

LPS stimulation²⁴ and the known importance of IL-6 and PGE₂ on *in vitro* hepatocyte proliferation indicate the central importance of the Kupffer cell in this unique mammalian response.

References

- Higgins GM, Anderson RM. Experimental pathology of the liver. *Arch Pathol* 1932; 12:186.
- Bucher NL, Swaffield MN. Regulation of hepatic regeneration in rats by synergistic action of insulin and glucagon. *Proc Natl Acad Sci USA* 1975; 72:1157.
- LaBrecque DR, Pesch LA. Preparation and partial characterization of hepatic regeneration stimulator substance (SS) from rat liver. *J Physiol* 1975; 248:273.
- Sigel B, Acevedo FJ, Dunn MR. The effect of partial hepatectomy on autotransplanted liver tissue. *Surg Gynecol Obstet* 1963; 117: 29–36.
- Moolten FL, Bucher NL. Regeneration of the rat liver: transfer of humoral agent by cross circulation. *Science* 1967; 158:272–274.
- Morley CG, Kingdon HS. The regulation of cell growth: I. Identification and partial characterization of a DNA synthesis stimulating factor from the serum of partially hepatectomized rats. *Biochim Biophys Acta* 1973; 308:260–275.
- Price JB Jr. Insulin and glucagon as modifiers of DNA synthesis in the regenerating rat liver. *Metabolism* 1976; 25:1426–1428.
- Starzl TE, Jones AR, Terblanche J. Growth-stimulating factor in regenerating canine liver. *Lancet* 1979; 1:127–130.
- Mayanskii DN, Scherbakoff VI, Mayanskaya NN. Lysosomal enzyme activity in hepatocytes and Kupffer cells from intact and partially hepatectomized rats. *Biochem Exp Biol* 1980; 16:309–314.
- Namasivayam A, Padmanabhan N. The possible role of reticuloendothelial system in hepatic regeneration. *Indian J Physiol Pharmacol* 1982; 26:105–112.
- Saba TM. Physiology and pathophysiology of the reticuloendothelial system. *Arch Intern Med* 1970; 126:1031.
- Bucher NL, Malt RA. *Regeneration of Liver and Kidney*, Boston: Little, Brown, 1971, p 17.
- West MA, Billiar TR, Curran RD, et al. Evidence that rat Kupffer cells stimulate and inhibit hepatocyte protein synthesis *in vitro* by different mechanisms. *Gastroenterology* 1989; 96:1572.
- West MA, Billiar TR, Mazuski JE, et al. Endotoxin modulation of hepatocyte secretory and cellular protein synthesis is mediated by Kupffer cells. *Arch Surg* 1988; 123:1400.
- Andus T, Bauer J, Gerok W. Effects of cytokines on the liver. *Hepatology* 1991; 13:364.
- Gauldie J, Richards C, Harnish S, Lansdrop P, Baumann H. Interferon beta-2/B-cell stimulatory factor type-2 shares identity with monocyte-derived hepatocyte-stimulating factor and regulates the acute phase protein response in liver cells. *Proc Natl Acad Sci USA* 1987; 84:7251.
- Callery MP, Kamei T, Mangino MJ, Flye MW. Kupffer cell prostaglandin E₂ production is amplified during hepatic regeneration. *Hepatology* 1991; 14:368.
- Callery MP, Kamei T, Flye MW. Kupffer cell tumor necrosis factor- α production is suppressed during liver regeneration. *J Surg Res* 1991; 50:515–519.
- Fausto N. New perspectives on liver regeneration. *Hepatology* 1986; 6:326.
- Mangino MJ, Brunt EM, Von Doerson P, Anderson CB. Effects of the thromboxane synthesis inhibitor CGS-12970 on experimental acute renal allograft rejection. *J Pharm Exp Ther* 1989; 248:23–28.
- Callery MP, Mangino MJ, Flye MW. A biological basis for Kupffer cell modification of immune reactivity to portal venous antigen. *Surgery* 1991; 110:221–230.
- Decker T, Lohmann-Matthes ML, Karck V, Peters T, Decker K. Comparative study of cytotoxicity, tumor necrosis factor and prostaglandin release after stimulation of rat Kupffer cells, murine Kupffer cells and urine inflammatory macrophages. *J Leukocyte Biol* 1989; 45:139.
- Barbul A. Arginine: Biochemistry, physiology and therapeutic implications. *JPEN* 1986; 10:227.
- Cornell RP. Restriction of gut-derived endotoxin impaired DNA synthesis for liver regeneration. *Am J Physiol* 1985; 249:R563.
- Ikebuchi K, Wong G, Clark S. Interleukin-6 enhancement of Interleukin-3 dependent proliferation of multipotential hemopoietic progenitors. *Proc Natl Acad Sci USA* 1987; 84:9035–9041.
- Takai Y, Wong GG, Clark SC. B cell stimulatory factor-2 is involved in the differentiation of cytotoxic T lymphocytes. *J Immunol* 1988; 140:508–512.
- Andreis PG, Whitfield JF, Armato U. Stimulation of DNA synthesis and mitosis of hepatocytes in primary cultures of neonatal rat liver by arachidonic acid and prostaglandins. *Exp Cell Res* 1981; 134:265.
- Miura Y, Fukui N. Prostaglandins as possible triggers for liver regeneration after partial hepatectomy. *Cell Mol Biol* 1979; 25: 179.

