

# A REVISED MAP OF THE EIGHT LINKAGE GROUPS OF *ASPERGILLUS NIDULANS*<sup>1</sup>

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THE homothallic ascomycete *Aspergillus nidulans* has become an important organism for genetic studies as a consequence of the basic research of PROFESSOR G. PONTECORVO and his associates. A preliminary linkage map was constructed nine years ago by KÄFER (1958). A revised genetic map is presented in the present paper, which complements the listing of *Aspergillus* stocks published by BARRATT, JOHNSON and OGATA (1965).

In 1966, *Aspergillus* workers were requested to send pertinent linkage data to this laboratory. Tables 1, 2 and 3 and Figure 1 were compiled from this information and that published in the literature; they are intended to serve as a reference for individuals who wish to use *A. nidulans* for research or teaching.

*Recommendations:* Hopefully, this paper will encourage the isolation and mapping of new genetic markers; a representative mutant of each new locus should be deposited in the Fungal Genetics Stock Center<sup>2</sup>. Furthermore, it is recommended that the system of genetic nomenclature for bacteria (DEMEREK, ADELBERG, CLARK and HARTMAN 1966) be employed in the assignment of symbols to new *Aspergillus* mutants. Recently, it has been shown (KÄFER 1965) that several of the genetic strains of *Aspergillus nidulans* contain one or more translocations. In crosses such strains produce a high frequency of inviable ascospores and give spurious linkage relations. In order to prevent the introduction of additional translocations into the stock strains, new mutants should so far as is possible be tested by mitotic analysis (PONTECORVO and KÄFER 1958; McCULLY and FORBES 1965) with translocation-free tester strains (available at the Fungal Genetics Stock Center [see Table 5 of BARRATT *et al* 1965]).

Over 140 different loci are listed in Table 1, which includes the genetic symbol, name and description of each mutant, together with the linkage group, list of allelic mutants, availability from the Fungal Genetics Stock Center, and origin of each mutant. The reference cited for each mutant does not necessarily refer to the person who isolated it; in several instances more than one investigator was involved in the isolation, characterization and mapping of a given mutant. For proper credit to the originator refer to the cited reference. The probable location of the centromeres is indicated in Figure 1 by a solid circle (●). Mutants placed

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to the right of a linkage group indicate that they have been assigned to that group by mitotic analysis but have not been further mapped.

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#### SUMMARY

One hundred forty-four mutant loci of *Aspergillus nidulans* are described, and a revised genetic map is presented.

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TABLE 1

*Located mutants of Aspergillus nidulans*

Symbol	Designation	Phenotype	Linkage Group	Allelic Mutants†	Available From FGSC††	Origin of Mutant		
						Strain	Mode	References
<u>VISIBLE</u>								
<u>bl1</u>	blue 1	blue ascospores	II	2,3,4	yes	<u>y</u> ; <u>w2</u> ; <u>s12</u>	nitrous acid	Apirion, 1963
<u>cha</u>	chartreuse	yellowish green conidia	VIII R	-	yes	-	spontaneous	" Kafer, 1961
<u>cl4</u>	colorless 4	colorless ascospores	IV	-	no	<u>y</u> ; <u>w2</u> ; <u>s12</u>	nitrous acid	} Apirion, 1963
<u>cl6</u>	colorless 6	colorless ascospores	I L	1,3,5	no	<u>y</u> ; <u>w2</u> ; <u>s12</u>	nitrous acid	
<u>co</u>	compact	compact colony	VIII R	-	yes	<u>paba1</u> <u>bi1</u>	spontaneous	Pontecorvo et al, 1953
<u>dil*</u>	dilute	dilute conidial color	III	-	yes	-	U.V.	G. Jansen, unpublished
<u>fw</u>	fawn	yellowish brown conidia	VIII L	-	yes	<u>bi1</u>	spontaneous	Clutterbuck, 1965
<u>mo96*</u>	compact 96	compact colony with grooves	III	-	yes	<u>y</u> ; <u>ad3</u> ; <u>s1</u>	β-propiolactone	Bainbridge, 1963
<u>p1*</u>	pale 1	light grayish green conidia	V	-	no	<u>an1</u> <u>bi1</u>	spontaneous	Van Arkel, 1962
<u>s1*</u>	slow	reduced growth rate	III	-	yes	<u>ad14</u> <u>bi1</u>	spontaneous	" Kafer, 1965
<u>sm</u>	small	colony compact	III L	-	yes	<u>bi1</u> ; <u>lys5</u>	spontaneous	Bainbridge, unpublished
<u>ve</u>	velvet	fluffy appearance	VIII L	-	yes	<u>bi1</u>	spontaneous	" Kafer, 1958
<u>w1</u>	white 1	colorless conidia, epistatic to <u>y</u>	II L	-	no	wild type	spontaneous	" Kafer, 1965
<u>y</u>	yellow	yellow conidia	I R	-	yes	<u>y</u>	x-rays	} Pontecorvo et al, 1953
<u>yg*</u>	yellow-green	yellow conidia and green conidia on complete; yellow conidia on minimal	II R	-	yes	<u>paba1</u> <u>bi1</u>	spontaneous	
<u>yg*</u>	yellow-green	yellow conidia and green conidia on complete; yellow conidia on minimal	II R	-	yes	<u>bi1</u>	U.V.	Dorn & Rivera, unpublished
<u>RESISTANT</u>								
		<u>Resistant to</u>						
<u>acr1</u>	Acriflavine 1	Acriflavine/malachite green dominant	II L	-	yes	<u>paba1</u> <u>y</u> ; <u>co</u>	Spontaneous	} Roper & Kafer, 1957
<u>acr2</u>	acriflavine 2	acriflavine/malachite green	II R	-	no	<u>bi1</u> ; <u>ad1</u> ; <u>s12</u> ; <u>pyro4</u>	Spontaneous	
<u>act1*</u>	Actidione 1	Actidione, dominant	III L	-	yes	<u>ribo1</u> <u>y</u> ; <u>nic8</u>	U.V.	Warr & Roper, 1965
<u>facA</u>	fluoroacetate A	fluoroacetate; acetate non-utilizer	V	303	yes	<u>bi1</u>	spontaneous	} Apirion, 1965
<u>facB</u>	fluoroacetate B	fluoroacetate; acetate non-utilizer	VIII R	101	yes	-	spontaneous	
				302,308,309	no	-	spontaneous	

