechanisms. *Nat Mach Intell* 2021;**3**:513–26. [https://doi.org/10.1038/](https://doi.org/10.1038/s42256-021-00325-y) [s42256-021-00325-y.](https://doi.org/10.1038/s42256-021-00325-y)
- [21.](#page-0-26) Kipf TN, Welling M. Semi-supervised classification with graph convolutional networks. arXiv preprint arXiv:1609.02907. 2016. [https://doi.org/10.48550/arXiv.1609.02907.](https://doi.org/10.48550/arXiv.1609.02907)
- [22.](#page-1-0) Peng W, Tang Q, Dai W. *et al*. Improving cancer driver gene identification using multi-task learning on graph convolutional network. *Brief Bioinform* 2022;**23**:bbab432. [https://doi.org/10.1093/](https://doi.org/10.1093/bib/bbab432) [bib/bbab432.](https://doi.org/10.1093/bib/bbab432)
- [23.](#page-1-1) Cui Y, Wang Z, Wang X. *et al*. SMG: self-supervised masked graph learning for cancer gene identification. *Brief Bioinform* 2023;**24**:bbad406. <https://doi.org/10.1093/bib/bbad406>.
- [24.](#page-1-2) Li Y, Lin B, Luo B. *et al*. Graph representation learning beyond node and homophily. *IEEE Trans Knowl Data Eng* 2022;**35**:4880–93.
- [25.](#page-1-3) Chanpuriya S, Rossi R, Rao AB. *et al*. Exact representation of sparse networks with symmetric nonnegative embeddings. In: *Advances in Neural Information Processing Systems*, pp. 21023–38. New Orleans, LA, USA: Curran Associates, Inc., 2023.
- [26.](#page-1-4) Sondka Z, Bamford S, Cole CG. *et al*. The cosmic cancer gene census: Describing genetic dysfunction across all human cancers. *Nat Rev Cancer* 2018;**18**:696–705. [https://doi.org/10.1038/](https://doi.org/10.1038/s41568-018-0060-1) [s41568-018-0060-1.](https://doi.org/10.1038/s41568-018-0060-1)
- [27.](#page-1-5) Zhang T, Zhang S-W, Xie M-Y. *et al*. A novel heterophilic graph diffusion convolutional network for identifying cancer driver genes. *Brief Bioinform* 2023;**24**:bbad137. [https://doi.org/10.1093/](https://doi.org/10.1093/bib/bbad137) [bib/bbad137.](https://doi.org/10.1093/bib/bbad137)
- [28.](#page-1-6) Repana D, Nulsen J, Dressler L. *et al*. The Network of Cancer Genes (NCG): a comprehensive catalogue of known and candidate cancer genes from cancer sequencing screens. *Genome Biol* 2019;**20**:1–12. [https://doi.org/10.1186/s13059-018-1612-0.](https://doi.org/10.1186/s13059-018-1612-0)
- [29.](#page-1-7) Kim J, So S, Lee H-J. *et al*. DigSee: disease gene search engine with evidence sentences (version cancer). *Nucleic Acids Res* 2013;**41**:W510–7. [https://doi.org/10.1093/nar/gkt531.](https://doi.org/10.1093/nar/gkt531)
- [30.](#page-1-8) McKusick VA. Mendelian inheritance in man and its online version, OMIM. *Am J Hum Genet* 2007;**80**:588–604. [https://doi.](https://doi.org/10.1086/514346) [org/10.1086/514346.](https://doi.org/10.1086/514346)
- [31.](#page-1-9) Kanehisa M, Goto S. KEGG: Kyoto Encyclopedia of Genes and Genomes. *Nucleic Acids Res* 2000;**28**:27–30. [https://doi.](https://doi.org/10.1093/nar/28.1.27) [org/10.1093/nar/28.1.27](https://doi.org/10.1093/nar/28.1.27).
- [32.](#page-1-10) Kamburov A, Wierling C, Lehrach H. *et al*. ConsensusPathDB a database for integrating human functional interaction networks. *Nucleic Acids Res* 2009;**37**:D623–8. [https://doi.org/10.1093/](https://doi.org/10.1093/nar/gkn698) [nar/gkn698](https://doi.org/10.1093/nar/gkn698).
- [33.](#page-1-11) Khurana E, Yao F, Chen J. *et al*. Interpretation of genomic variants using a unified biological network approach. *PLoS Comput Biol* 2013;**9**:e1002886. <https://doi.org/10.1371/journal.pcbi.1002886>.
- [34.](#page-1-12) Huang JK, Carlin DE, Yu MK. *et al*. Systematic evaluation of molecular networks for discovery of disease genes. *Cell Syst* 2018;**6**:484–495.e5. <https://doi.org/10.1016/j.cels.2018.03.001>.
- [35.](#page-1-13) Szklarczyk D, Gable AL, Nastou KC. *et al*. The string database in 2021: customizable protein–protein networks, and functional characterization of user-uploaded gene/measurement sets. *Nucleic Acids Res* 2021;**49**:D605–12. [https://doi.org/10.1093/](https://doi.org/10.1093/nar/gkaa1074) [nar/gkaa1074](https://doi.org/10.1093/nar/gkaa1074).
