

A Method of Estimating the Numbers of Human and Mouse Immunoglobulin V-Genes

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ABSTRACT

Mutations in immunoglobulin V-genes can be due to gene multiplication, allelic variations, mutations induced by antigens or somatic mutations, etc., and various combinations of these. Since the number of different mouse lambda light V-gene nucleotide sequences is relatively small, a pairwise comparison between these sequences can provide a rough idea as to the contributions of the above mechanisms to the number of nucleotide differences between sequences. A plot of occurrences against the number of differences suggests that differences between one to five can be attributed to somatic mutations. Six to 12 differences can be allelic. Thirteen to 17 may be due to allelic variations together with somatic mutations. Differences >17 appear to be derived from gene multiplication. Although these numbers are most likely somewhat different in humans, they can nevertheless provide a rough guide to sort out the effect of gene multiplication. Estimations of human heavy, kappa and lambda light chain immunoglobulin V-genes are in reasonably good agreement with recent experimental studies. For mouse kappa light and heavy chains, our estimations can provide some insight to future analyses by direct sequencing of these gene segments.

SUBGROUPS were initially used to estimate the number of genes of the variable region of human kappa light chains (MILSTEIN 1967). Various subgroups have subsequently been defined for mouse kappa (POTTER 1967), human heavy (KABAT *et al.* 1976), mouse heavy (DILDROP 1984), and human lambda (KABAT *et al.* 1979) chains, using different subgroup definitions (see, for example, KABAT *et al.* 1991). Unlike the above systems, where subgrouping is used as an estimation of gene number, the mouse immunoglobulin lambda light chain V-genes is relatively small, and the gene number has reportedly been identified (SANCHER *et al.* 1987).

Several basic mechanisms are believed to be responsible for the differences between immunoglobulin V-gene sequences. Antigen stimulation, for example, causes somatic mutations in the variable region, but the number of such mutations is probably relatively small (CHEN *et al.* 1992). Divergence of strains within a species, such as mice, presumably would result in larger numbers of nucleotide differences between homologous genes (SERRANO *et al.* 1994). Finally, gene multiplication, which occurred a long time ago, would generate even larger numbers of nucleotide differences (ANDERSSON *et al.* 1991). Since these processes are probably occurring concurrently, a complicated pattern of mutations has been observed.

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To decipher the contributions from each of these mechanisms from the resulting mutation patterns in human and mouse immunoglobulin V-genes, pairwise comparisons of known complete nucleotide sequences of all human and mouse V-genes were generated and analyzed. The mouse lambda light chains provide a simple pattern of mutational differences that can be correlated with the three above-mentioned genetic processes. The resulting correlation can then be used to estimate the number of different genes for human and other mouse chains. While this approach is somewhat different from the conventional method of amino acid sequence comparison, our result, in principle, should provide a reasonable estimation of the number of different immunoglobulin V-genes.

MATERIALS AND METHODS

We have been collecting amino acid and nucleotide sequences of immunoglobulins and related proteins (KABAT *et al.* 1991). The complete nucleotide sequences of mouse and human lambda and kappa light and heavy chain variable regions coded by V-genes, *i.e.*, codons 1–95 for light chains and codons 1–94 for heavy chains, were used. Generally, cDNA sequences differ from genomic sequences in these regions by only a few nucleotides. The detailed references of these sequences are listed in the Kabat Database of Sequences of Proteins of Immunological Interest (JOHNSON *et al.* 1996). Each sequence is designated by a Kabat database identification number (KADBID), and can be retrieved by sending an e-mail to:

seqhunt2@immuno.bme.nwu.edu

with the following message:

TABLE 1
Nucleotide differences from pairwise comparison of distinct complete mouse lambda light chain V-genes

	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17
	007 665	007 671	007 674	007 675	007 683	007 684	007 685	007 687	007 688	007 689	007 690	007 691	007 692	007 695	007 697	007 700	007 703
1. 007665	0	1	2	1	2	5	7	2	2	2	3	8	4	7	9	8	8
2. 007671		0	3	2	3	6	8	3	3	3	3	9	5	8	10	9	8
3. 007674			0	3	4	7	9	4	4	4	5	10	6	9	11	10	10
4. 007675				0	3	6	8	3	3	3	4	9	5	8	10	9	9
5. 007683					0	7	9	4	4	4	5	10	6	7	9	8	10
6. 007684						0	12	7	7	7	8	13	9	12	14	13	13
7. 007685							0	8	9	9	8	1	11	14	14	15	15
8. 007687								0	4	4	4	9	6	9	11	10	10
9. 007688									0	4	5	10	4	9	11	10	10
10. 007689										0	5	10	6	9	11	10	10
11. 007690											0	9	7	10	12	11	10
12. 007691												0	12	15	15	16	16
13. 007692													0	11	13	12	10
14. 007695														0	2	1	15
15. 007697															0	3	17
16. 007700																0	16
17. 007703																	0
18. 007704																	
19. 007707																	
20. 007710																	
21. 007711																	
22. 007714																	
23. 007715																	
24. 007716																	
25. 007717																	
26. 007718																	
27. 007719																	
28. 007720																	
29. 007721																	
30. 007725																	
31. 007728																	
32. 007746																	
33. 007748																	
34. 007750																	
35. 019435																	
36. 020130																	

