Estimates of Linkage Disequilibrium and the Recombination Parameter Determined From Segregating Nucleotide Sites in the Alcohol Dehydrogenase Region of Drosophila pseudoobscura

Stephen W. Schaeffer and Ellen L. Miller

Department of Biology and Institute of Molecular Evolutionary Genetics, The Pennsylvania State University, University Park, Pennsylvania 16802

> Manuscript received January 13, 1993 Accepted for publication July 2, 1993

ABSTRACT

The alcohol dehydrogenase (Adh) region of Drosophila pseudoobscura, which includes the two genes Adh and Adh-Dup, was used to examine the pattern and organization of linkage disequilibrium among pairs of segregating nucleotide sites. A collection of 99 strains from the geographic range of D. pseudoobscura were nucleotide-sequenced with polymerase chain reaction-mediated techniques. All pairs of the 359 polymorphic sites in the 3.5-kb Adh region were tested for significant linkage disequilibrium with Fisher's exact test. Of the 74,278 pairwise comparisons of segregating sites, 127 were in significant linkage disequilibrium at the 5% level. The distribution of five linkage disequilibrium estimators D_{ii} , D^2 , r_{ii} , r^2 and Dij were compared to theoretical distributions. The observed distributions of D_{ij} , D^2 , r_{ij} and r^2 were consistent with the theoretical distribution given an infinite sites model. The observed distribution of Dij differed from the theoretical distribution because of an excess of values at -1 and 1. No spatial pattern was observed in the linkage disequilibrium pattern in the Adh region except for two clusters of sites nonrandomly associated in the adult intron and intron 2 of Adh. The magnitude of linkage disequilibrium decreases significantly as nucleotide distance increases, or a distance effect. Adh-Dup had a larger estimate of the recombination parameter, 4Nc, than Adh, where N is the effective population size and c is the recombination rate. A comparison of the mutation and recombination parameters shows that 7-17 recombination events occur for each mutation event. The heterogeneous estimates of the recombination parameter and the inverse relationship between linkage disequilibrium and nucleotide distance are no longer significant when the two clusters of Adh intron sites are excluded from analyses. The most likely explanation for the two clusters of linkage disequilibria is epistatic selection between sites in the cluster to maintain premRNA secondary structure.

The observation of significant linkage disequilibrium that is consistent between populations is a very sensitive detector of natural selection (LEWONTIN 1974).

L INKAGE disequilibria or nonrandom associations among genetic markers may result from forces other than selection such as low recombination rates, random genetic drift, or population subdivision (KI-MURA and OHTA 1971; NEI 1987). LEWONTIN (1974) suggests that neutral forces fail to explain a consistent pattern of linkage disequilibrium in geographic populations unless the migration rate is sufficiently large to homogenize gene frequencies between populations. With extensive migration, population sizes would be too large for random genetic drift to maintain consistent patterns of nonrandom associations between linked loci.

Theoretical studies of the sampling distribution of linkage disequilibrium have focused on nonrandom associations between pairs of loci or sites to determine the importance of random genetic drift and selection as forces that generate linkage disequilibrium (HILL and ROBERTSON 1968; KARLIN and MACGREGOR 1968; SVED 1968; OHTA and KIMURA 1969a,b, 1971; SVED 1971; HILL 1975, 1977; STROBECK 1983; GOLD-ING 1984; HUDSON 1985). The expected value of linkage disequilibrium for a pair of selectively neutral loci in finite equilibrium populations is zero, although the variance of linkage disequilibrium can be quite large between populations (HILL and ROBERTSON 1968; OHTA and KIMURA 1969a). The alcohol dehydrogenase (Adh) region of Drosophila pseudoobscura is a model system to determine how much linkage disequilibrium exists in a selectively neutral region of the genome.

The Adh region is composed of two closely linked genes, Adh and Adh-Dup (SCHAEFFER and AQUADRO 1987), whose pattern and organization of nucleotide diversity failed to suggest the past action of positive Darwinian selection (SCHAEFFER and MILLER 1992b). In addition, evidence from nucleotide data indicate that geographic populations of *D. pseudoobscura* are not significantly substructured (SCHAEFFER and MILLER 1992a). We present here an analysis of linkage disequilibrium for all segregating sites in the Adh region of D. pseudoobscura to determine how much association exists among polymorphic sites in a genomic sequence that has not been influenced by selection or population subdivision.

MATERIALS AND METHODS

Strains and nucleotide sequences: Ninety-nine sequences were used in this study of linkage disequilibrium. Ninetyone of the nucleotide sequences were determined previously (SCHAEFFER and MILLER 1991, 1992a,b). The population location, strain names, and the EMBL/GenBank Data Library accession numbers for these 91 nucleotide sequences are: Stemwinder Provincial Park, British Columbia, Canada: BN16, BC86, BC93, BC414, PS101, PS102, PS108, PS126, PS139, PS163, PS164, PS170 and PS192 (X62181-X62183, X62185, X62230-X62234, X64468-X64471); Rainbow Orchard, Apple Hill, California: AH43, AH162, AH135, AH133, AH122, AH69, AH100, AH144, AH54 and AH165 (M60979-M60988); Gundlach-Bundschu Winery, Sonoma, California: GB12, GB13, GB24, GB26, GB27, GB32, GB33, GB41, GB54, GB59, GB66, GB68, GB69, GB78, GB82, GB91, GB92, GB96, GB103, GB104, GB109, GB112, GB114, GB116, GB118 and GB119 (X62191-X62216); Bryce Canyon National Park, Utah: BR3F, BR5F, BR7F, BR8F and BR10F (X62186-X62190); Mesa Verde National Park, Colorado: MV21, MV27, MV28, MV36 and MV43 (X62220-X62224); Kaibab National Forest, Arizona: PS214, PS219, PS220, PS224, PS230, PS243, PS245, PS261, PS262, PS265, PS274, PS279, PS281, PS282, PS283, PS287, PS289 and PS297 (X64472-X64489); San Bernadino Mountains, California: SB2, SB6, SB14 and SB18 (X62235-X62238); Mexico: MT350, MT228, MXA31, MXA64 and MXZ26 (X62218, X62219, X62225-X62227); Brookings, Oregon: OREG3 and OREG4 (X62228, X62229); MacDonald Ranch, California: DPSE (Y00602); Baja California, Mexico: BJ855 (X62184); and Gregory Canyon, Colorado: GC4 (X62217).

