LINKAGE STUDIES IN RICE' *

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INTRODUCTION

According to the chromosome theory, all genes on the same chromosome are linked, unless crossing over takes place. If this postulate is true, we should expect to find no more linkage groups than there are haploid chromosomes in the variety or species in question. Thus far, *Drosophila melanogaster* is the only species in the animal kingdom in which all known genes have been found to fall into one or another of the four linkage groups, each probably corresponding to one of the four haploid chromosomes. In the plant kingdom, *Zea mays* is the only species in which the linkage relations of Mendelian characters have been studied extensively.

In rice, the common species, *Oryza sativa L.,* has twenty-four chromosomes (KUWADA 1910, NAKATOMI 1923). We should expect, therefore, to find twelve linkage groups in the common races of rice. Investigations in this respect, however, have just begun. PARNELL (1917) observed the

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association between purple lining of the internode and purple glumes, and also between purple stigma and purple axil, whereas green internode and glumes are associated with purple stigma and axil. He further observed (1922) that purple pericarp color belongs to the same pattern. No satisfactory data, however, have been published regarding the chromosomal relations of the genes responsible for the characters just mentioned. **NAGAI** (1921) observed close association between the purple awn and the reddish-brown testa. **HECTOR** (1922) showed that the color of the pericarp is due either to the same factor which is responsible for ligule color or to a factor completely linked with it, and that the fact that a few plants were found with colored ligules and white grains is evidence in favor of the latter view. HECTOR (1922) further found that certain color characters on the vegetative parts of the plant are grouped in patterns or systems which are inherited together, segregating as if they were due to a single factor or due to the same interacting factors. In four cases, however, he observed that some patterns were altered. (1) In the cross Noachur \times Pookhi, the pattern "colored internode and stigma'' was altered three times out of 1,199 plants examined, giving three plants in F_2 with "colored stigma and green internode." (2) In the same cross, the pattern "colored leaf-sheath and apiculus" changed twice out of 1,199 plants giving two plants in **Fz** with "colored apiculus and green leaf-sheath." *(3)* In the cross BailabkriXPookhi the pattern "colored leaf-sheath, internode and stigma" was altered twenty-four times out of 4,687 plants examined, giving twenty-four plants with "colored leaf-sheath and stigma but green internode." (4) In the cross Agartollah $\times C_{25}$, the pattern "colored leaf-sheath, pulvinus, auricles, internode, glumes, apiculus" changed once out of 4,669 plants examined, namely, one plant was found with color in the glumes and apiculus, but with green leaf-sheath, internode, pulvinus and auricles. In all the cases thus far reviewed, while there is some indication of association between certain characters, no linkage group is definitely established.

In three instances, however, the linkage relation seems to be clear. The first of these is between the factor for awn color and the glutinous gene. In the cross, Tamanishiki XShinriki, **TAKAHASHI** (1923) found that the dominant factor for awn color (R) is coupled with the non-glutinous factor (U) , giving about twenty-one percent crossing over. The exact percentage of crossing over, however, is not certain, for **NAGAI** (1926) found 21.7 percent crossing over in one cross and 14.3 percent crossing over in another cross. Of course, the latter case may be concerned with a different factor. The second instance of linkage is given by YAMAGUCHI

(1926) who found that the factor for apiculus color (S) is coupled with the non-glutinous gene giving about **20-22** percent crossing over. Just recently, YAMAGUCHI (1927) found that the factor (F) for flowering time is also linked with the glutinous gene. The exact locus, however, is not certain.

The present studies were started in the winter of **1924.** Some twentyfive factors were studied and their chromosomal relations determined so far as practicable. Since it is not practical to make backcrosses with this plant, the linkage relations were analyzed exclusively from data obtained from the F_2 generation. The varieties used have been briefly described in an earlier paper by the author **(CHAO),** but a few important characters are pointed out in the following list.

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Characters

100 Glutinous endosperm; awnless; colorless apiculus and glumes.

- **200** Non-glutinous endosperm; long awn; tawny apiculus and glumes.
- **300** Glutinous; colorless apiculus, stigma and leaf-sheath.

400 Non-glutinous; colored apiculus, stigma and leaf-sheath.

- *600* Non-glutinous; colorless apiculus, stigma and leaf-sheath; red brown pericarp; colorless ligule, auricle, and internode; light hull with brown furrows.
- 800b Glutinous; colored apiculus, stigma and leaf-sheath; purple pericarp; purple ligule, auricle and internode; light hull without brown furrows.
- **4269** Glutinous; long spikelet and long glumes; colorless apiculus, stigma and leaf-sheath; colorless ligule, auricle and pulvinus; purple pericarp.
- **4957** Non-glutinous; short spikelet and short glumes; red apiculus; purple stigma; leafsheath purple lined; colorless ligule, auricle and pulvinus; white pericarp.

EXPERIMENTS AND RESULTS

Relation between the awn and the glutinous character

Concerning the inheritance of the awned and awnless character in rice, **YAMAGUCHI (1926)** reported a simple Mendelian **3** : **1** ratio. **NAGAI (1926)** reported three cases, one segregating in a ratio of three awned: one awnless; another segregating in a ratio of one awned: three awnless: and the third, a ratio of fifteen awned: one awnless.

The writer found a case similar to the one last mentioned. The F_1 was fully awned like the awned parent. The F_2 population consisted of four types. One type was fully awned and another fully awnless like the original parents. Of the two new types, one had awns on most of the spikelets, while the other had awns on **a** few spikelets only, as shown in figure **1.** These four types were designated as *fully awned, mostly awned, rarely awned,* and *fully awnless,* respectively. They occurred in a ratio of **GENETICS 13: Mr** *1928*

FIGURE 1.-Showing P_1 , F_1 , and F_2 generations of 101×205 , with particular reference to the **segregation of the awn character.**

12:1:2:1. When all the awned types were classified together, however, **the ratio of awned to awnless approaches very closely to 15** : **1, as shown in table 1.**

PHENOTYPES	OBSERVED	Calculated 12:1:2:1	OBSERVED	Calculated 15:1	DEVIATION
Fully awned	319	326.16			
Mostly awned	29	27.18	406	407.70	-1.7
Rarely awned	58	54.36			
Fully awnless	29	27.18	29	27.18	1.82

TABLE 1 Segregation of fifteen awned: one awnless in the F_2 generation of (101 \times 205) D.

Dev. 1.82 P.E. 3.41 $= 0.53$

These facts are explained on the basis of two pairs of factors $A_{n_1}a_{n_1}$ and $A_{n_2}a_{n_2}$, both being concerned with the production of the awn character. A_{n_1} is considered to be weaker in its action than A_{n_1} . So, A_{n_1} can produce the fully awned type with or without A_{n+1} . A_{n+2} in double dose may produce the mostly awned class, but in single dose, it will only produce the rarely awned type. The fully awnless type is due to the double recessive constitution. The interpretation that one of these two factors is weaker in its action than the other is supported by the fact that among the mostly awned and rarely awned types of F_2 plants the frequency of awned spikelets on the early or late panicles of the same plant may vary according to environmental conditions, while among the fully awned or fully awnless types the phenotypic expression is not so easily subject to environmental influence.

These two pairs of factors $A_{n_1}a_{n_1}$ and $A_{n_2}a_{n_2}$ are independent of the glutinous pair, $G_{i}g_{i}$, as shown in table 2. It is to be noted that among the **F2** population of **434** plants, **349** were non-glutinous and *85* glutinous, showing a deficiency of glutinous grains that is about *3.86* times the probable error.

	TABLE 2		
		Independent segregation between factors $A_{n1}a_{n1}$, $A_{n2}a_{n2}$ and G_1g_1 .	
PHENOTYPES	OBSERVED	Calculated 45:3:15:1	$(O-C)$ C
Non-glutinous awned	327	305.10	1.57
Glutinous awned	78	101.70	5.52
Non-glutinous awnless	22	20.34	0.13
Glutinous awnless		6.78	0.01
	434	434	$7.23 = X^2$

TABLE *2 Independent segregation between factors* $A_{n1}a_{n1}$ *,* $A_{n2}a_{n2}$ *and* G_1g_1 *.*

P=0.0718

The deviation from expectation on the basis of independent segregation is not large, if we take into consideration the significant deficiency of the glutinous plants involved.