TABLE 1—Continued

Symbol	Designation	Phenotype	Linkage Group	Allelic Mutants†	Available From FGSC††	Origin of Mutant		Reference
						Strain	Mode	
<u>RESISTANT</u>		<u>Resistant to</u>						
<u>facC</u>	fluoroacetate C	fluoroacetate; acetate non-utilizer	VIII R	102	yes	<u>w3</u> ; <u>pyro4</u>	spontaneous	} Apirion, 1965
<u>fanA</u>	fluoroacetate A	fluoroacetate	V	1,3,4,10,11,12,14,16	no	-	spontaneous	
<u>fanB</u>	fluoroacetate B	fluoroacetate	VII	52,54,60	no	-	spontaneous	
<u>fanC</u>	fluoroacetate C	fluoroacetate	VI	101	no	-	spontaneous	
<u>fanD</u>	fluoroacetate D	fluoroacetate	VIII	151	no	-	spontaneous	
<u>fanE</u>	fluoroacetate E	fluoroacetate	VI	5,7	no	-	spontaneous	
<u>fpA</u>	fluorophenyl-alanine A	fluorophenyl-alanine	I L	1	yes	<u>bi1</u>	spontaneous	} McCully, unpublished
<u>fpB</u>	fluorophenyl-alanine B	fluorophenyl-alanine	I	37	yes	<u>ribo1</u> <u>bi1</u>	spontaneous	
<u>fpC</u>	fluorophenyl-alanine C	fluorophenyl-alanine	VIII	43	no	<u>ribo1</u> <u>bi1</u>	spontaneous	
<u>fpD</u>	Fluorophenyl-alanine D	Fluorophenyl-alanine, dominant	III	11	yes	<u>ribo1</u> <u>bi1</u>	Spontaneous	
<u>Iod1*</u>	Iodoacetate 1	Iodoacetate, dominant	II L	-	yes	-	Spontaneous	Warr & Roper, 1965
<u>Sul1*</u>	Sulfanilamide 1	Sulfanilamide, dominant	I L	-	yes	-	U.V.	G. Jansen, unpublished
<u>teo6*</u>	teoquil 6	teoquil	III	-	no	-	spontaneous	Warr & Roper, 1965
<u>NUTRITIONAL</u>		<u>Growth response to</u>						
<u>adaA*</u>	adenine A	adenine	I L	AM74,AM3	yes	wild type	U.V.	} Foley et al, 1965
<u>adaB*</u>	adenine B	adenine	VIII R	AM57	yes	wild type	U.V.	
<u>ad1</u>	adenine 1	adenine/hypoxanthine	II R	-	yes	<u>y</u>	x-rays	} Pontecorvo et al, 1953 & Foley et al, 1965
<u>ad3</u>	adenine 3	adenine/hypoxanthine	II R	-	yes	<u>y</u> ; <u>thi1</u>	x-rays	
<u>ad8</u>	adenine 8	adenine/hypoxanthine	I R	10,11,12,16,19,20,21,22	yes	<u>bi1</u>	U.V.	Pritchard, 1955 & Foley et al, 1965
<u>ad9</u>	adenine 9	adenine/hypoxanthine	I R	13,17	no	<u>bi1</u>	U.V.	} Pritchard, unpublished, Calef, 1957 & Foley et al, 1965
				15	yes	<u>bi1</u>	U.V.	
				32	no	<u>bi1</u> ; <u>w3</u>	U.V.	
<u>ad14</u>	adenine 14	adenine/hypoxanthine	I L	-	yes	<u>bi1</u>	U.V.	} Pontecorvo & Kafer, 1958 & Foley et al, 1965
				18	no	<u>bi1</u>	U.V.	
<u>ad23</u>	adenine 23	adenine/hypoxanthine	II L	-	yes	<u>bi1</u>	U.V.	"Kafer, 1958 & Foley et al, 1965
<u>ad50*</u>	adenine 50	adenine/hypoxanthine	III L	-	no	<u>bi1</u>	U.V.	Dorn & Rivera, 1965
<u>sax*</u>	allantoic acid X	urea/ammonium	VI	1	yes	<u>bi1</u>	diethyl sulfate	Darlington & Scazzocchio, unpublished
<u>abi</u>	aminobutyric acid 1	aminobutyric acid	II R	-	yes	<u>bi1</u>	U.V.	Forbes, 1959
<u>an1</u>	aneurin 1	thiamine (aneurin)	I L	-	yes	<u>bi1</u>	U.V.	Pontecorvo & Kafer, 1958
<u>an2</u>	aneurin 2	thiamine (aneurin)	II R	-	yes	<u>bi1</u> ; <u>Acr1</u> <u>w3</u>	U.V.	"Pontecorvo & Kafer, unpublished

