Brain maps of general cognitive function and spatial correlations with neurobiological cortical profiles

Joanna E. Moodie[⊠]1,², Colin Buchanan^{1,2}, Anna Furtjes¹, Eleanor Conole¹, Aleks Stolicyn³, Janie Corley¹, Karen Ferguson^{1,3}, Maria Valdes Hernandez^{3,7}, Susana Munoz Maniega^{1,3}, Tom C. Russ^{1,3,4,5}, Michelle Luciano⁸, Heather Whalley³, Mark E. Bastin¹, Joanna Wardlaw^{3,6,7}, Ian Deary¹ and Simon Cox[⊠]1,2.

- ¹ Lothian Birth Cohorts, Department of Psychology, The University of Edinburgh, UK,
- ² Scottish Imaging Network, A Platform for Scientific Excellence (SINAPSE) Collaboration,

Edinburgh, UK

- ³ Centre for Clinical Brain Sciences, University of Edinburgh, UK
- ⁴Alzheimer Scotland Dementia Research Centre, University of Edinburgh, UK
- ⁵ Dementia Network, NHS Research Scotland
- ⁶ UK Dementia Research Institute
- ⁷ Row Fogo Centre for Research into Small Vessel Diseases
- ⁸Department of Psychology, University of Edinburgh, UK

[™] Corresponding authors:

Joanna E. Moodie, Lothian Birth Cohorts, Department of Psychology, The University of Edinburgh, Edinburgh EH8 9JZ, UK. Email: Joanna.moodie@ed.ac.uk

Simon R. Cox, Lothian Birth Cohorts, Department of Psychology, The University of Edinburgh, Edinburgh EH8 9JZ, UK. Email: simon.cox@ed.ac.uk.

Conflicts of interest:

None

CRediT statement

Joanna E. Moodie: Conceptualization, Methodology, Writing - Original Draft, Formal analysis, Visualization; Colin Buchanan: Writing - Review & Editing; Eleanor Conole: Writing - Review & Editing; Anna Furtjes: Writing - Review & Editing; Aleks Stolicyn: Data Curation, Writing - Review & Editing; Janie Corley: Writing - Data Curation, Review & Editing; Karen Ferguson: Writing - Review & Editing; Maria Valdes Hernandez: Data Curation, Writing -Review & Editing; Susana Munoz Maniega: Data Curation, Writing - Review & Editing; Tom C. Russ: Writing - Review & Editing; Michelle Luciano: Data Curation, Writing - Review & Editing; Heather Whalley: Data Curation, Funding Acquisition, Writing - Review & Editing; Mark E. Bastin: Data Curation, Funding Acquisition, Writing - Review & Editing; Joanna Wardlaw: Data Curation, Funding Acquisition, Writing - Review & Editing; Ian Deary: Funding Acquisition, Writing - Review & Editing; Simon Cox: Conceptualization, Data Curation, Project Administration, Resources, Funding Acquisition, Methodology, Writing -Original Draft, Supervision

Acknowledgements

We thank the participants of the three cohorts (UKB, Generation Scotland and LBC1936) for their participation and the research teams for their work in collecting, processing and giving access to these data for analysis. The UKB research was conducted using the UK Biobank Resource under Application number 10279. We are also thankful to the participants of all the studies involved in generating the neurobiological profiles, and to the people who collected and processed the data and made it openly available. SRC and JEM were supported by a Sir Henry Dale Fellowship, jointly funded by the Wellcome Trust and the Royal Society (221890/Z/20/Z). The LBC1936, supported by the BBSRC & ESRC (BB/W008793/1) (which also supports SMM, JC, and IJD), Age UK (Disconnected Mind project), the Medical Research Council (MR/M01311/1; MR/K026992/1), the US National Institutes of Health (R01AG054628) and the University of Edinburgh. CRB, MEB, IJD and SRC were supported by a National Institutes of Health (NIH) research grant R01AG054628. AF is supported by National Institutes of Health (NIH) grant R01AG073593. TCR is a member of the Alzheimer Scotland Dementia Research Centre funded by Alzheimer Scotland. MCVH and JW are funded by The Row Fogo Charitable Trust Centre for Research into Aging and the Brain (BRO-D.FID3668413). JW is also funded by the UK Dementia Research Institute which receives its funding from UK DRI, funded by the UK Medical Research Council, Alzheimer's Society, and Alzheimer's Research UK. AS was funded as part of the Generation Scotland study (Wellcome Trust reference 104036/Z/14/Z), which is led by HW and AM. ELSC is Junior Research Fellow in Applied AI at Lady Margeret Hall, University of Oxford (EPT-AI). For the purpose of open access, the author has applied a CC-BY public copyright licence to any Author Accepted Manuscript version arising from this submission.

Abstract

In this paper, we attempt to answer two questions: 1) which regions of the human brain, in terms of morphometry, are most strongly related to individual differences in domaingeneral cognitive functioning (q)? and 2) what are the underlying neurobiological properties of those regions? We meta-analyse vertex-wise *g*-cortical morphometry (volume, surface area, thickness, curvature and sulcal depth) associations using data from 3 cohorts: the UK Biobank (UKB), Generation Scotland (GenScot), and the Lothian Birth Cohort 1936 (LBC1936), with the meta-analytic N = 38,379 (age range = 44 to 84 years old). These *g*-morphometry associations vary in magnitude and direction across the cortex ($|\beta|$ range = -0.12 to 0.17 across morphometry measures) and show good cross-cohort agreement (mean spatial correlation r = 0.57, SD = 0.18). Then, to address (2), we bring together existing - and derive new - cortical maps of 33 neurobiological characteristics from multiple modalities (including neurotransmitter receptor densities, gene expression, functional connectivity, metabolism, and cytoarchitectural similarity). We discover that these 33 profiles spatially covary along four major dimensions of cortical organisation (accounting for 65.9% of the variance) and denote aspects of neurobiological scaffolding that underpin the spatial patterning of MRI-cognitive associations we observe (significant |r| range = 0.21 to 0.56). Alongside the cortical maps from these analyses, which we make openly accessible, we provide a compendium of cortex-wide and within-region spatial correlations among general and specific facets of brain cortical organisation and higher order cognitive functioning, which we hope will serve as a framework for analysing other aspects of behaviour-brain MRI associations.

1 Introduction

1 Individual differences in human cognitive function have well-established though modest 2 associations with individual differences in brain structure. For example, larger total brain 3 volumes are reliably associated with higher general cognitive function (g) scores (e.g., 1, N 4 = 18,363, r = 0.275, 95% C.I. = [0.252, 0.299]). The strength of associations between g and 5 brain volume varies by brain region $(^{2,1,3})$, and brain-cognition associations also vary by region for other morphometry measures, such as surface area (4,5), cortical thickness (6,5), 6 7 curvature (7) and sulcal depth (8, 9). The parieto-frontal integration theory (P-FIT, 2) 8 provides a theoretical basis for the involvement of parieto-frontal brain regions over others 9 in cognition, and there have been expansions and additions to that framework (e.g., ¹⁰, ⁹², 10 ¹¹). However, explanations of what regional morphometry-phenotypic association patterns 11 tell us are far from complete. Interpretations are complicated because measures of 12 morphometry from brain MRI are a conflation of multifarious underlying biological 13 properties which also vary by brain region. Thus, in the current paper, we aim to 14 characterise the spatial concordance between two types of brain map, i.e., 1) q-15 morphometry associations and 2) neurobiological profiles. We argue that this could help to 16 decode the neurobiological principles of cortical organisation that facilitate our complex 17 cognitive skills. Formally quantifying that spatial concordance, in turn, might further inform 18 a mechanistic understanding of how cognitive functioning differs between individuals in 19 health and dysfunction.

20 Until recently, inferences about the underlying biology of brain morphometry-behaviour 21 associations have been predominantly descriptive or indirect, reliant upon findings from 22 unrelated studies to draw together narrative conclusions. This is mainly due to practical 23 limitations in directly relating in vivo MRI findings to information taken postmortem, 24 limitations in the number of biological properties that can be measured in the same 25 individuals, and generally low participant numbers in instances that combine imaging and 26 post-mortem work. However, group-level summary data brain maps for several 27 neurobiological measures are increasingly being made open-source (¹², ¹³), and can now be 28 straightforwardly registered to the same common brain space as association maps (1^2) , 29 allowing for direct quantitative comparisons. Royer et al. (2024) provide a detailed perspective paper discussing the recent rise in the creation and use of cortical profiles to 30 31 make discoveries about brain organisation (14). A landmark study tested spatial associations 32 between neurotransmitter receptor distributions and cortical patterns from case/control analyses of 13 disorders, including depression, obsessive-compulsive disorder, 33

schizophrenia and Parkinson's disease; and identified spatial co-patterning between
neurotransmitter receptors and functional imaging significance patterns derived from
Neurosynth for a general factor of cognitive terms (including terms such as attention, stress,
and planning) (¹⁵).

38 Brain structural differences related to general cognitive functioning have been robustly 39 established and have wide-reaching associations with important life outcomes including everyday function, health, illness, dementia, and death (^{16,17,18}). There are increasingly 40 41 robust analyses which have established cognitive and brain structural associations (e.g., ³, 42 ¹⁹, ²⁰, ²¹), yet there remain no large-scale meta-analytic estimates of general cognitive-MRI 43 associations at the level of the cortical vertex. The measure of general cognitive function, as 44 a principal component or latent factor 'q', offers several relevant properties that make it a 45 behavioural measure of suitable quality for such analyses. It captures the general tendency 46 for cognitive test scores to be positively correlated, and is somewhat invariant to cognitive 47 test content, provided that multiple cognitive domains are captured (²², ²³, ²⁴). It is one of the most replicated phenomena in psychological science (25,26); and its individual 48 49 differences tend to be quite highly stable across the healthy lifespan (²⁷). We bring together 50 *q*-brain structure associations with biological cortical profiles to allow direct (quantitative) 51 inferences about the organising principles of the brain that underlie the cognitive-MRI 52 signals which we observe. Moreover, we produce an extension to prior analytic approaches 53 whereby we go beyond cortex-level spatial correlations (e.g., ¹⁵, ²⁸, ²⁹, ³⁰, ³¹), to additionally include regional-level spatial correlations. These regional-level spatial correlations (here, 54 55 using the Desikan-Killiany atlas, with 34 left/right paired cortical regions) provide nuanced 56 information about 1) the relative strengths of the spatial correlations in different regions 57 and 2) the homogeneity of co-patterning across regions.

58 In the current paper, we ask two main questions: 1) which regions of the human brain, in 59 terms of their morphometry, are most strongly related to individual differences in domain-60 general cognitive function? and 2) what are the underlying neurobiological properties of 61 those regions? We address these two important gaps in our knowledge by (see Figure 1), 62 first, conducting the largest vertex-wise (298,790 cortical vertices) analysis of q-cortex 63 associations across 3 cohorts with 5 morphometry measures (volume, surface area, 64 thickness, curvature, and sulcal depth) in community-dwelling adults (meta-analytic N =65 38,379). Then we quantitatively test how those brain regions that are associated with q are 66 spatially correlated with the patterning of 33 of the brain's neurobiological properties 67 across the human cerebral cortex (including neurotransmitter receptor densities, 68 cytoarchitectural, microstructural and functional connectivity similarity gradients, and

- 69 metabolism) (³²). We assemble open-source brain maps and derive novel ones; registering
- them to the same common brain space as our brain morphometry-*q* meta-analytic results;
- and then we quantitatively test their spatial concordance. Additionally, we identify four
- 72 principal components that explain the majority of the variance (65.9%) across the 33 maps
- 73 of the brain's neurobiological properties, which indicate major dimensions of fundamental
- brain organisation, and we test their associations with *g*-morphometry cortical profiles.
- 75 These analyses implement methods for uncovering principles of cortical organisation that
- 76 are associated with individual differences in our complex cognitive skills.



77

78 Figure 1 Overview of the methodological approach.

79 Figure 1 note A) Associations between g and 5 measures of brain morphometry (volume, surface area, 80 thickness, curvature and sulcal depth) were estimated for each of three cohorts of community-dwelling 81 adults (UKB, GenScot and LBC1936). These vertex-wise association maps were then meta-analysed, 82 which is the primary outcome of the first step. B) We curated and derived new maps of 33 83 neurobiological characteristics that vary across the cortex, and registered them to the same anatomical 84 space as the vertex-wise meta-analyses described in A. We also conduct a principal components analysis 85 which identifies four major dimensions of neurobiological organisation across the cortex. C) Finally, we 86 calculate the spatial correlations between g-morphometry profiles and neurobiological profiles, to 87 identify which principles of cortical organisation are most likely candidates for supporting complex 88 cognitive skills.

2 Methods

89 2.1 Methods for identifying individual differences

90 2.1.1 Participants

Data from three cohorts were used to calculate associations between general cognitive functioning (*g*) (and age and sex) and 5 measures of vertex-wise morphometry (volume, surface area, thickness, curvature, and sulcal depth) – the UK Biobank (UKB), Generation Scotland: Scottish Family Health Study (GenScot), and the Lothian Birth Cohort 1936 (LBC1936). They were also used to calculate meta-analysed means for the 5 morphometry measures. These maps will be openly available on publication in fsaverage space at github.com/JoannaMoodie/moodie-brainmaps-cognition.

98 The UKB (http://www.ukbiobank.ac.uk, ³³) is a study of ~500,000 participants, and the data 99 of 40,383 participants who attended the first neuroimaging visit (which included collection 100 of cognitive test data and brain MRI scans) are used in the present analyses. Participants 101 were excluded from the present analysis if their self-reported medical history, taken by a 102 nurse at the data collection appointment, recorded a diagnosis of, for example, dementia, 103 Parkinson's disease, stroke, other chronic degenerative neurological problems or other 104 demyelinating conditions, including multiple sclerosis and Guillain-Barré syndrome, and 105 brain cancer or injury (a full list of exclusion criteria is provided in *Table S1*). For the global 106 and subcortical brain structures analysis (see Supplementary Analysis 2), the sample 107 consisted of N = 39,250 (53% female, mean age = 63.91 years, SD = 7.67 years, and range = 108 44 to 83 years). For the vertex-wise analyses, participants were included if gcaching in 109 FreeSurfer ran successfully for all 5 morphometry measures. The final N for vertex-wise 110 analyses was 36,744 participants (53 % female, mean age = 63.71 years, SD = 7.63 years, 111 and range = 44-83 years). The UKB was given ethical approval by the NHS Research Ethics 112 Committee (REC reference 11/NW/0382) and the current analyses were conducted under UKB application number 10279. All participants provided informed consent. More 113 114 information on the consent procedure can be found at 115 https://biobank.ctsu.ox.ac.uk/crystal/label.cgi?id=100023.

116The GenScot imaging sample is a population-based study, developed from the Generation117Scotland: Scottish Family Health Study (34). Data are available for a maximum of N = 1188118participants. Cognitive and MRI data are available for N = 1043 participants (60% female,119mean age = 59.29 years, SD = 10.12 years, and range = 26 to 84 years). All 1043 participants120were used in the current global and subcortical brain structures analyses (see

121 *Supplementary Analysis 2*). For the vertex-wise analysis, qcaching in FreeSurfer ran 122 successfully for all measures for N = 1013 participants (60% female), mean age = 59.22 123 years (*SD* = 10.12 years), age range = 26 to 84 years. GenScot received ethical approval from 124 the NHS Tayside Research Ethics Committee (14/SS/0039), and all participants provided 125 informed consent.

126 The LBC1936 is a longitudinal study of a sample of community-dwelling older adults who 127 were born in 1936, most of whom took part in the Scottish Mental Survey of 1947 when 128 they were ~ 11 years old, and who volunteered to participate in this cohort study at ~ 70 129 years old (35,36) https://lothian-birth-cohorts.ed.ac.uk/. The current analysis includes data 130 from the second wave of data collection, which is the first wave at which head MRI scans 131 are available. In total, 731 participants agreed to MRI scanning. After image collection and 132 processing, N = 636 participants were included in the specific brain structures analyses 133 conducted in Supplementary Analysis 2 (47% female, mean age = 72.67 years, SD = 0.71134 years, and range = 70 to 74 years). Qcaching was unsuccessful for 14 participants, leaving a 135 final N for vertex-wise analyses of 622 (47% female, mean age = 72.66 years, SD = 0.73 years, 136 and range = 71 to 74 years). The LBC1936 study was given ethical approval by the Multi-137 Centre Research Ethics Committee for Scotland, (MREC/01/0/56), the Lothian Research 138 Ethics Committee (LREC/2003/2/29) and the Scotland A Research Ethics Committee 139 (07/MRE00/58). All participants gave written informed consent.