Linkage between the T_y t_y pair and the G, g, pair

In this particular material, when the panicle first emerges from the leaf-sheath, the apiculus, the glumes, and the awn, if present, are green like the other parts of the spikelet. **As** the spikelet is filled up by the developing grain, the three parts concerned gradually develop color through successive shades from Pale Orange Yellow, Light Orange Yellow up to **GENETICS 13:** Mr **1928**

Tawny (Ridgway) or even brighter and more glassy than Tawny at maturity. This is designated as the "tawny character," and the absence of it is non-tawny or colorless.

In inheritance, the tawny character is completely dominant over nontawny in the F_1 generation. In the F_2 , the writer obtained a simple 3:1 ratio, as shown in table *3.*

Dev. 10.5 P.E. **6.11**

Since the tawny color appeared on three parts of the spikelet and no crossing over was noticed in a population of 438 F_2 plants, it is very probable that the tawny character is due to one allelomorphic pair of factors rather than to several different genes completely linked. This allelomorphic pair is designated as *Tu tu.*

Breeding data presented in table 4 show that the T_y factor is coupled with the *G₁* factor. On the basis of independent segregation, we should

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Coupling between G_1 *and* T_y *(Gametic ratio r:s:s:r=4.48:1:1:4.48)*

$$
X^2 = 5.05
$$

P = 0.17

expect **247.5** non-glutinous tawny plants, **82.5** non-glutinous colorless plants, **82.5** glutinous tawny plants, and **27.5** glutinous colorless plants.

But the observed data deviated very widely from the expectation. By EMERSON'S (1916) method, the gametic ratio is found to be:

$$
r:s:s:r=4.48:1:1:4.48
$$

From this, the zygotic ratio is calculated. The percentage of crossing over is about 16.59 percent. Considering the significant deficiency of the glutinous plants involved, the calculated ratios are fairly close to the observed ones.

Relation between the pericarp color and the glutinous character

The pericarp color of rice varies from pure white, grey-brown, red to purple. Several investigators notably THOMPSTONE (1915), PARNELL and AYYANGAR (1917), IKENO (1918), NAGAI (1921), and HECTOR (1922), have reported that red and white colors form a simple Mendelian pair segregating in a 3:1 ratio in F_2 , red being dominant over white. PARNELL *et a2* (1917), however, found a case where red and white segregated in **a** 9:7 ratio. PARNELL (1922) reported that purple and white also form a Mendelian pair segregating in a 3:1 ratio and that purple \times red gave a ratio of 12 purple: 3 red: 1 white in F_2 . KATO and ISHIKAWA (1921) found a case where red \times white gave a ratio of 9 red: 3 yellow: 4 white. All these facts seem to indicate that there are two factors concerned with the production of pericarp color.

I found that the Chinese Imperial rice has a red pericarp color which also behaved as a simple Mendelian dominant when crossed with a white variety. This allelomorphic pair is designated as P_{r_1} , p_{r_1} . Further data show that P_{r_1} is independent of the factor G_l , as shown in table 5.

	NON-GLUTINOUS RED PERICARP	NON-GLUTINOUS WHITE PERICARP	GLUTINOUS RED PERICARP	GLUTINOUS WHITE PERICARP	TOTAL
Observed Calculated	96	31	37	15	179
9:3:3:1 Deviation	100.62 -4.62	33.54 -2.54	33.54 3.46	11.18 3.82	179

TABLE 5 *Independent segregation between* G_1 *and* P_{r1} (600 \times 100).

In another cross between 4269 and 4957, the former being purple and the latter white, a ratio of 15 colored:1 colorless was obtained, showing that there are two factors concerned. One of these two genes in this case

is presumably the same as P_{r_1} and the other is designated as P_{r_2} . Both of them are independent of the glutinous gene, as shown in table 6.

Independent segregation between P_{r1} and P_{r2} and g_1 from the cross 4269×4957.						
	NON-GLUTINOUS COLORED	NON-GLUTINOUS COLORLESS	GLUTINOUS COLORED	GLUTINOUS COLORLESS	TOTAL	
Observed	515	36	147	11	709	
Calculated~45:15:3:1	498.15	33.21	166.05	11.07	709	
Deviation	16.85	2.79	-9.05	-0.07		

TABLE 6

Inheritance of glume length and its relation with the g_1 *,* p_{r_1} *, and* p_{r_2} *factors*

At the base of the spikelet there are two small lance-shaped structures called glumes. In common varieties, the glumes are very short, about one-third as long as the lemma and palea. But there are some varieties in which the glumes are as long as the lemma and palea. In inheritance, **PARNELL** *et a1* (1917) and **NAGAI** (1921) have reported that the short glume is dominant to the long glume giving a simple $3:1$ ratio in \mathbf{F}_2 .

The writer found a case where the short glume \times long glume gave a **¹⁵**: l ratio, long glume being recessive, as shown in table **7.**

	SHORT GLUME	LONG GLUME	TOTAL
Observed	674	43	717
Calculated 15:1	672.15	44.81	717
Deviation		-1.81	

TABLE 7

Segregation for glume length in the cross 4269 (long) \times 4957 (short).

$$
\frac{\text{Dev.}}{\text{P.E.}} = \frac{1.81}{4.37} = 0.41
$$

Since the observed ratio is remarkably close to **15** : 1, undoubtedly there are two duplicate factors concerned with the production of the glume length. These two pairs of duplicate genes are designated as G_1g_1 and G_2g_2 .

Further data clearly indicate that g_1 and g_2 are independent of the glutinous gene *gl,* as shown in table 29. The observed data approach the calculated ratios very ciosely, considering the deficiency of the glutinous plants involved. Furthermore, the duplicate genes G_1 and G_2 have no chromosomal relations with factors P_{r_1} and P_{r_2} , as shown in table 30. *Inheritance of spikelet length and its relation with other characters*

The spikelet length varies with different varieties. The variety, **4957,** has spikelets varying from **3.5-4.9** mm in length with an average of **4.13** mm. The variety **4269** has spikelets varying from **7.3-10.3** mm in

FIGURE 2. $-$ Showing segregation for spikelet length in F_2 of cross 4269×4957 .

length with an average of **8.81** mm. The variation within each of these varieties follows a normal frequency curve.

In the cross, 4269×4957 , made by the writer at the CROWLEY RICE **EXPERIMENT** STATION, 1925, ten F₁ plants were obtained, each showing a similar intermediate type of spikelet ranging from **3.9-6.4** mm long with an average **of 5.33** mm in length. In the **Fz** generation, segregation for the **GENETICS 13: Mr 1928**

spikelet length took place, but the spikelets on the same plant were uniform, as expected. Six spikelets taken from the different parts of a panicle in a random fashion were measured and their average length was taken to represent the spikelet length of the plant in question. That this method of sampling actually gives a representative value is proved by the fact that when all the 95 spikelets of a panicle from the plant (Hg_i) were measured, their average was 5.66 mm approaching very closely to the respective average lengths of several samples of six spikelets each taken at random from that same panicle, namely 5.86, *5.3,* 5.75, 5.64, and 5.66. In a population of 718 F_2 plants, the spikelet length ranged from 4.7 to 9.7 mm, as shown in table 8. When the different class values were plotted against the frequency of each class, we obtained a distinctly bimodal curve (figure 2). Since the dividing point of the two groups on the curve

is clearly at the class center, 7.2 , which is exactly the length of the shortest spikelet of the long parent, and since the longest spikelet of either the short parent or of the F_1 is never over 6.5 mm in length, it appears legitimate to place the nine plants of the class (7.2) in the long group. In so doing, the **Fz** population is divided into two phenotypes, one with short spikelets and the other with long spikelets in almost exactly a *3:* 1 ratio, as shown in table 9.