This study also included eight new nucleotide sequences from strains collected from the Kaibab National Forest. This increased the sample size of the Kaibab population to 26 strains and the overall survey to 99 strains. SCHAEFFER and MILLER (1991, 1992a,b) describe the methods used to determine the nucleotide sequence of the Adh region of D. pseudoobscura. The names of the new strains and their GenBank/EMBL Data Library accession numbers are: PS298, PS299, PS302, PS306, PS309, PS314, PS315 and PS316 (X68159-X68166).

DNA sequence alignment: Figure 1 shows the fine structure of the *Adh* region as well as the fragment of DNA that was sequenced. The 99 nucleotide sequences were aligned manually with the Eyeball Sequence Editor [ESEE, version: 2.00a; (CABOT and BECKENBACH 1989)]. The alignments were determined by minimizing the number of mismatches and gaps assumed in the sequences. Any nucleotide site with two or more bases present in any population was defined as a segregating site or polymorphism. All segregating sites were used in linkage disequilibrium analyses. Insertion and deletion polymorphisms were excluded from all analyses and will be considered in later papers. A segregating site found within an insertion or deletion was also excluded from analyses of linkage disequilibrium.

Measures of linkage disequilibrium: We used five measures to estimate the degree of association between all pairs



FIGURE 1.—Fine structure of the Adh region of D. pseudoobscura. The region is subdivided into 17 sequence domains: 5' flanking, Adh adult leader, Adh adult intron, Adh larval leader, Adh exon 1, Adh intron 1, Adh exon 2, Adh intron 2, Adh exon 3, Adh 3' leader, intergenic, Adh-Dup exon 1, Adh-Dup intron 1, Adh-Dup exon 2, Adh-Dup intron 2, Adh-Dup exon 3, and 3' flanking.

of segregating sites i and j in the Adh region D_{ii} , D^2 , r_{ij} , r^2 and D_{ij} [HUDSON (1985); see equations 1 through 5]. The number of independent estimates of linkage disequilibrium between two segregating sites is equal to $(n_i - 1)(n_i - 1)$ where n_i is the number of nucleotides segregating at site iand n_j is the number of bases segregating at site j (WEIR 1990). Fisher's exact test of independence was used to determine if the estimated linkage disequilibrium values were significantly different from zero (WELLS and KING 1980; SOKAL and ROHLF 1981). An association was considered significantly different from zero if the probability of the exact test was less than 0.05. This analysis had the potential to find significant nonrandom associations in excess of the significance level of the exact test. We adjusted our significance level with the sequential Bonferroni test procedure (RICE 1989), to overcome the multiple comparison problem.

Detection of clustered polymorphic sites: The STEPHENS (1985) test was used to detect sets of segregating sites that form congruent phylogenetic partitions. Sites that had the same phylogenetic information and were significantly clustered may indicate intragenic recombination or gene conversion. Lack of clustered polymorphisms suggests free recombination in the region. SAWYER (1989) also has a test to detect gene conversion, however, this method's test statistic may indicate that gene conversion has occurred but does not locate which sites are involved in the conversion event. Thus, Stephens test was used as a method to determine which segregating sites were involved in potential recombination or conversion events.

Estimates of nucleotide distance between segregating sites: The distance separating two segregating nucleotides may vary for different pairs of sequences due to the presence of insertions and deletions. The simplest method to estimate distance between a pair of sites is to subtract the 5' nucleotide position number from the 3' nucleotide position number which would include any insertions or deletions that occurred among all sequences. A more accurate method is to estimate the average number of nucleotides that separates the segregating sites across all sequences (Figure 2). These two measures of nucleotide distance are highly correlated (R = 0.989; P < 0.01). We used the former distance because it requires less time to estimate in computer simulations.

Relationship between linkage disequilibrium and nucleotide distance: Nucleotide sites that are tightly linked might be expected to be more associated than sites separated by larger nucleotide distances, *i.e.*, the distance effect. We determined if significant nonrandom associations decrease

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A	G	С	T	-	G	A	A	r	G	A	G	С	T	τ	T	С	C	-	-	G	С	Т	A	-	T	T	T	A	G	22BP
A	G	С	T	-	G	A	A	T	-	-	-	-	-	-	T	C	C	G	C	G	С	Т	A	-	T	T	T	С	G	18BP
A	T	С	т	т	G	A	A	T	-	-	-	-	-	-	Т	С	C	-	-	G	С	т	A	-	T	T	T	C	G	17BP
A	G	С	T	-	G	A	A	T	-	-	-	-	-	-	T	C	С	-	-	G	С	T	A	A	T	T	T	С	G	17BP
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FIGURE 2.—Illustration of two methods to estimate distance between a pair of polymorphic sites. The top estimate of distance counts all positions between the pair of segregating sites including insertions and deletions. The distance estimate shown in the lower right is an average of the actual number of bases that separate the polymorphic sites in each sequence, shown along the right edge of the sequences.

as nucleotide distances increase by estimating correlation coefficients between nucleotide distance and each of the linkage disequilibrium estimators D_{ij} , D^2 , r_{ij} , r^2 and D_{ij} (SOKAL and ROHLF 1981). We used the absolute value of D_{ij} , r_{ij} and D_{ij} in analyses of the relationship of linkage disequilibrium and distance. The statistical significance of each correlation coefficient was determined with two random permutation methods that shuffled the sites as columns in the aligned sequence. Let m be the total number of nucleotides in the aligned set of sequences and S be the number of segregating sites. Also, let m_i be the position of nucleotide site i in the aligned sequence $(m_i, i = 1, 2, 3, \ldots, m)$ and S_i be the position of segregating site i in the aligned sequence $(S_i, i = 1, 2, 3, \ldots, S)$. In this study, (m = $3,618; m_1 = 1, m_2 = 2, m_3 = 3, \ldots, m_{3618} = 3,618)$ and (S = $359; S_1 = 2, S_2 = 3, S_3 = 5, \ldots, S_{359} = 3,591)$.

