

A cDNA cloned from pregnant mouse uterus exhibits temporo-spatial expression and predicts a novel protein

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A cDNA was cloned from a pregnant mouse uterus cDNA library. On conceptual translation, the cDNA has one long open reading frame that predicts a novel protein of 606 amino acids. This protein is principally composed of two CUB domains and a ZP domain; motifs found in proteins implicated in egg-sperm recognition. Probes derived from the cDNA were used to conduct Northern hybridizations. The expression of this mRNA is temporal; message first appears in the uterus 6 days prior to birth, it increases each subsequent day to attain maximal levels at 3 days prior to birth and then abruptly decreases during the

last 3 days of pregnancy. The expression of this mRNA is restricted; message is abundant in the uterus during late pregnancy, but it is not found in non-pregnant uterus or in a variety of adult or fetal tissues. The temporo-spatial expression of this pregnant uterus specific mRNA and the consolidation in the predicted protein of two motifs implicated in early pregnancy events suggests that the product of the gene represented by this mRNA may play an important role in events that transpire during late pregnancy.

INTRODUCTION

To identify molecular tools that will facilitate the investigation of events surrounding the induction of parturition, a differential screen was conducted to isolate selectively expressed mRNAs using a cDNA library constructed from late pregnancy mouse uterus. The reported cDNA was selected for evaluation because the expression of this mRNA decreased during late pregnancy, suggesting that the product of the gene represented by this mRNA may possess an inhibitory influence on the induction of parturition.

Conceptual translation of the cDNA predicts a new and distinct protein that incorporates two recently defined motifs; the CUB motif (complement subcomponents C1r/C1s, Uegf protein and bone morphogenetic protein) [1] and the ZP motif (zona pellucida) [2]. The CUB motif has been associated with developmental phenomena [1] and is found in several developmentally associated proteins to include the neuronal A5 antigen [3], the spermadhesins [4–8] and two developmentally linked members of the transforming-growth-factor beta superfamily; tolloid and bone morphogenetic protein [9,10]. However, it is also found in many other proteins not currently associated with development such as the tumour necrosis factor inducible protein TSG-6 (tumour necrosis factor stimulated gene-6) [11] or members of the complement family (C1s and C1r) [12–14]. The motif comprises 120 amino acids with four conserved cysteines; its function is currently unknown. The ZP motif is a peptide of 260 amino acids characteristically positioned near a carboxyl membrane-spanning region that includes eight conserved cysteines as well as invariant hydrophobic or aromatic amino acids suggesting a role in binding functions [2]. It is found in a heterogeneous group of proteins that includes the type III transforming-growth-factor beta receptor; betaglycan [15,16]. Binding studies conducted using a prokaryotic expression vector containing the betaglycan ZP domain confirm TGF β binding to this motif [17]. Although these data were not corroborated by constructs employing a eukaryotic expression vector [18], the disparity may simply reflect differences resulting from post-translational modification.

These two motifs are present in two separate families of proteins implicated in events surrounding egg-sperm recognition; the spermadhesins and the ZP protein(s) [19,20]. The presence of two motifs implicated with early pregnancy events and the temporal and tissue restricted expression of this mRNA in the pregnant uterus suggest that this new protein may possess an important role in uterine events in late pregnancy.

EXPERIMENTAL

Materials

The study was approved by an animal use committee and conforms with established guidelines for the use of laboratory animals. The cDNA library was constructed from mouse uterus collected during the last 2 days of pregnancy (strain CF-1, birth = 20 days from mating) [21]. RNA used in Northern hybridizations was purified from reliably dated tissue (strain CD-1) by the acid guanidine technique [22]. A cDNA previously isolated and identified by this laboratory consisting of 800 bp of 3' end of the mouse ubiquitin gene was used as a control for Northern blots. Probes for the differential screen were prepared using the same oligo(dT)-selected RNA employed in construction of the library and comparable RNA collected 3 days before birth [23]. A 1054 bp *Bam*HI fragment from the cDNA was used for probe preparation in Northern hybridizations as well as in a second screen of the library to isolate additional full-length cDNAs. Transcript size was determined using an RNA ladder (cat. no. 5620SA, Bethesda Research Laboratories, Gaithersburg, MD, U.S.A.).