140 2.1.2 Cognitive tests

141 All three cohorts have data collected across several cognitive tests, covering several 142 cognitive domains (e.g. memory, reasoning and processing speed), which enables the 143 estimation of a latent factor, g. The cognitive tests in each cohort have been described in 144 detail elsewhere: UKB (37, 10 tests included: Reaction time, Number span, Verbal and 145 numerical reasoning, Trail making B, Matrix pattern, Tower task, Digit-symbol substitution, 146 Pairs matching, Prospective memory, and Paired associates), GenScot (³⁴, 5 tests included: 147 Matrix reasoning, Verbal fluency, Mill Hill vocabulary, Digit-symbol substitution, and Logical 148 memory), and LBC1936 (35,38,39, 13 tests included: Matrix reasoning, Block design, Spatial 149 span, National Adult Reading Test (NART), Weschler Test of Adult Reading (WTAR), Verbal 150 fluency, Verbal paired associates, Logical memory, Digit span backwards, Symbol search, 151 Digit-symbol substitution, Inspection time, and Four-choice reaction time), see 152 *Supplementary Tables* S2 to S7 for more details. The cognitive tests from each cohort cover 153 various cognitive domains, including Crystallised (verbal) Ability, Reasoning, Processing 154 speed, and aspects of Memory.

155 A latent factor of q – capturing shared variance in performance across all cognitive tests – 156 was estimated for each cohort in a structural equation modelling framework. For UKB and 157 GenScot, no residual covariances between individual cognitive tests were included. For the 158 LBC1936, which has a larger cognitive battery that includes multiple tests for each cognitive 159 domain, g has previously been modelled with a hierarchical confirmatory factor analysis 160 approach, to incorporate defined cognitive domains (^{38, 39}). Here, in keeping with these 161 previous models, within-domain residual covariances were added for four cognitive 162 domains (visuospatial skills, crystallised ability, verbal memory and processing speed). 163 Latent g model fits were assessed using the following fit indices: Comparative Fit Index 164 (CFI), Tucker Lewis Index (TLI), Root Mean Square Error of Approximation (RMSEA), and 165 the Root Mean Square Residual (SRMR). All models had CFI > 0.95, TLI > 0.88, RMSEA < 0.08 and SRMR < 0.04. For specific details of the model fits, see *Table S9*. Results of the g166 167 measurement models are summarised in Figure S1 and Table S8. For all cohorts, all 168 estimated paths to latent g were statistically significant at the p < .001 level. To be clear, a g 169 factor was found in each of the three cognitive test batteries (that is, a model with a g factor 170 had a good fit to the data [i.e., the cognitive tests' covariance matrices]) and was not imposed 171 upon them.

The latent *g* scores were extracted for all participants (these were calculated with the slightly larger samples that were included in the global and subcortical structures analysis, see *Supplementary Analysis 2*, and these same scores were used for the vertex-wise analysis, which had a slightly smaller sample size due to qcaching failures). All *g* scores were scaled so that higher scores reflected better cognitive performance.

177 2.1.3 MRI protocols

178 Detailed information for MRI protocols in all three cohorts are reported elsewhere: UKB 179 (⁴⁰), LBC1936 (⁴¹) and GenScot (³⁴) but are briefly summarised here. In the present sample, 180 UKB participants attended one of four testing sites: Cheadle ($\sim 60\%$), Reading ($\sim 14\%$), Newcastle (~25%), and Bristol (~0.13%). The same type of scanner was used in all four 181 182 testing sites, a 3T Siemens Skyra, with a 32-channel Siemens head radiofrequency coil. 183 The UK Biobank MRI protocol includes various MRI acquisitions (more details available 184 here https://www.fmrib.ox.ac.uk/ukbiobank/protocol/V4_23092014.pdf) but in this 185 work we exclusively used the T1-weighted MPRAGE volumes. For T1-weighted images, 208 186 sagittal slices were acquired with a field view of 256 mm and a matrix size of 256 x 256 187 pixels, giving a resolution of $1 \ge 1 \ge 1 \ge 1$ mm³. The repetition time was 2000 ms and the echo 188 time was 2.01 ms (⁴⁰).

189 GenScot had 2 testing sites: Aberdeen (in the present sample, N = 528, 51% of the total 190 sample) and Dundee (N = 515, 49% of the total sample). Detailed information about the 191 GenScot structural image acquisitions is available here 192 https://wellcomeopenresearch.org/articles/4-185. For the current analysis, we used the 193 T1-weighted fast gradient echo with magnetisation preparation volume. The Aberdeen 194 site used a 3T Philips Achieva TX-series MRI system (Philips Healthcare, Best, 195 Netherlands) with a 32-channel phased-array head coil and a back facing mirror (software 196 version 5.1.7; gradients with maximum amplitude 80 mT/m and maximum slew rate 100 197 T/m/s). For T1-weighted images, 160 sagittal slices were acquired with a field of view of 198 240 mm and a matrix size of 240 x 240 pixels, giving a resolution of 1 x 1 x 1 mm³. 199 Repetition time was 1968 ms, echo time was 3.8 ms and inversion time was 1031 ms. In 200 Dundee, the scanner was a Siemens 3T Prisma-FIT (Siemens, Erlangen, Germany) with 20 201 channel head and neck phased array coil and a back facing mirror (Syngo E11, gradient 202 with max amplitude 80 mT/m and maximum slew rate 200 T/m/s). For T1-weighted 203 images 208 sagittal slices were acquired with a field of view of 256 mm and matrix size 204 256 x 256 pixels giving a resolution of 1 x 1 x 1 mm³. Repetition time was 1740 ms, echo 205 time was 2.62 ms, and inversion time was 900 ms ³⁴.

206 All LBC1936 participants were scanned in the same scanner in the same clinic, using a GE 207 Signa LX 1.5T Horizon HDx clinical scanner (General Electric, Milwaukee, WI) with a 208 manufacturer supplied 8-channel phased array head coil. More information on the 209 structural image acquisitions for the LBC1936 cohort is available in (⁴¹). For T1-weighted 210 images (3D IR-Prep FSPGR), 160 coronal slices were acquired, with a field of view of 256 211 mm and a matrix size of 192 x 192 pixels (zero filled to 256 x 256) giving a resolution of 1 x 212 1 x 1.3 mm³. The repetition time was 10 ms, echo time was 4 ms and inversion time was 213 500 ms.

214 For all cohorts, the FreeSurfer image analysis suite (http://surfer.nmr.mgh.harvard.edu/) 215 was used for cortical reconstruction and volumetric segmentation. The 46 global and 216 subcortical structures (including grey matter, white matter and ventricles), used in 217 Supplementary Analysis 2, were available for each cohort in the aseg FreeSurfer outputs. 218 Vertex-wise surface values for 5 morphometry measures (volume, surface area, thickness, 219 curvature and sulcal depth) were available at 9 smoothing tolerances (0, 5, 10, 15, 20, 25, 220 30, 35 and 40 mm FWHM, full width half maximum Gaussian kernel) by running the -gcache 221 flag.

222 Each cohort used a different version of FreeSurfer: UKB = v6.0, GenScot = v5.3, LBC1936 = 223 v5.1. The LBC1936 and GenScot parcellations have previously undergone quality control, 224 with manual editing to rectify parcellation issues including skull stripping, tissue 225 identification and regional boundary lines. The UKB regional data were extracted from the 226 bulk-downloaded aseg files provided by the UK Biobank. For the current study, UKB values 227 more than 4 standard deviations from the mean for any global or subcortical brain structure 228 volume were excluded (corresponding to < 1.2% of the data per variable; M = 87.97, SD =229 121.75, range = 0 to 474 participants) – note, outlier values were excluded by region, rather 230 than participant-wise exclusions

231 2.1.4 Morphometry measures

The morphometry measurements are illustrated in *Figure 1A* (middle panel). Volume is the 232 233 amount of three-dimensional space of a vertex, surface area is the total area of the cortical 234 sheet section of the vertex, and thickness is the distance between the pial and white matter 235 cortical surfaces. If thickness were uniform across the vertex, volume would be the product of surface area and thickness, but this relationship is more complex in practice. For 236 curvature, a value of zero represents no curvature "- "; those with negative values are 237 curving up (convex) " \frown "; those with positive values are curving down (concave) " \bigcirc ". The 238 239 sulcal depth is a measure of how removed a vertex is from a theoretical mid-surface that is estimated between the gyri and sulci (vertices on the mid surface receive a value of 0). A 240 241 more positive sulcal depth suggests a deeper location (i.e., away from the scalp) and a more 242 negative value is shallower (i.e., towards the scalp). Deep sulci tend to have more concave 243 curvature, shallower regions tend to have curvature magnitudes nearer to zero, and gyri 244 (defined here as regions with negative values for "sulcal depth") tend to correspond to 245 convex curvature. The measure of curvature provides information about how the cortex 246 folds at the local level, while sulcal depth provides a more macroscopic perspective on the 247 depth of sulci relative to the midpoint of the cortical surface, offering insights into the 248 overall brain folding complexity.

249 2.1.5 Meta-analyses

We chose a 20 mm FWHM smoothing tolerance for our main cohort meta-analyses, in line with our previous work (42 , 43). For each cohort, a standardised β was estimated between *g* and each vertex for the 5 vertex-wise morphometry measures. Each participant's cortical surface was aligned to the fsaverage template. Out of 327,684 initial vertices along the fsaverage surface, there are 298,790 vertices labelled as "cortex", and these vertices are analysed here. We first checked the cross-cohort agreement of the means of the five measures of cortical morphometry across the three cohorts. Spatial variations in measures of cortical volume, surface area, thickness, curvature and sulcal depth were highly stable between cohorts - all r > 0.843 (see *Table S*10, for the global means see *Figure S*3, and for the meta-analysed mean cortical maps see *Figure S*2). From these analyses, the meta-analysed mean profiles for surface area and thickness were included in the spatial correlation analyses in section 3.4.

For cohort-based association analyses, all brain measures were controlled for head position in the scanner (X, Y and Z coordinates, from UKB codes 25756, 25757, and 25758; and estimated in FreeSurfer for GenScot and LBC1936), testing site (for UKB and GenScot only) and, for LBC1936 only, time lag (because it was the only cohort with a time lag between cognitive and MRI appointments, *M* lag =65.08 days, *SD* = 37.77). Age and sex were included as covariates in models when they were not the variable of interest.

268 To characterise which regions of the human brain, in terms of morphometry, are most 269 strongly related to individual differences in q, we then meta-analysed the vertex-wise q270 associations between the three cohorts with random effects models. This type of model was 271 deemed the most appropriate due to the differing characteristics (e.g., age range) between 272 the cohorts. Vertex-wise brain maps for age and sex associations were meta-analysed in the 273 same way. For vertex-wise analyses of age, only GenScot and UKB cohorts were included 274 due to the narrow age range of the LBC1936 cohort (mean age = 72.67 years, SD = 0.71). For 275 all meta-analyses, between-cohort age moderation analyses were additionally conducted 276 (i.e., mean age for each cohort was included as a moderator in the *rma* function in the 277 *metafor* package ⁹⁰). UKB and GenScot have larger age ranges, and lower mean age (M =278 63.91 years, range = 44-83; M = 59.29 years, range = 26-84 respectively) compared to the 279 LBC1936 (M = 72.67 years, range = 70-74). Therefore, although we included age as a 280 covariate within cohorts, it remains possible that between-cohort age differences affect the 281 brain associations. Any between-cohort age moderation analyses significant at the $\alpha < .05$ 282 level are reported below.

283 2.2 Neurobiological cortical profiles

The 33 included neurobiological cortical profiles were derived from several modalities, including: in vivo MRI, rsfMRI (resting state functional MRI), fcMRI (resting-state functional connectivity MRI), PET (positron emission tomography) scans and postmortem tissue. Several of these cortical profiles are openly available online through neuromaps (¹²), and BigBrainWarp (¹³), and we registered all profiles to fsaverage space using neuromaps. We

289 include maps of metabolism (we calculated a principal component, derived from previously

- 290 published, open source maps of cerebral blood flow, oxygen metabolism, and glucose
- 291 metabolism, ⁷⁹); similarity gradients of cytoarchitecture (staining intensity), functional
- 292 connectivity, and microstructure (¹³); the first principal component of gene expression from
- the abagen toolbox (⁴⁴); cortical myelination T1/T2 ratio (⁴⁵) and 19 neurotransmitter
- receptor densities (12). These maps are described in more detail in Table 1 and in the
- subheadings below.

296 *Table 1* Description of the neurobiological cortical profiles. Descriptive statistics of all

297 vertex-wise cortical profiles in this paper are available in *Table S*13.

Мар	Source	Original	Data source	Participants	Category	Туре	Higher value
		source					=
Gene expression	Neuromaps	abagen	Postmortem	N = 6 adult	Microstructure	Principal	A higher
PC1		toolbox 44	(gene	human donors,		component 1	positive
			expression)	1 female, ages		of gene	component
				24 to57 years		expression	score for PC1
Microstructural	BigBrainWarp	Paquola et al.	In Vivo (qT1)	N = 50 healthy	Microstructure	Eigenvectors	A higher
similarity		13,77		adults ⁴⁶ , 23		1 and 2	positive
				women, age			eigenvector
				mean = 29.54			score
				(SD = 5.62)			
Cytoarchitectural	BigBrainWarp	Paquola et al.	Postmortem	<i>N</i> = 1 donor, 65-	Microstructure	Eigenvectors	A higher
similarity		13	(BigBrain,	year-old male		1 and 2 from	positive
			histology)			BigBrain	eigenvector
						staining	score
						intensity	
			-			profiles	
Myelination (T1/T2	Neuromaps	Glasser ⁴⁵	In vivo	N = 449 young	Microstructure	T1w to T2w	Higher
contrast)			(11/12)	adults (ages		ratio	myelin
				22–35) from			density
				the Human			
				Connectome			
A11	Colordate d Con	C	to show	Project (HCP)	M	L l	TT'shaa
Allometric scaling	calculated for	Current paper	In vivo	N = 38,379	Macrostructure	Log-log	Higner
	the current		(11, MRI)	UKP ConSect		regression	Coefficient
	paper			and I PC1026		vortov surface	deviation
				anu LBC1950,		area predicted	from
				84		by total	isometry)
				01		surface area	isometry)
Mean surface area	Calculated for	Current paper	In vivo (T1.	N = 38.379	Macrostructure	Mean	Larger
adult	the current		MRI)	adults, from			surface area
	paper		,	UKB, GenScot			
	P-P-			and LBC1936;			
				age range: 26-			
				84			
Mean thickness	Calculated for	Current paper	In vivo (T1,	N = 38,379	Macrostructure	Mean	Thicker
adult	the current		MRI)	adults, from			
	paper			UKB, GenScot			
				and LBC1936;			
				age range: 26-			
				84			
Intersubject	Neuromaps	Mueller et al. 47	In vivo	N = healthy	Functional	Variability in	More
variability			(fcfMRI)	subjects (age		fcMRI data	variability in
				51.8±6.99, 9			rsfMRI
				female)			
Functional	BigBrainWarp	Paquola et al.	In vivo	50 healthy	Functional	Eigenvectors	A higher
connectivity		13	(rsfMRI)	adults ⁴⁶ , 23		1 and 2 from	positive
similarity				women, age		rsfMRI data	eigenvector
				mean = 29.54			score
				(SD = 5.62)			
Cognition PC1	Neuromaps	Yarkoni et al.	In vivo (fMRI)	Unknown	Functional	Principal	A higher
Neurosynth		48				component 1	positive
						ot tMRI	component
						patterns	score for PC1
						associated	
						with various	
						terms in	
						Neurosynth	