The first method randomly shuffled the position of each nucleotide site, m_i , up to the total of m positions then the correlation between linkage disequilibrium and nucleotide distance was estimated for the permutation replicate. The second method randomly shuffled the positions of each segregating site, S_i , up to the maximum of S positions then the correlation between linkage disequilibrium and nucleotide distance was estimated for the permutation replicate. The fraction, P, of 1,000 permutation replicates with correlation coefficients more extreme than the observed value determined the statistical significance of the relationship. The first random permutation method assumes that all nucleotide sites are equally likely to mutate, while the second method assumes that not all nucleotide sites are free to mutate (i.e., regulatory sequences or nonsynonymous sites in coding sequences).

Our linkage disequilibrium analysis showed two clusters of linkage disequilibrium (CLD) in the Adh region. These clusters add a large number of sites separated by short distances. To determine if these clustered sites generate a distance effect, we removed these sites from our analysis and repeated the correlation analysis described above for the five linkage disequilibrium estimators and two permutation methods.

Linkage disequilibrium and population structure: We examined the effect of population structure on levels of linkage disequilibrium by OHTA's (1982) method. OHTA defined variance components of linkage disequilibrium $(D_{ls}^2, D_{ST}^2, D_{ls}^{r2}, D_{ST}^{r2}$ and D_{TT}^2 ; see equations 10 through 14, respectively) which are analogous to WRIGHT's (1940) inbreeding statistics. OHTA suggests that limited migration explains the observed linkage disequilibrium between populations if D_{ls}^{r2} > D_{ST}^{r2} and $D_{ST}^2 > D_{ls}^{r2}$. On the other hand, epistatic selection

is more apt to explain the observed linkage disequilibrium between populations if $D_{IS}^{r2} < D_{ST}^{r2}$ and $D_{ST}^{r2} < D_{IS}^{r3}$. We examined the variance components of linkage disequilibrium for the Gundlach-Bundschu and Kaibab populations for the two clusters of linkage disequilibria in the introns of Adh. These two populations were used because the number of sequences collected from each location is 26. All other populations had sample sizes less than 10 individuals, thus, chi-square tests of homogeneity across all populations were not feasible.

Estimates of the mutation $(4N\mu)$ and recombination (4Nc) parameters: We estimated the mutation parameter, $\hat{\pi} = 4N\mu$, by NEI's (1987; see equation 10.5) method, where N is the effective population size and μ is the neutral mutation rate. BROWN, FELDMAN and NEVO (1980) designed a set of summary statistics to describe levels of multilocus associations between genetic markers. They suggested that the moments of the distribution of K, the number of nucleotide differences between pairs of sequences, summarize levels of multilocus association. In particular, the variance of the random variable K can be represented as a function of pairwise linkage disequilibrium, \dot{D} and D^2 , summed over all pairs of sites. The variance of K may be used to estimate the recombination parameter, $\hat{C} = 4Nc$, from nucleotide sequence data if the variation in the region is selectively neutral (HUDSON 1987; see equations 1, 3 and 4), where Nis the effective population size and c is the recombination rate between the ends of the region sequenced. \hat{C} is inversely proportional to the average level of D and D^2 in the region.

We estimated \hat{C} , $\hat{\pi}$ and $\hat{C}/\hat{\pi}$ separately for Adh and Adh-Dup to determine if levels of recombination, mutation and the ratio of recombination and mutation differ between the two genes. Adh-Dup is a functional gene that resulted from an ancient duplication of the Adh gene or vice versa (SCHAEF-FER and AQUADRO 1987; KREITMAN and HUDSON 1991). All segregating nucleotide sites were used to estimate \hat{C} on a per locus basis. \hat{C} and $\hat{\pi}$ were weighted by the number of base pairs in Adh or Adh-Dup to determine \hat{C} and $\hat{\pi}$ on a per nucleotide basis. We used a bootstrap approach to determine a 95% confidence interval (CI) on the estimates of \hat{C} , $\hat{\pi}$, and $C/\hat{\pi}$. For each replicate, a sample of 99 nucleotide sequences were drawn at random with replacement from the observed set of sequences. Sequences rather than sites were sampled at random because we wanted to simulate random draws of sequences from natural populations. We estimated \hat{C} , $\hat{\pi}$ and $\hat{C}/\hat{\pi}$ from the random draw of sequences in Adh and Adh-Dup. We randomly sampled the 99 sequences 1,000 times. The estimates of \hat{C} , $\hat{\pi}$ and $\hat{C}/\hat{\pi}$ for the 1,000 bootstrap replicates were each rank ordered and the 95% confidence interval was determined from the distributions of \hat{C} , $\hat{\pi}$ and $\hat{C}/\hat{\pi}$.

RESULTS

The distributions of linkage disequilibrium estimators: We observed 359 segregating sites in the 99 Adh sequences collected from the geographic range of *D. pseudoobscura*. Of the 359 polymorphic sites, 332 and 27 sites had two and three nucleotides segregating, respectively (Figure 3). We generated 74,278 independent values for each of the linkage disequilibrium estimators (54,946 estimates for all comparisons of 332 sites with two nucleotides; 1,404 estimates for all comparisons of 27 sites with three nucleotides; and 17,928 estimates for all comparisons of the 332 sites with two nucleotides and 27 sites with three nucleotides). 544

Α				
	a 	b c 51111111111111111	111111111111122222222222222222222222222	44444456
	33311- 875534	122345555789990000012333345556 009481268593893478937034580695	7778888899999900000122333333444455577790222333333444455566789999122333334444 189024593456781268916902567802346013255012312456737890591423567947802480135	455577702
DPSE GC4	CTGAGC	ATGOCOCTTOCAAACACOGATCAACTGOOG	GGTTAAACTCGTTGGCGTTAGGTGAATTCAATTATTTCTAGTATTCCCATTATGTTCCAATCTGTTTGTGAAGGA	CGGAACGGA
BJ855 OFFC3	•••••	T	۸G	
OREG4		T		
582 586		T	Α	
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BR3F BR5F	c	Τ	G.A	T
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BRIOF		I	Α	c
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AH162			AA	
AH133		r	C A	
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G854 G859		Г	ΑΑΑ	
G866		ſGAG.G	AA	G
C869		r	Α	
G878 G882		ГСGА	ΑΑCΑC	
GB91 GB92	·	f r		
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G8103		r		• • <i>· ·</i> · • • • • • •
G8109 G8112	c	r	ΑG	T
G8114 G8116	. c	Г Г Т А		••••••
G8118		ΓΤ	ACCC.	•••••
GB119 BN16		Γ	A	c
8C93 8C86		r		
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P5102	.c.	ſG	AG	T
PS108 PS126	.c	ſ ſ		• • • • • • • • • • • • • • • • • • •
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PS192		[c
PS214 PS219		Г.АG Гсс	CA	• • • • • • • • • • • •
P5220	.c	TTTT	Α	
P5230		T		
PS243 PS245		r		• • • • • • • • • • • • • • • • • • •
PS261 PS262	.c	ΤΑΤ ΤΑ	АGGGGG, С. ЛАGАCAAAC АG	
PS265 PS274		TA	Α	т
PS279		T	GAAG.A	
PS281		TAGAA	۸	G
PS283 PS287		T	۸	
P\$289	. c	TG	٨٨	
PS298	••••••	т ГС.	Α.CΤ	
PS299 PS302		ГG Г		. T
PS306 PS309		ГАТ ГССАТ	A	
PS314		TG		c.T
P5316		ΤΑΤ	AG.CAT. TACAAAC.	