Methods

Total RNA was purified from uterus as well as an assortment of adult and fetal tissues (strain CD-1); 20 μ g aliquots were separated on 1% agarose gels and transferred to nylon membranes for use in Northern blotting. The quantity, integrity and

efficacy of transfer of RNA was confirmed by viewing both the ethidium bromide-stained gels and membranes under UV light. [α - 32 P]dCTP labelled cDNA probes were prepared by random priming (USB, Cleveland, OH U.S.A.) using standard protocols [24]. Northern hybridizations were performed under high stringency conditions (hybridization at 68 °C and final wash = 0.1 \times SSC and 0.5% SDS at 68 °C) and the blots exposed to Kodak XAR film for 2 days at -70 °C. One clone was completely sequenced (Sequenase, USB) in both directions using custom primers constructed at successive 250–300 bp intervals; two other full-length cDNAs were also sequenced extensively. The nucleotide sequence was conceptually translated (pc gene, Intelligenetics, San Jose, CA) and resultant nucleotide, as well as predicted amino acid sequence, compared to non-redundant nucleotide and protein databases in GenBank [25,26]. Linear full-length cDNA in Bluescript II KS (BSUTCZPfl, restriction digest with *Xho*I) as well as a truncated cDNA (BSUTCZPbgl, digest with *Bgl*II) were subjected to coupled *in vitro* transcription and translation with rabbit reticulocyte lysate in the presence of [35 S]methionine using a commercially available kit (TNT, Promega, Madison, WI, U.S.A.). The products were electrophoresed on an SDS/PAGE gel and the dried gel was exposed for 2 days to Kodak XAR film.

RESULTS

Comparison of the nucleotide sequence against the non-redundant nucleotide database of GenBank established the cDNA as novel. The cDNA (GenBank accession U69699) contains only one long open reading frame that conceptually translates a 606 amino acid protein with a predicted molecular weight of 67956 Da (Figure 1). Translation is presumed to start at the methionine-labelled amino acid 1. Consistent with the presence of a signal peptide, a majority of the first 20 amino acids are hydrophobic and potential cleavage sites are predicted between amino acids 18–19, 19–20, 20–21 and 21–22 [27]. The presence of this putative signal peptide reinforces the proposed location of the translational start site.

The protein was evaluated against the non-redundant protein database of GenBank [26]. Within the first 275 amino acids of the protein, two regions were identified that consistently displayed similarity to what was subsequently identified as the CUB motif. Analysis of these regions in the context of published CUB domains confirms that the protein contains two contiguous CUB domains; the first begins at Cys-32 and the second at Cys-154 (Figure 1).

At Cys-276, a region was identified that is similar to TGF betaglycan, uromodulin [28] and members of the ZP protein family; proteins conspicuous for the presence of a ZP motif [2]. Comparison of this region to published ZP domains confirms that this gene also contains a ZP domain (Figure 1).

The remainder of the protein displayed weak similarity to the protein ebnerin [29]. Analogous to ebnerin, there is a putative transmembrane domain present near the carboxyl end [30]. The presence of this putative transmembrane domain suggests that this protein may also function as an integral membrane-associated protein analogous to ebnerin or TGF betaglycan. However definitive evidence for this speculation requires confirmation from additional experiments. The protein terminates in a short 18 amino acid polypeptide presumably positioned within the cytoplasm.

