

GENETIC MAPPING OF THE *pho2*, *PHO82-pho4* AND *pho85* LOCI OF YEAST

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ABSTRACT

The *PHO82-pho4* (*PHOO-phoD*) locus was mapped on the right arm of chromosome VI, 6.5 cM centromere-distal to *met10*. A *pho85* (*phoU*) mutant was newly isolated, and its map location was determined on the left arm of chromosome XVI, 15 cM centromere-distal to *rad1*. A mutant gene that causes temperature-sensitive growth on nutrient medium was found very near (or at) the *pho85* locus. The supertriploid method was successfully applied to locate the *pho2* (*phoB*) locus on chromosome IV. The *pho2* locus was 40 cM from *rna11* on the left arm of chromosome IV.

GENETIC analyses of the production of repressible acid phosphatase of *Saccharomyces cerevisiae* (SCHURR and YAGIL 1971; TOH-E *et al.* 1973; TOH-E and OSHIMA 1974; UEDA, TOH-E and OSHIMA 1975; UEDA and OSHIMA 1975) revealed that at least six chromosomal genes, *pho2* (*phoB*)* *PHO82-pho4* (*PHOO-phoD*), *pho81* (*phoS*), *pho80* (*phoR*), *pho85* (*phoU*) and *pho84* (*phoT*), regulate the synthesis of the repressible acid phosphatase coded for by *pho5* (*phoE*). Among the *pho* genes described above, *pho5* and *pho80* have been mapped. HANSCHKE, BERES and LANGE (1978) mapped *pho5* between *tsm134* and *lys2* on chromosome II. The *pho80* gene was mapped on the right arm of chromosome XV (BERES, personal communication). This report describes the location on the yeast chromosome map of the *pho2*, *PHO82-pho4* and *pho85* loci that were mapped by using the supertriploid method (WICKNER 1979), as well as by standard tetrad analysis. Map distances were calculated by PERKINS (1949) equation, $x = 100 \frac{T + 6NPD}{2(PD + NPD + T)}$, and shown as centimorgans (cM).

The *PHO82* strain OC-6A (*apho3 PHO82*) was crossed with the multiply labelled strain, AT201. The segregation of markers in cross W221 indicated the linkage between *PHO82* and *his2* (Table 1). To determine the location of *PHO82* with respect to the other markers on the right arm of chromosome VI, two more crosses (W275 and W424) were carried out. Data shown in Table 1 locate the position of *PHO82* at 6.5 cM centromere-distal to *met10*.

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* The nomenclature system of yeast genetic markers recommended by PLISCHKE *et al.* (1975) is used throughout this work and the conventional name is shown in parenthesis as appropriate.

TABLE 1

Mapping of pho genes

Interval tested	Cross*	PD	No. of asci NPD	T	cM
<i>his2-PHO82</i>	W221	13	0	6	
	W275	52	0	13	
	W424	37	0	19	
	Total	102	0	38	13.6
<i>cdc14-PHO82</i>	W275	55	0	11	
	W424	43	0	12	
	Total	98	0	23	9.5
<i>met10-PHO82</i>	W424	47	0	7	6.5
<i>cdc14-met10</i>	W424	56	0	6	4.8
<i>his2-met10</i>	W424	48	0	15	11.9
<i>his2-cdc14</i>	W424	55	0	8	7.3
<i>rad1-pho85</i>	W465	29	0	14	
	W480	52	0	18	
	W486	31	0	16	
	Total	112	0	48	15.0
<i>trp1-pho2</i>	W320**	33	6	81	
	W382	1	8	1	
	W425	3	0	6	
	W458	27	3	61	
	W471	10	5	37	
	Total	74	15	193	49.8
<i>mak21-pho2</i>	W382	2	0	9	
	W425	3	0	6	
	W458	17	5	50	
	W471	12	4	40	
	Total	34	9	105	53.7
<i>rna11-pho2</i>	W425	4	0	5	
	W458	31	3	53	
	Total	35	3	58	39.6
<i>rna11-trp1</i>	W425	7	0	2	
	W458	57	0	31	
	Total	64	0	33	17.0
<i>mak21-trp1</i>	W425	7	0	5	
	W458	45	2	39	
	Total	52	2	44	28.6
<i>rna11-mak21</i>	W425	5	0	6	
	W458	32	3	41	
	Total	37	3	47	37.4

