Letters to the Editor

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In Vitro Fertilization May Increase the Risk of Beckwith-Wiedemann Syndrome Related to the Abnormal Imprinting of the *KCNQ1OT* Gene

To the Editor:

"Parental imprinting" refers to an epigenetic marking of genes that results in monoallelic expression. This phenomenon plays a critical role in embryogenesis and development. The epigenetic modification of the genome involves methylation changes and the remodeling of chromatin-associated proteins (Li 2002). Imprints are established during the development of germ cells, and the reprogramming of imprinting occurs within the first days after fertilization (Reik and Walter 2001). The alteration of normal imprinting patterns is implicated in a number of human genetic diseases. Among them, the Beckwith-Wiedemann syndrome (BWS [MIM 130650]) is an overgrowth syndrome secondary to the dysregulation of the imprinted 11p15 region (Maher and Reik 2000). Numerous mechanisms are involved in BWS, and ~70% of cases of BWS are related to epigenetic abnormalities at the 11p15 locus, mostly demethylation of the *KvDMR1* region of the *KCNQ1OT* (previously called "*LIT1*") gene (MIM 604115) (Engel et al. 2000; Bliek et al. 2001; Gaston et al. 2001; Weksberg et al. 2001; DeBaun et al. 2002). *KCNQ1OT* encodes a noncoding antisense transcript within intron 10 of the *KCNQ1* gene (MIM 192500) (Lee et al. 1999; Mitsuya et al. 1999; Smilinich et al. 1999) and might be involved in the regulation of parental imprinting of the centromeric domain of the 11p15 region (Fitzpatrick et al. 2002).

In sheep and cattle, epigenetic abnormalities have been shown to be involved in large offspring syndrome (LOS) (Young et al. 1998). Affected animals exhibit various phenotypes, including large size at birth. In both species, the syndrome is caused by the in vitro exposure of embryos, between fertilization and the blastocyst stage, to various unusual environments. LOS is related to the loss of imprinting of the IGF2 receptor gene (MIM 147280), which ensures internalization and degradation of IGF2 and displays an antiproliferative function (Young et al. 2001). In vitro preimplantation procedures in mice are also responsible for overgrowth, owing to the abnormal expression of various imprinted genes, particularly the genes located at distal chromosome 7 (*h19* [MIM 103280] and *igf2* [MIM 147470] genes), orthologous to the human 11p15 region (Humpherys et al. 2001; Rideout et al. 2001). In humans, a case of BWS was recently described after in vitro fertilization (IVF) (Olivennes et al. 2001). Moreover, two recent papers (DeBaun et al. 2003; Maher et al. 2003) described an increase in prevalence of assisted reproductive technologies (ART) in patients with BWS. De Baun et al. (2003) reported a sixfold increase (4.6% vs. 0.76% in the general population) and showed that four of the six patients for whom DNA was available exhibited an isolated demethylation of KvDMR1 in the KCNQ1OT gene. Maher et al. (2003) reported a threefold increase (4% vs. 0.997% in the general population) and demonstrated that two of the six patients on whom molecular analysis could be done also exhibited an isolated demethylation of KvDMR1.

Our department is a reference center in France for molecular diagnosis of BWS, and patients are referred from various medical departments (neonatology, pediatrics, genetics, and fetopathology). We studied a series of 149 patients referred for overgrowth syndromes and diagnosed as BWS, since all of them exhibited genetic or epigenetic defects at the 11p15 locus. According to the inclusion criteria described elsewhere (Gaston et al. 2001), 102 patients exhibited a complete form of BWS, and 47 exhibited an incomplete form of BWS. The techniques used to analyze the 11p15 region have been described elsewhere (Gaston et al. 2000, 2001). Epigenetic changes concerned 104 (70%) patients, most of whom ($n = 90$) exhibited a loss of *KvDMR1* methylation. Fourteen patients (9.4%) exhibited isolated hypermethylation of the H19 gene. Forty-two patients exhibited a genetic defect: 11p15 uniparental disomy ($n = 35$; 23.5%) and germline *CDKN1C* (MIM 600856) mutation ($n = 7$; 4.7%). Three patients (2%) had a chromosomal abnormality.

Six of the 149 patients were born following ART. Of note, these six patients exhibited the same epigenetic abnormality (isolated demethylation of *KvDMR1* with a demethylation index varying 72%–100%) (fig. 1). All of them were sporadic cases, and one was a DZ twin. The clinical features of these patients and the procedures of ART used for their conception are summarized in table 1. As shown in table 1, the phenotypes of patients

Table 1

Clinical Characteristics of the Six Patients with BWS Born Following ART

	Characteristics of Patient						Characteristics of Other Patients with Demethylation of KvDMR1 (n = 84)	P[a]
	15	94	98	115	131	137		
ART procedure:								
Sperm	Ejaculated	Ejaculated	Ejaculated	Ejaculated	Ejaculated	Ejaculated		
ICSI	Yes	No	No	No	No	Yes		
Frozen embryo	No	No	No	Yes	No	No		
Day of transfer	Day 2	Day 3	Day 2	Day 2	Day 2	Day 5[b]		
Phenotype:								
Sex	F	F	M	F	M	F	42F/42M	
Delivery (weeks)	40	33.5	38.5	37	20[c]	32/DZ twin[d]		
Macrosomia	Yes	Yes	Yes	Yes	Yes	No	72.3%	NS[e]
Birth weight (g)/Birth length (cm)	4090/51.5	2770/48.5	4460/53.5	4400/55	?/480	1765/43		
Macroglossia	Yes	Yes	Yes	Yes	Yes	Yes	96.4%	NS
Organomegaly	No	No	No	Liver	Pancreas	No	48.7%	NS
Abdominal wall	No	Exomphalos	No	Exomphalos	No	Exomphalos	72.3%[f]	NS
Hemihyperplasia	No	No	No	No	No	No	26.9%	NS
Ear abnormalities	No	No	Yes	No	No	No	68.9%	P = .02
Hypoglycemia	Yes	No	No	Yes	…	No	45.6%	NS
Facial naevus	Yes	No	No	Yes	No	Yes	54.5%	NS
Other	Macrocephalia, cystic fibrosis	Developmental delay, pyelic dilatation	Inguinal hernia		Adrenal cytomegaly, placental chorioangioma			

[a] χ^2 test.
[b] Transfer of three embryos, two at the morula stage and one at the blastocyst stage.
[c] Spontaneous abortion.
[d] DNA from the normal twin was not available.
[e] NS = not significant.
[f] 43.4% exomphalos, 24.1% umbilical hernia, 4.8% diastasis recti.