To determine the pattern of expression of this gene in the uterus and at other potential sites in the mouse, Northern hybridizations were conducted against total RNA purified from late gestation uterus as well as a variety of other tissues. Message

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28 CTG GCT GGC TTA TAG CCC TCT GAG ATG TGA CCT GGA AGA CTG ACC CCA GTG CCA
82 GGA ACT GCT GGC CAG GTT GCA GCT CCT CAG CTG TGG AGT TCT CAC CAG CAC CAC
136 CCC TGG GAT GGC CAA ACT GAC ACC TTT CCC TAG ATA CCA GGA AGC TTA CAA AAC
190 TCG GAG AAT TAT AAG AAG CCT GAG GGT GAG GGC CCT GAG GAC CCT GCG AAG ATG
M 1

244 GAG GTC ACT GGA AGG CTG TTC ATC TGG GCT ATT TTA GCT GTC TCC TGT GGG GCA
E V T G R L F I W A I L A V S C G A A
298 CAG CTG AAT TCT ACA GAG GCT GAA GGC AAA TCA AGA TCC AGA GCC AGT CTG GGA
Q L N S T E A E G K S R C T A S L G
37
352 GGG GCC AAT CTG GGA GAG ACC CAT AAA GCC CTG GTC CTA GAG CTC AGT GCG AAT
G A N L G E T H K A L V L Q L S A N
55
406 GAG AAC TGC ACT TGG ACC ATA GAG AGA CCT GAA AAC AGG AGC ATC AGA ATC ATC
E N C T W T I E R P E N R S I R I I
73
460 TTC TCC TAC ATC AAG TTG GAT CCA GGC ACC AGG TGT GAA ACT GAA AAC ATT AAG
F S Y I K L D P G C R C E T E N I K
91
514 GTG TTT GAT GGA AGC TCC ACC AGT GGT CCT CTG CTA GGG AAG GCC TGC AGC AGA
V F D G S S T S G P L L G K A C S R
109
568 AAT GAT TTC GTG CTT GTG TTT GAA TCA TCA TCC AAC TCG ATG ACA TTT CAG ATA
N D F V P V F E S S S N S M T F Q I
127
622 GTC ACT GGC TCG ACA AAG TTC CCA AGG AGT GTC TTT ATC TTC TAC TAT TTT TTC
V T G L T K F P R V I P R V Y F F
145
676 TCC GCT GCT ACC GTT ATT CCA AAC TGC GGT GGT GAC CTG CGA GCA TTG GAA GGG
S A A T V I P N C G G D L R A L E G
163
730 TCC TTC ACC AGC CCC AAT TAT CCA AAG CCG CAC CCT GAG CTG GCA TAT TGT GTC
S F S S P N Y P K P E H P E L A Y C V
181
784 TGG CAC ATA CAA GTG GGG AAA GGC TAT AAG ATA CAA TTG AAG TTT ACA GAT CTC
W H I Q V G K G Y K I Q L K F T D L
199
838 TTA CTA GAG ATG GAT GAA AAC TGC AAG TTT GAT TTC ATC GCA GTC TAC GAT GGC
L L E H D E N C K F D F I A V Y D G
217
892 CCC TCC ACC ACG GCA GGC CTG CTC AAA CAA TGC GGC GTG GAA CCT ACC TTA
P S T T A G L L K Q L C G V E P T L
235
946 GAA TCT TCC TCT GAC GCC ATG ACT GTC GTG CTA TCT ACA GAT TAT GCC AAT TCC
E S S S D A M T V I F I S K S F
253
1000 TAC AAA GGC TTT TCT GCT TCC TAC ACT TCA ATT TAC TAC CAC GAT GTC AAC ACT
Y K G F S A S Y T S I Y I H D V N T
271
1054 ACA TCT CTA AGT TGT GTT TCT GAC AAG ATG AGA GTC ATC ATA AGC AAA TCA TAC
T S L S C V S D R K M R V I T S K S F
289
1103 CTA CCG GCA CTC AAC TAC AAT GAG AGC AAT TTG CAG CTA AAT GAC CCA ACT TGC
L P A L N Y N E S N L O L N D P T C
307
1162 AGA CCG AAT GTA TCA AAT GTC ATA GAG TTC TCT ACT CCT CTG CAC GAA TGC GGT
R P N V S N V I E F M R V I T H E C F
325
1216 ACA GTC AAA AAG ATA GAG GAC CAC GCG ATC AGC TAT ACC AAC AGG ATC ACC TTC
T V K K I E D H A I S Y T N R I T F
343
1270 ATC GAG TCT CCG TCT GCT GTG ATC ACC CGA CAG AAG CTC CTC CAG ATC GTG
I E S P V S A V I T R O K L L O F V
361
1324 GTG ACC TGT GAG ATG GAG TAT AAC TCT ACC GTG GAG ATT ATG TAC ATA ACT GAA
V T C E M E Y N S T V E I N Y I T E
379
1378 GAC GAC ATC ATA CAG AAC CAG AGT GTC CTG GGC AAA TAC AAC ACC AGC CTG GCT
D D I I O N O S V I G C K Y N T S R L
397
1432 CTC TAT GAG TCC GAC TCA TTT GAA AAC CTC GTA CAG GAG TCA CCA TAT TAT GTA
L Y E S D S F E N L V O E S P Y Y V
415
1486 GAC TTG AAC CAG ACT CTC TTT CGT CCA AGC CAC CTT GCA CAC CTC GGA TCC AAG
D L N O T L F R P S H L A H L G S K
433
1540 CCT GGT GGT GTT TCT GGA TAC TTG CAG AGC TCG CCT ACG TCT GAC TTT GCA TCT
P G G V S G Y L O S S P T S D F A S
451
1594 CCA ACC TAC GAC TTA ATC AGC AGC GGA TGT TGT CAA GAT GAG ACC TGT AAG GTG
P T Y D L I S S G C C O D E T C K V
469
1648 TAT CCC TTA TTT GGG CAC TAT GGA AGG TTC CAG TTT AAT GCC TTT AAA TTC TTG
V P L F G H Y G R F O F N A F K F L
487
1702 AAA CAT CTC AAC TCT GTG TAT CTC AAG TGT AAG ATT TTG ATA TGT GAT AAC AAT
K H L N S V Y I K C K I L I C D N N
505
1756 GAC CAA ACA TCT CGC TGC AAT CAA GGC TGT GTC CCA AGA AGG AAA CGG GAT ATT
D O T S R C N O G C V P R R K R D I
523
1810 CCT TCC TAC AAA TGG AAG ACT GAC TCT GTT ATA GGA CCC ATT CGC CTG AAG AGG
P S Y K W K T D S V I G P I R L K R
541
1864 GAC CGA AGT GCA AGT AGA GAT TCA GGA CTT CTG CCT CAA ATA CAT GAA GCA GAA
D R S A S R D S G L L P Q I H E A E
559
1918 ATT TCA AAC CAG CCC CTC AGT CGC CTG TAC CTG TTT TCC TTC ATG GTT CTT GCG
I S N Q P L S R L Y L F S F M V L A
577
1972 CTA AAT GTG GTG ATT GTG GCG ATA ACC ACA GTG AAG CAT TTC CTA AAT CCG TGG
L N V V I V A I T T V K H F L N R W
595
2026 ATG GAT CAC AGA TAC CAG AAG CTG CAG GTC TAC TAG CTG TGG ACC CCA AGA AAG
M D H R Y Q K L Q V Y
2080 GCC TCT TGT CTT CTC CTG GGT GCC TGA TGA AGC AGC TCC CTG TGC CTA CAA GGG
2134 TTC AGA ATA AAC CAG GAA GGC CTA AAA AGC AAC ATG TGT ACC ATC ATG TTC AAA
2188 TCA CAT CGA CAG TGT GTT TCA TTA CCG AAT AGC TGA AAT GAA TAA ATA GTA CGT
2242 CTG GTC AAA AAA AAA AAA AAA AAA