TABLE 9 S egregation for short and long spikelet in F_2 *from* 4269×4957 .

	SHORT SPIKELET	LONG SPIKELET	TOTAL
Observed Calculated 3:1 Deviation	538 538.5	180 179.5 0.5	718 718

The data clearly indicates that the spikelet length, in this material at least, is due to one allelomorphic pair of factors which may be designated as S_p s_p .

The S_p s_p pair of genes are not linked with the G , g_i pair as shown in table 31, nor with P_{r_1} p_{r_1} and P_{r_2} p_{r_2} as shown in table 32.

We have seen, in the preceding pages, that glume length depends upon two independent duplicate factors G_1g_1 and G_2g_2 , the short glume being dominant and that the spikelet lengthdepends on one allelomorphic pair of genes, $S_p s_p$. When the variety, 4957, having short spikelet and short glumes, was crossed with the variety 4269, having long spikelet and long glumes, all **F1** plants had short glumes and intermediate spikelets. In the \mathbf{F}_2 generation, two new types occurred in addition to the two grandparental types, as illustrated in figure *3.*

The **Fz** population consists of 538 plants having short spikelet and short glumes, one plant having short spikelet and long glumes, 134 plants having long spikelet and short glumes, and 45 plants having long spikelet and long glumes, as shown in table IO. On the basis of three independent factors, we should expect the corresponding classes of F_2 plants to be 504.45, 33.63, 168.15, and 11.21, respectively. But this is not the case; the observed data show a great excess of the two parental types. If we assume that one of the glume factors is the same as the spikelet factor, the calculated ratios would be 12 **:0:** ³: 1, giving the class frequencies 538.50, having short glumes and short spikelet, none having short spikelet with long glumes, 134.625, having long spikelet with short glumes, and 44.875 having long spikelet and long glumes. The expectation on this assumption fits the observed data very well, except that the single plant with short spikelet and long glumes is not accounted for, as shown in table 10.

PROGENIES	SHORT SPIKELET SHORT GLUME	SHORT SPIKELET LONG GLUME	LONG SPIKELET SHORT GLUME	LONG SPIKELET LONG GLUME	TOTAL
4269×4957A	37	0	3	6	
ц в	48	0	11	4	
μ C	52	(CSTG) 1	13		
$\boldsymbol{\mu}$ D	58		14		
$\pmb{\mu}$ E	45	0	6		
$\pmb{\alpha}$ F	76	0	22	8	
и G	68	0	16		
$\pmb{\mathcal{U}}$ н	4	0	4		
ϵ	73	O	18	6	
$\pmb{\mathcal{U}}$	77		27		
Observed	538		134	45	718
Independent $45:3:15:1$	504.45	33.63	168.15	11.21	717.44
Independent $12:0:3:1$	538.50	0	134.625	44.875	718

TABLE **10** *Relation between the ghme length (15:l) nnd the spikelet length (3:l).*

FIGURE 3.-P₁ ss = The paternal parent having short spikelet and short glumes. $P_1 L L$ = The maternal parent, having long spikelet and long glumes. F₁ ss = Hybrid, having short or intermediate spikelet and short glumes. F_2 *ss* = One of 538 F_2 plants, having short spikelet and short glumes. **F,** sL=The only **F2** plant **which had** short spikelets and long glumes. This combination is a result of crossing over. F_2 Ls = One of the 134 F_2 plants having long spikelets and short glumes. $F_2 LL =$ One of the 45 plants having long spikelets and long glumes.

The same difficulty occurs if we assume complete linkage between one of the glume factors and the spikelet factor.

So far as the evidence goes, the best explanation, in the writer's opinion, may be obtained on the basis of close linkage between one of the duplicate factors for glume length and the factor for spikelet length. Here, it may be arbitrarily assumed that g_2 is closely linked with s_n . On this assumption, we can calculate the gametic ratio.

If only the factors G_1g_1 and $S_n s_n$ had been concerned in this cross, the $F₂$ distribution would be represented by the general formula,

$$
\frac{a}{3r^2+2(s^2+2rs)};\frac{b}{s^2;2rs};\frac{c}{s^2+2rs};\frac{d}{r^2}
$$
 (1)

Where $r: s: s: r$ is any gametic series and a, b, c , and d are the phenotypes, G_1S_p , G_1S_p , g_1S_p , and g_1S_p , respectively.

But when the G_2g_2 pair of genes is also involved, as is the case in this cross, r = the non-crossover gametes $G_1G_2S_p$, $g_1G_2S_p$, $G_1g_2s_p$, and $g_1g_2s_p$; and $s=$ crossover gametes $G_1G_2s_p$, $g_1G_2s_p$, $G_1g_2S_p$, and $g_1g_2S_p$. Combinations of these gametes in all possible ways grouped according to phenotypes may be represented by the general formula, $\frac{a}{2(s^2+2rs)}$: $\frac{b}{s^2:2rs}$: $\frac{c}{s^2+2rs}$: $\frac{a}{r^2}$ (1)

metic series and *a*, *b*, *c*, and *d* are the phenotypes,
 p, respectively.

ir of genes is also involved, as is the case in this

ver gametes G_1G_2S

$$
\frac{a}{12r^2+11(s^2+2rs)}:\frac{b}{3r^2+4(s^2+2rs)}:\frac{c}{s^2+2rs}:\frac{d}{r^2}
$$
(2)

From formula (2), we get the following four equations:

 $a = 12r^2+11(s^2+2rs)$ = Short spikelet and short glumes $b = 3r^2 + 4(s^2+2rs) =$ Long spikelet and short glumes $d = r^2$ = Long spikelet and long glumes $c = 2$ $rs + s^2$ = Short spikelet and long glumes

From these four equations, we can determine the gametic ratio directly from the observed zygotic series. Thus, we get $a+b+c+d=16r^2+16c$

$$
16r^{2} = a + b + d - 15c
$$
\n
$$
r^{2} = \frac{a + b + d - 15c}{16}
$$
\n
$$
r = 0.25\sqrt{a + b + d - 15c}
$$
\n
$$
16(s^{2} + 2rs) = a + b + c - 15r^{2}
$$
\n
$$
s^{2} + 2rs = \frac{a + b + c - 15r^{2}}{16}
$$
\n(3)

Also,

Adding *r2* (or *d)* to both sides, we get,

$$
r^{2}+2rs+s^{2} = \frac{a+b+c+d}{16}
$$

$$
r+s = 0.25\sqrt{a+b+c+d}
$$

$$
s = 0.25\sqrt{a+b+c+d-r}
$$
 (4)

By substituting the observed data of the four phenotypes for *a,* b, **c,** and *d.* in the formulae (3) and (4) , we get,

If we take
$$
s = 1
$$
, the ratio of $\frac{r}{s} = \frac{88.3}{1}$

By substituting the values of *r* and **S** in the four equations, we obtain the expected frequencies of the four phenotypes, which fit the observed data unusually closely, as shown in table 11.

Showing coupling between g_2 and s_p (crossover = 1.11 percent).

 $X^2 = 0.0533$. When $X^2 = 1$, $P = .801253$

Since this is a coupling phase, the crossover between g_2 and s_p will be,

$$
\frac{s}{r-s} \times 100 = 1.11 \text{ percent.}
$$

Inheritance of the apiculus color and its relation with other characters

The apex of the lemma and palea is colored in many varieties. This localized color spot at the upper tip of the spikelet is here spoken of as the apiculus color. The inheritance of this character has been studied by several investigators. HECTOR obtained a ratio **of** three colored apiculus to one colorless in 1913 and another ratio of 27:35 in 1916. Besides, **HECTOR** (1922) reported two new conditions, one of which segregated in a 9:7 ratio and the other in a **15:** 1 ratio.