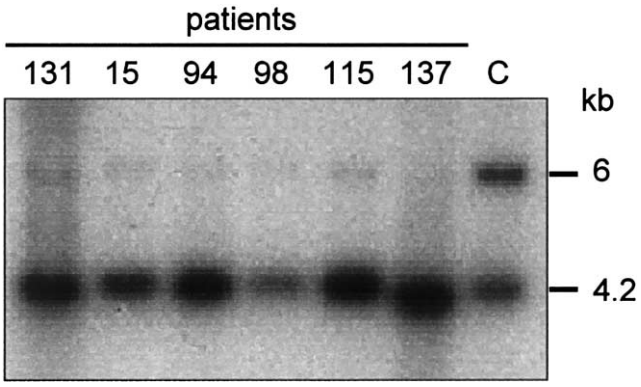

Figure 1 Methylation analysis of KvDMR1 in liver tissue (patient 131) and leukocytes (patients 15, 94, 98, 115, and 137) from the six patients with BWS born after ART and in leukocytes from a normal control (C). Genomic DNA was digested with *Bam*HI and the methylation-sensitive enzyme *Not*I. Digested samples were subjected to electrophoresis in a 0.7% agarose gel, blotted onto Hybond XL membranes, and hybridized with the HLHAY79 KvDMR1 probe corresponding to EST 68627 (ATCC; Manassas). The upper band (6 kb) is methylated and corresponds to the maternal allele. The lower band (4.2 kb) is unmethylated and corresponds to the paternal allele.

born after ART were not different from phenotypes of the other patients ($n = 84$) with isolated demethylation of KVDMR1, with the exception that only one patient born after ART exhibited ear abnormalities. These children were issued from various ART procedures: classical IVF, intracytoplasmic sperm injection (ICSI), embryo freezing, and transfer on day 2, day 3, and day 5. More recent procedures, like ICSI (two of six patients) or blastocyst transfer (one of six patients), did not prevail over other techniques. The representation of ART (4%) in our series is three times higher than that in the general population (1.3%), according to the national report of the French Ministry of Health (9,930 of 770,000 live births resulting in 1,999 from IVF, ICSI, or frozen embryo transfer). On the basis of this report, we would have expected 1.94 of the 149 patients with BWS to be born as a result of ART. To test the significance of this difference of frequencies, we used the Fisher's exact test ($P = .01$) as well as the Poisson approximation (two-tailed $P = .018$; 95% CI 1.5–8.7). Strength of the association between exposure to ART and risk of BWS is expressed by an odds ratio of 3.2 (95% CI 1.4–7.3). This rate is the same as that described by Maher et al. (2003) but lower than that described by DeBaun et al. (2003), which addressed a prospective study. In our series and in Maher's series, this rate is probably underestimated, as specific questions regarding ART have only been asked systematically in the past year.

Although the analysis of the imprinting status at chromosome 15q11-13 in children born after ICSI did not reveal an imprinting defect (Manning et al. 2000), two recent papers reported three patients with Angelman syndrome (MIM 105830) born after ICSI (Cox et al. 2002; Ørstavik et al. 2002). All three patients exhibited an imprinting defect, which is a rare cause of Angelman syndrome.

As in the previous two reports (DeBaun et al. 2003; Maher et al. 2003), our data suggest that ART may favor imprinting alterations at the centromeric imprinted 11p15 locus and, consequently, the incidence of BWS. These data highlight the need to carefully follow up children born after ART to test for BWS and other diseases related to imprinted regions. Although no specific procedures of ART appear to be associated with a risk of BWS in our series, these data lend support to the importance of precisely recording these different procedures of ART, particularly the stimulation protocol, the biological technique, the stage of maturation of the gametes, the culture media used at each step, and the timing of embryo transfer.

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Electronic-Database Information

The URL for data presented herein is as follows:

Online Mendelian Inheritance in Man (OMIM), http://www.ncbi.nlm.nih.gov/Omim (for BWS, KCNQ1OT, KCNQ1, IGF2 receptor, H19, IGF2, CDKN1C, and Angelman syndrome)

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To Trust or Not to Trust an Idiosyncratic Mitochondrial Data Set

To the Editor:

In a recent report, Silva et al. (2002) provided partial (8.8 kb) information on the mtDNA coding region (within the region 7148–15946, in the numbering of the Cambridge reference sequence [CRS]; Anderson et al. [1981]) in 40 individuals from Brazil. On the basis of the similarity in nucleotide diversity and age estimates of the four founder haplogroups A, B, C, and D, they claimed to have added new evidence for a single early entry of the founder populations into America. However, a site-by-site audit of the data reveals that their sequences are not of high enough quality to justify such statements. The authors failed to realize that a large number of mutations associated with basal branches of the worldwide mtDNA phylogeny (Finnilä et al. 2001; Maca-Meyer et al. 2001; Torroni et al. 2001; Derbeneva et al. 2002; Herrnstadt et al. 2002; Kivisild et al. 2002) were not correctly scored in their data set.