DISCUSSION

DR. ROBERT ZEPPA (Miami, Florida): Dr. Bland, Dr. Jones, colleagues, I am indebted to Dr. Flye for his invitation to discuss this paper under the assumption that I know something about cytokines. That is in error. I think Wayne asked me to comment on this because 20 years ago Joe Levi and I were studying the regenerative problem of rodent liver. And, of course, in paired *in situ* perfused animals, we discovered that at 24 hours, as Dr. Flye has pointed out, there came into the circulation media some substance that we were never able to characterize, which had an effect on normal liver if they were cross-circulated, in that DNA synthesis went up at that 24-hour period in both livers. That is by way of some historical background. But what I would like to ask Dr. Flye about these experiments is, if in fact the animals are pretreated with indomethacin, what happens then to your pellet of Kupffer cells? This is fascinating and reminds me a little of the presentation that Dick Simmons gives in which he calls it "pillow talk in the liver," that is, the communications between the Kupffer cells and the hepatocytes, which appear now more and more, thanks to the work of Dr. Flye and his group, to be one of the most important factors that we have in terms of how the liver responds to a variety of stimuli, not merely regeneration. Thank you.

DR. COURTNEY M. TOWNSEND, JR. (Galveston, Texas): Dr. Bland, Dr. Jones, Fellows and Guests, Dr. Flye and his colleagues have long been interested in the problem of hepatic regeneration and the interaction of immunologically competent cells with hepatocytes. This study, I think, provides more information into the mechanisms by which Kupffer cell functions are regulated. It appears that there is a tight autocrine control, at least of prostaglandin, on interleukin-6 (IL-6) production.

What are the signals that are at play here? That is, what turns the Kupffer cells on to begin increasing their responsiveness to make IL-6 in response to lipopolysaccharide? Also, does a reciprocal relationship exist? That is, if you were to decrease IL-6, would you increase prostaglandin production? And is there any evidence that exogenously added prostaglandin would further affect the level of IL-6 produced? And, finally, do you know whether the mechanism of action of prostaglandin on IL-6 production is direct or indirect? And if it is indirect, what possible second messengers are involved? Thank you very much.

DR. GEORGE PARKER (Richmond, Virginia): Dr. Bland, Dr. Jones. Like Dr. Zeppa, I am not sure why I was asked to discuss this paper. I think the only reason is that 19 years ago Wayne Flye was my first senior

resident when I was an intern, and he still treats me the same. I have a couple of questions and a couple of comments.

The comment would be I think it is a very elegantly designed and carried out study with all the proper controls, so I think the data are incontrovertible.

The first question I would have is, do you have any idea if the amount of interleukin-6 that is being produced would be sufficient to induce hepatocyte stimulation and mitosis? Second, you mentioned about the factors of the Kupffer cell inducing regeneration or helping in the regenerative process. What is different about certain injuries where there is just fibrosis *versus* certain injuries where there is regeneration? And would you speculate on the possible role of the Kupffer cell there? Thank you very much.

DR. WAYNE FLYE (Closing discussion): I would like to thank the discussants. Dr. Zeppa, you and Dr. Bernard Fisher earlier performed pioneering work in liver regeneration. Obviously, this is a much more complex phenomenon than can simply be explained by the effect of cytokines. Cytokines appear to regulate, more than initiate, regeneration. If you add PGE₂ to hepatocytes in culture, you can get increased DNA proliferation. Addition of indomethacin blocks prostaglandin synthesis and retards the course of liver regeneration. High doses of PGE₂ in vivo will result in diarrhea and animal death.

Dr. Townsend, you ask what signals are important for regeneration. In vitro addition of interleukin-6 downregulates prostaglandin E₂ production, as does tumor necrosis factor and interleukin-1. This may be effected by activation of phospholipase A2 and the mobilization of fatty

acids and arachidonic acid from the cell surface. We know that PGE₂ is an important regulator of cell proliferation, probably by increasing cyclic adenosine monophosphate (cAMP) levels. When we infuse the long-acting 16,16 dimethyl PGE₂, which is not metabolized on first passage through the lung, as are most natural prostaglandins, into a heterotopic heart allograft continuously for 2 weeks, we completely prevent graft rejection. We know that the early recognition phase of rejection is related to how the macrophage presents antigen to effector lymphocytes. We have evidence that the migration of lymphocytes and macrophages through the graft are impaired by PGE₂. Although liver regeneration is a different phenomenon, the mechanism of regulation of second signals is probably similar. Changes in interleukin-6 levels do not cause changes in PGE₂ production, whereas exogenous PGE₂ does decrease interleukin-6 levels.

Although the Kupffer cells are probably normally exposed to low levels of endotoxin, it is only during exposure to higher levels that Kupffer cells significantly increase PGE₂ and cytokine production, *i.e.*, during bacteremia and during the stress of liver regeneration. However, R. P. Cornell has shown that neutralization of the lipid A portion of endotoxin with polymyxin B in normal animals slows liver regeneration. The levels of PGE₂ produced by the Kupffer cells appear adequate to regulate the adjacent hepatocytes.

Dr. Parker, the fibrotic liver does not regenerate well. This is clinically important in the patient with cirrhosis undergoing liver resection. The hepatocyte within the milieu of the fibrotic tissue behaves differently than when it can function in a normal hepatic environment.

I would like to thank the discussants for their remarks and the Association for the privilege of presenting this work.