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In crosses between different varieties, the writer obtained various ratios in the F_2 generations, namely, 3:1, 9:7, 15:1, 27:37, and 162:94, indicating that there are at least four genetic factors responsible for the production of the apiculus color. The data are presented in table 12.

That variety 300, though colorless, actually carries some factor is shown by the fact that when it was crossed with another colorless variety 600 (see cross No. 2), the F_1 had colored apiculus. In the F_2 , a ratio of 27:37 was observed, showing at least three factors were involved. This situation may be explained, if the two parental types have the following genetic constitutions:

 $300 = g_1 g_1 CC a_{p_1} a_{p_1} a_{p_2} a_{p_2} a_{p_3} a_{p_4}$ (glutinous colorless)

 $600 = G_1 G_1$ cc $A_{p_1} A_{p_1} A_{p_2} A_{p_2} A_{p_3} A_{p_4}$ (non-glutinous colorless Where A_{p_1} , A_{p_2} and C are complimentary factors for apiculus color.

In cross No. 1 (300 \times 400) the 9:7 ratio clearly indicates that two factors are concerned in the production of the apiculus color, these being designated as A_{p_1} and A_{p_2} . The situation can be explained by assuming the genetic constitutions of the parents as follows:

Parent 300 = $g_1 g_1 CC a_{p_1} a_{p_1} a_{p_2} a_{p_2}$

Parent $400 = G_1 G_1 C C A_{p_1} A_{p_1} A_{p_2} A_{p_3}$

Where $C =$ chromogen, and A_{p_1} and A_{p_2} are complementary for the apiculus color.

That the variety.600 actually carries at least one apiculus factor differing from those in the variety 400 is proved by cross No. 3, which gave a ratio of 162:94, showing that four factors are involved. The situation may be explained on the bases of the foregoing genetic constitutions assumed for varieties **400** and 600, respectively.

Cross No. 4 gave a ratio of 15:1, showing that there are two factors involved, each of them alone producing apiculus color. These factors may be designated as A_{v_k} and A_{v_k} . As no further crosses were made, the assumption must be considered as a tentative one.

Cross No. 5 gave a simple *3:* 1 ratio, showing that only one factor pair was involved. This factor pair is different from A_{p_1} , A_{p_2} , and A_{p_3} , in that none of the latter alone can produce the apiculus color. Furthermore, this new factor is linked with the glutinous gene as will be shown later. This gene is designated as A_{p_4} . It may be that one of the two duplicate factors involved in cross No. 4 is the same as A_{p_4} .

Factors A_{p_1} and A_{p_2} are independent of the glutinous gene as shown in table 13.

	COLORED APEX NON-GLUTINOUS	COLORLESS APEX NON-GLUTINOUS	COLORED APEX GLUTINOUS	COLORLESS APEX GLUTINOUS	TOTAL
Observed	65	51	12	14	142
Calculated 27:21:9:7	59.67	46.41	19.89	15.47	142
Deviation	5.33	4.59	-7.89	-1.47	

TABLE 13 *Showing independent segregation between factors* A_{p_1} *,* A_{p_2} *and* g_l *(307* \times *410).*

As mentioned above, A_{ν} is linked with the glutinous gene as shown in table **14.** On the independent Mendelian basis, the expected frequencies should be **417.87** colored non-glutinous, 139.29 colored glutinous, and **46.43** colorless glutinous. But this is far from the observed data which clearly indicate coupling between a_{p_4} and g_i . On the latter basis, the

TABLE 14 *Showing coupling between* a_{p_4} *and* g_1 *from* F_2 *of* 4269×4957 *.*

	COLORED APEX NON-GLUTINOUS	COLORLESS APEX NON-GLUTINOUS	COLORED APEX GLUTINOUS	COLORLESS APEX GLUTINOUS	TOTAL
Observed	491	90	76	86	743
Calculated	474.17	83.16	83.16	102.62	743
Deviation	16.83	6.84	-7.16	-16.62	

 $X^2 = 4.46$, $P = 0.22$.

gametic ratio is found from EMERSON's (1916) formula, $r=10.13$, and $s=3.5$. The crossover is, therefore, about 22.34 percent.

The large deviation is clearly due to the deficiency of the glutinous plants, the latter being 2.98 times the probable error.

Further data show that A_{p4} is independent of the factors P_{r_1} and P_{r_2} , G_1 and G_2 , and S_p , as is seen in tables 33, 34, and 35 respectively.

Relation between stigma color and other characters

In some varieties, the stigma is colorless, and in others it is colored with an intensity varying from pale red to dark purple. In inheritance, colored and colorless stigma segregate in different ratios according to the material used. HECTOR (1916, 1922) reported cases of 3:1, 9:7, 27:37, and 81:175 ratios, showing that there are at least five factors responsible for the production of stigma color.

The writer obtained two cases, one segregating in a **3:** 1 ratio, and the other, 9:7, as shown in table 15.

CASE	CROSSES COLORLESS \times	${\bf F}_{1}$		F: SEGREGATION	CLOS- EST	F ₂ EXPECTED	DEVIA-	DEV. ------	SYMBOLS
	PURPLE					Colored Colorless RATIO Colored Colorless	TION	P. B.	
	307×410	Colored	101	41		$3:1$ 106.5 35.5	5.5	1.58	
2	4269×4957 (ABFGI)	Colored	242	179		9:7 236.79 184.17 5.17		0.75	$S_{a_1} S_{a_2}$

TABLE 15 *Showing segregation of stigma* **color.**

In the first case, the factor is probably the same as A_{p_1} (or A_{p_2}), because in the same **F,** population arising from the same cross, the apiculus **color** alone segregated in a 9:7 ratio (see table 12) whereas a 9:3:4 ratio was observed when both apiculus and stigma were considered at the same time, as shown in table 16.

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Showing F₂ segregation for apex and stigma color (307 \times *410).*

In the second case, there are clearly two separate factors for the production of the stigma color. These are designated as S_{a_1} and S_{a_2} . They are complementary to each other. One of them is linked with the glutinous gene, g_i . For the sake of convenience, the linkage may be arbitrarily assumed to be between s_{a_1} and g_1 . The data are presented in table 17.

 $X^2 = 1.4094$, $P = 0.707564$.

BRUNSON'S (1924) modified formulae

$$
r = \sqrt{\frac{(AB+3ab)-(Ab+aB)}{18}}
$$

$$
s = \frac{1}{4}\sqrt{AB+Ab+ab+ab} - r
$$

were used in calculating the intensity of the linkage between s_{a_1} and g_i . The gametic ratio is $r = 4.18$ and $s = 0.95$. Since this is a coupling phase, the percentage of crossing over is obtained from the formula $\frac{s}{s+s}$, namely 18.51 percent.

Data from the same cross show factors s_{a_1} and s_{a_2} are independent of p_{r_1} , p_{r_2} , g_1 , g_2 and $s\dot{p}$ as shown in tables 36, 37, and 38.

The exact relation between the stigma color and the apiculus color in this particular cross is not determined. As described above, a_{p_4} is coupled with g_{ν} giving about 22.34 percent crossing over; and s_{a_1} is also linked with g_{ν} giving 18.51 percent crossing over. When the apiculus color and the stigma color were involved at the same time, a new situation arose, as shown in table 18.

The zygotic ratios calculated on the basis of three independent factors do not fit the observed ratio at all. There are, then, only two alternative explanations for the situation, namely, complete linkage between a_{p_4} and s_{q_1} , or a_{p_1} being the same factor as s_{q_1} . Since both alternatives give identical

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zygotic ratio $(9:3:0:4)$, it is impossible to decide which alternative is correct. It may be mentioned, however, that one plant (FSt 68) which is not counted in table 18 had purple stigma with a doubtful apiculus color, because the latter was under a question mark (?) in the original notebook.		TABLE 18			
	COLORED APEX PURPLE STIGMA	<i>Relation between</i> a_{p_4} and s_{q_1} . COLORED APEX COLORLESS STIGMA	COLORLESS APEX PURPLE STIGMA	COLORLESS APEX COLORLESS STIGMA	TOTAL
Observed	243	64	1 (FSt 68?)	117	424
	178.74	139.02	59.58	46.34	424
Calculated 27:21:9:7 Complete linkage 9:3:0:4	238.5	79.5	0	106.0	424

TABLE 18 *Relation between* a_{p_4} *and* s_{q_1} *.*

Although a_{p_4} and s_{q_1} have different crossover values with the glutinous gene, the difference is only **3.83** percent. Since both alternatives are possible, the question must be left open for the present.