TABLE 1—Continued  
 Located mutants of *Aspergillus nidulans*

Symbol	Designation	Phenotype	Linkage Group	Allelic Mutants†	Available From FGSC††	Origin of Mutant		
						Strain	Mode	References
<u>NUTRITIONAL</u>								
		<u>Growth response to</u>						
<u>arg1</u> *	arginine 1 (argino-succinase)	arginine	VI	-	yes	<u>bi1</u>	x-rays	Bainbridge, Dalton & Walpole, 1966
<u>arg2</u>	arginine 2	arginine	III L	-	yes	<u>bi1</u>	U.V.	} Forbes, 1959
<u>arg3</u>	arginine 3	arginine	VIII R	-	yes	<u>bi1</u>	U.V.	
<u>bi1</u>	biotin 1	biotin	I R	-	yes	wild type	x-rays	} Roper, 1950
				2,3	no	<u>y; thi1</u>	x-rays	
<u>cho</u>	choline	choline	VII	-	yes	<u>bi1</u>	U.V.	"Kafer, 1958
<u>hisA</u> *	histidine A	histidine	IV R	10	yes	-	-	} M. Berlyn, 1966
<u>hisB</u> *	histidine B (IGF dehydrase)	histidine	I	-	no	-	-	
<u>hisC</u> *	histidine C (IAP transaminase)	histidine	VIII R	38	yes	-	-	
<u>hisD</u> *	histidine D (histidine dehydrogenase)	histidine	VIII R	-	no	-	-	
<u>hisE</u> *	histidine E (PR-ATP pyrophosphohydrolase)	histidine	VIII R	-	no	-	-	
<u>hisF</u> *	histidine F	histidine	VII	-	no	-	-	
<u>hisG</u> *	histidine G (PR-ATP pyrophosphorylase)	histidine	II	-	no	-	-	
<u>hisH</u> *	histidine H	histidine	VIII	13	yes	-	-	
<u>hisI</u> *	histidine I	histidine	VIII R	-	no	-	-	
<u>his122</u> *	histidine 122	histidine	VII	-	no	-	U.V.	
<u>hxA</u> *	hypoxanthine A	uric acid/allantoin/allantoate/urea/ammonium	V	1	yes	<u>bi1</u>	diethyl sulfate	} Darlington & Scazzocchio, unpublished
<u>hxB</u> *	hypoxanthine B	uric acid/allantoin/allantoate/urea/ammonium	VII	13	no	<u>bi1</u>	diethyl sulfate	
<u>ile1</u> *	isoleucine 1	isoleucine	VII	-	no	-	U.V.	Pees, 1966
<u>lu</u>	leucine	leucine	I L	-	yes	<u>bi1</u>	U.V.	Forbes, 1959
<u>lys1</u>	lysine 1	lysine	VI	2	yes	<u>w1</u>	x-rays	Pontecorvo et al, 1953
				8	no	-	U.V.	Pees, 1966
<u>lys5</u>	lysine 5	lysine	V	-	yes	<u>bi1</u>	U.V.	"Kafer, 1958
				16	no	-	U.V.	} Pees, 1966 & 1965
<u>lys7</u> *	lysine 7	lysine	VII	6	no	-	U.V.	
<u>lys10</u> *	lysine 10	lysine	V	-	no	-	U.V.	
<u>lys51</u> *	lysine 51	lysine	I R	-	no	-	U.V.	
<u>meth1</u>	methionine 1	methionine	IV L	-	yes	<u>bi1</u>	U.V.	
<u>meth2</u>	methionine 2	methionine	III L	-	yes	<u>bi1</u>	U.V.	Forbes, 1959
<u>meth3</u>	methionine 3	methionine	V	-	yes	<u>y; pyro4</u>	U.V.	Roberts, unpublished
<u>ni3</u>	nitrite 3	nitrite	II R	-	yes	<u>bi1; w3</u>	U.V.	"Kafer, 1958
<u>ni50</u> *	nitrite 50	nitrite/proline/arginine/ammonium	VIII R	-	yes	<u>bi1</u>	U.V.	} Dorn & Rivera, 1965
<u>ni51</u> *	nitrite 51	proline/arginine/ammonium	VIII R	-	yes	<u>bi1; phen3</u>	spontaneous	
				52	no	<u>bi1; phen3</u>	spontaneous	

TABLE 1—Continued

Symbol	Designation	Phenotype	Linkage Group	Allelic Mutants†	Available From FGSC††	Origin of Mutant		References
						Strain	Mode	
<u>NUTRITIONAL</u>		<u>Growth response to</u>						
<u>nic2</u>	nicotinic 2	nicotinic acid/ anthranilic acid	V	-	yes	wild type	x-rays	} Pontecorvo et al, 1953 & Kafer, 1958
<u>nic8</u>	nicotinic 8	nicotinic acid/ anthranilic acid/ tryptophan	VII	-	yes	<u>bi1</u>	U.V.	
<u>nic10</u>	nicotinic 10	nicotinic acid/ anthranilic acid	VI	-	yes	<u>bi1</u> ; <u>Acr1</u> <u>w3</u>	U.V.	" Kafer, 1958
<u>orn4</u>	ornithine 4	ornithine/arginine	IV R	-	yes	<u>bi1</u>	U.V.	} Pontecorvo et al, 1953 & Kafer, 1958
<u>orn7</u>	ornithine 7	ornithine/arginine	VIII R	8	no	<u>bi1</u>	U.V.	
				9	yes	<u>bi1</u>	U.V.	} Forbes, 1959
				20	yes	<u>bi1</u>	U.V.	
								Dorn & Rivera, 1965
<u>paba1</u>	p-aminobenzoic acid 1	p-aminobenzoic acid	I R	-	yes	<u>bi1</u>	x-rays	} Roper, 1953, Siddiqi, 1962 & Putrament, 1964
				2,3	no	<u>bi1</u>	U.V.	
				4,5	no	<u>bi1</u>	x-rays	
				6	yes	<u>bi1</u>	U.V.	
<u>paba22</u>	p-aminobenzoic acid 22	p-aminobenzoic acid	IV R	-	yes	<u>bi1</u>	U.V.	} Siddiqi, unpublished
				21	no	<u>bi1</u>	U.V.	
<u>panto</u>	pantothenic acid	pantothenic acid	III R	-	no	<u>y</u> ; <u>thi1</u>	x-rays	Pontecorvo et al, 1953
<u>phen2</u>	phenylalanine 2	phenylalanine/ phenylpyruvic acid	III L	-	yes	<u>bi1</u>	U.V.	" Kafer, 1958
				3	yes	-	-	Pontecorvo, unpublished
<u>pro1</u>	proline 1	arginine/proline	I R	2,5	yes	<u>bi1</u>	U.V.	} Forbes, 1956
				6,7,8	no	<u>bi1</u>	U.V.	
<u>pro3</u>	proline 3	arginine/proline	I R	4	no	<u>bi1</u>	U.V.	
<u>pu</u>	putrescine	putrescine/spermidine	II R	-	yes	<u>bi1</u> ; <u>w3</u>	U.V.	Sneath, 1955
<u>pyro4</u>	pyridoxine 4	pyridoxine	IV R	-	yes	<u>bi1</u>	U.V.	} " Kafer, 1958
				1,2	no	<u>bi1</u>	x-rays	
				5	no	<u>bi1</u>	U.V.	
				6,7,8	no	<u>bi1</u>	U.V.	
				9,10	no	<u>bi1</u> ; <u>Acr1</u> <u>w3</u>	U.V.	} C. Martin-Smith, unpublished
					yes	<u>bi1</u>	U.V.	
<u>ribo1</u>	riboflavin 1	riboflavin	I L	-	yes	<u>bi1</u>	U.V.	Pontecorvo & Kafer, 1958
<u>ribo2</u>	riboflavin 2	riboflavin	VIII R	-	yes	<u>bi1</u> ; <u>Acr1</u> <u>w3</u>	U.V.	" Kafer, 1958
				4	no	<u>bi1</u> ; <u>Acr1</u> <u>w3</u>	U.V.	} Forbes & Sundaram, unpublished
<u>ribo5</u>	riboflavin 5	riboflavin	V	-	yes	<u>y</u> ; <u>pyro4</u>	U.V.	
<u>ribo6</u>	riboflavin 6	riboflavin	II R	-	yes	<u>bi1</u> ; <u>w3</u>	U.V.	
<u>s1</u>	sulfite 1	sulfite	III R	4	yes	<u>bi1</u>	U.V.	} " Kafer, 1958
				2	no	<u>bi1</u>	U.V.	
				5,6,8,9	no	<u>bi1</u> ; <u>w3</u>	U.V.	
				-	yes	<u>bi1</u>	U.V.	
<u>s3</u>	sulfite 3	sulfite	VI	-	yes	<u>bi1</u>	U.V.	} " Kafer, 1958
				10,11	no	<u>bi1</u> ; <u>w3</u>	U.V.	
<u>s12</u> ( <u>w30</u> )	sulfite 12	sulfite	III R	-	yes	wild type	nitrogen mustard	} Pontecorvo et al, 1953
				7	no	<u>bi1</u> ; <u>w3</u>	U.V.	
<u>s50*</u>	thiosulfate 50	cysteic acid/ cysteine/thiosulfate/ methionine	VIII R	-	yes	<u>bi1</u>	U.V.	Dorn & Rivera, 1965