Metabolism	Neuromaps, and PC1 calculated for the current paper	Vaishnavi et al. 79	In vivo (PET)	N = 33 neurologically normal young adults, 19 women, 14 men, 20-33 years old	Functional	Principal component 1 of cerebral blood flow, oxygen metabolism and glucose metabolism maps	Higher positive values on PC1 (higher values on the maps for CBF, CMRO2 and CMRGlu)
5HT1a (Serotonin)	Neuromaps, (Hansen et al.)	Savli et al. ⁴⁹	In vivo (PET)	N = 35, age mean = 26.3 (SD = 5.2)	Receptor density	Group averaged PET data	Higher density of each receptor
5HT1b (Serotonin)	Neuromaps, (Hansen et al.)	Gallezot et al. 50	In vivo (PET)	N = 65, age mean = 33.7 (SD = 9.7)	Receptor density	Group averaged PET data	Higher density of each receptor
5HT2A (Serotonin)	Neuromaps, (Hansen et al.)	Beliveau et al. ⁵¹	In vivo (PET)	N = 29, age mean = 22.6 (SD = 2.7)	Receptor density	Group averaged PET data	Higher density of each receptor
5HT4 (Serotonin)	Neuromaps, (Hansen et al.)	Beliveau et al. 51	In vivo (PET)	N = 59, age mean = 25.9 (SD = 5.3)	Receptor density	Group averaged PET data	Higher density of each receptor
5HT6 (Serotonin)	Neuromaps, (Hansen et al.)	Radhakrishnan et al. ⁵²	In vivo (PET)	N = 30, age mean = 36.6, SD = 9)	Receptor density	Group averaged PET data	Higher density of each receptor
5HTT (Serotonin)	Neuromaps, (Hansen et al.)	Beliveau et al.51	In vivo (PET)	N = 100, age mean = 25.1 (SD = 5.8)	Receptor density	Group averaged PET data	Higher density of each receptor
D1 (Dopamine)	Neuromaps, (Hansen et al.)	Kaller et al. ⁵³	In vivo (PET)	N = 13, age mean = 33 years (SD = 13)	Receptor density	Group averaged PET data	Higher density of each receptor
D2 (Dopamine)	Neuromaps, (Hansen et al.)	Smith et al. and Sandiego et al. ⁵⁴ , ⁵⁵	In vivo (PET)	N = 37, age mean = 48.4 years (SD = 16.9); N = 55, age mean = 32.5 years (SD = 9.7)	Receptor density	Group averaged PET data	Higher density of each receptor
DAT (Dopamine)	Neuromaps, (Hansen et al.)	Dukart et al. ⁵⁶	In vivo (PET)	N = 174, age mean = 61 years (SD = 11)	Receptor density	Group averaged PET data	Higher density of each receptor
NAT (Norepinephrine)	Neuromaps, (Hansen et al.)	Ding et al. ⁵⁷	In vivo (PET)	N = 77, age mean = 33.4 (SD = 9.2)	Receptor density	Group averaged PET data	Higher density of each receptor
H3 (Histamine)	Neuromaps, (Hansen et al.)	Gallezot et al. ⁵⁸	In vivo (PET)	N = 8, age mean = 31.7 (SD = 9.0)	Receptor density	Group averaged PET data	Higher density of each receptor
A4B2 (Acetylcholine)	Neuromaps, (Hansen et al.)	Hillmer et al. ⁵⁹	In vivo (PET)	N = 30, age mean = 33.5 years (SD = 10.7)	Receptor density	Group averaged PET data	Higher density of each receptor

M1	Neuromaps,	Naganawa et	In vivo (PET)	N = 24, age	Receptor density	Group	Higher
(Acetylcholine)	(Hansen et al.)	al. 60		mean = 40.5		averaged PET	density of
				(SD = 11.7)		data	each
							receptor
VAChT	Neuromaps,	PIs: Tuominen,	In vivo (PET)	N = 4, age mean	Receptor density	Group	Higher
(Acetylcholine)	(Hansen et al.)	L. & Guimond,		= 37 (DF =		averaged PET	density of
		S.;		10.2);		data	each
		Aghourian et		N = 18 (age			receptor
		al. ⁶¹ ;		mean = 66.8, SD			
		Bedard et al. 62		= 6.8);			
				N = 5, age mean			
				= 68.3 (SD =			
				3.1)			
CB1	Neuromaps,	Normandin et	In vivo (PET)	N = 77, age	Receptor density	Group	Higher
(Cannabinoid)	(Hansen et al.)	al. 63		mean = 30		averaged PET	density of
				years (SD = 8.9)		data	each
							receptor
MU	Neuromaps,	Kantonen et al	In vivo (PET)	<i>N</i> = 204, age	Receptor density	Group	Higher
(Opioid)	(Hansen et al.)	64		mean = 32.3		averaged PET	density of
		-		years (SD =		data	each
				10.8)			receptor
NMDA	Neuromaps,	Galovic et al.65,	In vivo (PET)	N = 29, age	Receptor density	Group	Higher
(Glutamate)	(Hansen et al.)	66		mean = 40.9		averaged PET	density of
				years (SD =		data	each
				12.7)			receptor
mGluR5	Neuromaps,	Smart et al. 67;	In vivo (PET)	N = 73, age	Receptor density	Group	Higher
(Glutamate)	(Hansen et al.)	PIs: Rosa-Neto,		mean = 19.9		averaged PET	density of
		P. &		(SD = 3.04);		data	each
		Kobayashi, E.;		<i>N</i> = 22, age			receptor
		DuBois et al. 68		mean = 67.9			
				(SD = 9.6);			
				N = 28, age			
				mean = 33.1			
				(SD = 11.2)			
GABAa-bz	Neuromaps,	Nørgaard et	In vivo (PET)	<i>N</i> = 16, age	Receptor density	Group	Higher
(GABA)	(Hansen et al.)	al. ⁶⁹		mean = 32.3		averaged PET	density of
				(SD = 10.8)		data	each
							receptor

298

299 2.2.1 Gene expression

The gene expression map was the first principal component of gene expression from the abagen toolbox (⁴⁴). It is available in neuromaps in fsaverage 10k space, and we resampled it to fsaverage 164k using the transforms function in neuromaps. Seemingly due to registration error, there were more vertices outside the cortical mask than for the association maps, and most of the other neurobiological profiles. There were 292076 vertices included in the cortical mask.

306 2.2.2 T1/T2 ratio derived myelination

The map of cortical myelin content was previously derived from T1w to T2w ratios (maps calculated in ⁴⁵, method described in detail in ⁷⁰). T1/T2 ratio is thought by some to be a good estimate of relative myelin content across the cortical surface ⁷⁰, although it is important to note that this method only provides a proxy for myelin content, and also reflects tissue microstructures other than myelin, such as axon density and dendrite density

and iron content ⁷¹. Indeed, in some contexts, T1/T2 ratios do not make a good proxy for
myelination ⁷². Nevertheless, we call this map "myelination", in line with the source paper.

314 2.2.3 Allometric scaling

315 To obtain a map of allometric scaling, we calculated associations between total surface area 316 and vertex-wise surface for UKB, GenScot and LBC1936 cohorts and then meta-analysed 317 them. Allometric scaling was calculated based on previous work (7^3) , with a log-log 318 regression coefficient for vertex surface area predicted by total surface area. Allometric 319 scaling shows which vertices have a disproportionately larger surface area in people with 320 bigger brains. Comparable maps of allometric scaling are available for younger cohorts 321 compared to the current sample (the Philadelphia Neurodevelopmental Cohort, PNC, and a 322 National Institutes of Health, NIH, sample). These maps were created with samples 8 to 23 323 years old (N = 1373) and 5 to 25 years old (N = 792). These previously calculated maps were 324 correlated at r = 0.679, with each other, and at r = 0.430 and r = 0.378, respectively with our 325 log vertex area $\sim \log$ total surface area maps (which are created with data on adults, age 326 range = 26-83 years old). As most maps of interest included in the current study are derived 327 from adult data, we use the allometry map that we created from our current samples in our 328 further analyses. In our calculations of allometric scaling, the standardised estimates were 329 strongly spatially correlated between cohorts (LBC1936-GenScot r = 0.764, GenScot-UKB r 330 = 0.773 and UKB-LBC1936, r = 0.730, all $p < 2.2 \times 10^{-16}$), showing that across cohorts, the 331 regions that tended to be larger with increasing brain size were consistent.

332 2.2.4 Mean surface area and mean thickness

333 Meta-analysed mean values for surface area and thickness were calculated using UKB,

GenScot and LBC1936 data (meta-analytic N = 38,379). These are mapped to the cortex in

- *Figure S2* (between cohort spatial correlations were all r > 0.843, see *Table S10*).
- 336 2.2.5 Intersubject variability

Intersubject variability in rsfMRI varies spatially across the cortex (⁴⁷). In other words, for some regions, rsfMRI is similar across participants, whilst in other regions, it is more variable. An openly available cortical map of intersubject variability was available at 1k density in fsaverage space in neuromaps (¹²), and we registered it to 164k density in fsaverage space.

342 2.2.6 Cognition PC1 from Neurosynth

343 Component 1 from a principal components analysis of cognitive terms in Neurosynth

344 (which is a database of task-based fMRI results) (48). It is available as a cortical map in

345 MNI152 2mm space, and we registered it to fsaverage_164k space (¹²).

346 2.2.7 Similarity gradients: cytoarchitecture, functional connectivity and347 microstructure

Cortical similarity gradients of cytoarchitecture, functional connectivity and microstructure 348 349 readily available in fsaverage_164k space BigBrainWarp are in 350 https://bigbrainwarp.readthedocs.io/en/latest/. The BigBrain data (N = 1) (⁷⁴) was used to 351 create the cytoarchitectural similarity map, and the microstructural and functional maps 352 were based on N = 50 healthy adults, for whom multiscale MRI data is openly available⁷⁵. 353 The cytoarchitectural gradients data are based on staining intensity profiles. The 354 microstructural gradients are based on qT1 intensity, a quantitative measure of longitudinal 355 relaxation time, which provides an in vivo proxy for cortical microstructure. The functional 356 connectivity gradients are based on rsfMRI-derived functional connectomes. The methods 357 used to obtain these maps are available in detail in the documentation for BigBrainWarp 358 https://bigbrainwarp.readthedocs.io/en/latest/ and micapipe 359 https://micapipe.readthedocs.io/en/latest/. Briefly, the cytoarchitectural and functional 360 gradients were calculated with diffusion map embedding, which is a nonlinear manifold 361 learning technique (⁷⁶), applied to cross-correlations of vertex-wise staining intensity 362 profiles (77), and the microstructural and functional connectivity axes are calculated using 363 a microstructural profile covariance (MPC) approach (78), which provides eigenvectors of 364 common variation. The percentage of variance explained by the first two eigenvectors for 365 each measure were: cytoarchitectural similarity 1 = 42% and 2 = 35%, for functional 366 connectivity 1 = 12.9% and 2 = 6.5%, and for microstructural similarity 1 = 59.0% and 2 =367 10.5%. In an attempt to make the cortical similarity gradients from BigBrainWarp of 368 comparable granularity to our individual difference association maps (20 mm FWHM 369 smoothing), we performed additional smoothing on the BigBrainWarp-sourced maps. The 370 cytoarchitectural gradients were previously smoothed by 2 mm FWHM Gaussian kernel (13), 371 and so we smoothed these with an additional 18 mm FWHM kernel. With the approximate 372 guideline that the rsfMRI data approximately has a smoothing kernel of 6 mm (1^3) , we 373 smoothed the functional connectivity gradients with an additional 14 mm kernel. To our 374 knowledge, the microstructural similarity gradients available in BigBrainWarp have not 375 previously been smoothed, and here we smoothed them with a 20 mm FWHM kernel.

376 2.2.8 Metabolism

Metabolism data, available in neuromaps (¹²), was originally collected in 2010 by Vaishnavi et al. (⁷⁹). These data are an average of the PET maps across 33 young adults at rest. Here, we looked at 3 measures of cortical metabolism, cerebral blood flow, oxygen metabolism and glucose metabolism. We registered them from fsLR_164k to fsaverage_164k in

- neuromaps (12). These maps were all highly spatially correlated with each other (all r > 0.8),
- 382 and the first principal component explained 88% of the variance, with all three loadings >
- 383 0.5, see *Figure S*17. It is this first principal component of cortical metabolism that we used
- 384 for the "metabolism" map, included in our spatial correlation analyses. More positive values
- 385 denote higher metabolic activity.
- 386 2.2.9 Neurotransmitter receptor densities

Hansen et al. recently collected receptor density maps for serotonin (5HT1A, 5HT1B, 5HT2A, 5HT4, 5HT6, and 5HTT), dopamine (D1, D2, DAT), norepinephrine (NAT), histamine (H3), acetylcholine (α 4 β 2, M1, VAChT), cannabinoid (CB1), opioid (Mu), glutamate (NMDA, mGluR5) and GABA (GABA A/BZ) neurotransmitter receptors (¹⁵). They are available through neuromaps and we registered them from MNI152 space to fsaverage_164k space, also using neuromaps (¹²). Due to the lower spatial resolution of PET data, no further smoothing was performed.

394 2.3 Spatial correlations

395 The *g*-morphometry maps described above (*Figure 1A*) were then spatially correlated with 396 1) 33 neurobiological maps, and 2) 4 PCs derived from the 33 neurobiological maps and 397 denoting core components of cortical neurobiological organisation (Figure 1B). Spatial 398 correlations were calculated using Pearson's r (e.g. for each g-morphometry map, the vector 399 of cortical vertices was correlated with each other map's vector of cortical vertices). 400 Alexander-Bloch's spin-based permutation test was used to calculate *p*-values⁸⁰. Each *q*morphometry map was spun randomly 10000 times, and from the resulting null 401 402 distributions of the correlations, p values were calculated. The Pearson's r and p spin values 403 for spatial correlations between all maps included in the main correlation analyses in the 404 current paper are available in the Supplementary Tabular Data file.

All 33 neurobiological maps were inputted into a PCA which was calculated in R using the prcomp function, with the aim to identify core components of neurobiological spatial cortical organisation. With all vertices in the cortical mask, across the 33 maps, the vertex count was N = 292,056 for the PCA. Four components were extracted, based on the variance they explained (together, 65.9%), and rotated with the varimax method.

We also calculated within-region vertex-wise spatial correlations for *g*-morphometry and neurobiological map correlations. To do this, we used the fsaverage annotation files to identify which vertices were included in each region according to the Desikan-Killiany atlas (34 left/right paired cortical regions), and then calculated the spatial correlations

414 separately for each region. These within-region analyses offer important nuance to the 415 cortex-wide spatial correlation statistics in the form of two additional features: 1) the 416 relative strength of spatial correlations within different regions, and 2) the homogeneity of

417 correlations among regions.

418 2.4 Supplementary analyses

419 We conducted three supplementary analyses. Supplementary Analysis 1 addresses the 420 current lack of consensus about optimal smoothing parameters. Noise in the data, due to 421 registration inaccuracies, is minimized when the cortex is parcellated into larger regions 422 (i.e., greater smoothing) but, when the cortex is parcellated into smaller regions (i.e., less 423 smoothing), the % variance explained increases (⁸¹) due to the additional information 424 provided. Thus, at the vertex-wise level, there is a balance to be struck between the benefits 425 of reducing noise in the data, and the problem that increasing to higher levels of smoothing 426 will, at a point, remove fine-grained spatial information and thus reduce the spatial 427 specificity of detected associations. Lerch and Evans (2005) analysed the effect of different 428 smoothing tolerances on cortical thickness measurement sensitivity, and they concluded an 429 optimal kernel size of 30 mm FWHM (82 , N = 25). Some studies use 30 mm (83,84), and other common choices are 5 mm (⁴), 10 mm (^{85,86}), 15 mm (^{87,88}) or 20 mm (^{42,89}). We investigated 430 431 the effects of smoothing tolerances on *g*-morphometry associations here (see 432 Supplementary Analysis 1), and the results suggest that generally, across morphometry 433 measures, 10-20 mm FWHM tends to maximise noise reduction while maintaining localised 434 effects. These results may aid future smoothing tolerance choices for similar analyses.

435 *Supplementary Analysis 2* focuses on global and sub-cortical associations with *g*, age, and 436 sex. Although much previous work on *g*-brain associations focuses on the cortex, sub-437 cortical structures are becoming increasingly recognized for their associations with 438 cognitive function.

439 *Supplementary Analysis 3* tests whether *g*-morphometry associations differ by sex when 440 they are calculated separately for each sex. The spatial correlations were all r > 0.753 for 441 UKB, suggesting that meaningful sex differences do not exist in the general population.