Table 1

Sequence Variation in 40 Samples Reported by Silva et al. (2002)

Sample ID	Haplogroup	Sequence on Region 7148–15976[a]	Basal Mutations Missed[b]	Accession Number
GRC0149	A	7369 7522G 8027 **8794** 8860 *11335* 12007 12705 15326 15524	11719	AF465949
KTN0130	A	**8794** 8860 11129 11288 11719 12007 12406 12705 14178 14755 14861	15326	AF465956
KPO0013	A	8764 **8794** 9392 9966 *11335* 11719 12007 12292G 12314G 12705 13708 *14566*	8860 15326	AF465957
PTJ0003	A	**8794** 11719 *11944* 12007 12705	8860 15326	AF465962
WTE1182	A	**8794** 8860 11617G 11719 12007 12292G 12618 12705 15326	...	AF465972
WPI0167	A	**8794** 8860 *10398* 11719 12007 12705 14978	15326	AF465974
YAN0623	A	**8794** 8860 10694 11719 12007 12705 13928C 15317 15326	...	AF465975
YAN0665	A	**8794** 8860 11719 12007 12705 13928C	15326	AF465976
KCR0029	A	**8794** 8860 9192 *10398* ***10400*** *11335* 12007 12314G 12705	11719 15326	AF465950
GRC0169	B4b	7626 8860 9950 *11335* 11719 11821 **13590** 15326 **15535**	8281–8289del	AF465953
KTN0209	B4b	8860 11150 11719 **13590** 14645 14647 **15535** 15914C	8281–8289del 15326	AF465955
KPO0001	B4b	7369 7522G **8281–8289del** 8860 9950 *11335* 11719 **13590 15535**	15326	AF465958
KPO0039	B4b	8736 8860 9950 10954 *11335* 11719 **13590 15535**	8281–8289del 15326	AF465959
KPO0023	B4b	8552 10604 11719 **13590** 13708 **15535**	8281–8289del 8860 15326	AF465960
QUE1876	B4b	8020 8860 *11335* 11719 12618 **13590**	8281–8289del 15326 15535	AF465964
QUE1881	B4b	8860 9950 *11335* 11719 **13590** *15043* **15535**	8281–8289del 15326	AF465965
YAN0637	B4b	8860 9950 11177 11719 12155C **13590** 13708 15106 **15535**	8281–8289del 15326	AF465980
KRC0033	B4b	7227T 7251 8860 9950 *10398* ***10400*** *11335* 11719 **13590**	8281–8289del 15535 15326	AF465951
QUE1880	B4b	7231C 8860 9950 *10398* ***10400*** *11335* 11719 12192 **13590** 15326	8281–8289del 15535	AF465968
JAP1044	B4c/B4a	*10115* **10238***del* 10398 *11335* 11719 15326 **15346**	8281–8289del 8860	AF465948
ARL0058	C	**7196A** 8078 **8584** 8701 9540 **9545** 10398 10400 10873 11719 **11914** 12705 **13263** 14783 15043 15301 15326	8860 14318 15487T	AF465945
PTJ0068	C	8701 8860 9540 **9545** 10873 11719 **11914** 12705 **13263** **14318** 14783 14788 15043 15914C	7196A 8584 10398 10400 15301 15487T 15326	AF465961
QUE1875	C	**7196A** **8584** 8701 8860 9540 **9545** *11335* 11719 **11914** 12705 **13263** 13656 14783 15043 15301	10398 10400 10873 14318 15487T 15326	AF465966
QUE1878	C	**8584** 8701 9540 **9545** 10873 *11335* 11719 **11914** 12705 **13263** 13545 14783 15043 15191	7196A 8860 10398 10400 14318 15301 15487T 15326	AF465967
YAN0669	C	8701 8848 8860 9540 **9545** 10310 10398 10400 11719 **11914** 12705 **13263** 13326 **14318** 14783 15043 15326	7196A 8584 10873 15301 15487T	AF465977
YAN0591	C	**8584** 8701 8848 8860 9540 **9545** 10873 11719 **11914** 12705 **13263** 13326 14783 15043	7196A 10398 10400 14318 15301 15487T 15326	AF465978

YAN0650	C	**7196A** 8701 8848 8860 9540 **9545** 10398 10873 11617G 11719 **11914** 12705 **13263** 13326 **14318** 14783 15043 15301	8584 10400 15487T 15326	AF465979
JAP1045	D4	8701 8860 8964 9296 9540 9824A *10115* 10398 10873 11719 12705 14783 15043 15301 15326	8414 10400 14668	AF465947
GRC0131	D4	8701 8860 9540 10816T 10873 *11335* 11914 12705 13059 13067 14783 15043	8414 10398 10400 11719 14668 15301 15326	AF465952
JAP1043	D4	8701 8860 9540 10398 10400 10873 11215 11719 12705 14783 15043 15301 15326 15874	8414 14668	AF465946
KTN0018	D	8701 8860 9540 10873 10874 12705 14687 14783 15043	10398 10400 11719 15301 15326	AF465954
PTJ0001	D	8701 8860 9540 10398 10400 10873 11150 11719 12705 14783 15043 15106 15301	15326	AF465963
TYR0004	D	8701 8860 9540 10398 10400 11719 12406 12705 12810 15301	10873 14783 15043 15326	AF465969
TYR0016	D	8701 8860 10398 10400 10819 10873 10874 11719 12406 12705 12810	9540 14783 15043 15301 15326	AF465970
NGR0524	L2a	**7175** 7256 **7274** 7521 *8047del* 8701 8860 **9221** 9540 **10115** 10398 10873 11719 **11914 11944** 12314G **12693** 12705 **13590** 13650	7771 8206 13803 14566 15301 15326 15784	AF465941
NGR0522	L2a	7256 **7274** 7521 **7771** 8701 8860 **9221** 9540 10873 10994C 11029T *11335* 11719 **11914 11944** 12292G **12693** 12705 **13590** 13650 **13803 15784** *15802del 15848del*	7175 8206 10115 10398 14566 15301 15326	AF465942
NGR0475	L2a	**7175** 7256 **7274** 7521 **7771** 8701 8860 **9221** 9540 10373 10873 11719 **11914 11944 12693** 12705 **13590** 13650 **13803** *14668* **15784**	8206 10115 10398 14566 15301 15326	AF465943
NGR0510	L2a	7256 **7274** 7521 **7771** 8701 8860 **9221** 9540 **10115** 10398 10873 11617G 11719 **11914 11944 12693** 12705 **13590** 13650 **13803 15784**	7175 8206 14566 15301 15326	AF465944
WTE1150	L2a	**7175** 7256 **7274** 7521 **7771** 8701 8860 **9221 10115** 10398 10873 *11335* 11719 **11914 11944 12693** 12705 13194 **13590** 13650 **13803** 15301 15326 **15784**	8206 9540 14566	AF465973
WTE1145	U	7220A 7227T 7642 8860 9668 **11467** 11719 **12308 12372** *13590* 15326	...	AF465971