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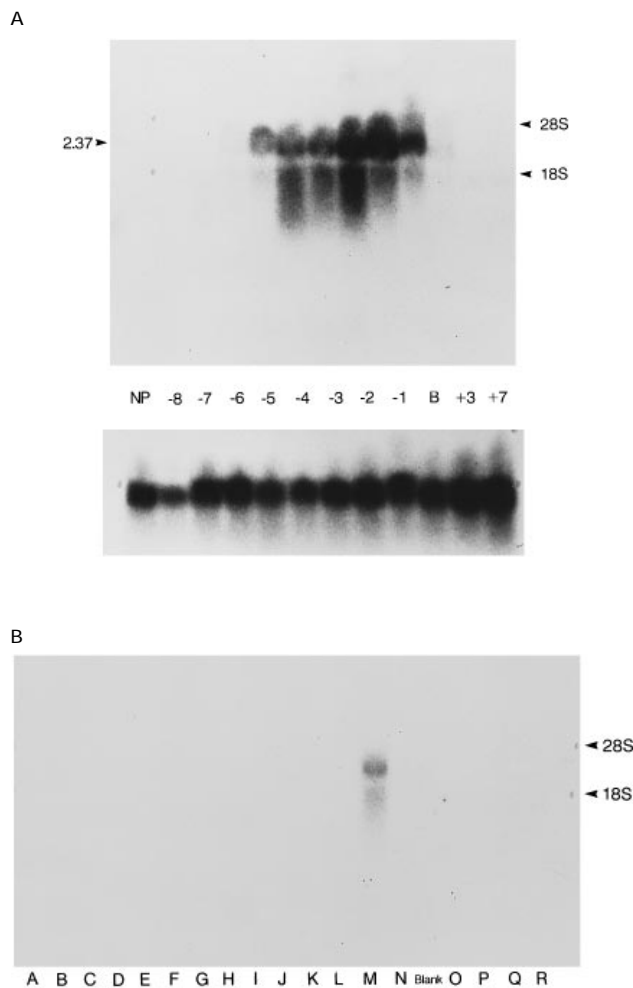