Relation between the leaj-sheath color and other characters

The leaf-sheath color when present may be either self red or purple, or merely consisting of colored stripes varying in intensity. There are several genetic factors responsible for its production. **PARNELL** (1917) found a case where colored and colorless leaf-sheath segregated in a simple **3:** 1 ratio. **HECTOR** (1916, 1922) reported four cases segregating in *3:* 1, 9:7 27:37, and 15:1, respectively.

The writer has obtained two kinds of ratios, namely, 9:7 and 15:1, as shown in table 19.

CASE	CROSSES	\mathbf{F}_1		F: SEGREGATION Colored Colorless	CLOSEST RATIO	Colored	Expected Colorless	DEVIA- TION	SYMBOLS
	4269×4957 (colorless X colored) Colored		204	139		9:7 $ 192.87 147.01 - 8.01 $			Ls_1 Ls_2
2	$800b_6 \times 625$ $\left(\text{colored} \times \text{colorless}\right)$	Colored	266						14 15:1 262.5 17.5 -2.05 Ls_8 Ls_4

TABLE 19 *Showing F2 segregation for leaf-sheath color.*

These data indicate that there are at least four factors which are concerned with the production of leaf-sheath color. In the first case, there must be two complementary factors, whose presence is necessary **for** the expression of color. These factors are designated as L_{s_1} and L_{s_2} . In the **GENETICS 13: MI 1928**

second case, there must be two duplicate factors each of which alone can produce color. These are designated as L_s , and L_s .

In the first cross, one of the complementary factors is linked with the glutinous gene. The linkage may be arbitrarily assumed to be between l_{s} , and g_{i} . The gametic ratio is calculated by BRUNSON's formulae.

$$
r = \sqrt{\frac{AB + 3ab - (Ab + aB)}{18}} = 3.73
$$

$$
s = \frac{1}{4} \sqrt{AB + Ab + aB + ab - r} = 0.90
$$

 $s = \frac{1}{4} \sqrt{AB + Ab + aB + ab} - r = 0.90$
The crossing over $= \frac{s}{r+s} \times 100 = 19.43$ percent. By substituting the values of *r* and **S** in the following equations, we get the zygotic ratios, **S** *r+s*

 $AB = 9r^2 + 12rs + 6s^2$ = colored sheath non-glutinous
 $Ab = 6rs + 3s^2$ = colored sheath glutinous $6rs+3s^2$ = colored sheath glutinous $aB = 3r^2 + 12rs + 6s^2$ = colorless sheath non-glutinous $ab = 4r^2 + 2rs + s^2 =$ colorless sheath glutinous

The calculated zygotic ratios fit the observed ratios very well considering the deficiency of glutinous plants, as shown in table 20.

	Showing coupling between ls_1 and g_1 .	I ADLE LU			
	COLORED SHEATH NON-GLUTINOUS	GLUTINOUS	COLORED SHEATH COLORLESS SHEATH NON-GLUTINOUS	COLORLESS SHEATH GLUTINOUS	TOTAL
Observed	179	25	80	59	343
Calculated 27:9:21:7	144.72	48.24	112.56	37.52	343
Linkage (19.43 $\%$ crossing over)	170.36	22.57	88.88	63.18	343
Deviation	8.64	2.43	-8.88	-4.18	

TABLE 20 *Showing coupling between Is1 and gl.*

Data obtained from the same cross indicate that l_{s_1}, l_{s_2} are independent of the factors $p_{r_1}, p_{r_2}, g_1, g_2,$ and s_p , as shown in tables 39, 40 and 41.

 l_{s_1} is closely linked with a_{p_4} as shown in table 21. Since a_{p_4} is coupled with g_i , giving 22.34 percent crossing over, and since $l_{i,j}$ is also linked with **gl,** giving **19.43** percent crossing over, the order of the three genes on the glutinous chromosome would appear to be,

$$
\begin{array}{c|c}\n & 22.34 \text{ percent} \\
\hline\na_{p_4} & l_{e_1} \\
\hline\n\end{array}\n\qquad\n\begin{array}{c}\n & 22.34 \text{ percent} \\
 & 19.43 \text{ percent} \\
 & g_i\n\end{array}
$$

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Linkage between stigma color and leaf-sheath color

As described above, in the F_2 generation of the cross, 4269×4957 , the stigma color alone segregated in a ratio of 9 colored:7 colorless, and the leaf-sheath color also segregated in a 9: 7 ratio. One of the twocomplemen-

tary factors for stigma color is linked with the glutinousgene, giving 18.71 percent crossing over. One of the two complementary genes for'the production of the leaf-sheath color is also linked with g_i , giving 19:43 percent crossing over. When the stigma color and the leaf-sheath color are considered at the same time, a very close linkage between the two characters is revealed, as shown in table 22a, where the calculated zygotic ratios on the basis of four independent factors are shown to be very far from the observed frequencies.

PROGENIES	PURPLE STIGMA PURPLE SHEATH	PURPLE STIGMA COLORLESS SHEATH PURPLE SHEATH		COLORLESS STIGMA COLORLESS STIGMA COLORLESS SHEATH	TOTAL
4269×4957B	34			23	eB.
4269×4957D	46			29	
4269×4957F	52			44	\mathcal{L}
4269×4957I	-57			37 ²	
Observed	189		15	133	346
Calculated on 4 independent					
factors	109.35	85.05	85.05	66.15	
3 independent factors	145.97	48.66	48.66	102.72	

TABLE 22a *Showing close linkage betweenihe stigma color and the leaf-sheath color.*

Since the stigma and the leaf-sheath give about the same percentage of crossing over with the glutinous gene, the difference being 0.72 percent, it is possible that the two characters have one factor in common. Most probably s_{a_1} is the same factor as l_{s_1} . For the sake of simplicity, this common factor is designated as *A.* Until further data demonstrate that the s_{a_1} and l_{s_1} are actually two separate entities, factor A will be considered **GENE~CS 13: Mr 1928**

as a common gene which has a manifold effect conditioning stigma color as well as leaf-sheath color. On this hypothesis, then, *A* is complementary with S_{a_2} for stigma color and *A* is also complementary with L_{a_2} for leafsheath color.

Since the calculated ratios on the basis that *A*, S_{a_2} and L_{a_2} are independent factors do not fit the observed frequencies, S_{a_2} and L_{s_2} must be coupled. The situtation may be represented by the following diagram:

$$
\begin{array}{c|c|c}\nA & a & b & b \\
\hline\nGl & g_1 & b_2 & b_3 \\
\hline\n\end{array}
$$

If *r*:s:s:*r* represents any gametic series between the two linked factors. s_{a_2} and l_{s_2} , the non-crossover classes may be represented by the following equations:
 $A S_{a_1} L_{a_2} = 3[3r^2 + 2(s^2 + 2rs)] = 9r^2 + 12rs + 6s^2$

$$
A S_{a_2} L_{s_2} = 3[3r^2 + 2(s^2 + 2rs)] = 9r^2 + 12rs + 6s^2
$$

\n
$$
a S_{a_2} L_{s_2} = 3r^2 + 2(s^2 + 2rs) = 3r\cdot 3^2 + 4rs + 2s^2
$$

\n
$$
A S_{a_2} L_{s_2} = (3r^2) = 3r^2
$$

\n
$$
a S_{a_2} L_{s_2} = (r^2) = r^2
$$

On the other hand, the cross over classes may be represented as follows

A
$$
s_{a_2} L_{a_2} = 3(s^2 + 2rs) = 6rs + 3s^2
$$

\nA $S_{a_2} l_{a_2} = 3(s^2 + 2rs) = 6rs + 3s^2$
\na $S_{a_2} l_{a_2} = (s^2 + 2rs) = 2rs + s^2$
\na $s_{a_2} L_{a_2} = (s^2 + 2rs) = 2rs + s^2$