FIGURE 3.—Segregating sites found in 99 strains of D. pseudoobscura in the Adh region used in an analysis of linkage disequilibrium. The numbers above the sequence are the position numbers of each segregating sites within the reference sequence DPSE (SCHAEFFER and AQUADRO 1987). The sequence domains of Adh are indicated by letters and are shown above the site numbers. The domains are abbreviated as follows: a. 5' Flanking ; b, Adh adult leader; c, Adh adult intron; d, Adh larval leader; e, Adh exon 1; f, Adh intron 1; g, Adh exon 2; h, Adh intron 2; i, Adh exon 3; j, Adh 3' leader; k, intergenic; l, Adh-Dup exon 1; m, Adh-Dup intron 1; n, Adh-Dup exon 2; o, Adh-Dup intron 2; p, Adh-Dup exon 3; and q, 3' flanking. The names of the sequences are given in MATERIALS AND METHODS.

The distributions for the five linkage disequilibrium estimators across all pairwise comparisons are shown in Figure 4. The theoretical distributions for D_{ij} , D^2 , r_{ij} , r^2 and D'_{ij} have been determined by Monte Carlo simulation for two loci with known estimates of the mutation and recombination parameters, $4N\mu$ and 4Nc (GOLDING 1984; HUDSON 1985). The comparison of the observed and theoretical distributions should be viewed with caution because the distribution is based on comparisons of sites that differ in allele

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OPSE GC4	CCAATAGGAAGCTGCAAAACTG	TACTO	GEGTAAATTEATETACEGGEACECECEGECE	COCATCACACTCTCCCC	TTGAACCCTTTT	CTGTCT	GCCTCCGGGTCAAGAGTGATCCGAGGC
BJ855 OREG3		A	ATA			ATC	
OREG4	TAC T	A	AGAG	CA. TTT TT			
586 5814	c	A	AAC	TA. TTT		A.C. TC	
5818 BR 3F	c.	A	A	TA.TTT	·····	TC	CGT A
BR5F BR7F		A	ΑΠ	TA. TTT	T	AC	
BRSF	π	Å	AA.T.			.T	
AT350		Â	A			ATC	
AXA 31		AG	A		G.	с	
AXA64		A/	AGTTAAA.		AAATT	Ac	
EV21 EV27		A/	A	TA. TTT	.	A.CTC	TACGT
AV28 AV36		A/	A			A.CTC	.τ
MV43		A	AA.TA	TA.TTT		C	c.cct
AH162		A	A	~~~~		тс	
AH133	TA	A	Α			A	
AH69		Ā	A		T	тс	
AH144		A	n		· · · · · · · · · · · · · · · · · · ·	ctc	
AH165		A/ A/	AGTT	TA.TTT		тс Атс	
G812 G813		A/	Απ		T	тс	
GB24 GB26	T	A	ΑC	T	сс	тс	C
GB 27 GB 32	G	AT . A	A	••••••	T	тс	
G833	c	A	AAC	TA.TTT	· · · · · <u>-</u> · · · · · ·	A.CTC	
G854		A /	ACG		•••••••••••••	тс	сссст
G866		Å I	AGAG		· · A · · · · · · · · · · · · · · · · · · ·	A.C.CTC	
G869		Å /	Α		AAATT	тс	
G882	•••••••	•••••	ATT	TA.TTT			.AA. .CGTA
G891	G	A/	ΑΤΤG	TA. TTT	G.	с	сст
G896 G8103	GC	A <i>I</i>	ΑΤΤΤΟ			.с. тс .сс	CGT
GB104 GB109		A/	ACA.			тс	
G8112 G8114	G.	A	AAGTT	CA	AAATT	TC	
G8116	·····	A/	ATTC			тс	
GB119		A /	ATTG		· · · · · · · · · · · · · · · · · · ·	тс	CGT
8093	G	AT . A/	ΑΤΤG	· · · TA · TIF · · · · · · · · ·	.ст		CGT
BC86 BC414	·····	A/	A.A	TA.TTTA		c	CGT
PS101 PS102		A) A)	ΑΑ.ΤΑ Α	TATTT	· · · · · · · · · · · · · · · · · · ·	С	C.CCT
PS108 PS126	G	AT.A/	ATTG		T	TC	
PS 139 PS 163	G	AT . A/	ATTG	CA TIT	T	тс	
PS164 PS170			GC	Th TTT	• • • • • • • • • • • • • • • • •	тс	.A
PS192			G			C	
PS219				ГТА.ТТТ			
P5224	•••••••••••••••••••••••••••••••••••••••		Λ		**************************************	тс	
PS230 PS243			Α			c	
PS245 PS261	·····		Ν	TA.TTT		C	
PS262 PS265	G		τ		· · · · · · · · · · · · · · · · · · ·	.cç	
PS274 PS279		T		T			
PS281 PS282	·····		G	· · · · · · · · · · · · · · · · · · ·		TC	
PS283	· · · · · · · · · · · · · · · · · · ·		π			TC	
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5299			c			тс	G
PS306 PS309	·····					.ctc	
PS314 PS315	· · · · · · · · · · · · · · · · · · ·					c	CCGT
P\$316	· · · · · · · · · · · · · · · · · · ·					тс	