- [36.](#page-1-14) Razick S, Magklaras G, Donaldson IM. iRefIndex: a consolidated protein interaction database with provenance. *BMC Bioinform* 2008;**9**:1–19.
- [37.](#page-1-15) Zhu J, Yan Y, Zhao L. *et al*. Beyond homophily in graph neural networks: current limitations and effective designs. *Adv Neural Inf Process Syst* 2020;**33**:7793–804.
- [38.](#page-1-16) Luan S, Hua C, Qincheng L. *et al*. Revisiting heterophily for graph neural networks. *Adv Neural Inf Process Syst* 2022;**35**: 1362–75.
- [39.](#page-2-1) Lee SY, Fanchen B, Yoo J. *et al*. Towards deep attention in graph neural networks: Problems and remedies. In: *International Conference on Machine Learning*, pp. 18774–95. Honolulu, Hawaii, USA: PMLR, 2023.
- [40.](#page-2-2) Cavallo A, Grohnfeldt C, Russo M. *et al*. 2-hop neighbor class similarity (2NCS): a graph structural metric indicative of graph neural network performance. arXiv preprint arXiv:2212.13202. 2022. [https://doi.org/10.48550/arXiv.2212.13202.](https://doi.org/10.48550/arXiv.2212.13202)
- [41.](#page-2-3) Bechler-Speicher M, Amos I, Gilad-Bachrach R. *et al*. Graph neural networks use graphs when they shouldn't. arXiv preprint arXiv:2309.04332. 2023. [https://doi.org/10.48550/arXiv.2309.](https://doi.org/10.48550/arXiv.2309.04332) 04332.
- [42.](#page-2-4) Fey M, Lenssen JE. Fast graph representation learning with pytorch geometric. arXiv preprint arXiv:1903.02428. 2019. <https://doi.org/10.48550/arXiv.1903.02428>.
- [43.](#page-2-5) Akiba T, Sano S, Yanase T. *et al*. Optuna: a next-generation hyperparameter optimization framework. In: *Proceedings of the 25th ACM SIGKDD International Conference on Knowledge Discovery and Data Mining*, pp. 2623–31. Anchorage, AK, USA: ACM, 2019.
- [44.](#page-3-0) Ying Z, Bourgeois D, You J. *et al*. GNNExplainer: generating explanations for graph neural networks. *Adv Neural Inf Process Syst* 2019;**32**:9240–51.
- [45.](#page-3-1) Defferrard M, Bresson X, Vandergheynst P. Convolutional neural networks on graphs with fast localized spectral filtering. *Adv Neural Inf Process Syst* 2016;**29**:1–9.
- [46.](#page-4-2) Chakravarty D, Gao J, Phillips S. *et al*. OncoKB: a precision oncology knowledge base. *JCO Precis Oncol* 2017;**1**:1–16.
- [47.](#page-4-3) Liu Y, Sun J, Zhao M. ONGene: a literature-based database for human oncogenes. *J Genet Genomics* 2017;**44**:119–21. [https://doi.](https://doi.org/10.1016/j.jgg.2016.12.004) [org/10.1016/j.jgg.2016.12.004.](https://doi.org/10.1016/j.jgg.2016.12.004)
- [48.](#page-5-2) Zhang X, Dong K, Zhang J. *et al*. GNB1 promotes hepatocellular carcinoma progression by targeting BAG2 to activate P38/MAPK signaling. *Cancer Sci* 2023;**114**:2001–13. [https://doi.org/10.1111/](https://doi.org/10.1111/cas.15741) [cas.15741](https://doi.org/10.1111/cas.15741).
- [49.](#page-5-3) Fan Z, Zheng J, Xue Y. *et al*. NR2C2-UORF targeting UCA1 miR-627-5P-NR2C2 feedback loop to regulate the malignant behaviors of glioma cells. *Cell Death Dis* 2018;**9**:1165. [https://doi.](https://doi.org/10.1038/s41419-018-1149-x) [org/10.1038/s41419-018-1149-x](https://doi.org/10.1038/s41419-018-1149-x).
- [50.](#page-5-4) Xie S, Yaru H, Jin J. *et al*. Regulation of the stem-like properties of estrogen receptor-positive breast cancer cells through NR2E3/NR2C2 signaling. *Exp Ther Med* 2023;**26**:1–11. [https://doi.](https://doi.org/10.3892/etm.2023.12173) [org/10.3892/etm.2023.12173](https://doi.org/10.3892/etm.2023.12173).
- [51.](#page-5-5) Huang T, He K, Mao Y. *et al*. Genetic variants in PPP2CA are associated with gastric cancer risk in a chinese population. *Sci Rep* 2017;**7**:11499. <https://doi.org/10.1038/s41598-017-12040-z>.