BEGIN

007665

for example. This message would retrieve entry 007665 from the database. Alternatively, the entries can be looked up through the database's WWW server at URL:

<http://immuno.bme.nwu.edu>

The Kabat database contains essentially all published nucleotide sequences of proteins of immunological interest currently available. To our knowledge, this is the largest and most complete aligned sequence database of these proteins.

A standard triangular table listing the number of nucleotide differences was generated by pairwise comparison of all distinct mouse lambda light chain variable region sequences. A second triangular table was obtained by removing any column or row containing differences of one to 17 from the first table. For other sequences, a similar second table was also created, together with a third table where columns and rows with differences of 18-34 were also removed.

The initial cutoff of 17 nucleotide differences was obtained from plotting the occurrences against the number of nucleotide differences from the first triangular table for mouse lambda light chains. For >17 differences, the mouse sequences being compared are most likely coded by two different genes.

RESULTS

The first triangular table for mouse lambda light chain V-genes (Table 1) contains 36 sequences each of which is identified by the KADBID as discussed in MATERIALS AND METHODS. Since many of these sequences are very similar, most the differences listed in the table are small. However, it is difficult to separate the relative contributions from various genetic mechanisms generating such differences, *e.g.*, gene multiplication, allelic variation, and somatic mutations induced

18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36
007	007	007	007	007	007	007	007	007	007	007	007	007	007	007	007	007	019	020
704	707	710	711	714	715	716	717	718	719	720	721	725	728	746	748	750	435	130
6	8	83	10	8	14	12	13	11	9	11	15	20	14	139	142	149	27	1
7	8	83	11	9	15	13	14	11	10	12	16	21	15	140	143	148	28	1
8	10	84	12	9	16	14	14	13	11	13	17	22	16	138	141	150	29	3
7	9	84	11	9	15	12	14	12	10	12	16	21	15	140	143	149	26	2
8	10	83	10	10	14	14	15	13	11	11	15	22	14	140	143	149	27	3
10	13	87	15	13	19	16	18	14	13	16	20	23	19	141	144	150	30	6
13	15	87	17	15	18	19	18	17	14	18	19	26	20	144	147	153	34	8
8	10	85	12	10	16	14	15	13	10	13	17	22	16	140	143	148	29	3
8	10	82	12	10	16	14	15	13	11	13	17	22	16	141	144	151	27	3
8	10	85	12	10	16	13	15	13	11	13	17	22	16	140	143	150	29	3
9	10	85	13	11	17	15	16	13	10	14	18	23	17	141	144	149	30	3
14	16	88	18	16	19	20	19	18	15	19	20	27	21	145	148	154	35	9
10	10	85	14	11	18	16	17	15	13	15	19	24	18	140	143	152	30	5
13	15	80	5	15	11	18	20	18	16	6	14	26	19	141	143	151	23	8
15	17	80	7	17	12	20	19	19	18	8	15	28	21	139	141	151	25	10
14	16	81	6	16	12	19	21	19	17	7	15	27	20	142	144	150	24	9
14	3	86	18	14	22	18	19	17	17	19	23	27	20	140	145	151	35	7
0	14	87	15	14	20	18	19	15	13	16	21	26	18	136	142	151	32	7
	0	87	18	15	22	19	19	17	17	19	23	27	20	142	145	149	35	7
		0	76	89	86	87	88	89	92	77	88	95	90	152	152	162	90	82
			0	18	14	21	23	21	19	1	17	29	22	139	142	151	26	11
				0	21	18	14	19	15	19	22	25	22	139	142	149	29	9
					0	24	25	25	23	15	4	30	24	144	145	154	30	15
						0	25	21	19	22	25	26	25	144	146	156	33	13
							0	23	20	24	26	27	23	137	139	146	36	14
								0	20	22	26	30	21	140	144	150	37	10
									0	20	24	27	23	139	142	151	33	10
										0	18	30	23	140	143	152	27	12
											0	31	25	245	146	155	33	16
												0	32	141	145	156	43	21
													0	143	146	152	37	15
														0	11	146	152	140
															0	145	150	143
																0	155	149
																	0	28
																		0

by antigens, etc. On the other hand, the origins of these sequences are well documented. Using this information, the contributions of genetic mechanisms to the observed sequence differences might be deciphered. Figure 1 illustrates a plot of occurrences against the number of differences between mouse lambda light chain sequences. Figure 1a shows all differences while Figure 1b only differences up to 40.