TABLE 1—Continued  
*Located mutants of Aspergillus nidulans*

Symbol	Designation	Phenotype	Linkage Group	Allelic Mutants†	Available From FGSC††	Origin of Mutant		Reference
						Strain	Mode	
<u>NUTRITIONAL</u> Growth response to								
<u>thi4</u>	thiazole	aneurin/4-methyl-5-hydroxyethyl-thiazole	II R	-	yes	<u>bi1</u>	U.V.	} "Kafer, 1958
				1	no	<u>y</u>	x-rays	
<u>trypA*</u>	tryptophan A (anth. synthetase)	tryptophan/indole/anthranilic acid	II	69	yes	<u>paba1 y</u>	U.V.	} Roberts, 1967 & Hutter & Demoss, 1967
				10 alleles	no	<u>paba1 y</u>	U.V.	
				21 alleles	no	<u>bi1; Acr1 w3; nic8</u>	U.V.	
<u>trypB*</u>	tryptophan B (tryp. synthetase)	tryptophan	I	403	yes	<u>paba1 y</u>	U.V.	
				18 alleles	no	<u>paba1 y</u>	U.V.	
				19 alleles	no	<u>bi1; Acr1 w3; nic8</u>	U.V.	
<u>trypC*</u>	tryptophan C (PRA isomerase InGP synthetase anth. synthetase)	tryptophan/indole	VIII R	801	yes	<u>paba1 y</u>	U.V.	
				12 alleles	no	<u>paba1 y</u>	U.V.	
				46 alleles	no	<u>bi1; Acr1 w3; nic8</u>	U.V.	
<u>trypD*</u>	tryptophan D (PR transferase)	tryptophan/indole	II	432	yes	<u>paba1 y</u>	U.V.	
				2 alleles	no	<u>paba1 y</u>	U.V.	
				16 alleles	no	<u>bi1; Acr1 w3; nic8</u>	U.V.	
<u>uY*</u>	urea Y	ammonium	VII	5	yes	<u>bi1</u>	diethyl sulfate	} Darlington & Scazzocchio, unpublished
<u>uaX*</u>	uric acid X	allantoin/allantoate/urea/ammonium	VI	1	yes	<u>bi1</u>	diethyl sulfate	
				10	no	<u>bi1</u>	diethyl sulfate	
<u>CARBON SOURCE</u> Fails to grow on								
<u>fr1</u>	fructose 1	fructose	IV R	-	yes	<u>y; pyro4</u>	U.V.	} Roberts, 1963
<u>gal1</u>	galactose 1 (not inducible kinase or transferase)	galactose	III L	6,15,17,23,24,27,29,30,32,35,36	yes	<u>bi1; w3</u>	U.V.	
<u>gal3</u>	galactose 3	galactose	II	-	yes	<u>bi1; w3</u>	U.V.	} Roberts, 1963 & 1964
<u>gal4</u>	galactose 4	galactose	VIII	-	no	<u>bi1; w3</u>	U.V.	
				7	yes	<u>bi1; w3</u>	U.V.	
<u>gal5</u>	galactose 5 (transferase)	galactose	I L	8,10,13,14,19,33	yes	<u>bi1; w3</u>	U.V.	
<u>gal9</u>	galactose 9 (kinase)	galactose	III L	-	yes	<u>bi1; w3</u>	U.V.	
<u>lac1</u>	lactose 1	lactose	VI	2,4	yes	<u>y; pyro4</u>	U.V.	} Roberts, 1963
				6,7	no	<u>bi1; w3</u>	U.V.	
<u>lac3</u>	lactose 3	lactose	II R	-	no	<u>bi1; w3</u>	U.V.	
<u>mal1</u>	maltose 1	maltose	VII	-	yes	<u>y; pyro4</u>	U.V.	
				2	no	<u>bi1; w3</u>	U.V.	
<u>sb3</u>	sorbitol 3	sorbitol	VI	-	yes	<u>bi1; w3</u>	U.V.	