442 2.5 Analysis software

Within-cohort vertex-wise analyses were conducted in surfstat
http://www.math.mcgill.ca/keith/surfstat/ in MATLAB. All meta-analyses (metafor, ⁹⁰),
structural equation models (lavaan, ⁹¹), and spatial correlations were conducted in R 4.0.2.

446 (R Core Team, 2020). Structural equation models were estimated with the full information447 maximum likelihood method.

448 2.6 Data Availability

449 All UKB data analysed here were provided under project reference 10279. A guide to access 450 UKB data is available from http://www.ukbiobank.ac.uk/register-apply/. To access data 451 from the GenScot study, see https://www.research.ed.ac.uk/en/datasets/stratifying-452 resilience-and-depression-longitudinally-GenScot-a-dep, and to access the Lothian Birth 453 Cohort data, see https://www.ed.ac.uk/lothian-birth-cohorts/data-access-collaboration. 454 The BigBrainWarp toolbox, released by Paquola et al. (13), is available for download at 455 https://bigbrainwarp.readthedocs.io/en/latest/. The neuromaps toolbox is available at 456 https://github.com/netneurolab/neuromaps. Analysis script templates and the vertex-457 wise β estimate cortical maps for *g*, age, sex and allometric scaling, along with the meta-458 analytic means, the principal component of the metabolism maps, and the four principal 459 components derived from the 33 neurobiological maps that are calculated in the current 460 paper will be available on publication here: github.com/JoannaMoodie/moodie-brainmaps-461 cognition.

3 Results

462 3.1 Associations between general cognitive functioning and brain morphometry:463 cross-cohort replicability and meta-analysis results

464 3.1.1 Global morphometry associations with *g*

At the global cortical level (measures summed across all vertices, associations calculated for 465 each cohort, then meta-analysed), participants with higher general cognitive function had a 466 467 greater total cortical volume ($\beta = 0.178$, SE = 0.035, $p = 3.18 \times 10^{-7}$), higher total cortical surface area ($\beta = 0.154$, SE = 0.021, $p = 5.90 \times 10^{-12}$), and (nominally) thicker cortex on 468 469 average ($\beta = 0.073$, SE = 0.037, p = .049). Higher *g* was marginally associated with greater overall concave curvature ($\beta = 0.080$, SE = 0.005, $p = 6.20 \times 10^{-60}$) although, as shown below, 470 471 the direction and magnitude of the association substantially depended on region (range 472 vertex-wise $\beta = -0.10$ to 0.09) Average sulcal depth was not associated with g ($\beta = 0.018$, SE 473 = 0.025, p = .472). This appears to be due to regional variation in the direction of effects, which cancel each other out (range vertex-wise $\beta = -0.12$ to 0.13; see *Figure* 2). 474

475 3.1.2 Vertex-wise *g*-morphometry associations: cross-cohort replicability

We ran vertex-wise *g*-morphometry analyses in each of the three cohorts. We used a 20 mm 476 477 FWHM smoothing tolerance, which provided a good balance between noise reduction and 478 loss of fine-grained cortical information (Supplementary Analysis 1). The patterning of 479 associations between general cognitive function and brain cortical measures showed good 480 between-cohort spatial agreement with moderate-to-strong correlations (see Table 2, 481 correlations ranged from Pearson's r = 0.174 to 0.581, all $p < 2.2 \times 10^{-16}$). The mean between-482 cohort spatial correlation for *g* profiles was r = 0.424, SD = 0.132, which provides further 483 evidence for the utility of *q* for replicable brain-behaviour analyses (24 , 23). Note that even for traits that have high reliability like sex, the between-cohort correlation is not r = 1 (r =484 485 0.710, SD = 0.073, see *Table* 1 for details). The lowest *q*-association agreements involved the 486 LBC1936, which has an older and narrower-age-range compared to the other two cohorts 487 (mean age = 72.67 years, SD = 0.71). Spatial correlations between GenScot and UKB were all 488 r > 0.345. Notably, the magnitude of the associations between vertex-wise cortical measures 489 and g did not change significantly across mean cohort age groups (there were no between-490 cohort age moderation effects, FDR Q > .05). The g-association maps for each cohort are 491 shown in *Figures S7* to 11 and the density distributions of the β values are summarised in Figure S12. Associations between subcortical and global volumes and g found in the current 492 493 work are presented and discussed in detail in *Supplementary Analysis 2*.

494 Table 2 Spatial agreement across cohorts in the patterning of vertex-wise associations with g, age, and 495 sex for cortical volume, surface area, thickness, curvature and sulcal depth. All $p < 2.2x10^{-16}$.

496 Table 2 Note Pearson's r is shown, indicating a spatial correlation of vectors between each pairwise

497 combination of cohorts (LBC1936, GenScot, and UKB). The vector for each cohort is a list of standardised

498 β at each cortical vertex, denoting the cortex-wide association between morphometry (volume, surface

499 *area, thickness, curvature and sulcal depth) and g, or age, or sex.*

Measure		g		Age		Sex	
Cohorts	LBC1936	GenScot-	UKB-	GenScot-	LBC1936	GenScot-	UKB-
	-GenScot	UKB	LBC1936	UKB	-GenScot	UKB	LBC1936
Volume	0.177	0.435	0.254	0.553	0.765	0.780	0.749
Surface area	0.292	0.579	0.391	0.785	0.627	0.723	0.630
Thickness	0.455	0.539	0.579	0.438	0.723	0.622	0.583
Curvature	0.273	0.345	0.297	0.625	0.681	0.781	0.677
Sulcal depth	0.581	0.538	0.452	0.663	0.718	0.847	0.742

500

501 3.1.3 Vertex-wise *g*-morphometry associations: meta-analysis results

502 We further meta-analysed the *g*-vertex associations with random effects models (mapped to the 503 cortex in *Figure* 2 and shown in extended detail in *Figures* S13 and S14). Qualitative summaries 504 of the cortical regions with the strongest positive and negative associations for *g* are in *Table* S12. 505 Across volume and surface area (respective β ranges = < 0.001 to 0.17, and 0.01 to 0.15), there 506 were positive associations in lateral temporal, lateral frontal and parietal regions of the cortex. 507 These loci are broadly consistent with the P-FIT (²) and other results from single-cohort analyses. 508 These results offer substantially greater spatial fidelity than prior ROI-based analyses.

509 These meta-analyses also provide novel information about cognitive-cortical associations. For 510 thickness, some regions had positive associations and others had negative associations. These 511 associations (β range = -0.08 to 0.13, M = 0.03, SD = 0.03) were most strongly positive in the 512 temporal pole and entorhinal cortex and were most strongly negative in the anterior cingulate. 513 medial orbitofrontal and medial occipital regions, where a thinner cortex predicted higher q. 514 Curvature and sulcal depth tended to be absent in prior ROI-based analyses, and so have not been 515 considered in detail in reviews and previously published meta-analyses (e.g., ⁹²). For curvature 516 (β range = -0.10 to 0.09, M = 0.02, SD = 0.02), higher g is associated with more concave vertices in 517 medial frontal and medial occipital regions and more convex vertices in the anterior cingulate. 518 Lastly, for sulcal depth itself, our vertex-wise results provide regional detail beyond the null 519 association found when only a global measure was used. There was substantial heterogeneity in 520 regional associations (β range = -0.12 to 0.13, M = <0.01, SD = 0.03). The results suggest that, 521 relative to the whole cortex, deeper vertices in the medial frontal, temporal pole and parieto-522 frontal regions are associated with higher q and less deep vertices in the cingulate and 523 hippocampal gyrus are associated with higher *g* (see Supplementary Table 12).



525 Figure 2 Vertex-wise g-morphometry associations.

Figure 2 note A) Associations between g and cortex-level means for all 5 morphometry measures, for the 3 cohorts (UKB, GenScot and LBC1936) and the meta-analysis. B) Vertex-wise g associations, mapped to the cortex. The lower scale limit for log Q maps is set at the minimum available value for any morphometry measure (which is -263.24, or FDR Q = 4.75×10^{-115}). C) Density distributions for the meta-analysed g ~ morphometry associations for the 5 measures of morphometry (volume, surface area, thickness curvature and sulcal depth). The vertical dotted line marks $\beta = 0$.

532 3.1.4 Vertex-wise *g*-morphometry associations: agreement between morphometry 533 measures

534 There were different regional association patterns for the 5 morphometry measures (see Table 535 3, and *Table S11* for the absolute β value correlations). For example, surface area and thickness 536 had negative and non-significant spatial agreement correlations with each other for both the g 537 and age analysis (see *Table 2, g: r* = -0.182, *p_spin* = .252; age: *r* = -0.265, *p_spin* = .462). This result 538 is consistent with previous findings that surface area and thickness associations are spatially, 539 phenotypically and genetically distinct (⁹³,⁹⁴,⁹⁵,⁹⁶). The current results agree with the previous 540 findings that patterns of g associations are not consistent between different morphometry 541 measures (e.g., 3, 4). This serves as a reminder that g-morphometry associations do not simply tell 542 us where g is in the brain; rather, they each index a conflation of multifarious biological properties 543 which also vary by brain region. The differential nature of these *g*-morphometry associations 544 might be explained by different underlying neurobiological factors of the brain (which we discuss 545 further in section 3.4).

546 3.1.5 Vertex-wise *g*-morphometry associations: within-region correlations

547 Within-region correlations show that 21/34 regions had a negative correlation between g-548 surface area and *q*-thickness and the top 5 regions with negative correlations, *r* range = -0.86 to -549 0.65 are: lateral orbitofrontal, caudal middle frontal, pericalcarine, rostral middle frontal, and 550 temporal pole (see *Figure* 4). There were 13/34 regions with positive correlations, and the top 5 551 regions for which g-surface area are positively associated with g-thickness are the inferior 552 parietal region, caudal anterior cingulate, lateral occipital, transverse temporal and frontal pole 553 (r range = 0.355 to 0.785). These results show that the concordance between these two maps has 554 a considerable amount of variation across the cortex, which is not possible to tell from the overall

555 cortical correlation (r = -0.182).

556 Table 3 Correlations (r) between directional (not absolute) g-associations for the 5 vertex-wise measures.

Permutation-based p-values are available for g correlations in Table S13, and correlation charts are shown in
 Figure S15. If the p-values are <.05, they are presented in bold font.

		Volume	Surface area	Thickness	Curvature
g	Volume				
	Surface area	0.656			
	Thickness	0.442	-0.182		
	Curvature	-0.230	-0.163	-0.069	
	Sulcal depth	-0.122	0.085	-0.109	0.176
Age	Volume				
	Surface area	0.602			
	Thickness	0.345	-0.309		
	Curvature	-0.266	-0.105	0.020	
	Sulcal depth	0.194	0.452	-0.342	-0.111
Sex	Volume				
	Surface area	0.771			
	Thickness	0.600	0.169		
	Curvature	-0.174	0.139	-0.298	
	Sulcal depth	0.041	0.237	-0.105	0.410

559 3.2 Brain regions most related to *g* are those most susceptible to ageing

In addition to the meta-analytic *g*-morphometry association maps, we similarly calculated metaanalytic maps of associations of age, and sex with cortical morphometry. *Figure 3* shows how these age and sex associations map to the cortex, and *Table S*12 provides a qualitative description of the cortical regions that have the most strongly positive, and most strongly negative β values for each measure for age and sex associations. The *g*, age and sex association maps are in the same analysis space (fsaverage), and so we can quantitatively compare their spatial patterning across the cerebral cortex.

The vertex-wise age associations show that older people tend to have a smaller cortex in termsof volume and surface area, and most of the cortex also thins with age; . Frontal, lateral temporal

and parietal regions are among those most strongly negatively associated with age. For curvature, vertices in the insula tend to be more concave with age, whilst most of the rest of the cortex sees an increase in convex vertices which is consistent with previous findings that show that cortical gyrification decreases with age (¹⁰⁵). For sulcal depth, the anterior cingulate gyrus, medial frontal region and insula become increased sulcal depth as age increases, and the medial orbitofrontal, posterior cingulate, and lateral orbitofrontal regions are less deep as age increases.

575 We use these data to quantitatively assess prior observations (which have arisen mostly from 576 qualitative inferences from disparate publications) that those parts of the brain most susceptible 577 to ageing are also those most strongly implicated in our most complex thinking skills (97). We 578 tested the spatial agreement of the vertex-wise associations for g with those for age (i.e., g-volume 579 with age-volume etc.). The results show that, as previously qualitatively observed (98, 19, 99), 580 regions of the brain most associated with q are also those that decline most with age: spatial 581 correlations range from r = -0.311 to -0.575. These overall spatial correlations broadly held across 582 most regions of the brain (mean number of negative correlations across measures = 28.8/34583 regions; Figure 3C): for most regions, vertices associated with higher g tend to exhibit more 584 ageing-related shrinkage and thinning. The 5 regions with the strongest negative correlations for 585 age-volume and *q*-volume comparisons were the transverse temporal, isthmus cingulate, frontal 586 pole, caudal anterior cingulate and superior temporal regions (r range -0.660 to -0.912). The 587 correlation of age-cortex and *q*-cortex associations across all 46 global and subcortical measures 588 was r = -0.860, $p = 2.86 \times 10^{-13}$ (see Supplementary Analysis 2 for extended analyses). These findings 589 are compatible with previous findings that present brain, age and g associations (e.g., ¹⁹, ¹⁰⁰, ¹⁰¹). 590 For example, this finding could be linked to the "last-in-first-out" hypothesis of ageing, whereby 591 the neocortical regions that are responsible for more complex cognition mature later in 592 development, and are also more vulnerable to ageing, which might be related to the high degree 593 of dendritic plasticity and remodelling required for successful functioning (¹⁰², ¹⁰³).

594 3.3 Sex differences paradox may be due to a compensatory volume-gyrification trade-595 off

The vertex-wise profiles for sex associations show that, across the cortex, males tend to have a larger volume and surface area of frontal regions than females. Females tend to have thicker superior frontal and parietal regions than males, although lateral temporal regions are thicker in males than females. Males tend to have generally more concave curvature across the cortex, compared to females, and increased sulcal depth, particularly in medial frontal regions.

Different brain regions have been shown to differentially mediate associations between sex and
 cognitive performance (e.g., for volume, the mediation % for verbal and numeric reasoning has

603 been shown to range from 0.9% in the cuneus to 29.1% in the superior temporal region, in a sample of 5216 UKB participants) (104). Here, we also tested the strength of spatial correlations 604 605 between sex and g vertex-wise cortical maps. The correlation between sex- and g-associations for 606 subcortical and global regions was r = 0.305, p = .0496 (details available in *Supplementary Analysis* 2). For vertex-wise cortical spatial correlations, regions that tend to be larger/more 607 608 concave/deeper in males than females also tend to be more associated with g (r = 0.310 to r =609 0.486), but this was a considerably weaker, non-significant result for g-thickness (r = 0.024, p spin 610 = .912). The weaker result for thickness is understood better by looking at the within-region 611 analysis. For the other 4 morphometry measures, the majority of *g*-sex brain map correlations 612 are in one direction (positive, see *Figure* 3, 2D). In contrast, for thickness, there is more of a 613 balance between positive and negative correlations. For some regions, a higher g is associated 614 more with a thicker cortex in males (top 5 correlations, r range = 0.50 to 0.62: pericalcarine, 615 isthmus cingulate, middle temporal, superior temporal, posterior cingulate), whereas in others, a 616 higher q score is associated with thicker cortex in females (top 5, r range = -0.24 to -0.54 : frontal 617 pole, paracentral, superior parietal, lateral occipital, superior frontal).



618

619 Figure 3 g-age and g-sex correlations. Figure 3 note A) Age (1) and sex (2) associations mapped to the cortex. 620 Some FDR Q values were estimated to be zero, and these have been set to the closest minimum that was 621 successfully calculated. For age and sex, the log Q limit was set at -704.3499, which is FDR Q = 1.273×10^{-306} 622 (Note that the log of Q = .05, a typical α significance threshold, is -2.9957). B) The cortex-level spatial 623 correlations between the g-morphometry associations with the age- and sex-morphometry associations. Note 624 g-sex volume and g-sex thickness had p_spin values > .05, but for all others p_spin < .05. C) Distributions of 625 regional correlations. D) Scattergraphs showing the cortex-wide correlations (representing the numbers in B) 626 in black and showing whether and where that overall spatial agreement holds for different regions (colours 627 represent the 34 paired left/right Desikan-Killiany regions) of the brain.