The peaks around 143 and 150 differences in Figure 1a are due to mouse sequence Y31 (SANCHER *et al.* 1990), a distinct lambda light chain variable region gene, and sequences CZ81 and SD 26 (REIDL *et al.* 1992) from wild mice, which show more resemblance to human lambda light chain sequences. The peak around 87 differences (Figure 1a) appears as a result of including a synthetic MOPC315 sequence (BALDWIN and SCHULTZ 1989) that is drastically different from other

sequences. The peaks in the region of <40 differences are more clearly shown in Figure 1b.

The peak around three differences (Figure 1b) is most likely due to somatic mutations induced by antigens or by other mechanisms, *e.g.*, radiation, etc. This peak is quite distinct, covering the range of one to five differences. It overlaps with the next peak around 10 differences. As shown in Table 1, this may be the result of sequences from different strains of mice, *i.e.*, allelic variations. Other peaks are present around 15 and 19 differences, and less distinct ones around 22, 27 and 30 differences. Since the mouse lambda 2 gene has been identified as E3-19 and it has two different sequences most likely from different strains or sources (MOTOYAMA *et al.* 1991, 1994), the peak around 19 differences can be due to gene duplication or multiplication. The peak around 15 differences may be the com-

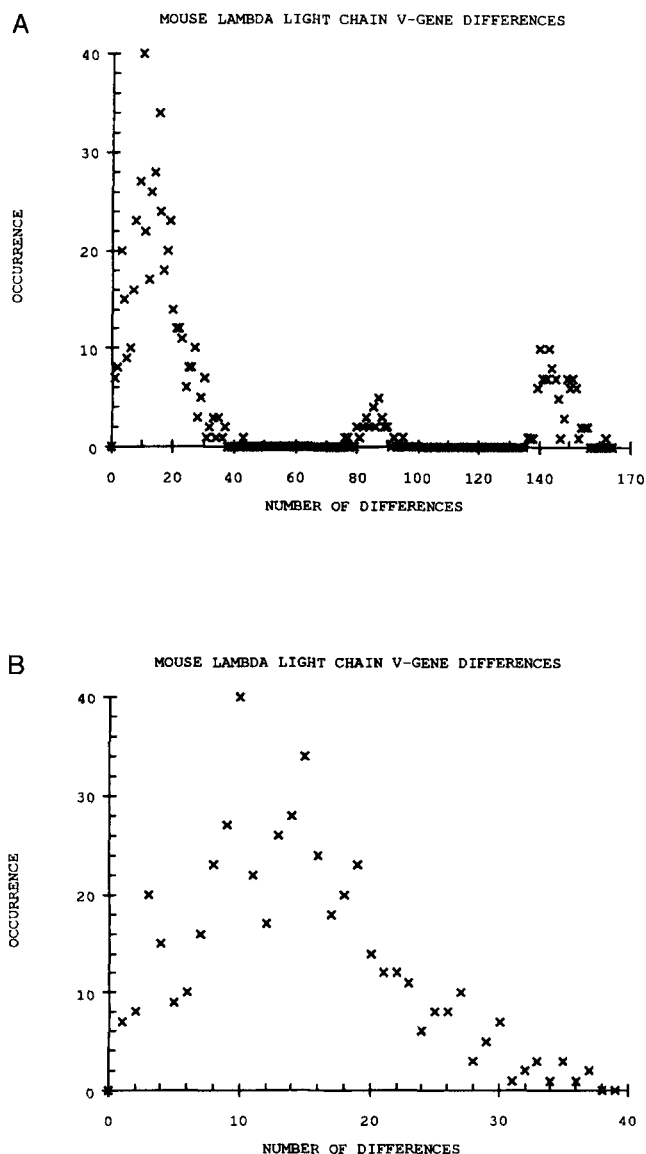


FIGURE 1.—Plot of occurrences against number of nucleotide differences listed in Table 1 for all differences (a) and for differences up to 40 (b).

bined effect of allelic variations and somatic mutations. The remaining peaks around 22, 27 and 30 differences are probably due to the various combinations of these genetic mechanisms.