NOTE.—Sites are numbered according to the revised reference sequence (Andrews et al. 1999); suffixes A, G, C, and T indicate transversions; "del" indicates a deletion. The mutations in boldface distinguish each sequence from the nearest mtDNA ancestor of haplogroups L2′3, M, N, and R. Potential reading errors or possible phantom mutations are italicized and underlined.

[a] All bear 14766 in addition.

[b] Basal polymorphisms that were undetected or omitted by Silva et al. (2002), including 11719 and the two rare mutations (8860 and 15326) in the CRS.

In the case of the hypervariable segments of the mtDNA control region, Bandelt et al. (2001, 2002) have highlighted lab-specific idiosyncrasies through comparative phylogenetic analysis. For the coding region, the task of identifying anomalies and reconstructing their potential causes is somewhat easier because the vast majority of sites there do not appear to undergo frequent mutations. The coding region well supports a basal nesting of (monophyletic) haplogroups, many of which had already been identified through RFLP analysis and sequencing of the hypervariable segments (Richards and Macaulay 2001). For example, the basal division of Eurasian mtDNAs into macrohaplogroups M and N is amazingly clear cut. The Eurasian mtDNA phylogeny that emerges from the phylogenetic analysis of the complete mtDNA database is detailed (for east Asia) in figure 1 of Kivisild et al. (2002), which attempts a reconstruction of the mutational history. The African mtDNA phylogeny has also been well documented in recent papers (Maca-Meyer et al. 2001; Torroni et al. 2001; Herrnstadt et al. 2002).

Silva et al. (2002) reported 40 mtDNAs, of which they assigned 31 to the Native American haplogroups A, B, C, and D (according to their fig. 1). The remaining nine mtDNAs can be assigned unambiguously to the Asian haplogroups B4 and D4, the Eurasian haplogroup U, and the African haplogroup L2a (table 1), as we will argue below. Figure 1 displays the truncation (relative to the 8.8-kb fragment under study) of the rooted phylogeny that is relevant for assigning these 40 mtDNAs to their respective haplogroups. This phylogeny is unanimously supported by the earlier publications. (However, note that mutations at 15301 and 11944 were not reconstructed most parsimoniously along the African mtDNA tree shown in fig. 1 of Herrnstadt et al. [2002]). The only instances of recurrent mutations (real or not) for the mutations and haplogroups highlighted in figure 1 are then as follows: the transversion 15487T is missing in the single haplogroup C lineage of Maca-Meyer et al. (2001); in the data of Herrnstadt et al. (2002), the B4b lineage 375 has experienced a transition at 14766, the L2a lineage 223 lacks the 7521 transition, and the 14566 transition is missing in the L2a lineage 165, which is closely related to another L2a lineage (bearing the 14566 mutation) from Torroni et al. (2001) in that they both share additional mutations at 3010 and 6663.

It is conspicuous that in all five haplogroup L2a mtDNAs of Silva et al. (2002), two of the basal transitions, 8206 and 14566, characteristic of L2 and L2a, respectively, are missed. Further L2a-diagnostic mutations, such as 7175, 7771, 13803, and 15784, are not always reported in the sequences (table 1). Moreover, the five L2a lineages have a total of only 11 other (private) mutations, comprising as many as five transversions, four deletions, and only two transitions. This pattern of private mutations differs from that in the three L2a lineages (nine transitions and no other mutations) of Ingman et al. (2000) and Torroni et al. (2001) in the same mtDNA region. It thus looks as though most of the real private mutations in the L2a mtDNAs were missed and that, instead, phantom mutations were scored.

The basal mutation 15487T of haplogroup M8 (which embraces haplogroups C and Z) is omitted in all seven C lineages of Silva et al.'s data (table 1). Other basal mutations for haplogroup C lineages are missing at sites 7196A, 8584, and 14318, in different combinations. It is remarkable that even deep mutations, such as 10400, 10873, and 15301 that distinguish macrohaplogroups M and N, were overlooked in six of the seven C lineages.

Among the seven D lineages in Silva et al. (2002), three sequences share mutations or motifs with D sequences reported elsewhere (Ingman et al. 2000; Derbeneva et al. 2002). The sequence JAP1045 (from an individual of Japanese origin) shares 8964, 9296, and 9824A with a Japanese mtDNA sequence from Ingman et al. (2000) and, therefore, definitely belongs to haplogroup D4, although the two characteristic D4 transitions (8414 and 14668) are not reported in the entire data set, except for one occurrence of 14668 in an L2a sequence! Similarly, the Japanese mtDNA sequence JAP1043 bears one of the mutations, 11215, found in Siberian mtDNAs of haplogroup D4 (Ingman et al. 2000; Derbeneva et al. 2002). The Guarani sequence GRC0131 of Silva et al. (2002) shares a rare transversion 10816T and a rare transition 13059 with the Guarani sequence of Ingman et al. (2000), but only the latter one has 8414 and 14668 and is thus confirmed as belonging to D4. These cases provide strong evidence for the systematic oversight of the basal mutations 8414 and 14668 in all haplogroup D lineages from Silva et al. (2002). Just as in the case of haplogroup C, several of the basal mutations that separate M and N are also missing in most of the D lineages.