Figure 2 Northern hybridization

Northern hybridization using 20 μ g (pooled from three mice) of total RNA obtained from various mouse tissues. **(A)** Data against RNA purified from mouse uterus collected during pregnancy. Days prior to (–) and following (+) birth **(B)** are indicated by numbers and NP = non-pregnant. Upper panel shows hybridization to probes from UTCZP and lower panel shows hybridization to control probes (ubiquitin). **(B)** Data from hybridization against RNA purified from a variety of adult and fetal mouse tissue. Lanes: A, brain; B, fetal brain; C, thymus; D, heart; E, lung; F, spleen; G, pregnant maternal liver; H, fetal liver; I, liver; J, pregnant maternal kidney; K, kidney; L, ovary; M, pregnant uterus; N, blank lane; O, placenta; P, skin; Q, fetal skin; R, gut.

first appears in the uterus at 6 days before birth (Figure 2A). The amount of mRNA (as reflected by signal strength) increases daily to reach a maximum at 3 days before birth. The level of mRNA then decreases on each subsequent day and by the first day

Figure 1 Nucleotide and predicted amino acid sequence of UTCZP

Nucleotides are numbered on the left and amino acids are numbered on the right. The putative signal peptide is shown as a dashed underline (amino acid 1-M E V T . . V S C G-18). The potential cleavage sites for the signal peptide are indicated by the inverted arrows. The CUB domains are shown as a labelled single underline (first CUB domain amino acid 32-C T A S . . V I P N-153 and the second CUB domain amino acid 154-C G G D . . T S L S-275). The ZP domain is shown as a labelled boxed region (amino acid 276-C V S D . . P R R K-520). The putative transmembrane domain is shown by a double underline (amino acid 568-L Y L F . . I T T V-588). Boxed regions in the 3' untranslated region indicate potential polyadenylation signals. *Bgl*II (nucleotide 831) and *Bam*HI (nucleotides 479 and 1533) restriction sites are as indicated.