TABLE 1—Continued

Symbol	Designation	Phenotype	Linkage Group	Allelic Mutants†	Available From FGSC††	Origin of Mutant		
						Strain	Mode	Reference
<u>MISCELLANEOUS</u>								
<u>Reduced ability to cleave</u>								
<u>pacA</u>	acid phosphatase A	$\alpha$ -naphthylphosphate at pH 4.8	IV	1	yes	<u>bi1</u> <u>rA1</u>	U.V.	} Dorn, 1965a
<u>pacB</u>	acid phosphatase B	$\alpha$ -naphthylphosphate at pH 4.8	VIII R	4	no	<u>bi1</u>	U.V.	
<u>pacC</u>	acid phosphatase C	$\alpha$ -naphthylphosphate at pH 4.8	VI	5	yes	<u>bi1</u>	U.V.	
<u>palA</u>	alkaline phosphatase A	$\alpha$ -naphthylphosphate at pH 8.2	III L	1	yes	<u>bi1</u> <u>rA1</u>	U.V.	
<u>palB</u>	alkaline phosphatase B	$\alpha$ -naphthylphosphate at pH 8.2	VIII R	7	yes	<u>bi1</u> <u>rA1</u>	U.V.	
<u>palC</u>	alkaline phosphatase C	$\alpha$ -naphthylphosphate at pH 8.2	IV R	4	yes	<u>bi1</u> <u>rA1</u>	U.V.	
<u>palD</u>	alkaline phosphatase D	$\alpha$ -naphthylphosphate at pH 8.2	VII	8	yes	<u>bi1</u> <u>rA1</u>	U.V.	
<u>palE</u>	alkaline phosphatase E	$\alpha$ -naphthylphosphate at pH 8.2	VIII R	11	yes	<u>bi1</u> <u>rA1</u>	U.V.	
<u>palF</u>	alkaline phosphatase F	$\alpha$ -naphthylphosphate at pH 8.2	VII	15	yes	<u>bi1</u> <u>rA1</u>	U.V.	
<u>palcA</u>	alkaline-acid phosphatase A	$\alpha$ -naphthylphosphate at pH 4.8 & 8.2	II R	1	yes	<u>bi1</u> <u>rA1</u>	U.V.	
<u>palcB</u>	alkaline-acid phosphatase B	$\alpha$ -naphthylphosphate at pH 4.8 & 8.2	III L	3	no	<u>bi1</u>	U.V.	
<u>palcC</u>	alkaline-acid phosphatase C	$\alpha$ -naphthylphosphate at pH 4.8 & 8.2	VIII R	4	no	<u>bi1</u>	U.V.	
<u>Enhanced ability to cleave</u>								
<u>rA</u>	phosphatase enhanced	$\alpha$ -naphthylphosphate at pH 8.2	I L	1,2,3	no	<u>bi1</u>	U.V.	Dorn, 1965a
<u>rB*</u>	phosphatase enhanced	$\alpha$ -naphthylphosphate at pH 4.8	II R	50	no	<u>bi1</u>	U.V.	} Dorn & Rivera, 1965
<u>rC*</u>	phosphatase enhanced	$\alpha$ -naphthylphosphate at pH 4.8	VIII R	51	no	<u>bi1</u>	U.V.	
<u>Suppressed mutants</u>								
<u>su1ad20</u>	suppressor 1 of <u>ad20</u>	<u>ad20</u>	I L	-	yes	<u>ad20</u> ; <u>pyro4</u>	spontaneous	Fritchard, 1955
<u>su1paba22</u>	suppressor 1 of <u>paba22</u>	<u>paba22</u>	IV R	4	no	<u>bi1</u> ; <u>paba22</u>	spontaneous	Luig, 1962
<u>suApalB7</u>	suppressor A of <u>palB7</u>	<u>palB7</u>	VIII R	1	no	<u>bi1</u> ; <u>palB7</u>	spontaneous	} Dorn, 1965a
<u>suBpalB7</u>	suppressor B of <u>palB7</u>	<u>palB7</u>	VI	2	no	<u>bi1</u> ; <u>palB7</u>	spontaneous	
<u>suCpalF15</u>	suppressor C of <u>palF15</u>	<u>palF15</u>	V	6	no	<u>bi1</u> ; <u>palF15</u>	spontaneous	Dorn, unpublished
<u>suDpalA1</u>	suppressor D of <u>palA1</u>	<u>palA1</u>	I	2	no	<u>bi1</u> ; <u>palA1</u>	spontaneous	Dorn & Rivera 1965
<u>suSpalA1</u>	suppressor 5 of <u>palA1</u>	<u>palA1</u>	VIII R	-	no	<u>bi1</u> ; <u>palA1</u>	spontaneous	Dorn, 1965b
<u>Su1pro</u>	Suppressor 1 of proline, dominant	<u>pro</u> 1-4, <u>pro7</u>	III R	-	no	<u>pro1</u> <u>paba1</u> <u>y</u> <u>X</u> <u>pro7</u> <u>bi1</u>	Spontaneous	} Forbes, unpublished
<u>Su4pro</u>	Suppressor 4 of proline, dominant	<u>pro</u> 1-5	III	-	yes	<u>pro5</u> <u>bi1</u>	Spontaneous	

TABLE 1—Continued  
*Located mutants of Aspergillus nidulans*

Symbol	Designation	Phenotype	Linkage Group	Allelic Mutants†	Available From FGSC††	Origin of Mutant		
						Strain	Mode	Reference
<u>MISCELLANEOUS</u>								
<i>ts</i>	temperature sensitive	grows at 25°C but not at 37°C	VIII	-	yes	-	-	Forbes, unpublished
<i>tsA</i>	temperature sensitive A	grows at 25°C but not at 37°C	II R	25	no	<i>bi1</i>	U.V.	} Forbes & Sinha, 1966
<i>tsB</i>	temperature sensitive B	grows at 25°C but not at 37°C	VI	5	no	<i>bi1</i>	U.V.	
<i>tsC</i>	temperature sensitive C	grows at 25°C but not at 37°C	II R	17	no	<i>bi1</i>	U.V.	
<i>uvs1*</i>	UV-sensitive 1	-	I R	-	no	-	U.V.	G. Jansen, unpublished

\* These mutants did not originate from the Aspergillus Center at The University, Glasgow, Scotland.