Anomalies are also found in the nine sequences belonging to haplogroup A, although it was claimed by Silva et al. (2002) to be "the most homogeneous and best characterized" cluster in figure 1. Sample KCR0029 contains basal mutations 10398 and 10400 for haplogroup M. Sample KPO0013 has the 14566 mutation that is characteristic of haplogroup L2a. Sample PTJ0003 bears the L2abc-specific mutation 11944. Moreover, site 8027 is found mutated in only one A lineage, whereas this mutation was present in all the A sequences in Herrnstadt et al. (2002) and in one Chukchi sequence reported by Ingman et al. (2000).

In the 11 B lineages, only sample KPO0001 has the 9-bp deletion in the COII/tRNA^{Lys} intergenic region, characteristic of haplogroup B. One or both of the basal mutations of B4b, 13590 and 15535, occur in all the samples (with the exception of JAP1044) and hint that they belong to B4b. It should be noted that in Herrnstadt

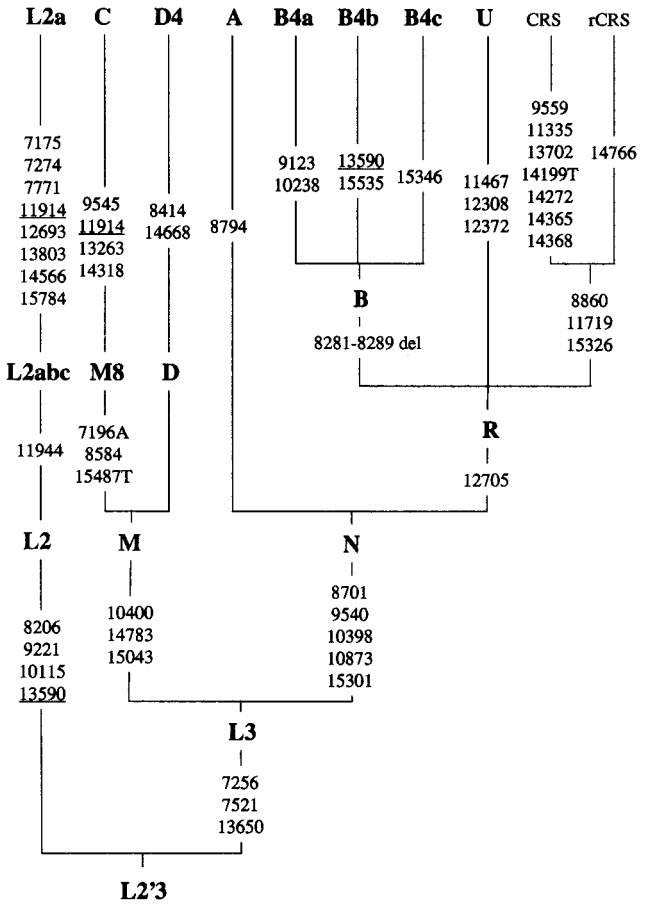

Figure 1 Skeleton of the basal mtDNA phylogeny for the haplogroups identified in the data of Silva et al. (2002). "CRS" and "rCRS" refer to the reference sequence of Anderson et al. (1981) and the revised reference sequence of Andrews et al. (1999), respectively. The suffixes A, G, C, and T indicate transversions, and "del" indicates a deletion. Parallel mutations in different branches are underlined.

et al. (2002), mutations 9950 and 11177 further defined a subhaplogroup of B4b that was baptized "B2." We suggest that the 11177 mutation could have been omitted by Silva et al. (2002) as well. The Japanese B lineage JAP1044 could belong to haplogroup B4c or, alternatively, to B4a, as judged by the 15346 mutation or the 10238 transition, respectively (if the latter was simply misreported as a deletion). Two samples, KRC0033 and QUE1880, bear the 10400 mutation of haplogroup M, whereas sample QUE1881 harbors the 15043 mutation of M.

The U sequence in Silva et al. (2002) contains the full motif of haplogroup U, plus two transversions and three transitions not previously found in the published U sequences (Ingman et al. 2000; Finnilä et al. 2001; Maca-Meyer et al. 2001; Herrnstadt et al. 2002).

Rare deletions are found in two L2a and one B lineage of Silva et al. (2002). The 15802delA and 15848delA in the cytochrome *b* gene of sample NGR0522, 8047delT in the COII gene of sample NGR0524, and 10238delT in the ND3 gene of sample JAP1044 generate premature stop codons in these genes. These rare deletions all occur at a 2-bp repeat of the deleted base and might be generated by the Sequencer reading program. It is clear that the sequences of Silva et al. (2002) harbor more rare transversions and fewer private transitions than other reported sequences (Ingman et al. 2000; Finnilä et al. 2001; Maca-Mayer et al. 2001; Torroni et al. 2001; Herrnstadt et al. 2002). One cannot exclude the possibility that true transitions were erroneously scored as transversions or deletions by Silva et al. (2002). The two rare mutations 8860 and 15326 of the CRS are also missed in most of the sequences. The mutation 11335 in the CRS, which was found to be a sequencing error (Andrews et al. 1999), was present in 16 mtDNAs.

Processes that could account for these anomalies include the following:

1. Only one strand of mtDNA was sequenced;
2. Sequences were aligned with some variant of the CRS (a likely source of problems in the past; see Macaulay et al. [1999]);
3. Sequences from different samples, especially those belonging to different haplogroups, were aligned together during the editing process (In this way, one might easily "borrow" a fragment of one sample into another when the sequences of the latter were not overlapping and, thus, introduce basal polymorphisms of one mtDNA lineage into another);
4. Possible sample crossover or contamination during data collection;
5. Relying just on the sequence scored by the Sequencer reading program without further manual checking of the chromatogram, especially relevant in the case of the rare deletions; and/or
6. PCR errors during amplification.

In summary, we have every reason to mistrust the mtDNA sequences published by Silva et al. (2002). One cannot escape the conclusion that these data are seriously flawed or, at least, are not mtDNA as we know it.