Since either stigma color or leaf-sheath color needs two complementary factors $(A S_{a_2}$ or $A L_{s_2})$ for its expression, the five classes $(a S_{a_2} L_{s_2}, A S_{a_2} l_{s_1})$ $u_{s_{a_1}}$, u_{s_1} , $u_{s_{a_2}}$, $u_{s_{a_2}}$, $u_{s_{a_1}}$, $u_{s_{a_2}}$, $u_{s_{a_2}}$ are thrown into the same phenotypic group By this regrouping, the four phenotypic classes are represented by the four equations:

$$
XY = 9r^2 + 12rs + 6s^2
$$
 Purple stigma and purple sheath
\n
$$
Xy = 6rs + 3s^2
$$
 Purple stigma and colorless sheath
\n
$$
xY = 6rs + 3s^2
$$
 Colorless stigma and purple sheath
\n
$$
xy = 7r^2 + 8rs + rs^2
$$
 Colorless stigma and colorless sheath

Where $X =$ purple stigma, $x =$ colorless stigma, $Y =$ purple sheath and $y =$ colorless sheath. From these equations, we get:

$$
XY + xy = 16r^2 + 20rs + 10s^2
$$
 (A)

$$
Xy + xY = 12rs + 6s^2
$$
 (B)

Dividing both **(A)** and (B) by 2, we get

$$
\frac{XY+xy}{2} = 8r^2 - 10rs + 5s^2
$$
 (C)

$$
\frac{XY+xY}{2} = 6rs + 3s^2
$$
 (D)

Multiplying (C) by **3,** and (D) by S, we get

$$
24r^{2} = \frac{3}{2}(XY+xy) - \frac{5}{2}(Xy+ xY)
$$

$$
r = \sqrt{\frac{\frac{3}{2}(XY+ xy) - \frac{5}{2}(Xy+ xY)}{24}}
$$
 (1)

Also,

 $XY+Xy+xY+xy=16(r^2+2rs+s^2)$

Taking the square roots, we get

$$
4(r+s) = \sqrt{XY+Xy+XY+xy}
$$

$$
s = \frac{1}{4}\sqrt{XY+Xy+XY+xy} - r
$$
 (2)

From (1) and (2), we can calculate the gametic ratio, this being:

$$
r = 4.19
$$

$$
s = 0.46
$$

Since this is a coupling case, the percentage of crossing $over =$

$$
\frac{0.46}{4.19 + 0.46} \times 100 = 9.8 \text{ percent}
$$

Substituting the calculated values of *r* and *S* in the four original equations representing the four **Fz** phenotypic classes, we get the frequencies quite similar *to* the observed ones, as shown in table 22b:

TABLE 22b *Showing close linkage between sa₂ and ls₂.*

	PURPLE STIGMA PURPLE LEAF-SHEATH	PURPLE STIGMA COLORLESS SHEATH	COLORLESS STIGMA PURPLE SHEATH	COLORLESS STIGMA COLORLESS SHEATH
Observed Calculated Deviation	189 182.68 6.32	12.19 -3.19	15 12.19 2.81	133 138.88 -5.88
		$X^2 = 1.947$		

$$
P=.584536
$$

Relation between ligule color and other characters

The ligule is a structure that projects out at the juncture of the leafblade and the leaf-sheath. In some varieties, this structure has a deep purple color. In inheritance, the ligule color depends on several factors for its expression. **HECTOR** (1922) reported two cases, one segregating in **9: 7** ratio and the other segregating in 27:37 ratio.

The writer has obtained a ratio of 27 purple: 37 green in the cross, 4269x4957, as shown in table **23.**

TABLE 23

The data clearly show that three factors are concerned in the production of ligule color in this case. These factors are designated as $l_{q_1}, l_{q_2},$ and l_{q_3} . These three genes are probably independent of the factors g_1 , g_2 , g_3 , and s_p , as shown in tables 24, 42, and 43.					
		TABLE 24			
	Showing independent segregation of $g_1, l_{g_1}, l_{g_2},$ and l_{g_3} .				
PROGENIES	NON-GLUTINOUS PURPLE LIGILE	NON-GLUTINOUS GREEN LIGULE	GLUTINOUS PURPLE LIGULE	GLUTINOUS GREEN LIGULE	TOTAL
4269×4957A	21	15	2	9	
4269×4957B	18	26	4	15	
4269×4957C	33	27	4	5	
4269×4957D	23	42	4	9	
Observed	95	110	14	38	257
Calculated 81:111:27:37	81.32	111.44	27.11	37.15	
Deviation	13.68	-1.44	-13.11	0.85	

Relation between ligule color and pericarp color

As mentioned above, the pericarp color in this cross is due to two duplicate factors which have been designated as P_{r_1} and P_{r_2} . When the ligule color and the pericarp color are considered at the same time, a new situation appears, as shown in table **25.**

As-shown in table 25, there is only one individual in the class "purple ligule and colorless pericarp" from four progenies. It is possible that some of the ligule factors are closely linked with the pericarp genes, and the rare occurrence of this class is due to linkage and the small population.

On the other hand, it is equaly possible that one of the ligule genes is the same as one of the pericarp factors, and the single individual may be due to contamination. Indeed, the calculated ratios (108:0:132:16) on the latter basis fit the observed ratios remarkably well. However, the question must be left open for the present. Earlier investigations (HECTOR 1922) have shown a similar situation.

Relation of hull color to other characters

The term "hull" here used includes the lemma and palea which enclose the grain within. There are different colors either extended entirely over the hull such as "dark gold, ripening gold, and ripening straw," or restricted to certain portions of the hull, such as "dark furrows, piebald gold, tipped gold, patchy gold, mottled gold, and granular furrows" (PARNELL 1922). The character with which we are immediately concerned here is the ripening black color. The hull is first green as usual, but when the grains reach maturity, the hull turns black. PARNELL (1917) reported two cases **of** a similar condition, one segregating in a **3** : 1 ratio and the other 9: **7.**

The writer crossed two non-black varieties $(800b_5 \times 625)$, the F_1 spikelets at first had green hull which later changed black or sooty black (Ridgway). The F_2 plants segregated in a ratio of approximately 9 black : 7 non-black, as shown in table 26, where the data for this and other characters involved in the same cross $(800b₅ \times 625)$ are presented together.

TABLE 26

Showing segregation for colors of hull, internode, leaf-sheath, a piculus, and pericarp.

The data clearly show that there are two complementary factors for the production of the black hull. These are designated as H_1 and H_2 . Both these genes are independent of the factors $I_{n_1}, I_{n_2}, L_{s_3}, L_{s_4}, A_{p_5}, A_{p_6}, P_{r_1}$ and $P_{,2}$, as shown in tables 27, 28, 44, and 45.

TABLE 27

	BLACK HULL COLORED INTER- NODE	BLACK HULL COLORLESS INTERNODE	NON-BLACK COLORED INTERNODE	NON-BLACK COLORLESS INTERNODE	TOTAL
Observed Calculated 81:63:63:49 Deviation	63 66.74 -3.74	47 51.91 -4.91	44 51.91 -7.91	57 40.38 16.62	211 210.94

Showing independent segregation between h_1 , h_2 , i_{n_1} , and i_{n_2} .

The large deviation of the observed frequencies may be due to the small population, because at least 256 individuals are necessary to make the ratios barely even.