† The numbers under the "Allelic mutants" column represent the isolation numbers and are to be appended to the symbol concerned, i.e.—*bi2*, *bi3*, *bi4*.

†† Fungal Genetics Stock Center, Department of Biological Sciences, Dartmouth College, Hanover, New Hampshire 03755.

TABLE 2  
*Centromere location*

Linkage group	Method	Location	Reference
I	Tetrad analysis	Between <i>ad14</i> and <i>pro3</i>	STRICKLAND 1958
II	Tetrad analysis	Between <i>w3</i> and <i>an2</i>	STRICKLAND 1958
IV	Tetrad analysis	18 units right of <i>meth1</i>	STRICKLAND 1958
VIII	Mitotic recombination	Between <i>fw</i> and <i>orn7</i>	DORN, unpublished

TABLE 3  
*Mutant strains of Aspergillus nidulans\**

Loci for each strain are listed in order of linkage groups and arms, beginning with I left, insofar as the order is known; linkage groups are separated by semicolons.

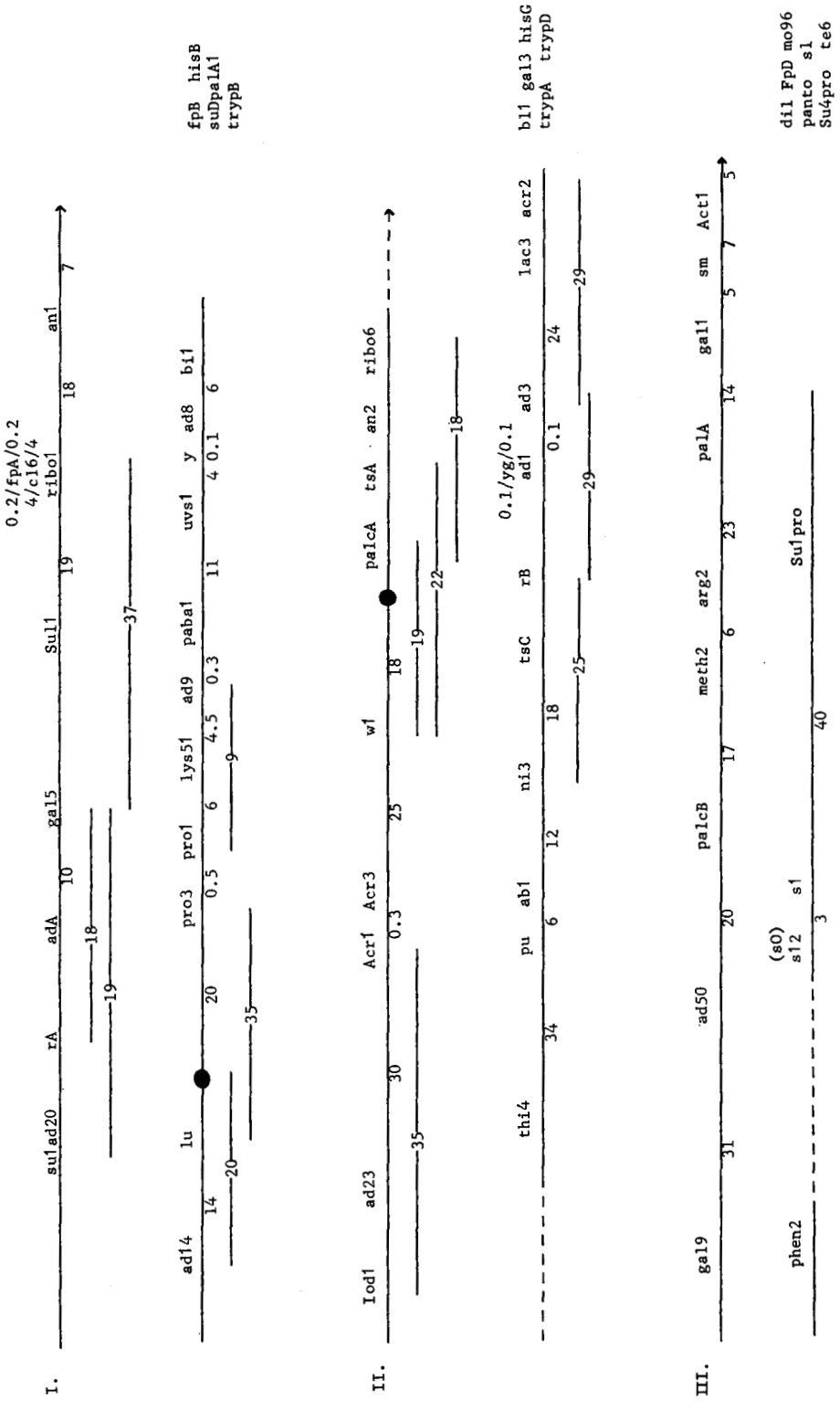
*Symbols:* C—cross; DE—diethyl sulfate; none—no translocation; S—spontaneous; T—translocation; T?—derived from strains with translocation, or irradiated (UV or X) strains or descendant from such strains and not tested; UV—ultraviolet; X—X-ray;  $\beta$ p— $\beta$ -propiolactone;  $\alpha$ —may contain *su1 ad20*.

FGSC No.	Genotype	Translocations	Origin
38	<i>w1; lys1 (ve<sup>+</sup>)</i>	T?	X of <i>w1</i> ; ( <i>ve<sup>+</sup></i> )
39	<i>bi1; w3 pu</i>	T(I,III;VIII) T(VI; VII)	UV of <i>bi1</i> ; <i>w3</i>
216	<i>ada (AM55)</i>	T?	UV of <i>ve</i>
217	<i>adB (AM57) ve</i>	T?	UV of <i>ve</i>
230	<i><math>\gamma</math>; w2; arg1; ve<sup>+</sup></i>	T?	C of <i>bi1</i> ; <i>arg1</i> $\times$ <i><math>\gamma</math>; w2 ad1; s12; ve<sup>+</sup></i>
231	<i>ribo1 <math>\gamma</math>; Act1 nic8</i>	T?	UV of <i>ribo1 <math>\gamma</math>; nic8</i>
232	<i><math>\gamma</math>; ad3; s1 mo96</i>	T?	$\beta$ p of <i><math>\gamma</math>; ad3; s1</i>
233	<i>bi1 w3 cys2 (=s12)</i>	T(III;VIII) T(VI; VII)	UV of <i>bi1</i> ; <i>w3</i>
235	<i>paba1 <math>\gamma</math> trypA69</i>	T?	UV of <i>paba1 <math>\gamma</math></i>
236	<i>trypB403 paba1 <math>\gamma</math></i>	T?	UV of <i>paba1 <math>\gamma</math></i>