FIGURE 3.—Continued

frequency. In addition, the disequilibria values are correlated especially when one segregating site is in common. The observed distributions D_{ij} and r_{ij} were consistent with theoretical distributions that had used large values of 4Nc (HUDSON 1985) because much of the probability distribution is located at D_{ij} or r_{ij} equal zero. The observed distribution of D'_{ij} resembled the theoretical distributions that are based on 4Nc near zero because 90% of the D'_{ij} are either -1 or 1. The observed distributions of D^2 and r^2 had more values 546

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6					
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	45578258566666788999	1124445334679900123345	5668990447991	12223333444556777811233	44555677999022233345667800144456789357891
OPSE	84532925702347423234 CTCCCTCAACCCAAATACTT	0380368398926902735846 TCACCTOCOCACTOCOCTOCOC	93505876904568 CCTCCTACACCC	82360456236790138228469	25148223349836725879251946013981001882597
GC4	····.c	CG.C	TTCG	NCC	GTC
B J855	····cc	ст	TT.G/	c	GA
OREG4		C	TTCG	ντcg	GCAA
582		ст	ГA.T.G	۱cc	G
585 5814	·····	c	ГCGAG4		GC
5818		C	ΓΤ	N C	G
BR3F	······································	C	ГТ.С	••••••••••••••••••••••••••••••••••••••	GC
BR7F	······································	CTG	Г	NC	GCA
BRSF		ст	ГА.Т	۱C	GC
BRIOF AT350	····A·····C····C·	cc	ΓΑ.ΤΤ/	\	GTG.T
#1228		CCG	TA.T.G	ι	GC
AXA31		c	Г А. ТСG И		GCC
ANZ26		C	E A. 10 /	· · · · · · · · · · · · · · · · · · ·	GG.AG
MV21		c	гт	۱	GTG
AV27 AV78		cc.	TA. T.G	G	GC
AV36	T	C	T		C.AG
AV43		<u>.</u>	ГТ.G		G
AH162		C	ГТТG	NCT	
AH135	· · · · · · · · · · · · · · · · · · ·	c	ГТ.G	۱œ	GC
AH133 AH177	······	CG	ГА.ТСА	G CC	GCAAATCGT
AH69	G	C	Г АТ. G1	ſ	
AH100	c	çc.c	т.с	cc	ст.
AH54			ΓΤ.G	кт	цКАТ.
AH165	T	CGA.1	ГТ	cc	
G812 G813	G	c		ГТ	
G824		C	ΓΤ	NG	GCAAT
G826		<u>.</u>	г А. ТС А	·····	GCAA
G827 G832	· · · · A · · · · · · · · · · · · · · ·	СТТ СТТ	Г А. Т. G 1 Г. А. Т. G 1	ГТ Г АС Т	
G833	c	CG	CGAG	· · · · · · · · · · · · · · · · · · ·	GC
GB41	····A······	CTT	ГА.Т.G 1	ſ 	
G859		C	ГАС		GC
G866		ccc	TT.G	cc	
G869	·····	CCG	ГТ.G/ ГТА./	N.C	GG
G878	c	стG	ГА.ТС		GCAAT
G882 G891	·····C	CG	Г А. ТСС Т А		GT
G892	····A. · · · · · · · · · · · · · · · · ·	ст т	Г А. Т. G Т	F	
G896		ç	[N.C	G
G8103	· · · · · · · · · · · · · · · · · · ·	C	ГТ.GИ		GA
G8109		C	Г А. Т. G		AA
G8112 G8114		CG	ГТ.GА Г.А.Т.С. П	ΛCC	GTCC.
G8116	C	C	ſ	G	GCAAT
G8118	·····	<u>.</u>	ГТ.G		ccc.
BN16	Τ	CT.A.GA		ν	G
BC93	· · · · A · · · · · · · · · · · · · · ·	CTT1	ГА.Т.G1	rt	
BC86 BCA14	····¢····c·	cc	Г А. Т Т А Г А. Т. С	······································	GTG.T
PS101		C	ГТ.G	G	G
PS102		cc	гт.с. <i></i>	c	GA
PS126	····A	un an faire an	ГА.Т.G1 ГА.Т.G1	ГТ Г.,Т.	
PS139	· · · · A · · · · · · · · · · · · · · ·	ст т	ſ	ſ	
PS163 PS164		C	ГТ	G	GGAAG
PS170		CCG	гт	N G	
PS192	· · · · · · · · · · · · · · · · · · ·	c.c	Г А. Т	c	GG
P5219		CCGT	1 A . 1		ч.,,лл
PS220		C	Г А. Т. G А	GCA	G
PS224 PS230		ECG	ГТСИ ГТ.СИ		G.,CAA
PS243	CC	CG	ГА. ТС	·····	
P5245	·····c	c	Г А. ТС	••••••	
PS262		C	ГТ.С	CA.C	······································
PS265	c	c	ГА.ТС	۰	GCAAGCT
PS274	· · · · · · · · · · · · · · · · · · ·	<u>.</u>	ГТ/ Г Т	۱cco	GGAAGG
P5281		CG	Г Т. G		GAAC
PS282	·····	c	гт.с	،c	GAAC
P5283 P5287		c			GCGA
PS289		CTCG	г.т. тс	·····	CAA
P5297		ççq	ГТ.GИ	······	GCAAATT
PS299	·······················	стс	ГТ/	v	
P\$302	TCA	c	гтси	c	G
PS306 PS309	·····c····	C	1Α.Τ	N	, . , , . , . , . , . , . , . , . , . ,
P\$314	GG	C	ГТ.Ğ/	۱ с	GC
P\$315	·····c.	<u>.</u>	TT	c	
73310		••••••			u,,unn

FIGURE 3.—Continued

near zero than the theoretical distributions with 4Nc equal to zero (GOLDING 1984; HUDSON 1985).