	BLACK HULL PURPLE SHEATH	NON-BLACK HULL PURPLE SHEATH	BLACK HULL	COLORLESS HULL COLORLESS SHEATH COLORLESS SHEATH	TOTAL
Observed	111				217
Calculated 135:105:9:7	114.75	89.25	7.65	5.95	217
Deviation	-3.75	7.75	-4.65	0.05	
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Showing independent segregation between h_1 , h_2 , l_{s_3} , and l_{s_4} .

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DISCUSSION OF RESULTS

In the foregoing pages, data have been presented to show the behavior and chromosomal relations of twenty-five or more genetic factors affecting one or another part of the plant. To facilitate further discussion, we may outline the established genes and their behavior as follows:

While the results outlined above are self-explanatory, a few of them may be briefly discussed with advantage. First of all, it is interesting to note that the awn, being a sporophytic character, should be either present or absent on all the spikelets of the same plant in the \mathbf{F}_2 generation. Contrary to this expectation, two new types have appeared. One type has most of the spikelets awned, and the other has only a few spikelets awned, the remaining spikelets of the same panicle being awnless, as shown in figure 1. These new types and the original grand-parental types occur in about a 12: 1 :2: 1 ratio. The observed frequencies have been explained on the basis of two duplicate factors. The awn may be conceived of as an extension of the central nerve of the lemma. The gene, A_{n_1} , extends the central nerve either in single or double dose with or without $A_{n_{\bullet}}$. The gene A_n , is of similar nature, but it is weaker in activity; so, the double doses may extend most of the central nerves, while the single dose may extend just a few of them. Environmental conditions may also enter in, thus influencing the action of A_{n_2} during the morphogenesis of the spikelets and particularly of the awn. As the spikelets on the top of the panicle and those on the lower part do not develop at the same time, we can easily see the differential action of the gene under different conditions in the extension of the awn. It is interesting to note that in the "mostly awned" and "rarely awned" classes, usually it is the spikelets at the lower part of the panicle whose awns are not extended.

Factorially, the case may be represented as follows:

 $A_{n_1} A_{n_1} A_{n_2} A_{n_2}$ $A_{n_1} A_{n_1} A_{n_2} a_{n_2}$ $\left.\frac{2 A_{n_1} A_{n_1} A_{n_2} A_{n_2}}{A_{n_1} A_{n_2} A_{n_2}}\right\} = 12 \text{ fully awarded}$ A_{n_1} a_{n_1} A_{n_2} a_{n_2} $A_{n_1}A_{n_1}$ a_{n_2} a_{n_2} A_{n_1} a_{n_1} a_{n_2} a_{n_3}

1 a_{n_1} a_{n_1} A_{n_2} A_{n_2} = 1 mostly awned 2 a_{n_1} a_{n_1} A_{n_2} $a_{n_2} = 2$ rarely awned **1** a_n , a_n , a_n , $a_n = 1$ wholly awnless

Several factors have been shown to lie on the glutinous chromosome. T_y is coupled with G_y , giving 16.59 percent crossing over. $a_{p₄}$ is linked with g_i , giving 22.34 percent crossing over. l_{s_i} is also coupled with g_i , giving 19.43 percent crossing over. These constitute the first linkage qroup which may be expressed in the following diagram:

$$
a_{p_4} \quad l_{s_1} \quad t_y(2) \quad g_1 \quad t_y(2)
$$

Whether t_{ν} is on the left or right hand side of g_{ν} is not known at present. It may be noted that s_a , is also linked with g_1 , giving about 18.51 percent crossing over. However, whether s_{a_1} is the same factor as l_{s_1} , or a separate gene closely linked with l_{s_1} remains to be determined. It may be further noted that TAKAHASHI'S factor *(R)* for awn color and **YAMAGUCHI'S** factor (S) for apiculus color are also in the same linkage group, though their exact loci cannot be stated. It is possible that (S) is the same factor as s_{p_4} .

The second linkage group constitutes two factors, namely, s_p and g_2 . The glume length depends on two duplicate factors, g_1 and g_2 , segregating in a 15:1 ratio long glume being recessive. The spikelet length depends on a simple factor pair, $S_n s_n$, long spikelet being recessive. The factors g_2 and s_p are coupled, giving 1.11 percent crossing over. That the short spikelet factor, S_p , is coupled with one of the duplicate glume genes (G_2) is beyond doubt. It is of interest to note, however, that on the basis of 1.11 percent crossing over, only 0.99 or one plant in a population of 718 individuals is expected to have short spikelets and long glumes, and one such individual has been obtained as shown in figure 3 (F_2 SL). That this individual is a crossover is proved by the fact that there is no such variety in my stock that has short spikelets and long glumes, thus eliminating any error through contamination. It may further be pointed out that if the observed crossover plant had not appeared, the data (see table 10) could be **ex**plained equally well on the basis of $12:0:3:1$ ratio by assuming that one of the duplicate glume factors is the same as the S_p factor for the spikelet length. The difficulty, however, is that on this assumption, it must follow that the long spikelet plant must necessarily have long glumes also. But this is not the case.

The third linkage group consists of s_{a_1} and l_{a_2} . The purple stigma in this case depends upon the presence **of** two dominant complementary factors, namely, S_{a_1} and S_{a_2} . The purple leaf-sheath also depends on two comple-**GENETICS 13: MI 1928**

mentary factors, L_{s_1} and L_{s_2} . Since both L_{s_1} and S_{s_1} are linked with G_{t_1} , giving about the same percentage of crossing over, L_{s_1} and S_{a_1} may be the same factor. For the sake of convenience, a common factor *(A)* is assumed to represent both L_{s_1} and S_{s_1} , though the assumption must await further verification. It is certain, however, that linkage does exist between the characters, no matter what assumption we may make. The observed frequencies cannot be explained on the independent segregation of either four or three factors, as shown in table 22a. It appears that only on the assumption of a close linkage between S_{a} , and L_{s} , can we explain the observed frequencies. In so doing, the crossing over value is found to be 9.8 percent between s_{a_2} and l_{s_2} . The calculated zygotic ratios on this basis fit the observed frequencies closely.

'Thus far, three linkage groups have been established beyond doubt. **A** fourth group is indicated by the data presented in table 25. The observed frequencies can be explained by two alternatives. One is that one of the pericarp factors is the same as one of the three ligule factors, thus giving a ratio of $108:0:132:16$ on the basis of four factors, that is the two characters have one factor in common. The other alternative is that one of the pericarp factors is completely or very closely linked with one of the three ligule factors. The occurrence of one individual having "purple ligule and colorless pericarp,'' which cannot be accounted for by the first alternative, favors the second view. In this connection, the writer takes the liberty to rearrange HECTOR'S (1922) data in the following table for comparison with his own results:

It must be noted, however, that though the two sets of observed frequencies are similar in nature, they are not exactly comparable. For in HECTOR'S case, the color of the pericarp depends on one factor pair segregating in a *3:* l ratio, while in the present case, the pericarp color is due to duplicate factors segregating in a 15: 1 ratio. The common feature in both cases is that on the assumption of a common factor for both pericarp and ligule color, these few exceptions having "purple ligule and colorless pericarp" can not be accounted for. On the other hand, these exceptional individuals tend to support the view that one of the ligule factors is closely linked with one of the factors responsible for the pericarp color.

Finally, it is interesting to note that so many of the characters studied are due to duplicate genes. This is of particular interest in view of the fact that all the varieties used have twenty-four chromosomes.

SUMMARY

1. Data have been presented to show the Mendelian behavior and chromosomal relations of twenty-five or more genetic factors which affect one or another part of rice plants.

2. Of the twelve characters studied, five are due to duplicate genes, each segregating in a ratio of 15:1.

3. Through the study of the interrelations between these twenty-five genes, three linkage groups have been established beyond doubt and possibly a fourth group is also indicated.

4. The first linkage group consists of four or five factors, namely, g_l, a_p, l_{s_l}, t_v , and possibly s_a .

5. There is 16.59 percent crossing over between g_l and t_g ; 18.51 percent between g_l and s_{a_1} , 19.43 percent between g_l and l_{a_1} , and 22.34 percent between g_l and a_{p_l} . In addition a_{p4} is very closely linked with l_{q_1} .