TABLE 3—(Continued)

FGSC No.	Genotype	Translocations	Origin
237	<i>paba1</i> $\gamma$ ; <i>trypC801</i>	T?	UV of <i>paba1</i> $\gamma$
238	<i>paba1</i> $\gamma$ <i>trypD432</i>	T?	UV of <i>paba1</i> $\gamma$
239	<i>bi1</i> ; <i>ad23</i> <i>Acr1</i> <i>w3</i> ; <i>nic8</i>	none	C of <i>pro1 paba1</i> $\gamma$ ; <i>ad23</i> <i>w3</i> ; ( <i>ve</i> <sup>+</sup> ) $\times$ <i>bi1</i> ; <i>Acr1</i> ; <i>phen2</i> ; <i>lys5</i> ; <i>s3</i> ; <i>nic8</i>
240	<i>bi1</i>	none	C of $\gamma$ <i>bi1</i> $\times$ $++$
241	<i>bi1</i> ; <i>pacA1</i>	T?	UV of <i>rA1</i> <i>bi1</i>
242	<i>bi1</i> ; <i>pacC5</i>	none	UV of <i>bi1</i>
243	<i>bi1</i> ; <i>palA1</i>	T?	UV of <i>rA1</i> <i>bi1</i>
244	<i>bi1</i> ; <i>palB7</i>	T?	UV of <i>rA1</i> <i>bi1</i>
245	<i>bi1</i> ; <i>palD8</i>	T?	UV of <i>rA1</i> <i>bi1</i>
246	<i>bi1</i> ; <i>palE11</i>	T?	UV of <i>rA1</i> <i>bi1</i>
247	<i>bi1</i> ; <i>palF15</i>	T?	UV of <i>rA1</i> <i>bi1</i>
248	<i>bi1</i> ; <i>palcA1</i>	T?	UV of <i>rA1</i> <i>bi1</i>
249	<i>bi1</i> ; <i>s50</i>	none	UV of <i>bi1</i>
250	$\gamma$ ; <i>fr1</i> <i>palC4</i> <i>paba22</i> <i>pyro4</i>	T?	UV of <i>rA1</i> <i>bi1</i>
253	<i>bi1</i> ; <i>lod</i> <i>w3</i> ; <i>nic8</i>		C of ?
254	<i>bi1</i> ; <i>Acr1</i> <i>w3</i> <i>ab1</i> <i>ni3</i> <i>ad3</i>	T?	C of <i>bi1</i> ; <i>ab1</i> $\times$ <i>ribo1</i> <i>bi1</i> ; <i>Acr1</i> <i>w3</i> <i>thi4</i> <i>ni3</i> <i>ad3</i>
255	<i>w3</i> ; <i>pyro4</i> <i>facC102</i>	T?	S in <i>w3</i> ; <i>pyro4</i>
256	<i>paba1</i> ; <i>w3</i> ; <i>arg3</i> <i>facB101</i> <i>ribo2</i>	T?	C of <i>bi1</i> ; <i>arg3</i> $\times$ <i>paba1</i> ; <i>w3</i> ; <i>facB101</i> <i>ribo2</i>
257	<i>bi1</i> ; <i>w</i> ; <i>Acr1</i> ; <i>nic8</i> ; <i>his38</i>	T?	C of ?
258	<i>nic2</i> <i>hxA1</i> <i>facA303</i> <i>ribo5</i>	T?	C of ?
259	<i>bi1</i> ; <i>ad1</i> ; <i>s12</i> ; <i>pyro4</i>	T?	C of <i>bi1</i> ; <i>pyro4</i> $\times$ $\gamma$ ; <i>w2</i> <i>ad1</i> ; <i>s12</i>
260	<i>bi1</i> ; <i>phen3</i> ; <i>ni51</i>	none	S in <i>bi1</i> ; <i>phen3</i>
261	<i>bi1</i> ; <i>Acr1</i> <i>w3</i> <i>an2</i>	T?	UV of <i>bi1</i> ; <i>Acr1</i> <i>w3</i>
262	<i>bi1</i> ; <i>uY5</i>	T?	DE of <i>bi1</i>
263	<i>bi1</i> ; <i>orn20</i>	none	UV of <i>bi1</i>
264	<i>bi1</i> ; <i>uaX1</i>	T?	DE of <i>bi1</i>
265	<i>bi1</i> ; <i>pu1</i>	T	UV of <i>bi1</i> ; <i>w3</i>
266	<i>bi1</i> ; <i>ni50</i>	none	UV of <i>bi1</i>
267	<i>fpB37</i> <i>an1</i> $\gamma$ <i>ad20</i> ; <i>w2</i>	T?	C of <i>ribo1</i> <i>bi1</i> <i>fpB37</i> $\times$ <i>an1</i> $\gamma$ <i>ad20</i> ; <i>w2</i>
268	$\gamma$ ; <i>w2</i> <i>thi4</i> <i>ni3</i> <i>ad3</i> <i>bl1</i>	T?	C of $\gamma$ ; <i>w2</i> <i>bl1</i> ; <i>s12</i> $\times$ <i>ribo1</i> <i>bi1</i> ; <i>Acr1</i> <i>thi4</i> <i>ni3</i> <i>ab1</i> <i>ad3</i>
269	<i>paba1</i> ; <i>fw</i> <i>facB101</i> <i>ribo2</i> <i>ga17</i> <i>ts</i>	T?	C of ?
270	<i>bi1</i> ; <i>yg</i>	none	UV of <i>bi1</i>
271	<i>Sul1</i> ; <i>ad3</i> ; <i>di1</i>	T?	C of <i>Sul1</i> ; <i>Acr1</i> <i>ad3</i> ; <i>s1</i> $\times$ $\gamma$ ; <i>dil</i> ; <i>pyro4</i>
272	$\gamma$ ; <i>pyro4</i> ; <i>meth3</i>	T?	UV of $\gamma$ ; <i>pyro4</i>
273	<i>ad17</i> <i>paba1</i> $\gamma$ ; <i>FpD11</i>	T?	C of <i>ribo1</i> <i>bi1</i> ; <i>FpD11</i> $\times$ <i>ribo1</i> <i>ad17</i> <i>paba1</i> $\gamma$
274	$\gamma$ <i>lu</i>	T?	C of <i>pro1</i> $\gamma$ $\times$ <i>a</i> <i>gal5</i> <i>an1</i> <i>lu</i> <i>bi1</i>
275	<i>fpA1</i> <i>ad14</i> <i>paba1</i> $\gamma$	T?	C of <i>fpA1</i> <i>bi1</i> $\times$ <i>ribo1</i> <i>paba1</i> <i>ad14</i> $\gamma$
276	$\gamma$ ; <i>fr1</i> <i>paba22</i> ; <i>pyro4</i>	T?	C of <i>bi1</i> ; <i>paba22</i> $\times$ $\gamma$ ; <i>fr1</i> <i>pyro4</i>
277	<i>bi1</i> $\gamma$ ; <i>Acr1</i> ; <i>hisA10</i> ; <i>ribo2</i>	T?	C of ?
278	$\gamma$ <i>bi1</i> ; <i>Acr1</i> ; <i>ribo2</i> <i>hisH13</i>	T?	C of ?
279	$\gamma$ ; <i>pyro4</i> ; <i>aaX1</i>	T?	C of ?