A total of 127 of the 74,278 (0.17%) pairwise comparisons of segregating sites were significantly different from zero when the Fisher's exact test was used with the sequential Bonferroni method (WELLS and KING 1980; SOKAL and ROHLF 1981; RICE 1989) (Figure 5). The overall significance level used was 6.7 $\times 10^{-7}$ (0.05 significance level/74,151 comparisons). Two clusters of segregating sites were significantly Linkage Disequilibrium in Adh



FIGURE 4.—A comparison of the distributions of five linkage disequilibrium estimators D_{ij} , D^2 , r_{ij} , r^2 and D'_{ij} . The distributions are based on the comparison of all pairs of the 359 segregating sites in the *Adh* region.

associated in the Adh region, sites in the adult intron (nucleotides 331, 332, 334, 337, 343, 347, 350 and 355) and intron 2 of Adh (nucleotides 1454, 1460, 1464, 1465, 1467, 1471, 1473, 1474, 1475, 1495, 1496, 1497, 1498 and 1500) (Figure 5). These sites will be referred to as the clustered linkage disequilibrium (CLD) sites. Comparisons of sites between the two clusters were independently associated when tested with Fisher's exact method. Similar patterns of linkage disequilibrium were observed in the Gundlach-Bundschu (n = 26) and Kaibab (n = 26) populations (S. W. SCHAEFFER, data not shown).

The STEPHENS (1985) test detected 29 phylogenetic partitions with segregating sites that were significantly clustered. We observed 21 partitions that discriminated a single strain from the 98 others (Table 1) and eight partitions that grouped multiple strains together (Table 2), single and multiple strain partitions, respectively. The number of segregating sites within any of the phylogenetic partitions varied from a minimum of 2 to a maximum of 10 (Tables 1 and 2). The majority of phylogenetic partitions were composed of 2 or 3 polymorphic sites. The Stephens test detected some of the segregating sites in the CLD sites in the adult intron and intron 2 of Adh (Table 2; partitions 2 and 1, respectively). Some sites within the two CLDs were undetected by the Stephens test because not all the CLD sites have the same phylogenetic information.

Correlation of linkage disequilibrium and distance: The 127 pairs of sites in significant linkage disequilibrium were tested for the distance effect. The five linkage disequilibrium estimators were significantly negatively correlated with nucleotide distance when tested with both random permutation methods (Table 3). The Gundlach-Bundschu and Kaibab populations do not show a significant correlation between



FIGURE 5.—Significant nonrandom associations among all pairs of 359 segregating sites in the *Adh* region in a sample of 99 strains of *D. pseudoobscura*. Each box in the matrix represents the comparison of two polymorphic sites. A black box represents a comparison where Fisher's exact test was significant at the 0.05 level. The location of the segregating sites relative to the two structural genes is shown on the diagonal. The only segregating sites included in the figure are those that show significant nonrandom associations.

nucleotide distance and any of the linkage disequilibrium estimators with either random permutation analysis (S. W. SCHAEFFER, unpublished data). We had little power to detect a distance effect in these populations because the number of strains and segregating sites was small. The distance effect disappears when the CLD sites were removed from the correlation analysis for four of the five estimates of linkage disequilibrium (Table 3). The distance effect still exists for D^2 when the CLD sites were removed.

Linkage disequilibrium and population structure: The two clusters of linkage disequilibrium may be due to either limited migration between demes or epistatic selection. We used OHTA's (1982) variance of linkage disequilibrium analysis to discriminate between the epistatic selection and limited migration hypotheses. All pairs of segregating sites within the adult intron and intron 2 were used to estimate D_{IS}^2 , D_{ST}^2 , $D_{IS}^{'2}$, $D_{ST}^{'2}$ and D_{IT}^2 from the Gundlach-Bundschu and Kaibab sequence data. The majority of comparisons showed D_{ST}^2 and $D_{IS}^{\prime 2}$ to be greater than D_{IS}^2 and $D_{ST}^{\prime 2}$, respectively, in the two intron sites (Figure 6). The data were more consistent with the epistatic selection hypothesis rather than the limited migration hypothesis. In other words, similar nonrandom associations were observed in the two populations. These data are consistent with the analysis of population structure by SCHAEFFER and MILLER (1992a).

				Statist signific	tical ance
Strain	\$	do	g.	$P(d \leq d_o)$	P
PS261	10	2653	2562	0.208	0.000
SB2	4	1725	1553	0.278	0.029
GB68	3	1943	1941	0.555	0.001
DPSE	3	2633	2179	0.818	0.001
PS315	3	3279	1958	0.975	0.001
PS214	3	1384	1095	0.327	0.001
BR8F	3	2801	1660	0.870	0.001
PS265	3	1391	994	0.330	0.001
MXA64	3	2726	1366	0.848	0.001
PS219	3	2984	2673	0.919	0.001
AH43	3	3195	3155	0.962	0.001
GB119	3	2840	2816	0.881	0.001
PS314	3	2020	1317	0.587	0.001
PS163	3	1789	1732	0.492	0.001
PS192	3	1572	1143	0.402	0.001
MXZ26	3	2157	1864	0.643	0.001
MXA31	3	867	801	0.145	0.002
PS164	3	1184	800	0.251	0.002
OREG3	2	1		0.001	
AH135	2	1		0.001	
PS306	2	40		0.022	

S, number of segregating sites; d_o , observed distance between the distal segregating sites; g_o , observed maximum distance between segregating sites for $(S \ge 3)$; $P(d \le d_o)$, probability of observing a distance less than d_o ; and P, probability that a random segment of length, $d_o - (S - 1)$, has at least one segment greater than g_o .

Estimates of the mutation and recombination parameters in Adh and Adh-Dup: The estimate of \hat{C} was significantly larger in Adh-Dup than in Adh. The estimate of $\hat{\pi}$ was significantly larger in Adh than in Adh-Dup. Our estimate of the ratio of the recombination and mutation rates, c/μ , was greater in Adh-Dup than in Adh (Table 4). The differences of \hat{C} , $\hat{\pi}$ and c/μ between Adh and Adh-Dup were statistically significant because the 95% confidence interval did not overlap for the two genes (Table 4).

HUDSON'S (1987) estimate of \hat{C} assumes that all the segregating sites are selectively neutral. OHTA'S (1982) analysis suggests that the two clusters of segregating sites in the *Adh* introns may be in linkage disequilibrium because of epistatic selection. Thus, we estimated \hat{C} , $\hat{\pi}$ and c/μ after removing the CLD sites from the data set. The estimates of \hat{C} , $\hat{\pi}$ and c/μ were not statistically different between *Adh* and *Adh-Dup* when the CLD sites were removed (Table 4). The estimates of the ratio of recombination and mutation for the *Adh* region suggest that 7–17 recombination events have occurred for each mutation event (Table 4).