628 These data offer a valuable new quantitative insight into a well-documented paradox: although 629 global brain volume exhibits clear sex differences, with mean brain volume differing significantly 630 between males and females, these structural disparities do not translate into measurable 631 differences in cognitive functioning between the sexes. One hypothesised explanation for this 632 paradox involves compensatory mechanisms that mitigate volumetric sex differences, such as 633 increased gyrification (104). Here, there appears to be direct quantitative evidence of that: we 634 found that brain regions that were largest in males were also more convex in females (r = -0.174, 635 $p_{\rm spin} = .016$ between volume and curvature for sex). Convex vertices are associated with greater 636 gyrification which is generally a sign of a younger, healthier brain (105). For this sex ~ volume, sex 637 \sim curvature comparison, there were negative within-region correlations for the majority of 638 regions (28/34 regions, see *Figure* 4B), with the top 5 correlations r range = -0.77 to -0.81 for the 639 caudal middle frontal, rostral anterior cingulate, pars triangularis, paracentral, and temporal pole 640 regions.



641

642 Figure 4 Examples of within-region spatial correlations.

643Figure 4 note Within-region vertex-wise spatial correlations for A) $g \sim$ surface area and $g \sim$ thickness644(overall r = -0.182) and B) sex \sim volume and sex \sim curvature (overall r = -0.174). The overall r is shown with645a black line, and the 34 paired Desikan-Killiany regions are plotted according to the colour legend on the646right-hand side of the plot-These results offer regional underpinnings of cortex-wide associations. Assessing647within-region correlations allows identifying relative strengths of spatial correlations in different regions648across the cortex, as well as the homogeneity of the effects.

649 3.4 Neurobiological correlations between g and brain profiles - what is distinctive650 about regions associated with g?

We found widespread cortex-wide spatial correlations between g's brain morphometry associations and 33 neurobiological cortical spatial profiles (*Figure* 5). This represents the most detailed compendium of shared spatial signatures between the structure of cognitively-relevant brain regions and, microstructural, macrostructural, functional and receptor densities to have been assembled at high regional fidelity. *g*-volume and *g*-surface area association maps were significantly correlated respectively with 14 and 15 neurobiological profile maps. The neurobiological correlations of *g*-volume and *g*-surface area are highly correlated (r = 0.919) 658 suggesting that these two measures are highly similar in their relationships to underlying spatial 659 characteristics. The results indicate that regions of the cortex where larger volume or surface area 660 is more strongly associated with better general cognitive functioning were also those regions that 661 show, for example, lower metabolic activity at rest (r = -0.41 and -0.29, respectively), lower 662 cortical myelination (T1/T2 contrast, r = -0.35 and -0.47, respectively), higher receptor densities 663 (5HT1a, 5HT2a, 5HT4, 5HT6, D1, D2 M1, VAChT, CB1, MU, NMDA, *r* range = 0.25 to 0.60), and also 664 show significant co-localisation with the primary axis of cortical gene expression (r = -0.44 and -0.48, respectively). The negative association for T1/T2 contrast-derived myelination might be 665 666 explained as the most highly myelinated areas of the brain are not those involved in higher order 667 cognition but rather are areas receiving large volumes of sensory input such as the primary 668 motor, somatosensory, auditory and visual cortices. Myelination decreases with distance from 669 these regions ¹⁰⁶. Additionally, higher T1/T2 ratios have previously been associated with poorer 670 outcomes such as Alzheimer's disease (107) and, as discussed earlier, this ratio is perhaps not a 671 good proxy for myelination. Neurobiological profiles without any cortex-wide correlations with 672 *g*'s brain associations were cytoarchitectural staining similarity gradient 1, functional 673 connectivity similarity gradient 2, 5HT1b, A4B2. However, this does not suggest that there are no 674 meaningful spatial correlations between *g*-morphometry profiles and these neurobiological 675 profiles at the regional level (see *Figure* S22 for an extended version of *Figure* 5, with distributions 676 of the within-region correlations, and *Figures* S26 to S30 for further detail).

677 As reported in the analyses in section 3.1.4 above, there was a negative and non-significant 678 cortex-wide spatial correlation between $g \sim$ surface area and $g \sim$ thickness (r = -0.182, p_spin = 679 .252). In the current analyses, our cortical map of mean surface area (i.e., one of the 680 neurobiological profile maps) was positively associated with $g \sim$ surface area (r = 0.29, p_spin > 681 .05) and negatively associated with $g \sim$ thickness (r = -0.31, $p_{-}spin > .05$) and these two 682 correlations appear to cancel each other out in mean surface area's association with $q \sim$ volume 683 (r = -0.02, p spin > .05). Neurobiological profiles might offer some further insights into these 684 relationships between $g \sim$ surface area and $g \sim$ thickness. While microstructure gradient 1 was 685 correlated with g-surface area (r = -0.59), microstructure gradient 2 was correlated with g-686 thickness (r = 0.48). A similar pattern occurred for functional connectivity similarity gradients 687 where the first one was significantly correlated with *q*-surface area (r = -0.44), and the second 688 had a moderate correlation with g-thickness (r = 0.29, although $p_spin = .205$ The tendency of g-689 surface area and g-thickness to align with different microstructural and functional similarity 690 gradients may help explain why their cortical spatial patterns are spatially distinct, potentially 691 reflecting their unique phenotypic and genetic characteristics (93, 94, 95, 96).

Regions of the brain where volume and surface areas were most strongly related to q were also

693 those that density show particularly high receptor density across multiple neurotransmitters 694 (5HT1a, D2, D1, 5HT4, CB1, VAChT, and 5HT2a; r range = 0.34 to 0.59). For the other g-695 morphometry associations, cortex-level spatial correlations were generally smaller and positive, 696 with some exceptions. For example, there is a relatively large negative correlation between g-697 curvature and MU (r = -0.31, suggesting that regions with a higher density of MU receptors are 698 those for which more convex vertices are associated with higher q) and between q-sulcal depth 699 and VACHT and mGluR5 (both r = -0.29, suggesting that regions with a higher density of VACHT 700 and mGluR5 receptors tend to be those for which a higher g is associated with a more gyral 701 vertex). At the cortex-wide level, there were no *p_spin* significant associations between the *q*-702 thickness map and any neurotransmitter receptor profiles, although, as shown in *Figure* S28 (and 703 reported in the Supplementary Tabular Data File), this appears to be because there was a mix of

692

positive and negative correlations at the regional level, which likely cancel each other out.



705

- Figure 5 Vertex-wise spatial correlations between g-morphometry associations and 33
- 707 neurobiological cortical profiles.
- Figure 5 note Spatial correlations with p_spin values < .05 are underlined. An extended version of
- this Figure is available in Figure S22, showing the underlying regional correlation summaries, like
- those in Figure 4.

711 3.5 Four major dimensions explain the majority of spatial variation across 33 cortical 712 properties

713 Since there were some qualitatively observable consistencies in spatial maps across the 33 714 cortical properties presented in *Figure* 5, we conducted a spatial component decomposition using 715 principal components analysis (PCA) to quantitatively identify any underlying spatial similarities 716 (i.e., statistical 'dimensions') that were shared across multiple maps (Figure 6). We conducted the 717 PCA across all 33 maps. It might be argued that one should reduce the dimensionality of the N =718 19 receptor maps first. However, there was no evidence that those receptor maps were more 719 similar to each other than to all other maps, i.e., the absolute correlations within 720 neurotransmitters, and those within other types of maps were not significantly different from 721 each other (t = -0.198, p = .843, neurotransmitter maps' mean |r| = 0.371, SD |r| = 0.237, other 722 maps' mean |r| = 0.367, SD |r| = 0.267).

723 Consistent with our qualitative observations, the 33 maps shared only four main spatial patterns: the first four components accounted for 65.9% of the variance, and there was a marked inflection 724 725 point in the variance explained after these four (Figure 6A and scree plot in Figure 6B). We 726 extracted the first four components with varimax rotation (loadings presented in *Figure 6C*). The 727 first component alone accounted for almost one third of the spatial variance (the loadings of PC1 728 were similar whether rotated or unrotated; coefficient of factor congruence = 0.900; Figure S18). 729 We describe these major dimensions as mapped onto the cortical surface (see Figure 6D), which 730 appear to reflect core organisational principles of the brain's neurobiology across multiple scales:

- 731 PC1 resembles previously reported latent variables of cortical macrostructure (¹⁰⁸, ⁷⁸). Its 732 cortical profile is characterized by a gradient from unimodal sensory input areas 733 (sensorimotor, primary auditory and visual / medial occipital regions) at one end of the 734 scale, and amodal association cortices (medial frontal and temporal regions) at the other 735 ⁽⁷⁸⁾. It captures multiple aspects of neurobiological information with high loadings across 736 microstructure, macrostructure, functional activity, and neurotransmitter receptor 737 density categories. The largest positive receptor loadings were for 5HT1a, MU, CB1, D1 738 and D2. NAT and GABAa-bz had negative loadings < -0.3.
- PC2 is medial temporal and is most strongly characterized by allometric scaling,
 intersubject variability and metabolism showing strong parahippocampal localisation.
 High-loading receptor maps were A4B2, M1, VAChT, CB1, MU, mGluR5 and GABA-bz.
- PC3 is an anterior-posterior component and is associated with functional activity (both
 FC similarity gradient 2 and CogPC1 Neurosynth) and the first principal component of
 cortical gene expression, as well as with cytoarchitectural staining and microstructural

similarity profiles. There is a large negative loading for GABAa-bz, and the largest positive
loadings for mGlur5, MU and H3.

PC4 is superior/inferior, with strikingly strong component scores in the insula. It is
largely a receptor-based component, with notable positive loadings for several serotonin
maps (5HT1b, 5HT2a, 5HT4, 5HTT), and all three dopamine maps (D1, D2 and DAT), as

- 749 maps (5HT1b, 5HT2a, 5HT4, 5HTT), and all three dopamine maps (D1, D2 and DAT), as
- 750 well the two glutamate maps (NMDA, and mGLuR5) and VAChT (acetylcholine).

751 We correlated these major dimensions of neurobiological organization with the *q*-morphometry 752 association maps (see Figure 6E). Notably, PC1 correlations were highest with g-volume (r = 0.39, 753 *p_spin* =.009) and *g*-surface area (*r* = 0.56, *p_spin* = .002). The strongest cortex-wide correlation 754 for PC2 is for *g*-thickness, although it is not significant in the spin test (r = 0.29, p spin = .074). The 755 only spin test p value < .05 for g-morphometry associations with PC3 is for sulcal depth (r = 0.21, 756 *p* spin = .049). PC4 had the strongest association with *q*-volume (r = 0.43, *p* spin = .010), which 757 appears to be led by g-surface area (r = 0.38, $p_{spin} = .079$) rather than g-thickness (r = 0.18, p =758 .364).

Within-region analyses results show that significant cortex-wide map correlations are underpinned by homogenous correlations at regional level (see Figure 6F). Neurobiological profiles with null correlations at the cortex level had both positive and negative correlations at the regional level, which cancel each other out but could still reveal important associations of *g*morphometry maps and core organisational principles of the human brain. These results are presented in detail in Figure S31 and the Supplementary Tabular Data File.

Together, the results show that multiple biological properties covary together in relatively few
spatial axes across the cortex. These dimensions represent multi-system neurobiological
foundations of individual differences in general cognitive functioning.



768

Figure 6 Many neurobiological profiles converge on four spatial dimensions which are spatially correlated
 with g-morphometry maps.

Figure 6 note A) A correlation plot between the 33 neurobiological profiles. B) A scree plot showing percentage
of explained variance by the first 10 PCs. The first four were extracted with varimax rotation. C) The loadings
of each neurobiological profile on the first four components. Loadings < |.3| are shown in a reduced alpha to
aid interpretations. D) PC scores mapped onto the cortex. E) Correlations of four PCs with g ~ morphometry
associations (with spin test < .05 underlined). F) The distributions of within-region g ~ morphometry
correlations. Note that this plot does not take into consideration the number of vertices in each region.

4 Discussion

This study provides the most definitive cross-cohort characterisation of regional morphometric 777 778 brain associations with g to-date. It demonstrates how such associations vary in strength and 779 direction across the volume, surface area, thickness, curvature and sulcal depth of the cortex. We 780 also provide a compendium of spatial associations between *q*-morphometry profiles and 33 781 neurobiological profiles and discover that these 33 profiles share four major dimensions of 782 spatial cortical organization. We look at spatial correlations between *q*-morphometry 783 associations and neurobiological profiles to provide insights into the neurobiological 784 mechanisms underpinning our complex cognitive functions.

785 Using the spatial correlations approach, the current results provide further detail about g and 786 brain-neurobiology relationships. Key findings were: 1) We provide the largest to-date meta-787 analytic vertex-wise associations between brain morphometry and g (i.e., morphometry maps) 788 and characterise how they vary across the cortex; 2) Vertices with larger *q* associations tend to 789 be more susceptible to age; 3) Vertices most strongly associated with *q* were largest in males and 790 also more convex in females; 4) *q*-morphometry associations maps substantially overlap with 791 maps of neurobiological properties; 5) The cortical spatial patterning of 33 neurobiological maps 792 can be concisely summarised in 4 PCs, and their correlations with *g*-morphometry patterns are 793 presented.

794 There were *p_spin* significant correlations between several *g*-morphometry and neurobiological 795 profiles, such as dopamine, serotonin, VACHT, CB1 and NMDA neurotransmitter receptor 796 densities. These neurotransmitter-cognition results are in line with previous reports e.g., 797 dopamine and serotonin have previously been shown to be important for cognitive processes 798 (e.g., ¹⁰⁹, ¹¹⁰, ¹¹¹), VACHT dysfunction has been shown to be related to intellectual disabilities and 799 Parkinson's Disease, as well as to prefrontal cortex functioning (^{112,113}), acute CB1 disruption 800 results in a decline in verbal learning and working memory performance ¹¹⁴, and NMDA has been 801 selected as a promising target for cognitive enhancement e.g., in dementia (¹¹⁵, ¹¹⁶). There was a 802 negative correlation between brain volume and metabolism, which might be explained as the 803 metabolism data were collected a rest, so one might expect these regions to be de-coupled from 804 the cognition-relevant regions we identify here. *g*-volume and *g*-surface area also had moderately 805 strong correlations with PC1 gene expression, in line with our previous work which characterised 806 g-morphometry and gene expression profile associations in more detail (3). Microstructural and 807 functional similarity gradients had differential correlations between g-surface area and g-808 thickness, which could give insights into the mechanisms behind why the spatial profiles of these 809 two *g*-morphometry profiles are different.

810 The spatial variation of the neurobiological profiles across the cortex was captured in four 811 dimensions which reflect multi-scale organisational principles of the brain that support general 812 cognitive functioning. Neurobiological PC1 resembles previously reported latent variables of 813 cortical macrostructure (¹⁰⁸, ⁷⁸). Its cortical profile is characterized by a gradient from unimodal 814 sensory input areas (sensorimotor, primary auditory and visual / medial occipital regions) at one 815 end of the scale, and amodal association cortices (medial frontal and temporal regions) at the 816 other. It captures multiple aspects of neurobiological information with high loadings across 817 microstructure, macrostructure, functional activity, and neurotransmitter receptor density 818 categories. The consistency with previous reports suggests that our applied methods are 819 promising and that the core dimensions of cortical organisation may be replicated across 820 different measures and analysis strategies. PC1 had moderate correlations with *g*-volume and *g*-821 surface area, suggesting that this dimension of neurobiological characteristics is important for 822 cognition-brain organisation.

823 In the current study we also offer a framework to extend the increasingly popular method of 824 calculating spatial correspondences between two cortical profiles (usually represented by a 825 single correlation of assumed linear correspondence). Our approach to examine vertex-wise 826 within-region spatial agreement offers important insights about the relative strength of 827 correlations for different regions and the extent of homogeneity of cortex-wide correlations 828 across regions. For correlations between age-brain and *q*-brain maps, the vast majority of within-829 region correlations are negative. For example, for cortical volume – across regions, vertices for 830 which a higher *q* is associated with higher volume tend to be the same vertices for which a higher 831 age is associated with lower volume (i.e., appears to decline more with age). On the other hand, 832 for example, for the $g \sim$ surface area and $g \sim$ thickness comparison, there are 13 regions with 833 positive associations (rs M = 0.27, SD = 0.24, range = 0.006 to 0.785) and 21 with negative 834 associations (rs M = -0.397, SD = 0.269, range = -0.012 to -0.859), showing that the concordance 835 between these two maps has a considerable amount of variation across the cortex, which is not 836 possible to tell from the overall corrical correlation (r = -0.182). Sometimes a corrical correlation 837 might be null because positive and negative within-region correlations cancel each other out. For 838 example, for $q \sim$ thickness, there are no p spin significant cortex-wise correlations with any 839 neurotransmitter receptor density maps, but at the regional level, there are both large positive 840 and negative associations for several receptor types. These within-region correlations cancel 841 each other out at the cortex-level and conceal potentially meaningful spatial correlations between 842 $g \sim$ thickness and neurotransmitters receptor densities.