\* This list contains recent additions to the Fungal Genetics Stock Center, Dartmouth College. See BARRATT *et al.* (1965) for the basic list of mutant strains.



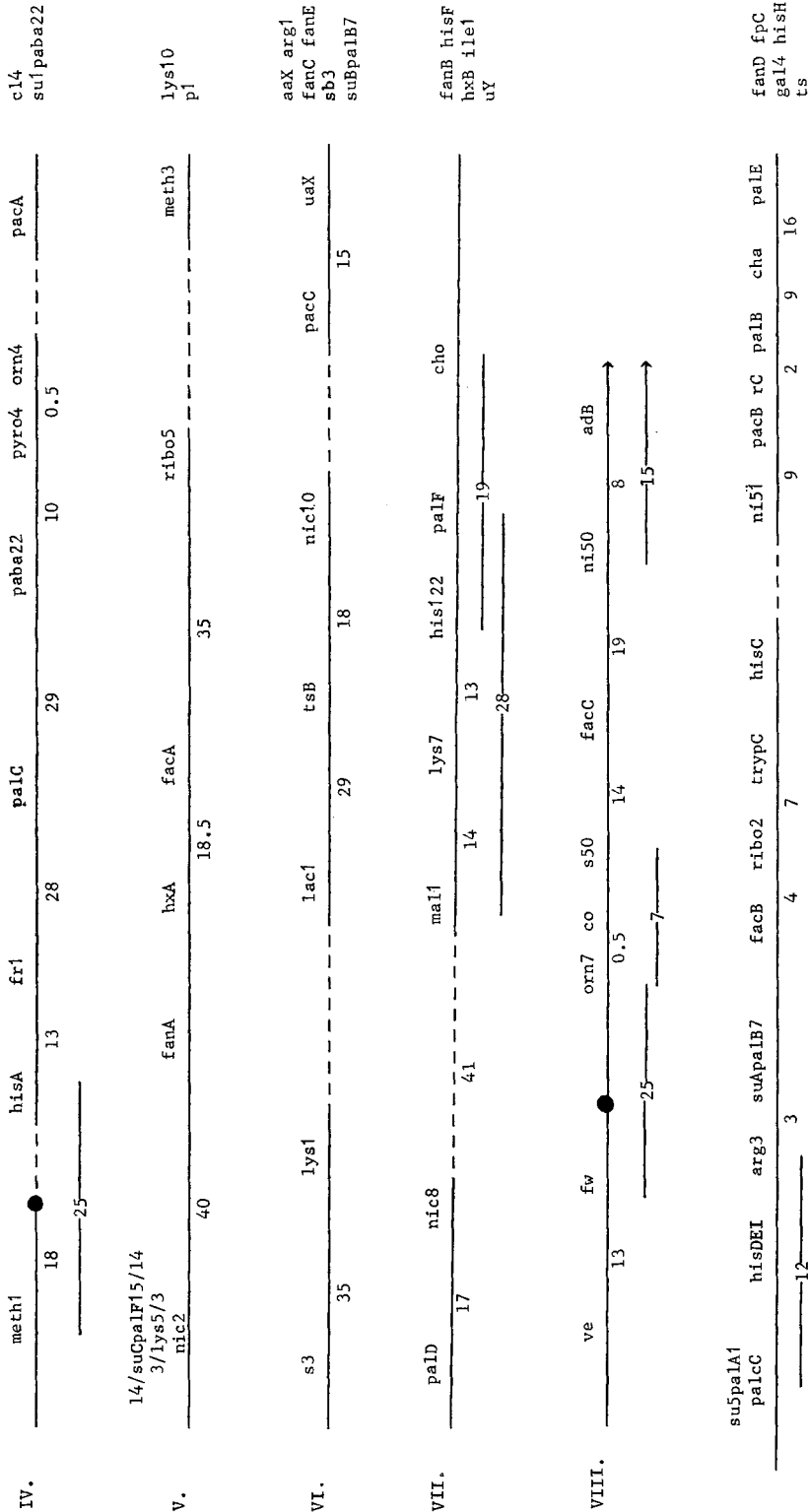


FIGURE 1.—Linkage map of *Aspergillus nidulans*. The linkage distances, given in percent meiotic recombination, are derived from a variety of sources and, hence, should only be considered as approximate values. Mutants placed to the right of a linkage group indicate that they have been assigned to that group by mitotic analysis but have not been further mapped.