Since the number of subgroups is related to gene multiplication, we are therefore able to separate its effect on nucleotide differences by constructing a second triangular table from the first one by eliminating all columns and rows containing differences between one and 17, *i.e.*, the valley between the peaks around 15 and 19 differences. This second table (Table 2) contained only six sequences. One representing the mouse lambda 1 gene, two for the two different sequences of mouse lambda 2 gene, one for the third mouse lambda gene, one from a wild mouse, and one for the synthetic MOPC315 sequence.

TABLE 2

Derived from Table 1 by eliminating all columns and rows containing one to 17 differences

	1	2	3	4	5	6
	007	007	007	007	007	019
	665	710	725	746	750	435
1. 007665	0	83	20	139	149	27
2. 007710		0	95	152	162	90
3. 007725			0	141	156	43
4. 007746				0	146	152
5. 007750					0	155
6. 019435						0

This simple result can then be applied to human lambda light chain sequences. A triangular table was constructed with all available complete distinct sequences for pairwise comparison. This table contains 151 sequences, which is too large to be included in this paper. The KADBIDs of these sequences are listed in the APPENDIX at the end of this paper. Unlike the mouse sequences (Table 1), the distribution of nucleotide differences is much more complicated, probably due to the large number of allelic variations and extensive gene multiplication. Although the cutoff value of 17 differences for mouse sequences may not be applicable to human sequences, it can nevertheless provide a rough estimation. Thus two additional tables are generated for human lambda light chains by eliminating all columns and rows containing one to 17 differences (Table 3) and one to 34 (*i.e.*, two times 17) differences (Table 4). These tables should give a range to the number of different human lambda light chain genes. Table 3 lists 58 sequences, while Table 4 contains 29 sequences.

Therefore, if we equate the number of subgroups to the number of distinct genes, human lambda light chains have at least 29 subgroups (Table 4), while the upper limit may be >58 (Table 3). This estimation is larger than most of previous studies (CH'ANG *et al.* 1995).

Recent experimental studies by direct nucleotide sequencing of the human lambda light chain gene region indicate that there are at least 52 genes (FRIPPIAT *et al.* 1995), which is in excellent agreement with our theoretical estimation.

The number of available nucleotide sequences for human and mouse, heavy and kappa light chains are much larger (JOHNSON *et al.* 1996). However, this simple method can also be used to estimate their gene numbers. Unfortunately, the triangular tables and KADBIDs listings are all too large to be included in this paper. Listings of currently available sequences can be obtained from the database mentioned in the MATERIALS AND METHODS section, and triangular table generated accordingly.

The results are listed in Table 5. The lower limits are based on a cutoff of 34 nucleotide differences, and the upper limits a cutoff of 17 nucleotide differences. For human heavy chains, we estimate that there are ~116–175 different genes. Experimental results of HONJO's group (MATSUMURA *et al.* 1994) indicate that there are >90 different human heavy chain V-genes. About one-third of them are pseudogenes. These pseudogenes can, however, be functional through the mechanism of gene conversion (BALTIMORE 1991) as in the case of chicken immunoglobulin system (REYNAUD *et al.* 1985, 1989). Thus, our estimation is only slightly higher than what has so far been found experimentally for human heavy chain V-genes.

For human kappa light chains, we estimated the gene number to be around 27–71, while experimentally this number is around 76 (SCHABLE *et al.* 1994). For mouse kappa light chains, our estimation is around 76–134, which is not an order of magnitude larger than the number of human kappa light chains as generally believed. Interestingly, the total number of human kappa and lambda light chain genes is, in our estimation, ~56–128; while that of mouse is around 79–139. Therefore, human and mouse appear to have the similar number of light chain V-genes.

For mouse heavy chains, our theoretical analysis gives around 99–199 genes, *i.e.*, again about the similar number as human heavy chain genes.

DISCUSSION

Subgroups (MILSTEIN 1967), sub-subgroups, families, etc. are used to classify variable regions sequences of human and mouse heavy and light chains, so that one can roughly estimate the numbers of V-genes coding for each of these chains. Such information can then provide some insight to the basic mechanisms of generating antibody diversity.

For human heavy chain variable region genes, HONJO's group (MATSUMURA *et al.* 1994) has sequenced the entire gene segment and concluded that there are over 90 different genes with one-third of these being pseudogenes. For human kappa light chain genes, ZACHAU's group (SCHABLE *et al.* 1994) has determined that there are ~76 different genes. Recently, the human lambda light chain gene segment has also been sequenced, and there are ~52 different genes (FRIPPIAT *et al.* 1995). Ideally, similar studies can soon be carried out experimentally for mouse heavy and kappa light immunoglobulin variable region genes as well as other multiple gene loci, *e.g.*, T cell receptors for antigens, MHC class I and II molecules, etc. However, in the absence of such studies, theoretical estimations of the numbers of these genes will be very valuable.