- [52.](#page-5-6) Lever J, Zhao EY, Grewal J. *et al*. CancerMine: a literaturemined resource for drivers, oncogenes and tumor suppressors in cancer. *Nat Methods* 2019;**16**:505–7. [https://doi.org/10.1038/](https://doi.org/10.1038/s41592-019-0422-y) [s41592-019-0422-y.](https://doi.org/10.1038/s41592-019-0422-y)
- [53.](#page-5-7) Abbott KL, Nyre ET, Abrahante J. *et al*. The candidate cancer gene database: a database of cancer driver genes from forward genetic screens in mice. *Nucleic Acids Res* 2015;**43**:D844–8. <https://doi.org/10.1093/nar/gku770>.
- [54.](#page-5-8) Cavallaro U, Christofori G. Cell adhesion and signalling by cadherins and Ig-CAMs in cancer. *Nat Rev Cancer* 2004;**4**:118–32. [https://doi.org/10.1038/nrc1276.](https://doi.org/10.1038/nrc1276)
- [55.](#page-5-9) Hryniewicz-Jankowska A, Augoff K, Biernatowska A. *et al*. Membrane rafts as a novel target in cancer therapy. *Biochim Biophys Acta-Rev Cancer* 2014;**1845**:155–65. [https://doi.org/10.1016/](https://doi.org/10.1016/j.bbcan.2014.01.006) [j.bbcan.2014.01.006.](https://doi.org/10.1016/j.bbcan.2014.01.006)
- [56.](#page-5-10) Stebbing J, Lit LC, Zhang H. *et al*. The regulatory roles of phosphatases in cancer. *Oncogene* 2014;**33**:939–53. [https://doi.](https://doi.org/10.1038/onc.2013.80) [org/10.1038/onc.2013.80](https://doi.org/10.1038/onc.2013.80).
- [57.](#page-5-11) He Y, Sun MM, Zhang GG. *et al*. Targeting PI3K/AKT signal transduction for cancer therapy. *Signal Transduct Target Ther* 2021;**6**:425. <https://doi.org/10.1038/s41392-021-00828-5>.
- [58.](#page-5-12) Liu C-J, Fei-Fei H, Xia M-X. *et al*. GSCALite: a web server for gene set cancer analysis. *Bioinformatics* 2018;**34**:3771–2. [https://](https://doi.org/10.1093/bioinformatics/bty411) [doi.org/10.1093/bioinformatics/bty411.](https://doi.org/10.1093/bioinformatics/bty411)
- [59.](#page-5-13) Liu C-J, Fei-Fei H, Xie G-Y. *et al*. GSCA: an integrated platform for gene set cancer analysis at genomic, pharmacogenomic and immunogenomic levels. *Brief Bioinform* 2023;**24**:bbac558. [https://](https://doi.org/10.1093/bib/bbac558) doi.org/10.1093/bib/bbac558.
- [60.](#page-5-14) Chen Y-J, Wang W-H, Wan-Yu W. *et al*. Novel histone deacetylase inhibitor AR-42 exhibits antitumor activity in pancreatic cancer cells by affecting multiple biochemical pathways. *PloS One* 2017;**12**:e0183368. [https://doi.org/10.1371/](https://doi.org/10.1371/journal.pone.0183368) [journal.pone.0183368.](https://doi.org/10.1371/journal.pone.0183368)
- [61.](#page-5-15) Lin C-Y, Huang K-Y, Kao S-H. *et al*. Small-molecule PIK-93 modulates the tumor microenvironment to improve immune checkpoint blockade response. *Sci Adv* 2023;**9**:eade9944. [https://](https://doi.org/10.1126/sciadv.ade9944) doi.org/10.1126/sciadv.ade9944.
- [62.](#page-6-1) Xie H, Lei Y, Mao Y. *et al*. FK866 inhibits colorectal cancer metastasis by reducing NAD+ levels in cancer-associated fibroblasts. *Genes & Genomics* 2022;**44**:1531–41. [https://doi.org/10.1007/](https://doi.org/10.1007/s13258-022-01318-w) [s13258-022-01318-w](https://doi.org/10.1007/s13258-022-01318-w).
- [63.](#page-6-2) Korpanty GJ, Graham DM, Vincent MD. *et al*. Biomarkers that currently affect clinical practice in lung cancer: EGFR, ALK, MET, ROS-1, and KRAS. *Front Oncol* 2014;**4**:204.
- [64.](#page-6-3) Drilon A, Cappuzzo F, Sai-Hong Ignatius O. *et al*. Targeting met in lung cancer: will expectations finally be met? *J Thorac Oncol* 2017;**12**:15–26. [https://doi.org/10.1016/j.jtho.2016.10.](https://doi.org/10.1016/j.jtho.2016.10.014) 014.
- [65.](#page-6-4) Xue D, Shao Y, Qin H-F. *et al*. ALK -rearrangement in non-smallcell lung cancer (NSCLC). *Thoracic cancer* 2018;**9**:423–30. [https://](https://doi.org/10.1111/1759-7714.12613) doi.org/10.1111/1759-7714.12613.
- [66.](#page-6-5) Li H, Zebei Han Y, Wang SF. *et al*. CGMega: explainable graph neural network framework with attention mechanisms for cancer gene module dissection. *Nat Commun* 2024;**15**:5997. [https://](https://doi.org/10.1038/s41467-024-50426-6) [doi.org/10.1038/s41467-024-50426-6.](https://doi.org/10.1038/s41467-024-50426-6)