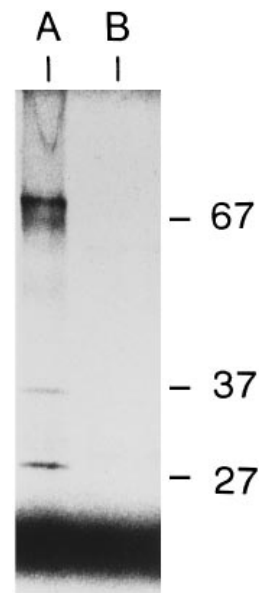


Figure 3 In vitro transcription and translation

Products of coupled *in vitro* transcription and translation from the plasmids BSUTCZPfl and BSUTCZPbgl are represented in lanes A and B. Three bands are prominent in lane A (BSUTCZPfl) located near the 67, 37 and 27 kDa markers representing transcription/translation from the full-length cDNA and transcription/translation from methionines carboxyl to the start site. As expected, all three bands are absent in products derived from the truncated BSUTCZPbgl plasmid (lane B).

following birth, it is almost undetectable. Signal is not detected in RNA collected from the uterus at 7 days before birth or in the absence of pregnancy to include days 3 and 7 following birth. Message was not found in the placenta or in any of the other fetal or adult tissues analysed (Figure 2B). The expression of this gene appears to be temporal and restricted to the gravid uterus. The RNA size marker on the Northern blot confirms the length of the cDNA. The Northern blot is also remarkable for the presence of prominent smears below the major band of hybridization. This does not represent degradation of the RNA as shown by the ubiquitin control; speculatively these smears may reflect rapid turnover of the message for this gene.

To confirm the presence of a functional open reading frame and the putative start site (methionine labelled 1), plasmids BSUTCZPfl and BSUTCZPbgl were subjected to coupled *in vitro* transcription and translation analysis. Three major bands near the 67, 37 and 27 kDa markers are evident (Figure 3) as well as a minor band in the vicinity of the 67 kDa band. The major 67 kDa band is consistent with translation from the putative start site and is compatible with the predicted molecular weight of the protein. The 37 and 27 kDa bands presumably represent translation from other methionines carboxyl to the start site. The minor band observed near the 67 kDa band conceivably represents transcriptional infidelity or post-translational modification of the protein. Translation from the truncated plasmid pBSUTCZPbgl results in loss of all bands as expected. The data confirm translation from the methionine labelled as 1.

DISCUSSION

In the absence of functional data, we have designated this cDNA as UTCZP (uterine cub motif zona pellucida motif). The mRNA encoded by this gene is relatively abundant within the uterus

during late pregnancy and expression is temporal and tissue restricted. The conceptually translated protein contains two motifs; the CUB motif and the ZP motif. Although these motifs are found in a variety of heterogeneous proteins, the presence of motifs that are prominent features of two distinct protein families implicated in egg-sperm recognition in a late pregnancy uterine-specific protein is intriguing. Collectively, the data suggest that this protein possesses an important role in late pregnancy uterine events.

The predicted protein is structurally similar to ebnerin; an integral membrane associated protein of unknown function that is found in von Ebner's glands [29] and ductin, a mouse protein found in the intestine, liver and pancreas [31]. Both of these proteins contain CUB and ZP motifs, however the arrangement of these domains in these proteins differs from the arrangement present in UTCZP. Analogous to these proteins, UTCZP has a putative transmembrane domain that is followed by a short cytoplasmic tail. Speculatively, TTCZP may also be an integral membrane-associated protein. To determine if a human counterpart exists, a search was made of the expressed sequence tag (EST) database (dbest) using peptide sequence derived from this protein. Several ESTs originating from human pancreas were found that exhibit a high degree of similarity to this protein. It is uncertain whether these ESTs represent the human counterpart to this protein or to ductin at present, but the peptides predicted from short open reading frames contained in these ESTs display more similarity to UTCZP than to ductin, suggesting that they may represent the human counterpart to UTCZP.

Bovine acidic seminal fluid protein is a member of the sperm-adhesin family that essentially consists of a single CUB domain [32–35]. *In vitro*, this protein functions as a mitogen, growth factor and it stimulates progesterone secretion in cultured ovarian cells [32]. If the CUB motifs found in UTCZP can be demonstrated experimentally to exhibit similar function, the presence of this protein in the uterus during late pregnancy has important implications in understanding the induction of parturition.

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