DISCUSSION

The distribution of linkage disequilibrium estimators: The expected value of linkage disequilibrium

TABLE 1

Clustered polymorphisms for single strain partitions in the Adh region

TABLE 2

Clustered polymorphisms for multiple strain partitions in the Adh region

				Statist signific	ical ance
Partitions	S	d_{o}	g.	$P(d \leq d_o)$	Р
1	4	7	4	0.000	0.187
2	3	12	6	0.000	0.174
3	2	6		0.003	
4	2	1		0.001	
5	2	2		0.001	
6	2	24		0.013	
7	2	3		0.002	
8	2	2		0.001	
All sites	359	3589	115	0.201	0.002

The symbols are defined as in Table 1. Strains that comprise each partition are as follows: Partition 1: DPSE, BR3F, BR7F, MT350, MXA31, MXA64, MXZ26, MV27, AH43, AH162, AH135, AH133, AH122, AH165, GB13, GB24, GB26, GB27, GB32, GB41, GB59, GB82, GB92, GB96, GB112, GB114, GB116, BC93, PS108, PS126, PS139, PS164, PS219, PS220, PS230, PS261, PS274, PS279, PS281, PS282, PS289, PS299, PS309, and PS316; Partition 2: MT350, MT228, MXZ26, MV43, AH69, AH54, GB12, GB24, GB27, GB32, GB33, GB82, GB91, GB92, GB114, GB116, BC93, BC414, PS101, PS108, PS126, PS139, PS192, PS214, PS243, PS245, and PS315; Partition 3: MXZ26 and GB119; Partition 4: SB2, AH100, GB109, and PS274; Partition 5: BR8F and PS306; Partition 6: AH135, AH165, and GB112; Partition 7: MXA31, MXA64; AH135, AH165, GB68, GB112, PS220, PS261, and PS279; and Partition 8: DPSE, SB2, AH43, PS224, and PS289.

TABLE 3

Correlation between nucleotide distance and five linkage disequilibrium estimators

Ent	All	sites inclu	ded	All sites with CLD removed					
mator	Cor- relation	P_1	P_2	Cor- relation	P_1	P_2			
D_{ij}	-0.32	< 0.004	< 0.005	-0.20	NS	NS			
r_{ii}	-0.34	< 0.004	< 0.002	-0.05	NS	NS			
D'_{ij}	-0.31	< 0.002	< 0.005	0.02	NS	NS			
D^2	-0.34	< 0.001	< 0.001	-0.22	< 0.041	< 0.046			
r^2	-0.38	< 0.001	< 0.002	0.00	NS	NS			

 P_1 , probability of observing this correlation coefficient or a larger value determined from 1000 random permutations that shuffled the positions of all *m* nucleotides at random; P_2 , probability of observing this correlation coefficient or a larger value determined from 1000 random permutations that shuffled the positions of the *S* segregating sites at random; and CLD, clusters of segregating sites in linkage disequilibrium in the adult intron and intron 2 of *Adh*.

for two loci in equilibrium populations is zero although its variance can be quite large (HILL and **ROBERTSON** 1968; OHTA and KIMURA 1969a). The comparison of all pairwise combinations of 359 segregating sites in the *Adh* region of *D. pseudoobscura* shows that 0.1% of the linkage disequilibria were significantly different from zero. The observed distributions of D_{ij} , D^2 , r_{ij} and r^2 were each consistent with their theoretical distributions given our estimates of



FIGURE 6.—Variance of linkage disequilibrium analysis for the Gundlach-Bundschu and Kaibab populations by OHTA's (1982) method. All pairwise comparisons of CLD sites within the adult intron and intron 2 of *Adh* were used to estimate D_{IS}^2 , D_{ST}^2 , D_{IS}^2 , D_{ST}^2 , and D_{TT}^2 . Each box in the figure represents a comparison of a pair of sites. The position of each site in the standard sequence DPSE is shown along the diagonal. If a pairwise comparison finds $D_{IS}^{2} < D_{ST}^{2}$ and $D_{ST}^2 < D_{IS}^2$ then the box is shown as an open box. If a pairwise comparison finds $D_{IS}^{2} > D_{ST}^{2}$ and $D_{ST}^2 < D_{IS}^2$ then the box is shown as a closed box. If a pairwise comparison finds $D_{IS}^{2} > D_{IS}^{2}$ then the box is shown as a hatched box.

the recombination parameter (Figure 4, Table 4) (GOLDING 1984; HUDSON 1985). The observed distribution of Dij, however, differs significantly from its theoretical distribution with $\hat{C} = 20$ or ∞ presented in Figure 4 of HUDSON (1985). We found that 90% of the pairwise comparisons of sites estimated to be either -1.0 or 1.0. The frequency spectrum for the Adh region suggests an explanation for the observed Ushaped distribution of Dij. Of the 359 segregating sites in the region, 262 sites have frequencies of rare variants ≤ 0.05 . These sites are likely to have estimates of Dij near -1.0 or 1.0 and to account for approximately 46% of the pairwise comparisons of the polymorphisms. Rare frequency sites are most likely to have arisen recently in the population by the introduction of new mutations. Therefore, the observed distribution of D'_{ii} may reflect linkage disequilibria due to recently introduced mutations.

The distributions of the linkage disequilibrium estimators determined in this study should be viewed with caution because the distributions were deterTABLE 4

Estimates of the recombination and mutation parameters for the Adh and Adh-Dup genes

Gene	4Nc (95% CI)	4Nµ (95% CI)	c/µ (95% CI)
Adh	0.083 (0.047-0.097)	0.013 (0.012-0.013)	6.558 (3.749-7.742)
Adh-Dup	0.383 (0.135-0.453)	0.010 (0.010-0.011)	37.202 (13.323-44.583)
Total	0.087 (0.044-0.092)	0.012(0.011 - 0.012)	7.453 (3.766-7.932)
Adh w/o CLD	0.184(0.071 - 0.274)	0.009(0.008-0.009)	21.283 (8.319-33.072)
Total w/o CLD	0.152 (0.061-0.155)	0.009 (0.009-0.010)	16.264(6.644 - 16.925)

4Nc, neutral recombination parameter; $4N\mu$, neutral mutation parameter; CI, confidence interval; CLD, sites in clustered linkage disequilibrium in the adult intron and intron 2 of Adh.

mined from many segregating sites with different allele frequencies. In addition, some of the linkage disequilibria may be correlated when one segregating site is held in common. HUDSON's (1983a,b) simulation approach is a method that may determine the neutral expectation of linkage disequilibrium for an assemblage of segregating sites given our estimates of the mutation and recombination parameters. This approach would allow us to determine if clusters of linkage disequilibria develop with regularity and if an excess of rare frequency variants is typical under an infinite sites model of molecular evolution. Our estimates of $4N\mu = 37.83$ and 4Nc = 281.68 (on a per locus basis) are prohibitively large for the computations in the HUDSON (1983a,b) simulation method to determine theoretical expectations.