For mouse lambda light chains (SANCHEZ *et al.* 1987), the number of genes is very small, usually assumed to

be three. In our present study, we have developed a method of analyzing available mouse lambda light chain nucleotide sequences by pairwise comparison (Table 1). The occurrences of various nucleotide differences can be used (Figure 1) to separate three possible genetic mechanisms of generating such differences, namely, gene multiplication, allelic variations, and somatic mutations due to antigen stimulation or due to other mechanisms. These effects can also be present simultaneously.

From the plots shown in Figure 1, there are several distinct peaks of nucleotide differences. Together with known origins of the mouse sequences, it is possible to assign these peaks to the effects of various genetic mechanisms. The peak around 19 differences seems to be due to gene multiplication. Thus, to estimate the number of different genes, we may use a cutoff of ~17 nucleotide differences. On the conservative side, we may use a cutoff value twice as large, *i.e.*, 34 nucleotide differences.

Our method assumes that human and mouse are under similar selective pressure so far antibody V-genes are concerned. We may underestimate the total number of immunoglobulin V-genes if after duplication, the two genes do not diverge. For example, in the case of human kappa light chains, some of the V-genes are transposed to other chromosomes and thus under different selective pressure (SCHABLE *et al.* 1994). However, in general, this is unlikely since new V-genes obtained by divergence can give a selective advantage for the survival of the organism.

The very good agreements between our theoretical estimations and experimental sequencing of V-genes for human heavy, kappa and lambda light chains, based on mouse lambda light chain V-gene numbers, suggest that evolutionary dynamics of these genetic loci in human and mouse are reasonably similar. However, when more sequences become available, our theoretical numbers are expected to increase.

Our method of estimating the number of different genes has also been applied to mouse kappa light and mouse heavy chain V-genes. The results are shown in Table 5. Roughly speaking, human and mouse have similar numbers of immunoglobulin heavy chain V-genes. The sum of their kappa and lambda light chain genes are also similar. The smaller number of mouse lambda light chain V-genes seems to be compensated by the larger number of kappa light chain V-genes. Recent experimental studies (KIRSCHBAUM *et al.* 1996) suggest that there are ~140 mouse kappa light chain V-genes, also in very good agreement with our estimation of 76–134.

This simple method of gene number estimation can easily be applied to T cell receptor for antigen alpha and beta chain genes, MHC class I and II molecules,

TABLE 5

Estimated numbers of human and mouse immunoglobulin V-genes, based on these of mouse lambda light chain V-genes

	Mouse	Human
Lambda light chain	3-5	29-58
Kappa light chain	76-134	27-71
Heavy chain	99-199	116-175

and other proteins from multi-gene families. We are currently analyzing these sequences.

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APPENDIX

The 151 distinct complete human lambda light chain V-gene sequences, positions 1-95, are (in KADBID) as follows:

007406; 007407; 007410; 007413; 007417; 007421;
 007422; 007424; 007425; 007426; 007427; 007428;
 007430; 007433; 007434; 007438; 007440; 007441;
 007444; 007447; 007449; 007450; 007453; 007459;
 007460; 007461; 007462; 007463; 007464; 007468;
 007471; 007473; 007474; 007480; 007484; 007488;
 007489; 007492; 007493; 007494; 007495; 007498;
 007502; 007505; 007508; 007509; 007510; 007511;
 007512; 007513; 007514; 007515; 007516; 007517;
 007518; 007519; 007520; 007522; 007523; 007526;
 007527; 007533; 007534; 007535; 007536; 007537;
 007538; 007539; 007540; 007542; 007544; 007546;
 007550; 007552; 007555; 007557; 007560; 007562;
 007568; 007572; 007574; 007575; 007576; 007678;
 007579; 007580; 007581; 019745; 019767; 020079;
 020080; 020081; 020098; 020102; 020104; 010205;
 020112; 020136; 020138; 020522; 020623; 020624;
 020625; 020634; 020640; 020642; 020644; 020646;
 020650; 021310; 021311; 021312; 021455; 021741;
 021742; 021744; 021745; 021746; 021747; 021748;
 021749; 021750; 021751; 021752; 021753; 021754;
 021755; 021756; 021757; 021758; 021759; 021760;
 021761; 021762; 021763; 021764; 021765; 021766;
 021767; 021768; 021769; 021770; 021771; 022264;
 022265; 022266; 022682; 023194; 023195; 026955;
 026956.