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Correction: Mitochondrial DNA Variation in Amerindians

To the Editor:

We thank Yao et al. (2003 [in this issue]) for calling our attention to inconsistencies in our data reporting mitochondrial DNA variations in Amerindians (Silva et al. 2002). We reviewed the original chromatograms and resequenced all the samples (forward and reverse). On the basis of the reanalysis of the initial data and sequencing that has been repeated, we conclude that most criticisms of Yao et al. are correct. We identified two sources of problems: (*a*) alignment with a variant CRS (Macaulay et al. 1999) and (*b*) mutations missed at regions of low-quality chromatograms in one (forward or reverse) of the first sequencing. Elimination of these two problems, by a second (and, in a few cases, a third) sequencing, careful manual checking of the chromatograms, and use of the correct rCRS reference sequence (MITOMAP) eliminated the discrepancies. A summary of all 40 corrected sequences is presented in figure 1, and the general pattern is similar to that recently reported by Herrnstadt et al. (2002). The presence of a private mutation in more than one individual or the absence of a basal mutation probably represent examples of homoplasy or of reverse mutations. Extensive homoplasy within the coding region of mtDNA has been documented (Eyre-Walker et al. 1999; Herrnstadt et al. 2002) and will probably be found more often as the number of mtDNA samples sequenced increases. For instance, the group C basal mutation 9545G was found in one individual from the haplogroup A, whereas private mutation 14460G was found in two individuals who belong to haplogroups A and D, and 15670C is present in one individual who belongs to haplogroup A and two who belong to haplogroup C (Herrnstadt et al. 2002). The finding of two similar private mutations (12406A) in two individuals of the same tribe (TYR0004 and TYR0016) is probably the consequence of a single mutational event, as is the occurrence of the reverse mutation 8584 in two individuals of another tribe (YAN0669 and YAN0650).

Recalculation of the age estimates for the four founder haplogroups on the basis of the reviewed data continues

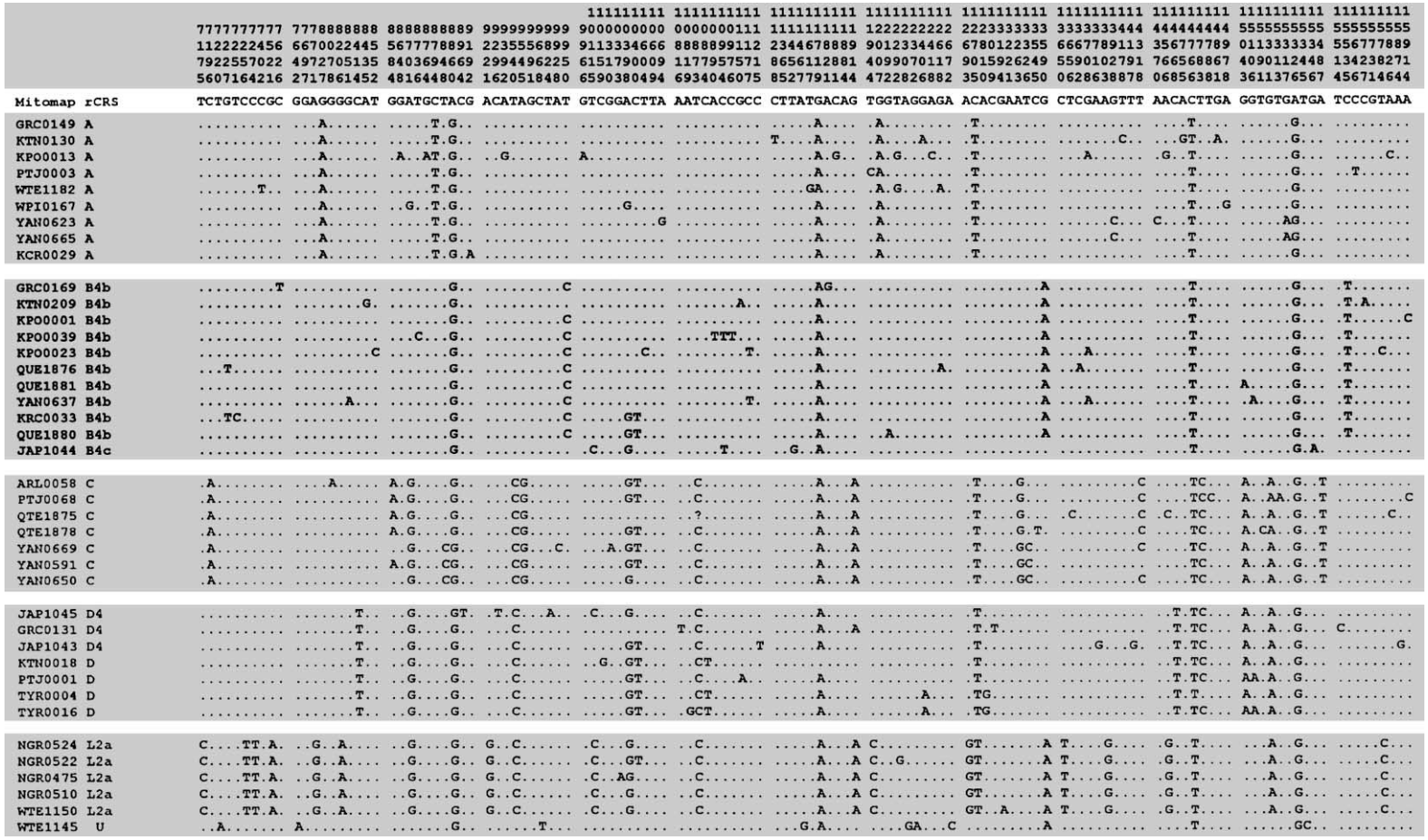

Figure 1 Data matrix showing the corrected informative nucleotide positions for the 8.8-kb mtDNA segment for 40 individuals sequenced by us

Table 1

Nucleotide Diversity and Age Estimates for mtDNA Belonging to the Four Founder Haplogroups of New World Natives

Haplogroup	No. of Sequences	Genetic Diversity[a] (SE)	Mean Age in Years[b] (95% CI)
A	10	0.73 (0.15)	15,398 (12,052–18,744)
B	11	0.75 (0.14)	15,819 (12,659–18,970)
C	9	0.64 (0.13)	13,520 (10,616–17,425)
D	5	0.86 (0.18)	18,144 (14,137–22,151)
Weighted mean		0.75 (0.15)	15,720 (12,366–19,074)

[a] $\pi(\times 10^{-3})$.
[b] Calculated as in Silva et al. (2002).

to show similarities between the four haplogroups and does not differ significantly from the previously published values (table 1). This supports our primary conclusion in favor of a single migration wave, with a mean age for the four haplogroups of 12,366–19,074 years before present.