The distance effect: We observed a significant distance effect. The distance effect disappears for four of the five linkage disequilibrium estimators when the two clusters of linkage disequilibrium in the *Adh* introns are removed from the analysis (Table 3). These data suggest that recombination is quite effective in breaking up associations among sites in this region. In fact, more recombination events occur relative to mutation events, 7 to 17. Thus, new linkage disequilibrium introduced by the mutation process is probably short lived in the *Adh* region of *D. pseudoobscura*.

The two clusters of linkage disequilibria in the introns of Adh: The only significant pattern of linkage disequilibrium observed in the Adh region were the two clusters of linkage disequilibria in the adult intron and intron 2 of Adh. Could the alignments generate these clusters of linkage disequilibrium? The adult intron cluster of 8 sites covers 25 base pairs of sequence; only 2 of the 8 segregating sites were consecutive and no gaps were assumed in the local alignment. The intron 2 cluster of 14 sites occupies 47 nucleotide positions; 2 and 4 segregating sites were in consecutive positions and one gap was assumed in the local alignment, however, the gap was not located near the consecutive bases. Commercially available sequence alignment programs (ALIGN DNASTAR, Madison, WI), with all their shortcomings, support the alignments that were used in these regions. In addition,

not all the segregating sites in the clusters were in absolute or complete linkage disequilibrium, intermediate types were found. Therefore, we do not think that sequence alignments of the two regions are artificially creating linkage disequilibrium.

Three possible explanations could account for the clusters of linkage disequilibrium: (1) low recombination rates, (2) population subdivision or (3) strong epistatic selection. We have shown that estimates of the recombination rate in the Adh region are larger than the mutation rate because the ratio of the recombination and mutation parameters is significantly different from one (Table 4). Recombination rates do vary across the Drosophila genome (AGUADE, MIYASH-ITA and LANGLEY 1989a,b; STEPHAN 1989; STEPHAN and LANGLEY 1989; BEGUN and AQUADRO 1992), however, there is no evidence that recombination rates vary within a 4-kilobase region. Thus, the recombination rates that we estimate for the Adh region seem sufficient to break up the associations in the two intron clusters.

Population subdivision could create nonrandom associations among the intron sites. Two analyses suggest that D. pseudoobscura populations are not subdivided because gene flow is extensive between populations. SCHAEFFER and MILLER (1992a) have shown that at least two migrants are exchanged between populations each generation which is sufficient to homogenize gene frequencies between all North American populations of D. pseudoobscura. OHTA's (1982) variance of linkage disequilibrium analysis shows that most comparisons of sites in the two introns are more consistent with an epistatic selection hypothesis than with a limited migration or population subdivision hypothesis (Figure 6). In other words, the major haplotypes or multisite genotypes detected in the two intron clusters are found across all populations. Thus, population subdivision is an unlikely explanation for the clustered linkage disequilibrium.

If low recombination rate and population subdivision are unlikely explanations for the two intron clusters of linkage disequilibria then strong epistatic selection is left as the most probable hypothesis. LEWONTIN (1974) suggests that: "The observation of significant linkage disequilibrium that is consistent between populations is a very sensitive detector of natural selection." Why would the linkage disequilibrium analysis suggest epistatic selection and previous studies show no evidence of the past action of positive Darwinian selection (Schaeffer and MILLER 1992a,b)? The TA-JIMA (1989) and HUDSON, KREITMAN and AGUADE (1987) tests of selective neutrality, performed in SCHAEFFER and MILLER (1992a,b), lumped nucleotide sites into two large regions Adh or Adh-Dup. Thus, tests of selective neutrality may have missed strong epistatic selection on two small local regions such as the intron clusters because the majority of sites in the region evolve under the assumptions of an infinite sites model of molecular evolution.

What is the nature of the epistatic selection? Analysis of pre-mRNA secondary structure of the Adh transcript indicates that the segregating sites in intron 2 form stem-loop structures (WOLFGANG STEPHAN, University of Maryland, personal communication). Ten of the segregating sites in intron 2 form three major haplotypes. Two of the haplotypes form stable stemloop structures. The strong linkage disequilibrium among sites appears to be due to compensatory nucleotide changes that maintain mRNA secondary structure. We do not know whether the cluster of linkage disequilibrium in the adult intron of Adh is due to compensatory changes that maintain premRNA secondary structure. Further theoretical and molecular analysis is necessary to confirm or refute the pre-mRNA hypothesis.

Analysis of linkage disequilibrium in the Adh region of D. pseudoobscura showed little evidence for nonrandom associations between segregating sites. Levels of recombination in the Adh region appear to be sufficient to rapidly breakup associations between nucleotide sites that are introduced by new mutations. The two clusters of linkage disequilibrium observed in the introns of Adh may result from strong epistatic selection to maintain secondary structure of the premRNA.

We would like to thank CHARLES F. AQUADRO, ANDREW G. CLARK, CHARLES H. LANGLEY, KIMBERLYN NELSON, WOLFGANG STEPHAN, BRUCE S. WEIR, THOMAS S. WHITTAM and three anonymous reviewers for thoughtful comments on the analyses presented in this manuscript. This work was supported in part by grant GM42472 to S.W.S. from the National Institutes of Health.

Note added in proof: The two clusters of linkage disequilibrium observed in the Adh introns of D. pseudoobscura are similar to a pattern observed at the white locus of D. melanogaster (MIYASHITA and LANGLEY 1988). A recent analysis of the population structure of the cluster of linkage disequilibrium in the white locus suggests that strong epistatic selection maintains the nonrandom associations among segregating sites (MIYASHITA, AGUADÉ and LANGLEY 1993). These data may indicate a general phenomenon of nonrandom association that may be found at other genetic loci in the genome.

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Communicating editor: B. S. WEIR