*W221 α + + + + + + + + *pho3 PHO82*

a *ade1 ura3 his2 leu1 trp5 ura1 aro7 met2 pho3* +

W275 a *his2 his7 cdc14* + + + *pho3* +

α + + + *leu1 trp5 ura1 pho3 PHO82*

TABLE 1—Continued

	Interval tested	Cross*	PD	No. of asci NPD	T	cM
W424	α <i>his2 cdc14 PHO82</i>	+ + + + +				<i>pho3</i>
	a + + + +	<i>met10 his4 leu2 ura1 ade2 pho3</i>				
W465	a <i>his7 + + + +</i>	<i>pho3 pho85</i>				
	α + <i>arg1 rad1 cdc4 gal4 pho3</i>	+				
W480	a <i>his7 cdc4 gal-</i>	+ + + +				<i>pho3 pho85</i>
	α + + +	<i>arg6 rad1 cyh2 mak3 pho3</i>				
W486	a <i>his7 arg6 cyh2 rad1</i>	+ +				<i>pho3 pho85</i>
	α + + + +	<i>leu1 arg1 pho3</i>	+			
W320	a + + +	<i>arg1 pho3 pho2</i>				
	α <i>cdc16 asp5 trp1</i>	+ <i>pho3</i> +				
W382	a + + +	<i>arg1 pho3 pho2</i>				[KIL-0]
	α <i>leu2 mak21</i>	+ + +				
W425	a + + +	<i>trp1 pho3 pho2</i>				[KIL-b]***
	α <i>arg6 pho3 pho2</i>	+ + +				[KIL-b]
W458	α + + +	<i>trp1 rna11 mak21</i>				[KIL-0]
	a <i>arg1 pho3 pho2</i>	+ + +				[KIL-b]
	α + + +	<i>trp1 rna11 mak21</i>				[KIL-0]

** Pooled tetrad data from crosses other than listed in this table are included in this line.

*** [KIL-b] is a mutant killer plasmid showing superkiller phenotype. This mutant plasmid bypasses some *mak* mutations, but not *mak21* (TOH-E and WICKNER, unpublished).

A new *pho85* mutant (B104) was isolated from AT199 (**a** *pho3 his7*) after ethyl methanesulfonate mutagenesis (LINDEGREN et al. 1965) and mapped. B104 showed two other mutant phenotypes: temperature-sensitive growth on nutrient medium and respiratory deficiency. Both of these phenotypes segregated 2+:2- in the cross of B104 \times wild type. The temperature sensitivity co-segregated with the *pho85* character, but the respiratory deficiency of B104 segregated independently of the *pho85* trait. No recombination could be detected between *pho85* and *ts* among 46 asci tested. A temperature-resistant revertant was isolated from the *pho85-ts* strain (AT275). The revertant still had the *pho85* phenotype. However, this reversion was due to an extragenic suppressor.

Since the *pho85* locus was known previously to be centromere-linked, AT275 (**a** *pho3 pho85-ts his7*) was crossed with strains having known centromere markers. Linkage between *pho85* and *rad1* on chromosome XVI was found in cross W465. The pooled tetrad ratio collected from the crosses (W465, W480, and W486) was PD : NPD : T = 112 : 0 : 48, and the distance between these two genes was calculated as 15 cM (Table 1). No close linkage between *pho85* and *mak3* could be observed. The sequence, *pho85-rad1-centromere*, was deduced by analyzing individual tetrads of cross W486. This arrangement is consistent with the fact that the frequency of second-division segregation of *pho85* (25.5%) is higher than that of *rad1* (15%).

The linkage group of *pho2* was analyzed by the supertriploid method described by WICKNER (1979). A *pho2* strain W273-14C (**a** *pho3 pho2 arg6*) was mated with spores of supertriploid 1234 *en masse* and protrophs were selected. Resulting

diploids having random trisomes were dissected, and the segregation patterns of *pho2* and that of standard markers were compared. Thus, the possible chromosomal location of *pho2* was narrowed to chromosome *IV*, *VI* or a new chromosome. The tetrad ratio data between *pho2* and *trp1* were collected from crosses in which both of these two markers segregated 2+ : 2-. The cumulative ratio was PD : NPD : T = 33 : 6 : 81. Since this ratio indicates loose linkage between *pho2* and *trp1*, the linkage between *pho2* and other markers on chromosome *IV* was tested. Cross W366, containing *aro1D* and *pho2*, gave the ratio PD : NPD : T = 0 : 2 : 3 with respect to these markers, which showed no linkage between *aro1D* and *pho2*. Then, the linkage between *pho2* and *rna11* and *mak21* was tested. The tetrad distribution shown in Table 1 indicates: (1) *pho2* is loosely linked to *trp1* and *mak21*, and (2) the linkage between *pho2* and *rna11* is closer than that between *pho2* and *trp1*. WICKNER and LEIBOWITZ (1979) mapped *mak21* between *trp1* and *aro1D* and explained the inconsistency of map distance between *mak21* and *rna11* by relocating *rna11* on the left arm of chromosome *IV*. The linkage data shown in Table 1 support this revision.

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