The revised versions of the sequences have been submitted to GenBank.

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Electronic-Database Information

The URL for data presented herein is as follows:

MITOMAP, http://www.mitomap.org (for a human mitochondrial genome database)

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Reply to Silva et al.

To the Editor:

Silva et al. (2003 [in this issue]) have certainly improved their data by eliminating many of the errors in the current version of the data matrix, and they have admitted most of their innocent mistakes. Their efforts and atti-

tude should be encouraged (cf. Forster 2003). However, we are still skeptical about the corrected results presented in figure 1, for some idiosyncrasies remain and others seem to have been newly introduced. For example, some sites (e.g., 8584, 14318 [YAN0591; C type] and 14783 [TYR0004; D type]), at which Silva et al. (2003 [in this issue]) have now corrected some of the entries in their original data table, still show back mutations. Homoplasy in the coding region is much less than in the control region and may have only a few hot spots (see, e.g., table 2 of Herrnstadt et al. [2002]); the reference to Eyre-Walker et al. (1999) is not really relevant, since those authors have taken quite problematic data at face value (Kivisild and Villems 2000). The recorded variation at 10400 remains highly suspicious. It is hard to believe that 10400 has actually mutated in two B types (KRC0033 and QUE1880) and one L2a type (NGR0522) and reverted in two C types (QTE1875 and YAN0650) and two D4 types (JAP1045 and GRC0131), because no single homoplasious change at this site has been observed in >900 coding-region sequences or fragments that cover site 10400 from Ingman et al. (2000), Maca-Meyer et al. (2001), Derbeneva et al. (2002), Herrnstadt et al. (2002), and Yao et al. (2002). Moreover, site 11177 is found in only 2 of 10 B4b mtDNAs of Silva et al., which contrasts to the co-occurrence of 11177 and 9950 in all 14 B4b mtDNAs of Herrnstadt et al. (2002). To thoroughly settle these anomalies, it is imperative that the authors take notice of the potential processes that might introduce errors, as listed in our letter (Yao et al. 2003 [in this issue]), especially sample crossover. We would encourage the authors to resequence some short fragments that cover the sites listed above.

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A Multicolor FISH Assay Does Not Detect DUP25 in Control Individuals or in Reported Positive Control Cells

To the Editor:

Gratacòs et al. (2001) reported recently that the co-occurrence of panic and phobic disorders with joint laxity was associated with an interstitial duplication of the chromosomal region 15q24-q26 (named "DUP25"). DUP25, which encompasses a region of the size of 17 Mb, was observed only as mosaicism in three different forms (designated as "direct telomeric," "inverted telomeric," and "centromeric"). In each reported case, cells with DUP25 represented the majority (>50%). In addition, DUP25 mosaicism was also observed in 7% of control individuals, indicating that it could represent a