The extent to which brain-behaviour associations are stable and replicable is a subject of current
debate (¹¹⁷, ¹¹⁸), and here we formally quantified the extent to which the patterning of associations

is stable across cohorts. The results show good cross-cohort agreement, suggesting that it is not
always the case that thousands of individuals are required to produce reproducible brain-wide
associations. We also provide a critical evaluation of smoothing tolerances for these associations,
which suggests that between 10-20 mm FWHM is a good choice across morphometry measures.
We further conducted cross-cohort meta-analytic data on subcortical associations with *g*, age, and
sex (described in detail in *Supplementary Analysis 2*) which contribute significantly to the
literature on this topic.

852 For our meta-analyses, methods were matched, where possible, between the UKB, GenScot and 853 LBC1936 cohorts (e.g., obtaining brain morphometric measures from FreeSurfer and including 854 multiple different cognitive test scores in our calculation of latent g scores). This consistency 855 allowed for direct quantitative comparison between cohorts and leads to improved confidence in 856 the final meta-analytic estimates of *q*-brain associations. With that said, some differences in MRI 857 data and processing protocols between the three cohorts might differentially affect the cortical 858 surface results: 1) each of the three cohorts used different scanners for MRI acquisition and, 859 although T1-weighted data provides consistent between-scanner measures $(^{119})$, we cannot rule 860 out between-cohort scanner-specific effects; 2) Desikan-Killiany parcellations were visually 861 inspected and manually edited for LBC1936 and GenScot, which would also affect the vertex-wise 862 surfaces, but manual inspections were not carried out for UKB; and 3) different FreeSurfer 863 versions were used for each cohort and are likely to have contributed to some differences in 864 estimations, alongside different types and quantity of cognitive tests. It is therefore encouraging 865 that the spatial correlations were fairly stable and that the meta-analytic results also show 866 significant associations with multi-modal biological data from independent sources.

867 All cortical maps included in the current analyses were registered to fsaverage space. Registration 868 differences might have had a small impact on the results – e.g., there were a few vertices around 869 the cortical mask that were present in some transformed fsaverage maps, and not in others (e.g. 870 there were fewer vertices included in the cortical mask for the gene expression PC1). However, 871 we only included vertices within the cortical mask across all maps in each analysis, and we would 872 expect the effects of such registration inaccuracies to be small. Additionally, efforts were made to 873 harmonise smoothing tolerances between maps for different data types, but the original sampling 874 density across the cortex was different between maps. Cortical data obtained at lower spatial 875 resolutions (e.g. neurotransmitter density maps, derived from PET data) may have contributed to some uncertainty in our vertex-wise analyses, particularly within smaller cortical structures. 876 877 The cortex-wide *r* values should thus be interpreted alongside the corresponding *p_spin* values, 878 as this limits the spatial autocorrelation effects that tend to increase with increased smoothing.

879 Although the spin test goes some way to improve the validity of spatial correlations, it is 880 important to note, too, that using Pearson's r to test spatial correlations of cortical profiles 881 assumes that we are working with linear spatial associations. This is not always the case (e.g., see 882 Figure S15), as non-linear patterns are often found. The spin test does not address the 883 heterogeneity of spatial autocorrelation effects across the cortex. Further methods are currently 884 being developed to address this issue and more accurately characterise spatial concordance 885 between cortical maps (120). The methods and cortical maps we provide here could aid more 886 detailed investigations into how and where spatial concordance/discordance between 887 neurobiological and brain structural profiles is relevant to brain-behaviour relationships.

The methods used in the current paper rely on all cortical profiles having results at all vertices. The publication and release of summary data for all vertices should be encouraged, not only those vertices that reach certain criteria (e.g., significance thresholds). While for example, cluster-based analyses can be useful for identifying the main regions of interest, presenting results for all vertices provides information about the relative patterning of effects across the whole cortex that can be directly compared with other whole-cortex profiles.

894 A limitation of this study is that participants were likely to be in relatively good health, as we 895 chose to exclude participants with self-declared neurological issues from the UKB sample. Having 896 said that, our UKB exclusion criteria did not include GP, hospital or death records, so it is likely 897 that some participants with such conditions remain in the UKB sample, and may influence the 898 findings, as some neurological diseases e.g. stroke or brain lesions are likely to affect brain-899 cognition associations. The GenScot imaging sample was biased to have more participants with 900 past or current depression than would likely be typical in the general population, in line with the 901 initial aims of their study. Depression status was not controlled for in the current analyses, and it 902 is possible that it could affect brain-cognition associations. All three cohorts used to calculate g-903 brain associations are also largely white and northern European, and so it is not clear whether 904 these findings apply in other world regions or ethnic groups. Additionally, whereas the cognitive-905 MRI data do not include childhood and adolescence (and therefore the results may not relate 906 directly to those parts of the life span), the good adulthood age coverage, absence of age 907 moderation of the meta-analytic estimates within-cohort, and clear agreement across cohorts 908 suggests that the well-powered results capture adulthood brain-*q* correlations. The open-source 909 neurobiological maps that we use here are also limited in terms of generalisation due to sample 910 characteristics, which are also not directly comparable with the cohorts used to calculate *q*-911 morphometry associations. For example, gene expression PC1 was calculated based on data from 912 6 donors, aged 24 to 57 years old, which is a small sample, and has a younger age range than the 913 current study (age range = 44 to 84 years old); the cytoarchitectural similarity maps were based

on just one donor, and the PET maps tended to be conducted on young healthy adults, with sample
sizes ranging from 8 to 204. At this stage, the results should be thought of as being at a high-level
overview stage, and be used for hypothesis generation, rather than being taken as direct evidence
of brain-behaviour relationships.

918 As further group-level brain-wide maps are made openly available, the correlational structures 919 between different cortical profiles should continue to be examined and updated. Moreover, the 920 future collection or release of individual-level data across different neurobiological 921 characteristics would enable individual differences analyses, for example, to test the relationship 922 between the density of certain receptor types and *q*-morphometry associations at the individual 923 level. This would allow for more direct associations between neurobiological characteristics and 924 g, and longitudinal studies with within-participant cognitive, brain imaging and neurobiological 925 data could also directly reveal whether which neurobiological characteristics underpin the clear 926 similarity between *g* and age-related brain patterns.

5 Conclusion

927 This study advances our understanding of how different neurobiological profiles in the human 928 cortex share spatial patterning with *g*-structural morphometry profiles. We discovered four 929 principal components, which explain 65.9% of the variance across 33 neurobiological profiles, 930 and represent major fundamental axes along which the human cortex is organised. These results 931 offer new perspectives on the neurobiological properties underlying observable brain-cognitive 932 associations. We provide important new data and a framework to study brain-behavioural 933 associations in the future.

6 References

¹ Cox, S. R., Ritchie, S. J., Fawns-Ritchie, C., Tucker-Drob, E. M., & Deary, I. J. (2019). Structural brain imaging correlates of general intelligence in UK Biobank. *Intelligence*, *76*, 101376. https://doi.org/10.1016/j.intell.2019.101376

² Jung, R. E., & Haier, R. J. (2007). The Parieto-Frontal Integration Theory (P-FIT) of intelligence: converging neuroimaging evidence. *The Behavioral and brain sciences*, *30*(2), 135–187. https://doi.org/10.1017/S0140525X07001185

³ Moodie, J. E., Harris, S. E., Harris, M. A., Buchanan, C. R., Davies, G., Taylor, A., Redmond, P., Liewald, D., Del C Valdés Hernández, M., Shenkin, S., Russ, T. C., Maniega, S. M., Luciano, M., Corley, J., Stolicyn, A., Shen, X., Steele, D., Waiter, G., Sandu-Giuraniuc, A., Bastin, M. E., ... Cox, S. R. (2023). General and specific patterns of cortical gene expression as substrates of complex cognitive function. *bioRxiv*, 2023.03.16.532915. https://doi.org/10.1101/2023.03.16.532915

⁴ Tadayon, E., Pascual-Leone, A., & Santarnecchi, E. (2020). Differential Contribution of Cortical Thickness, Surface Area, and Gyrification to Fluid and Crystallized Intelligence. *Cerebral cortex (New York, N.Y.: 1991)*, *30*(1), 215–225. https://doi.org/10.1093/cercor/bhz082

⁵ Nyberg, L., Andersson, M., & Lundquist, A. (2023). Longitudinal change-change associations of cognition with cortical thickness and surface area. *Aging brain*, *3*, 100070. https://doi.org/10.1016/j.nbas.2023.100070

⁶ Narr, K. L., Woods, R. P., Thompson, P. M., Szeszko, P., Robinson, D., Dimtcheva, T., Gurbani, M., Toga, A. W., & Bilder, R. M. (2007). Relationships between IQ and regional cortical gray matter thickness in healthy adults. *Cerebral cortex (New York, N.Y.: 1991)*, *17*(9), 2163–2171. https://doi.org/10.1093/cercor/bhl125

⁷ Gregory, M. D., Kippenhan, J. S., Dickinson, D., Carrasco, J., Mattay, V. S., Weinberger, D. R., & Berman, K. F. (2016). Regional Variations in Brain Gyrification Are Associated with General Cognitive Ability in Humans. *Current biology: CB*, *26*(10), 1301–1305. https://doi.org/10.1016/j.cub.2016.03.021

⁸ Im, K., Choi, Y. Y., Yang, J. J., Lee, K. H., Kim, S. I., Grant, P. E., & Lee, J. M. (2011). The relationship between the presence of sulcal pits and intelligence in human brains. *NeuroImage*, *55*(4), 1490–1496. https://doi.org/10.1016/j.neuroimage.2010.12.080

⁹ Liu, T., Wen, W., Zhu, W., Kochan, N. A., Trollor, J. N., Reppermund, S., Jin, J. S., Luo, S., Brodaty, H., & Sachdev, P. S. (2011). The relationship between cortical sulcal variability and cognitive performance in the elderly. *NeuroImage*, *56*(3), 865–873. https://doi.org/10.1016/j.neuroimage.2011.03.015

¹⁰ Gur, R. C., Butler, E. R., Moore, T. M., Rosen, A. F. G., Ruparel, K., Satterthwaite, T. D., Roalf, D. R., Gennatas, E. D., Bilker, W. B., Shinohara, R. T., Port, A., Elliott, M. A., Verma, R., Davatzikos, C., Wolf, D. H., Detre, J. A., & Gur, R. E. (2021). Structural and Functional Brain Parameters Related to Cognitive Performance Across Development: Replication and Extension of the Parieto-Frontal Integration Theory in a Single Sample. *Cerebral cortex (New York, N.Y.: 1991), 31*(3), 1444–1463. https://doi.org/10.1093/cercor/bhaa282 ¹¹ Deary IJ, Cox SR, Hill WD. Genetic variation, brain, and intelligence differences. Mol Psychiatry. 2021:27:335–353. https://doi.org/10.1038/s41380-021-01027-y

¹² Markello, R.D., Hansen, J.Y., Liu, ZQ. *et al.* neuromaps: structural and functional interpretation of brain maps. *Nat Methods* 19, 1472–1479 (2022). https://doi.org/10.1038/s41592-022-01625-w

¹³ Paquola, C., Royer, J., Lewis, L. B., Lepage, C., Glatard, T., Wagstyl, K., DeKraker, J., Toussaint, P. J., Valk, S. L., Collins, L., Khan, A. R., Amunts, K., Evans, A. C., Dickscheid, T., & Bernhardt, B. (2021). The BigBrainWarp toolbox for integration of BigBrain 3D histology with multimodal neuroimaging. *eLife*, *10*, e70119. https://doi.org/10.7554/eLife.70119

¹⁴ Royer, J., Paquola, C., Valk, S. L., Kirschner, M., Hong, S. J., Park, B. Y., Bethlehem, R. A. I., Leech, R., Yeo, B. T. T., Jefferies, E., Smallwood, J., Margulies, D., & Bernhardt, B. C. (2024). Gradients of Brain Organization: Smooth Sailing from Methods Development to User Community. *Neuroinformatics*, *22*(4), 623–634. https://doi.org/10.1007/s12021-024-09660-y

¹⁵ Hansen, J.Y., Shafiei, G., Markello, R.D. *et al.* Mapping neurotransmitter systems to the structural and functional organization of the human neocortex. *Nat Neurosci* **25**, 1569–1581 (2022). https://doi.org/10.1038/s41593-022-01186-3

¹⁶ Jonas, K., Lian, W., Callahan, J., Ruggero, C. J., Clouston, S., Reichenberg, A., Carlson, G. A., Bromet, E. J., & Kotov, R. (2022). The Course of General cognitive function in Individuals with Psychotic Disorders. JAMA psychiatry, 79(7), 659–666. https://doi.org/10.1001/jamapsychiatry.2022.1142

¹⁷ van der Lee, S. J., Teunissen, C. E., Pool, R., Shipley, M. J., Teumer, A., Chouraki, V., Melo van Lent, D., Tynkkynen, J., Fischer, K., Hernesniemi, J., Haller, T., Singh-Manoux, A., Verhoeven, A., Willemsen, G., de Leeuw, F. A., Wagner, H., van Dongen, J., Hertel, J., Budde, K., Willems van Dijk, K., ... van Duijn, C. M. (2018). Circulating metabolites and general cognitive function and dementia: Evidence from 11 cohort studies. Alzheimer's & dementia: the journal of the Alzheimer's Association, 14(6), 707–722. https://doi.org/10.1016/j.jalz.2017.11.012

¹⁸ Deary, I. J., Corley, J., Gow, A. J., Harris, S. E., Houlihan, L. M., Marioni, R. E., Penke, L., Rafnsson, S. B., & Starr, J. M. (2009). Age-associated cognitive decline. *British medical bulletin*, *92*, 135–152. https://doi.org/10.1093/bmb/ldp033

¹⁹ Cox, S. R., Harris, M. A., Ritchie, S. J., Buchanan, C. R., Valdés Hernández, M. C., Corley, J., Taylor, A. M., Madole, J. W., Harris, S. E., Whalley, H. C., McIntosh, A. M., Russ, T. C., Bastin, M. E., Wardlaw, J. M., Deary, I. J., & Tucker-Drob, E. M. (2021). Three major dimensions of human brain cortical ageing in relation to cognitive decline across the eighth decade of life. *Molecular psychiatry*, *26*(6), 2651–2662. https://doi.org/10.1038/s41380-020-00975-1

²⁰ Yan, J. Iturria-Medina, Y., Bezgin, G., Toussaint, P-J., Hilger, K., Genç, E., Evans, A., Karama, S. Association between Brain Morphometry and Cognitive Function during Adolescence: Insights from a Comprehensive Large-Scale Analysis from 9 to 15 Years Old. bioRxiv 2024.06.18.599653; doi: https://doi.org/10.1101/2024.06.18.599653

²¹ Bachmann, T., Schroeter, M. L., Chen, K., Reiman, E. M., Weise, C. M., & Alzheimer's Disease Neuroimaging Initiative (2023). Longitudinal changes in surface-based brain morphometry measures in amnestic mild cognitive impairment and Alzheimer's Disease. *NeuroImage. Clinical*, *38*, 103371. https://doi.org/10.1016/j.nicl.2023.103371 ²² Salthouse T. A. (2005). Relations between cognitive abilities and measures of executive function. *Neuropsychology*, *19*(4), 532–545. https://doi.org/10.1037/0894-4105.19.4.532

²³ Johnson, W., Bouchard, T. J., Jr., Krueger, R. F., McGue, M., & Gottesman, I. I. (2004). Just one *g*: Consistent results from three test batteries. *Intelligence*, *32*(1), 95–107. <u>https://doi.org/10.1016/S0160-2896(03)00062-X</u>

²⁴ Johnson, W., Nijenhuis, J.T., & Bouchard, T.J. (2008). Still just 1 g: Consistent results from five test batteries. *Intelligence, 36*, 81-95. https://doi.org/10.1016/j.intell.2007.06.001.