6. The second linkage group consists of two genes, namely, s_p and g_a . The glume length depends on duplicate factors, g_1 and g_2 , segregating in ^a**15** : **1** ratio, long glume being recessive. The spikelet length depends on a simple factor pair, $S_p s_p$, long spikelet being recessive. One of the duplicate genes, presumably g_2 , is coupled with $s\phi$, giving 1.11 percent crossing over.

7. The third linkage group consists of s_{a_2} and l_{s_2} . Purple stigma in this case depends upon the presence of two dominant complementary factors, namely, S_{a_1} and S_{a_2} . Purple leaf-sheath also depends on two complementary factors, L_{s_1} and L_{s_2} . Between these two characters, there is close linkage. Only on the assumption of coupling between S_{a_2} and L_{a_2} can the observed frequencies be explained. BRUNSON'S method for calculating linkage intensities involving complementary factors is further modified for the present case. The crossing over value is found to be **9.8** percent between s_a , and l_s .

8. A fourth linkage group is indicated between p_{r1} or p_{r2} and one of the three complementary factors for purple ligule color.

9. Factors g_1 and g_2 are independent of g_1 , p_r , and p_{r_2} .

10. The factor s_p is independent of g_l , p_{r_1} , p_{r_2} .

11. Factors a_{p_1} and a_{p_2} are independent of g_i .

12. Gene a_{p_4} is independent of $p_{r_1}, p_{r_2}, q_1, q_2,$ and s_p .

13. Factors s_a , and s_a , are independent of p_r , p_r , g_1 , g_2 , and s_a .

14. Factors l_{ϵ_1} and l_{ϵ_2} are independent of $p_{\tau_1}, p_{\tau_2}, g_1, g_2$, and s_{ν} .

15. Factors $l_{q_1}, l_{q_2},$ and l_{q_3} are independent of $g_l, g_1, g_2,$ and s_p .

16. Factors h_1 and h_2 are independent of $i_{n_1}, i_{n_2}, l_{n_3}, l_{n_4}, a_{p_5}, a_{p_6}, p_{r_1}$ and p_{r-1} .

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APPENDIX-TABLES 29-45

TABLE 29

Independent segregation between G_1 , G_2 and G_1 from the cross (4269×4957).

 $P=0.2810$

TABLE 30

 $P=0.495$

TABLE 31

	SHORT SPIKELET NON-GLUTINOUS	SHORT SPIKELET GLUTINOUS	LONG SPIKELET NON-GLUTINOUS	LONG SPIKELET GLUTINOUS	TOTAL
Observed	417	118	138	40	713
Calculated 9:3:3:1	-400.5	133.5	133.5	$+4.5$	713
Deviation	15.5	-15.5	4.5	-4.5	

Independent segregation between $S_p s_p$ and G_{1g_1} in F_2 .

 $X^2 = 4.34.$ $P = 0.2309$.

The deviation is apparently due to the large deficiency of glutinous grains.

Independent segregation between factors s_p , p_{r_1} , and p_{r_2} (Ratio 45:4:15:1).

When $X^2=1$, $P=0.8013$.

TABLE 33

Showing independent segregation between A_{p_A} and p_{r_1} and p_{r_2} .				

 $X^2=0.982$, $P=0.801253$ when $X^2=1$.

TABLE 34

Showing independent segregation between a_{p_4} and g_1 and g_2 .									
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 $X^2 = 7.839$. $P = 0.0418.$

The cause of the deviation is unknown.

TABLE 35

	COLORED APEX SHORT SPIKELET	COLORLESS APEX SHORT SPIKELET	COLORED APEX LONG SPIKELET	COLORLESS APEX LONG SPIEELET	TOTAL
Observed	428	133	139	43	743
Calculated 9:3:3:1	417.87	- 139.29	139.29	46.43	
Deviation	10.13	-6.29	-0.29	-3.43	

Showing independent segregation between A_{p_4} and s_p .

 $P = 0.801253$ when $X^2 = 1$. $X^2 = 0.788$

TABLE 36

Showing independent segregation between s_{a_1} , s_{a_2} and p_{r_1} , p_{r_2} .

The deviation is probably due to the small number of plants.

TABLE 37

 $X^2 = 2.92.$ $P = 0.405996$.

TABLE 38

Showing independent segregation between s_{a_1} , s_{a_2} and s_p .

	PURFLE STIGMA SHORT SPIKELET	COLORLESS STIGMA, SHORT SPIKELET	PURPLE STIGMA LONG SPIKELET	COLORLESS STIGMA, LONG SPIKELET	TOTAL
Observed	189	130	53	49	421
Calculated 27:21:9:7	177.39	137.97	59.13	45.99	421
Deviation	11.61	-7.97	-5.13	3.01	

 $X = 1.85$. $P = 0.6067$.

TABLE 39

	COLORED PERICARP COLORED SHEATH	COLORED PERICARP COLORLESS SHEATH	COLORLESS PERICARP COLORED SHEATH	COLORLESS PERICARP COLORLESS SHEATH	TOTAL
Observed Calculated 135:105:9:7 Deviation	179 177.12 1.88	134 137.76 -3.76	19 11.80 7.20	9.18 -5.18	336 336

Showing independent segregation between L_{s_1} , l_{s_2} and p_{r_1} , p_{r_2} (4269×4957).

The deviation is probably due to the small number of plants.

TABLE 40

 \bar{z}

Showing independent segregation for l_{s_1} , l_{s_2} , g_1 and g_2 .

 $X^2 = 2.7$. $P = 0.4458$.

TABLE 41

	COLORED LEAF SHEATH SHORT SPIKELET	COLORLESS SHEATH, SHORT SPIKELET	COLORED SHEATH LONG SPIKELET	COLORLESS SHEATH, LONG SPIKELET	TOTAL
Observed Calculated 27:21:9:7	154 144.72	100 112.56	50 48.24	39 37.52	343 343
Deviation	9.28	-12.56	1.76	1.48	

Showing independent segregation for l_{s_1} , l_{s_2} and s_p .

 $X^2 = 2.1$, $P = 0.5543$.

TABLE 42

	PURPLE LIGULE SHORT GLUMES	PURPLE LIGULE LONG GLUMES	GREEN LIGULE SHORT GLUMES	GREEN LIGULE LONG GLUMES	TOTAL
Observed	104		134	13	256
Calculated (5 factors)	101.25	6.75	138.75	9.25	256
Deviation	.2.75	-1.75	-4.75	3.75	

Showing independent segregation of g_1 , g_2 , l_g , l_{g_2} and l_{g_3} .

Ratio=405:27:555:37

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TABLE 43

	PURPLE LIGULE SHORT SPIKKLET	PURPLE LIGULE LONG SPIKELET	GREEN LIGULE SHORT SPIKELET	GREEN LIGULE LONG SPIKELET	TOTAL
Observed	87	21	108	40	256
Calculated 81:27:111:37	81	27	111	37	256
Deviation		—ი	-3		

Showing independent segregation between $l_{g_1}, l_{g_2}, l_{g_3}$ and s_p .

X2=2.09, **P=0.5561.**

TABLE 44

 $\label{eq:2.1} \frac{1}{2} \int_{\mathbb{R}^3} \frac{1}{\sqrt{2}} \left(\frac{1}{\sqrt{2}} \sum_{i=1}^3 \frac{1}{\sqrt{2}} \right) \frac{1}{\sqrt{2}} \, \mathrm{d} \mathcal{L} \, \mathrm{d}$

Showing independent segregation between h_1 , h_2 , a_{p_5} and a_{p_6} .

The deviation is probably due to the small number **of** plants.

Showing independent segregation between h_1 , h_2 , p_{r_1} and p_{r_2} .

The deviation is probably due to the small number **of** plants.