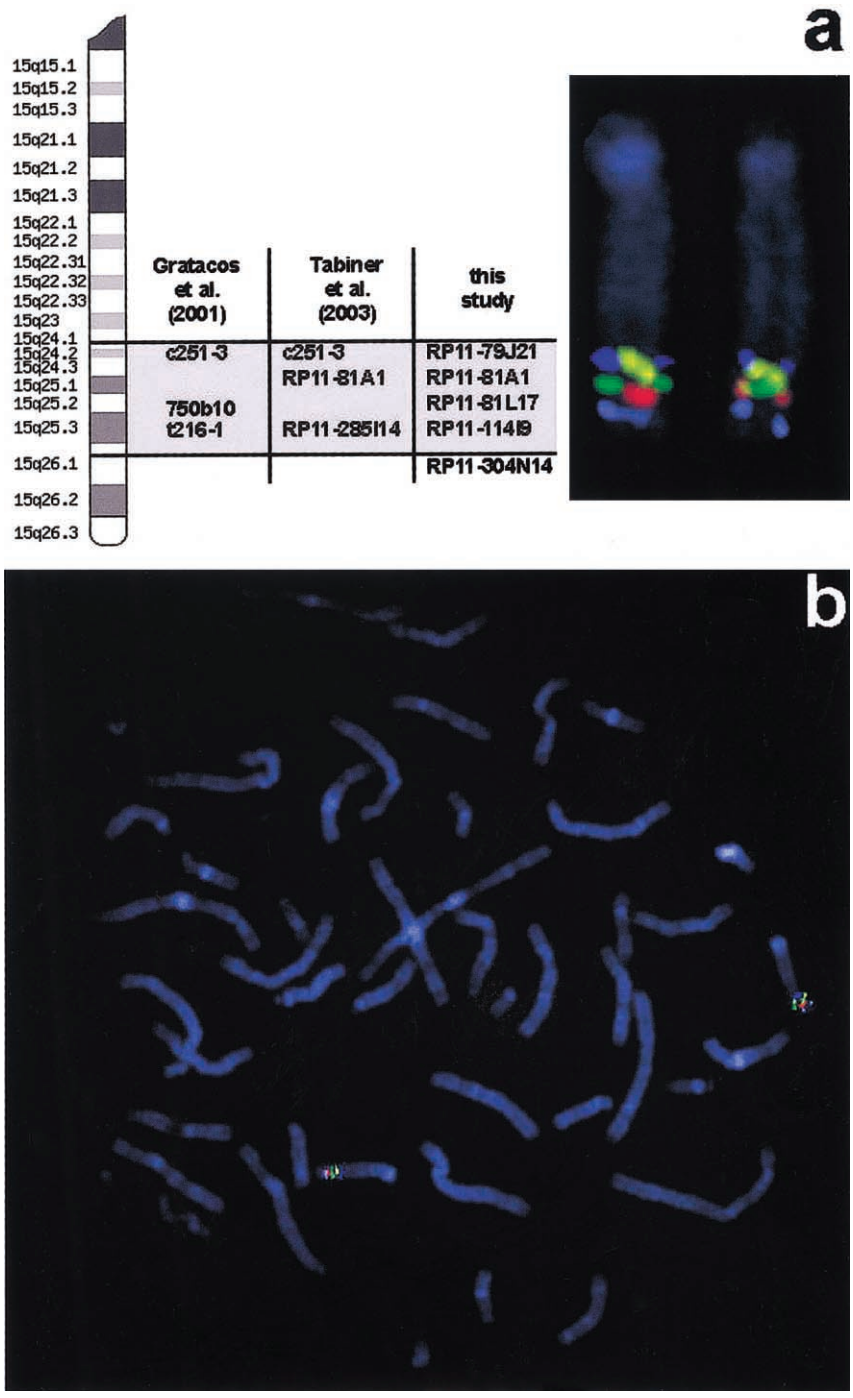

Figure 1 *a,* Ideogram of the distal part of the long arm of chromosome 15. The DUP25 region is gray shaded. For comparison, the probes used by Gratacòs et al. (2001), by Tabiner et al. (2003), and by us are shown. The FISH image shows a representative chromosome 15 pair with the normal sequence of signals (from proximal to distal): RP11-79J21 (*dark blue*), RP11-81A1 (*yellow*), RP11-81L17 (*green*), RP11-114I9 (*red*), RP11-304N14 (*light blue*). *b,* Representative metaphase spread of case B33. Ninety percent of cells were supposed to have a DUP25. In our analysis, all metaphase spreads showed a normal hybridization pattern, as depicted here.

common polymorphism. Another important implication was a proposed new, non-Mendelian mode of inheritance for DUP25.

Tabiner et al. (2003) reported in the *Journal* analyses of 40 control individuals, 16 patients with anxiety disorders, and three reported positive control cell lines. However, the authors could not find DUP25 in any sample, including the previously reported positive control cell lines. Consequently, Tabiner et al. (2003) demanded "that other groups should try to confirm or refute the presence of a polymorphic large mosaic duplication involving chromosome band 15q25 and to determine its association with anxiety disorder."

Here, we used a multicolor FISH assay to screen samples for DUP25. We assembled a FISH probe panel consisting of five different BAC probes, each labeled with a different color. The probe set includes four probes that, according to Gratacòs et al. (2001), map within DUP25 (RP11-79J21, RP11-81A1, RP11-81L17, and RP11-114I9) and one probe that is distal to DUP25 (RP11-304N14) (fig. 1*a*). Probe labeling and hybridization was done essentially according to protocols that we previously published in the *Journal* (Uhrig et al. 1999; Azofeifa et al. 2000). The cells were analyzed by use of single fluorescence filters for each fluorochrome. After image acquisition, the gray-scale images were pseudocolored and overlayed.

We analyzed slides from 70 randomly selected anonymous individuals from the southern part of Germany (Bavaria). If we assume that, indeed, 7% of the general population should have DUP25 mosaicism, the likelihood to find none among 70 randomly selected individuals is 6×10^{-3}. In each case, we evaluated at least 25 metaphase spreads, which should be sufficient, since DUP25 was reported to occur in an average of 59% of cells (Gratacòs et al. 2001). All control individuals showed a normal hybridization pattern for all probes (fig. 1*a*). We never observed duplicated probes or a change in the order of the hybridization signals.

In the next step, we requested positive control slides from the Barcelona laboratory for confirmation. Lluis Armengal kindly provided us with slides from an established cell line (P3) and a case, which was prepared directly from blood of a patient (B33). According to the analysis done in the Barcelona laboratory, DUP25 should have been present in up to 90% of cells in either case. However, our FISH assay could not identify any hint of duplication in all analyzed cells (fig. 1*b*). The slides were also evaluated by GTG-banding analysis. The average resolution was 450–500 bands/metaphase spread. As the draft human genome had been significantly improved since the publication of the Gratacòs et al. (2001) paper, we used the Ensembl Genome Browser (release 10.30.1, last updated on January 30, 2003) of the Sanger Institute for a precise and updated assessment of the DUP25 size. The distance between the genes *LOXL1* and *IQGAP1,* which corresponds to DUP25, is 16.81 Mb. Although a duplication of such a size present in 90% of cells should be detectable, experienced cytogeneticists did not find any evidence of any structural rearrangement in the respective region.

Our results indicate a difference in the frequency of DUP25 as reported by Gratacòs et al. (2001) and confirm the observations made by Tabiner et al. (2003). Most importantly, there are now two groups, which achieved different results on reported positive control cell lines as the Barcelona group.

It is difficult to explain the differences between the laboratories in scoring the positive controls. It is known that this chromosome 15 region is rich in low copy repeats (Pujana et al. 2001). Therefore, this region may be prone for structural rearrangements. However, although our multicolor FISH assay should have a high sensitivity (four clones map in DUP25), we did not find any evidence for an increased rate of structural rearrangements in the distal part of chromosome 15 in all analyzed metaphases. The differences cannot be explained by different hybridization protocols, since these are basically identical in all three laboratories (Nadal et al. 1997; Uhrig et al. 1999; Tabiner et al. 2003). Therefore, additional data by other groups should add to the clarification of the proposed causative role of large genomic duplications involving chromosome bands 15q24-q26 in panic and phobic disorders.

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Electronic-Database Information

Accession numbers and URLs for data presented herein are as follows:

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