²⁵ Deary I. J. Intelligence. Annual review of psychology, 63, 453–482. 10.1146/annurev-psych-120710-100353 (2012).

²⁶ Panizzon, M. S. et al. Genetic and Environmental Influences of General Cognitive function: Is *g* a valid latent construct? *Intelligence*, *43*, 65–76. <u>10.1016/j.intell.2014.01.008</u> (2014).

²⁷ Deary, I. J. (2014). The Stability of Intelligence from Childhood to Old Age. Current Directions in Psychological Science, 23(4), 239-245. https://doi.org/10.1177/0963721414536905

²⁸ Hansen, J. Y., Markello, R. D., Tuominen, L., Nørgaard, M., Kuzmin, E., Palomero-Gallagher, N., Dagher, A., & Misic, B. (2022). Correspondence between gene expression and neurotransmitter receptor and transporter density in the human brain. *NeuroImage*, *264*, 119671. https://doi.org/10.1016/j.neuroimage.2022.119671

²⁹ Huang, T., Hua, Q., Zhao, X., Tian, W., Cao, H., Xu, W., Sun, J., Zhang, L., Wang, K., & Ji, G. J. (2024). Abnormal functional lateralization and cooperation in bipolar disorder are associated with neurotransmitter and cellular profiles. *Journal of affective disorders*, *369*, 970–977. Advance online publication. https://doi.org/10.1016/j.jad.2024.10.108

³¹ Kim, C. Y., Park, Y., Namgung, J. Y., Park, Y., & Park, B. Y. (2024). The macroscale routing mechanism of structural brain connectivity related to body mass index. *Human brain mapping*, *45*(13), e70019. https://doi.org/10.1002/hbm.70019

³² Hansen, J.Y., Shafiei, G., Markello, R.D. *et al.* Mapping neurotransmitter systems to the structural and functional organization of the human neocortex. *Nat Neurosci* **25**, 1569–1581 (2022). https://doi.org/10.1038/s41593-022-01186-3

³³ Sudlow, C., Gallacher, J., Allen, N., Beral, V., Burton, P., Danesh, J., Downey, P., Elliott, P., Green, J., Landray, M., Liu, B., Matthews, P., Ong, G., Pell, J., Silman, A., Young, A., Sprosen, T., Peakman, T., & Collins, R. (2015). UK biobank: an open access resource for identifying the causes of a wide range of complex diseases of middle and old age. *PLoS medicine*, *12*(3), e1001779. https://doi.org/10.1371/journal.pmed.1001779

³⁴ Habota, T., Sandu, A. L., Waiter, G. D., McNeil, C. J., Steele, J. D., Macfarlane, J. A., Whalley, H. C., Valentine, R., Younie, D., Crouch, N., Hawkins, E. L., Hirose, Y., Romaniuk, L., Milburn, K., Buchan, G., Coupar, T., Stirling, M., Jagpal, B., MacLennan, B., Priba, L., ... McIntosh, A. M. (2021). Cohort profile for the STratifying Resilience and Depression Longitudinally (GenScot) study: A depression-focused investigation of Generation Scotland, using detailed clinical, cognitive, and neuroimaging assessments. *Wellcome open research*, *4*, 185. https://doi.org/10.12688/wellcomeopenres.15538.2

³⁵ Deary, I. J., Gow, A. J., Taylor, M. D., Corley, J., Brett, C., Wilson, V., Campbell, H., Whalley, L. J., Visscher, P. M., Porteous, D. J., & Starr, J. M. (2007). The Lothian Birth Cohort 1936: a study to examine influences on cognitive ageing from age 11 to age 70 and beyond. *BMC geriatrics*, *7*, 28. https://doi.org/10.1186/1471-2318-7-28

³⁶ Taylor, A. M., Pattie, A., & Deary, I. J. (2018). Cohort Profile Update: The Lothian Birth Cohorts of 1921 and 1936. *International journal of epidemiology*, *47*(4), 1042–1042r. https://doi.org/10.1093/ije/dyy022

³⁷ Fawns-Ritchie, C., & Deary, I. J. (2020). Reliability and validity of the UK Biobank cognitive tests. *PloS* one, 15(4), e0231627. https://doi.org/10.1371/journal.pone.0231627

³⁸ Ritchie, S. J., Tucker-Drob, E. M., Cox, S. R., Corley, J., Dykiert, D., Redmond, P., Pattie, A., Taylor, A. M., Sibbett, R., Starr, J. M., & Deary, I. J. (2016). Predictors of ageing related decline across multiple cognitive functions. *Intelligence*, *59*, 115–126. https://doi.org/10.1016/j.intell.2016.08.007

³⁹ Tucker-Drob, E. M., Briley, D. A., Starr, J. M., & Deary, I. J. (2014). Structure and correlates of cognitive aging in a narrow age cohort. *Psychology and aging*, *29*(2), 236–249. https://doi.org/10.1037/a0036187

⁴⁰ Miller, K. L., Alfaro-Almagro, F., Bangerter, N. K., Thomas, D. L., Yacoub, E., Xu, J., Bartsch, A. J., Jbabdi, S., Sotiropoulos, S. N., Andersson, J. L., Griffanti, L., Douaud, *g.*, Okell, T. W., Weale, P., Dragonu, I., Garratt, S., Hudson, S., Collins, R., Jenkinson, M., Matthews, P. M., ... Smith, S. M. (2016). Multimodal population brain imaging in the UK Biobank prospective epidemiological study. *Nature neuroscience*, *19*(11), 1523–1536. https://doi.org/10.1038/nn.4393

⁴¹ Wardlaw, J. M., Bastin, M. E., Valdés Hernández, M. C., Maniega, S. M., Royle, N. A., Morris, Z., Clayden, J. D., Sandeman, E. M., Eadie, E., Murray, C., Starr, J. M., & Deary, I. J. (2011). Brain aging, cognition in youth and old age and vascular disease in the Lothian Birth Cohort 1936: rationale, design and methodology of the imaging protocol. *International journal of stroke: official journal of the International Stroke Society*, *6*(6), 547–559. https://doi.org/10.1111/j.1747-4949.2011.00683.x

⁴² Cox, S. R., Lyall, D. M., Ritchie, S. J., Bastin, M. E., Harris, M. A., Buchanan, C. R., Fawns-Ritchie, C., Barbu, M. C., de Nooij, L., Reus, L. M., Alloza, C., Shen, X., Neilson, E., Alderson, H. L., Hunter, S., Liewald, D. C., Whalley, H. C., McIntosh, A. M., Lawrie, S. M., Pell, J. P., ... Deary, I. J. (2019). Associations between vascular risk factors and brain MRI indices in UK Biobank. *European heart journal*, *40*(28), 2290–2300. https://doi.org/10.1093/eurheartj/ehz100

⁴³ Page, D., Buchanan, C. R., Moodie, J. E., Harris, M. A., Taylor, A., Valdés Hernández, M., Muñoz Maniega, S., Corley, J., Bastin, M. E., Wardlaw, J. M., Russ, T. C., Deary, I. J., & Cox, S. R. (2024). Examining the neurostructural architecture of intelligence: The Lothian Birth Cohort 1936 study. *Cortex; a journal devoted to the study of the nervous system and behavior, 178,* 269–286. https://doi.org/10.1016/j.cortex.2024.06.007

⁴⁴ Markello, R. D. et al. Standardizing workflows in imaging transcriptomics with the abagen toolbox. Elife 10, e72129 (2021). <u>https://doi.org/10.7554/eLife.72129</u>

⁴⁵ Glasser MF, Coalson TS, Robinson EC, Hacker CD, Harwell J, Yacoub E, Ugurbil K, Andersson J, Beckmann CF, Jenkinson M, Smith SM, Van Essen DC. A multi-modal parcellation of human cerebral cortex. Nature. (2016) 11;536(7615):171-178. doi: 10.1038/nature18933.

⁴⁶ Royer, J., Rodríguez-Cruces, R., Tavakol, S. *et al.* An Open MRI Dataset for Multiscale Neuroscience. *Sci Data* **9**, 569 (2022). https://doi.org/10.1038/s41597-022-01682-y

⁴⁷ Mueller, S. et al. Individual variability in functional connectivity architecture of the human brain. *Neuron* **77**, 586–595 (2013). https://doi.org/10.1016/j.neuron.2012.12.028

⁴⁸ Yarkoni, T., Poldrack, R. A., Nichols, T. E., Van Essen, D. C. & Wager, T. D. Large-scale automated synthesis of human functional neuroimaging data. *Nat. Methods* **8**, 665–670 (2011). https://doi.org/10.1038/nmeth.1635

⁴⁹ Savli, M. et al. Normative database of the serotonergic system in healthy subjects using multi-tracer PET. *Neuroimage* **63**, 447–459 (2012). Doi: 10.1016/j.neuroimage.2012.07.001

⁵⁰ Gallezot, J.-D. et al. Kinetic modeling of the serotonin 5-HT_{1B} receptor radioligand [¹¹C] P943 in humans. *J. Cereb. Blood Flow Metab.* **30**, 196–210 (2010). https://doi.org/10.1038/jcbfm.2009.195

⁵¹ Beliveau, V. et al. A high-resolution in vivo atlas of the human brain's serotonin system. *J. Neurosci.* **37**, 120–128 (2017). doi: <u>10.1523/JNEUROSCI.2830-16.2016</u>

⁵² Radhakrishnan, R. et al. Age-related change in 5-HT₆ receptor availability in healthy male volunteers measured with ¹¹C-GSK215083 PET. *J. Nucl. Med.* **59**, 1445–1450 (2018). doi: <u>10.2967/jnumed.117.206516</u>

⁵³ Kaller, S. et al. Test-retest measurements of dopamine D₁-type receptors using simultaneous PET/MRI imaging. *Eur. J. Nucl. Med. Mol. Imaging* **44**, 1025–1032 (2017). https://doi.org/10.1007/s00259-017-3645-0

⁵⁴ Smith, C. T. et al. Partial-volume correction increases estimated dopamine D2-like receptor binding potential and reduces adult age differences. *J. Cereb. Blood Flow Metab.* **39**, 822–833 (2019). https://doi.org/10.1177/0271678X17737693

⁵⁵ Sandiego, C. M. et al. Reference region modeling approaches for amphetamine challenge studies with [¹¹C] FLB457 and PET. *J. Cereb. Blood Flow Metab.* **35**, 623–629 (2015). DOI: 10.1038/jcbfm.2014.237

⁵⁶ Dukart, J. et al. Cerebral blood flow predicts differential neurotransmitter activity. *Sci. Rep.* **8**, 4074 (2018). https://doi.org/10.1038/s41598-018-22444-0

⁵⁷ Ding, Y.-S. et al. PET imaging of the effects of age and cocaine on the norepinephrine transporter in the human brain using (S, S)-[11C] O-methylreboxetine and HRRT. Synapse 64, 30–38 (2010). DOI: 10.1002/syn.20696

⁵⁸ Gallezot, J.-D. et al. Determination of receptor occupancy in the presence of mass dose: [¹¹C] GSK189254 PET imaging of histamine H³ receptor occupancy by PF-03654746. *J. Cereb. Blood Flow Metab.* **37**, 1095– 1107 (2017). doi: <u>10.1177/0271678X16650697</u> ⁵⁹ Hillmer, A. T. et al. Imaging of cerebral $\alpha_4\beta_2^*$ nicotinic acetylcholine receptors with (–)-[¹⁸F] flubatine PET: implementation of bolus plus constant infusion and sensitivity to acetylcholine in human brain. *Neuroimage* **141**, 71–80 (2016). DOI: 10.1016/j.neuroimage.2016.07.026

⁶⁰ Naganawa, M. et al. First-in-human assessment of ¹¹C-LSN3172176, an M1 muscarinic acetylcholine receptor PET radiotracer. *J. Nucl. Med.* **62**, 553–560 (2021). DOI: 10.2967/jnumed.120.246967

⁶¹ Aghourian, M. et al. Quantification of brain cholinergic denervation in Alzheimer's disease using PET imaging with [¹⁸F]-FEOBV. *Mol. Psychiatry* **22**, 1531–1538 (2017). DOI: 10.1038/mp.2017.183

⁶² Bedard, M.-A. et al. Brain cholinergic alterations in idiopathic REM sleep behaviour disorder: a PET imaging study with ¹⁸F-FEOBV. *Sleep Med.* **58**, 35–41 (2019). DOI: 10.1016/j.sleep.2018.12.020

⁶³ Normandin, M. D. et al. Imaging the cannabinoid CB1 receptor in humans with [¹¹C] OMAR: assessment of kinetic analysis methods, test-retest reproducibility, and gender differences. *J. Cereb. Blood Flow Metab.* **35**, 1313–1322 (2015). DOI: 10.1038/jcbfm.2015.46

⁶⁴ Kantonen, T. et al. Interindividual variability and lateralization of μ-opioid receptors in the human brain. *Neuroimage* **217**, 116922 (2020). <u>https://doi.org/10.1016/j.neuroimage.2020.116922</u>

⁶⁵ Galovic, M. et al. In vivo NMDA receptor function in people with NMDA receptor antibody encephalitis. Preprint at <u>https://www.medrxiv.org/content/10.1101/2021.12.04.21267226v1</u> (2021).

⁶⁶ Galovic, M. et al. Validation of a combined image derived input function and venous sampling approach for the quantification of [¹⁸F]GE-179 PET binding in the brain. *Neuroimage* **237**, 118194 (2021). DOI: 10.1016/j.neuroimage.2021.118194

⁶⁷ Smart, K. et al. Sex differences in [¹¹C] ABP688 binding: a positron emission tomography study of mglu5 receptors. *Eur. J. Nucl. Med. Mol. Imaging* **46**, 1179–1183 (2019). https://doi.org/10.1007/s00259-018-4252-4

⁶⁸ DuBois, J. M. et al. Characterization of age/sex and the regional distribution of mglur5 availability in the healthy human brain measured by high-resolution [¹¹C] ABP688 PET. *Eur. J. Nucl. Med. Mol. Imaging* **43**, 152–162 (2016). https://doi.org/10.1007/s00259-015-3167-6

⁶⁹ Nørgaard, M. et al. A high-resolution in vivo atlas of the human brain's benzodiazepine binding site of GABA_A receptors. *Neuroimage* **232**, 117878 (2021). DOI: 10.1016/j.neuroimage.2021.117878

⁷⁰ Glasser MF, Van Essen DC. Mapping human cortical areas in vivo based on myelin content as revealed by T1- and T2-weighted MRI. J Neurosci. 2011 Aug 10;31(32):11597-616. doi: 10.1523/JNEUROSCI.2180-11. 2011..

⁷¹ Sandrone, S. *et al.* Mapping myelin in white matter with T1-weighted/T2-weighted maps: Discrepancy with histology and other myelin MRI measures. *Brain Struct. Funct.* **228**, 525–535 (2023). DOI: <u>10.1007/s00429-022-02600-z</u>

⁷² Mühlau M. (2022). T1/T2-weighted ratio is a surrogate marker of demyelination in multiple sclerosis:
 No. *Multiple* sclerosis (Houndmills, Basingstoke, England), 28(3), 355–356.
 https://doi.org/10.1177/13524585211063622

⁷³ Reardon, P. K., Seidlitz, J., Vandekar, S., Liu, S., Patel, R., Park, M. T. M., Alexander-Bloch, A., Clasen, L. S., Blumenthal, J. D., Lalonde, F. M., Giedd, J. N., Gur, R. C., Gur, R. E., Lerch, J. P., Chakravarty, M. M., Satterthwaite, T. D., Shinohara, R. T., & Raznahan, A. (2018). Normative brain size variation and brain shape diversity in humans. *Science (New York, N.Y.)*, *360*(6394), 1222–1227. https://doi.org/10.1126/science.aar2578

⁷⁴ Amunts, K., Lepage, C., Borgeat, L., Mohlberg, H., Dickscheid, T., Rousseau, M. É., Bludau, S., Bazin, P. L., Lewis, L. B., Oros-Peusquens, A. M., Shah, N. J., Lippert, T., Zilles, K., & Evans, A. C. (2013). BigBrain: an ultrahigh-resolution 3D human brain model. *Science (New York, N.Y.)*, *340*(6139), 1472–1475. https://doi.org/10.1126/science.1235381

⁷⁵ Royer, J., Rodriguez-Cruces, R., Tavakol, S., Lariviere, S., Herholz, P., Li, Q, Vos de Wael, R., Paquola, C., Benkarim, O., Park, B., Lowe, A. J., Margulies, D., Smallwood, J., Bernasconi, A., Bernasconi, N., Frauscher, B., Bernhardt, B. C. (2021). An open MRI dataset for multiscale neuroscience. bioRxiv. https://doi.org/10.1101/2021.08.04.454795

⁷⁶ Coifman RR, & Lafon S. (2006) Diffusion maps. *Applied and Computational Harmonic Analysis* 21:5–30. https://doi.org/10.1016/j.acha.2006.04.006

⁷⁷ Paquola C, Vos De Wael R, Wagstyl K, Bethlehem RAI, Hong S-J, Seidlitz J, et al. (2019) Microstructural and functional gradients are increasingly dissociated in transmodal cortices. PLoS Biol 17(5): e3000284. https://doi.org/10.1371/journal.pbio.3000284

⁷⁸ Margulies, D. S., Ghosh, S. S., Goulas, A., Falkiewicz, M., Huntenburg, J. M., Langs, G., Bezgin, G., Eickhoff, S. B., Castellanos, F. X., Petrides, M., Jefferies, E., & Smallwood, J. (2016). Situating the default-mode network along a principal gradient of macroscale cortical organization. *Proceedings of the National Academy of Sciences of the United States of America*, *113*(44), 12574–12579. https://doi.org/10.1073/pnas.1608282113

⁷⁹ Vaishnavi, S. N., Vlassenko, A. G., Rundle, M. M., Snyder, A. Z., Mintun, M. A., & Raichle, M. E. (2010). Regional aerobic glycolysis in the human brain. *Proceedings of the National Academy of Sciences of the United States of America*, *107*(41), 17757–17762. https://doi.org/10.1073/pnas.1010459107

⁸⁰ Alexander-Bloch AF, Shou H, Liu S, Satterthwaite TD, Glahn DC, Shinohara RT, Vandekar SN, Raznahan A. On testing for spatial correspondence between maps of human brain structure and function. Neuroimage. (2018). 178:540-551. doi: 10.1016/j.neuroimage.2018.05.070.

⁸¹ Fürtjes, A. E., Cole, J. H., Couvy-Duchesne, B., & Ritchie, S. J. (2023). A quantified comparison of cortical atlases on the basis of trait morphometricity. Cortex: a journal devoted to the study of the nervous system and behavior, 158, 110–126. https://doi.org/10.1016/j.cortex.2022.11.001

⁸² Lerch, J. P., & Evans, A. C. (2005). Cortical thickness analysis examined through power analysis and a population simulation. *NeuroImage*, *24*(1), 163–173. https://doi.org/10.1016/j.neuroimage.2004.07.045

⁸³ Buchy, L., Barbato, M., Makowski, C., Bray, S., MacMaster, F. P., Deighton, S., & Addington, J. (2017). Mapping structural covariance networks of facial emotion recognition in early psychosis: A pilot study. *Schizophrenia research*, *189*, 146–152. https://doi.org/10.1016/j.schres.2017.01.054

⁸⁴ Chung, M. K., Worsley, K. J., Robbins, S., Paus, T., Taylor, J., Giedd, J. N., Rapoport, J. L., & Evans, A. C. (2003). Deformation-based surface morphometry applied to gray matter deformation. *NeuroImage*, *18*(2), 198–213. https://doi.org/10.1016/s1053-8119(02)00017-4

⁸⁵ Nho, K., Risacher, S. L., Crane, P. K., DeCarli, C., Glymour, M. M., Habeck, C., Kim, S., Lee, G. J., Mormino, E., Mukherjee, S., Shen, L., West, J. D., Saykin, A. J., & Alzheimer's Disease Neuroimaging Initiative--ADNI (2012). Voxel and surface-based topography of memory and executive deficits in mild cognitive impairment and Alzheimer's disease. *Brain imaging and behavior*, *6*(4), 551–567. https://doi.org/10.1007/s11682-012-9203-2

⁸⁶ Lamballais, S., & Muetzel, R. L. (2021). QDECR: A Flexible, Extensible Vertex-Wise Analysis Framework in R. *Frontiers in neuroinformatics*, *15*, 561689. https://doi.org/10.3389/fninf.2021.561689

⁸⁷ Holla, B., Bharath, R. D., Venkatasubramanian, G., and Benegal, V. (2019) Altered brain cortical maturation is found in adolescents with a family history of alcoholism, *Addiction Biology*, 24, 835–845. doi: <u>https://doi.org/10.1111/adb.12662</u>.

⁸⁸ Wang, Y., Jiang, Y., Lu, H., Tian, W., Li, P., Xu, K., Fan, M., Zhao, X., Dong, Q., Jin, L., Chen, J., Cui, M., & Chen, X. (2022). Cross-sectional associations between cortical thickness and independent gait domains in older adults. *Journal of the American Geriatrics Society*, *70*(9), 2610–2620. <u>https://doi.org/10.1111/jgs.17840</u>

⁸⁹ Cox, S. R., Dickie, D. A., Ritchie, S. J., Karama, S., Pattie, A., Royle, N. A., Corley, J., Aribisala, B. S., Valdés Hernández, M., Muñoz Maniega, S., Starr, J. M., Bastin, M. E., Evans, A. C., Wardlaw, J. M., & Deary, I. J. (2016). Associations between education and brain structure at age 73 years, adjusted for age 11 IQ. Neurology, 87(17), 1820–1826. https://doi.org/10.1212/WNL.00000000003247

⁹⁰ Viechtbauer, W. (2010). "Conducting meta-analyses in R with the metafor package." *Journal of Statistical Software*, 36(3), 1–48. https://doi.org/10.18637/jss.v036.i03.

⁹¹ Rosseel, Y. (2012). lavaan: An R Package for Structural Equation Modeling. Journal of Statistical Software, 48(2), 1-36.

⁹² Basten, Ulrike & Hilger, Kirsten & Fiebach, Christian. (2015). Where smart brains are different: A quantitative meta-analysis of functional and structural brain imaging studies on intelligence. Intelligence. 51. 10-27. 10.1016/j.intell.2015.04.009.

⁹³ Panizzon, M. S., Fennema-Notestine, C., Eyler, L. T., Jernigan, T. L., Prom-Wormley, E., Neale, M., Jacobson, K., Lyons, M. J., Grant, M. D., Franz, C. E., Xian, H., Tsuang, M., Fischl, B., Seidman, L., Dale, A., & Kremen, W. S. (2009). Distinct genetic influences on cortical surface area and cortical thickness. *Cerebral cortex*, 19(11), 2728–2735. https://doi.org/10.1093/cercor/bhp026

⁹⁴ Storsve, A. B., Fjell, A. M., Tamnes, C. K., Westlye, L. T., Overbye, K., Aasland, H. W., & Walhovd, K. B. (2014). Differential longitudinal changes in cortical thickness, surface area and volume across the adult

life span: regions of accelerating and decelerating change. *The Journal of neuroscience: the official journal of the Society for Neuroscience, 34*(25), 8488–8498. https://doi.org/10.1523/JNEUROSCI.0391-14.2014

⁹⁵ Dickerson, B. C., Feczko, E., Augustinack, J. C., Pacheco, J., Morris, J. C., Fischl, B., & Buckner, R. L. (2009). Differential effects of aging and Alzheimer's disease on medial temporal lobe cortical thickness and surface area. *Neurobiology of aging*, *30*(3), 432–440. https://doi.org/10.1016/j.neurobiolaging.2007.07.022

⁹⁶ Eyler, L. T., Chen, C. H., Panizzon, M. S., Fennema-Notestine, C., Neale, M. C., Jak, A., Jernigan, T. L., Fischl, B., Franz, C. E., Lyons, M. J., Grant, M., Prom-Wormley, E., Seidman, L. J., Tsuang, M. T., Fiecas, M. J., Dale, A. M., & Kremen, W. S. (2012). A comparison of heritability maps of cortical surface area and thickness and the influence of adjustment for whole brain measures: a magnetic resonance imaging twin study. *Twin research and human genetics: the official journal of the International Society for Twin Studies*, *15*(3), 304–314. https://doi.org/10.1017/thg.2012.3

⁹⁷ Glisky EL. Changes in Cognitive Function in Human Aging. In: Riddle DR, editor. Brain Aging: Models, Methods, and Mechanisms. Boca Raton (FL): CRC Press/Taylor & Francis; 2007. Chapter 1. Available from: https://www.ncbi.nlm.nih.gov/books/NBK3885/

⁹⁸ Madole, J. W., Ritchie, S. J., Cox, S. R., Buchanan, C. R., Hernández, M. V., Maniega, S. M., Wardlaw, J. M., Harris, M. A., Bastin, M. E., Deary, I. J., & Tucker-Drob, E. M. (2021). Aging-Sensitive Networks Within the Human Structural Connectome Are Implicated in Late-Life Cognitive Declines. *Biological psychiatry*, 89(8), 795–806. https://doi.org/10.1016/j.biopsych.2020.06.010

⁹⁹ Brito, D.V.C., Esteves, F., Rajado, A.T. *et al.* Assessing cognitive decline in the aging brain: lessons from rodent and human studies. *npj Aging* **9**, 23 (2023). https://doi.org/10.1038/s41514-023-00120-6

¹⁰⁰ Raz, N. (2024). Ageing and the Brain. In eLS, John Wiley & Sons, Ltd (Ed.). <u>https://doi.org/10.1002/9780470015902.a0003375.pub3</u>

¹⁰¹ Johansson, J., Wåhlin, A., Lundquist, A., Brandmaier, A. M., Lindenberger, U., & Nyberg, L. (2022). Model of brain maintenance reveals specific change-change association between medial-temporal lobe integrity and episodic memory. *Aging brain*, *2*, 100027. https://doi.org/10.1016/j.nbas.2021.100027

¹⁰² Douaud, G., Groves, A. R., Tamnes, C. K., Westlye, L. T., Duff, E. P., Engvig, A., Walhovd, K. B., James, A., Gass, A., Monsch, A. U., Matthews, P. M., Fjell, A. M., Smith, S. M., & Johansen-Berg, H. (2014). A common brain network links development, aging, and vulnerability to disease. *Proceedings of the National Academy of Sciences of the United States of America*, *111*(49), 17648–17653. https://doi.org/10.1073/pnas.1410378111

¹⁰³ Fjell, A. M., McEvoy, L., Holland, D., Dale, A. M., Walhovd, K. B., & Alzheimer's Disease Neuroimaging Initiative (2014). What is normal in normal aging? Effects of aging, amyloid and Alzheimer's disease on the cerebral cortex and the hippocampus. *Progress in neurobiology*, *117*, 20–40. https://doi.org/10.1016/j.pneurobio.2014.02.004

¹⁰⁴ Ritchie, S. J., Cox, S. R., Shen, X., Lombardo, M. V., Reus, L. M., Alloza, C., Harris, M. A., Alderson, H. L., Hunter, S., Neilson, E., Liewald, D. C. M., Auyeung, B., Whalley, H. C., Lawrie, S. M., Gale, C. R., Bastin, M. E., McIntosh, A. M., & Deary, I. J. (2018). Sex Differences in the Adult Human Brain: Evidence from 5216 UK Biobank Participants. *Cerebral cortex (New York, N.Y.: 1991)*, *28*(8), 2959–2975. https://doi.org/10.1093/cercor/bhy109 ¹⁰⁵ Madan C. R. (2021). Age-related decrements in cortical gyrification: Evidence from an accelerated longitudinal dataset. *The European journal of neuroscience*, *53*(5), 1661–1671. https://doi.org/10.1111/ejn.15039

¹⁰⁶ Nieuwenhuys, R., & Broere, C. A. (2017). A map of the human neocortex showing the estimated overall myelin content of the individual architectonic areas based on the studies of Adolf Hopf. *Brain structure & function*, *222*(1), 465–480. https://doi.org/10.1007/s00429-016-1228-7

¹⁰⁷ Pelkmans, W., Dicks, E., Barkhof, F., Vrenken, H., Scheltens, P., van der Flier, W. M., & Tijms, B. M. (2019). Gray matter T1-w/T2-w ratios are higher in Alzheimer's disease. *Human brain mapping*, *40*(13), 3900–3909. https://doi.org/10.1002/hbm.24638

¹⁰⁸ Mesulam, M. M. From sensation to cognition. *Brain* **121**, 1013–1052 (1998).

¹⁰⁹ Rogers, R. The Roles of Dopamine and Serotonin in Decision Making: Evidence from Pharmacological Experiments in Humans. *Neuropsychopharmacol* 36, 114–132 (2011).
 https://doi.org/10.1038/npp.2010.165

¹¹⁰ Cools, R., Nakamura, K. & Daw, N. Serotonin and Dopamine: Unifying Affective, Activational, and Decision Functions. *Neuropsychopharmacol* **36**, 98–113 (2011). https://doi.org/10.1038/npp.2010.121

¹¹¹ González-Burgos, I., & Feria-Velasco, A. (2008). Serotonin/dopamine interaction in memory formation. *Progress in brain research*, *172*, 603–623. https://doi.org/10.1016/S0079-6123(08)00928-X

¹¹² Falace, A., Volpedo, G., Scala, M., Zara, F., Striano, P., & Fassio, A. (2024). V-ATPase Dysfunction in the Brain: Genetic Insights and Therapeutic Opportunities. *Cells*, *13*(17), 1441. https://doi.org/10.3390/cells13171441

¹¹³ Dembrow, N., & Johnston, D. (2014). Subcircuit-specific neuromodulation in the prefrontal cortex. *Frontiers in neural circuits*, *8*, 54. https://doi.org/10.3389/fncir.2014.00054

¹¹⁴ Zhornitsky, S., Pelletier, J., Assaf, R., Giroux, S., Li, C. R., & Potvin, S. (2021). Acute effects of partial CB₁ receptor agonists on cognition - A meta-analysis of human studies. *Progress in neuropsychopharmacology & biological psychiatry*, *104*, 110063. https://doi.org/10.1016/j.pnpbp.2020.110063

¹¹⁵ Collingridge, G. L., Volianskis, A., Bannister, N., France, G., Hanna, L., Mercier, M., Tidball, P., Fang, G., Irvine, M. W., Costa, B. M., Monaghan, D. T., Bortolotto, Z. A., Molnár, E., Lodge, D., & Jane, D. E. (2013). The NMDA receptor as a target for cognitive enhancement. *Neuropharmacology*, *64*, 13–26. https://doi.org/10.1016/j.neuropharm.2012.06.051

¹¹⁶ Chang, CH., Liu, CY., Chen, SJ. *et al.* Effect of *N*-methyl-d-aspartate receptor enhancing agents on cognition in dementia: an exploratory systematic review and meta-analysis of randomized controlled trials. *Sci Rep* **11**, 22996 (2021). https://doi.org/10.1038/s41598-021-02040-5

¹¹⁷ Marek, S., Tervo-Clemmens, B., Calabro, F.J. *et al.* Reproducible brain-wide association studies require thousands of individuals. *Nature* **603**, 654–660 (2022). https://doi.org/10.1038/s41586-022-04492-9

¹¹⁸ Liu, S., Abdellaoui, A., Verweij, K.J.H. *et al.* Replicable brain–phenotype associations require large-scale neuroimaging data. *Nat Hum Behav* **7**, 1344–1356 (2023). https://doi.org/10.1038/s41562-023-01642-5

¹¹⁹ Buchanan, C. R. et al. Comparison of structural MRI brain measures between 1.5 and 3 T: Data from the Lothian Birth Cohort 1936. *Human Brain Mapping*, 42(12), 3905–3921. <u>10.1002/hbm.25473</u> (2021).

¹²⁰ Leech, JS Smallwood, R Moran, EJH Jones, N Vowles, D Leech, EM Viegas, FE Turkheimer, F Alberti, D M argulies, E Jefferies, B Bernhardt, F Váša. The impact of heterogeneous spatial autocorrelation on comparisons of brain maps. bioRxiv 2024.06.14.598987.
 (2024). doi: <u>https://doi.org/10.1101/2024.06.14.598987</u>