Corrections

COMMENTARY. For the article "On the sequencing and assembly of the human genome," by Eugene W. Myers, Granger G. Sutton, Hamilton O. Smith, Mark D. Adams, and J. Craig Venter, which appeared in number 7, April 2, 2002, of *Proc. Natl. Acad. Sci. USA* (**99,** 4145–4146; First Published March 19, 2002; 10.1073/pnas.092136699), in ref. 7, the journal name "*Nat. Rev. Neurosci.*" should be "*Nat. Neurosci.*"

www.pnas.org/cgi/doi/10.1073/pnas.132227099

BIOPHYSICS. For the article "Tryptophan zippers: Stable, monomeric β-hairpins," by Andrea G. Cochran, Nicholas J. Skelton, and Melissa A. Starovasnik, which appeared in number 10, May 8, 2001, of *Proc. Natl. Acad. Sci. USA* (**98,** 5578–5583; First Published May 1, 2001; 10.1073/pnas.091100898), the authors note the following. Quantitative analysis of the ^{1}H chemical shifts using the Sander module of AMBER 6.0 reveals that the frequencies of the Hβ and Hε3 resonances of Trp4 and Trp11 (for trpzips 1 and 2) and Trp5 and Trp14 (for trpzip4) are inconsistent with the side chain orientations previously determined. Instead, refinement of the structures based not only on NOE-derived distance restraints and dihedral angle restraints, but also on ^{1}H chemical shift-based restraints, indicates that the side chains for these residues actually reside primarily in the 180° χ1 rotamer well, not the −60° rotamer previously indicated. Refined coordinates are available in the Protein Data Bank, www.rcsb.org [PDB ID codes 1LE0 (trpzip1), 1LE1 (trpzip2), and 1LE3 (trpzip4)]. The updated coordinates are very similar to those determined previously; as before, for all three peptides, the two strands are highly twisted. The important difference, however, is that each pair of cross-strand tryptophan rings now shows edge-to-face packing against one another (see Fig. 1) that is conserved among all three trpzip peptides regardless of turn-type.

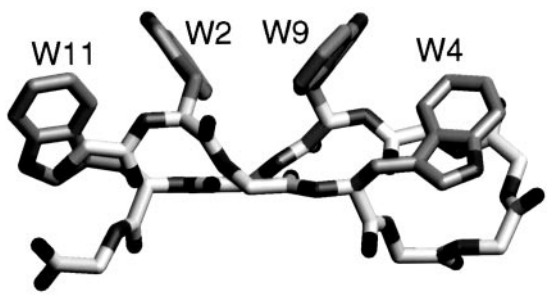

Fig. 1. Representative structure of trpzip1 refined using ^{1}H chemical shift-based restraints. Edge-to-face packing is observed for Trp11/Trp2 and Trp4/Trp9 cross-strand pairs.

www.pnas.org/cgi/doi/10.1073/pnas.132249099

BIOPHYSICS. For the article "Fluorescence polarization of green fluorescence protein," by Shinya Inoué, Osamu Shimomura, Makoto Goda, Mykhailo Shribak, and P. T. Tran, which appeared in number 7, April 2, 2002, of *Proc. Natl. Acad. Sci. USA* (**99,** 4272–4277), the last closing parenthesis on Eqs. **3** and **4** should appear at the end of the equations. The correct equations appear below.

$$I/I_0 = 0.5(\alpha_{ss} + (\alpha_{ss} + \alpha_{sp} - 2\alpha_{ss})\cos^2\phi + (\alpha_{pp} + \alpha_{ss} - \alpha_{ps} - \alpha_{sp})\cos^4\phi). \quad [3]$$

$$I/I_0 = 0.5(\alpha_{ps} + (\alpha_{pp} + \alpha_{ss} - 2\alpha_{ps})\cos^2\phi - (\alpha_{pp} + \alpha_{ss} - \alpha_{ps} - \alpha_{sp})\cos^4\phi). \quad [4]$$

www.pnas.org/cgi/doi/10.1073/pnas.132280599

MEDICAL SCIENCES. For the article "HLTF gene silencing in human colon cancer," by Helen R. Moinova, Wei-Dong-Chen, Lanlan Shen, Dominic Smiraglia, Joseph Olechnowicz, Lakshmeswari Ravi, Lakshmi Kasturi, Lois Myeroff, Christoph Plass, Ramon Parsons, John Minna, James K. V. Willson, Sylvan B. Green, Jean-Pierre Issa, and Sanford D. Markowitz, which appeared in number 7, April 2, 2002, of *Proc. Natl. Acad. Sci. USA* (**99,** 4562–4567; First Published March 19, 2002; 10.1073/pnas.062459899), the author name "Wei-Dong-Chen" appeared incorrectly due to a printer's error. The correct name is "Wei-Dong Chen." The online version has been corrected.

www.pnas.org/cgi/doi/10.1073/pnas.122187999

NEUROBIOLOGY. For the article "Wild-type and mutated presenilins 2 trigger p53-dependent apoptosis and down-regulate presenilin 1 expression in HEK293 human cells and in murine neurons," by Cristine Alves da Costa, Erwan Paitel, Marc P. Mattson, Robert Amson, Adam Telerman, Karine Ancolio, and Frédéric Checler, which appeared in number 6, March 19, 2002, of *Proc. Natl. Acad. Sci. USA* (**99,** 4043–4048), the authors note that the author name "Marc P. Mattson" should be "Mark P. Mattson." The online version has been corrected.

www.pnas.org/cgi/doi/10.1073/pnas.122187899

NEUROBIOLOGY. For the article "Sustained stimulation shifts the mechanism of endocytosis from dynamin-1-dependent rapid endocytosis to clathrin- and dynamin-2-mediated slow endocytosis in chromaffin cells," by Cristina R. Artalejo, Abdeladim Elhamdani, and H. Clive Palfrey, which appeared in number 9, April 30, 2002, of *Proc. Natl. Acad. Sci. USA* (**99,** 6358–6363; First Published April 16, 2002; 10.1073/pnas.082658499), a label at the top of Fig. 3 was omitted due to a printer's error. The complete figure and its legend appear below.

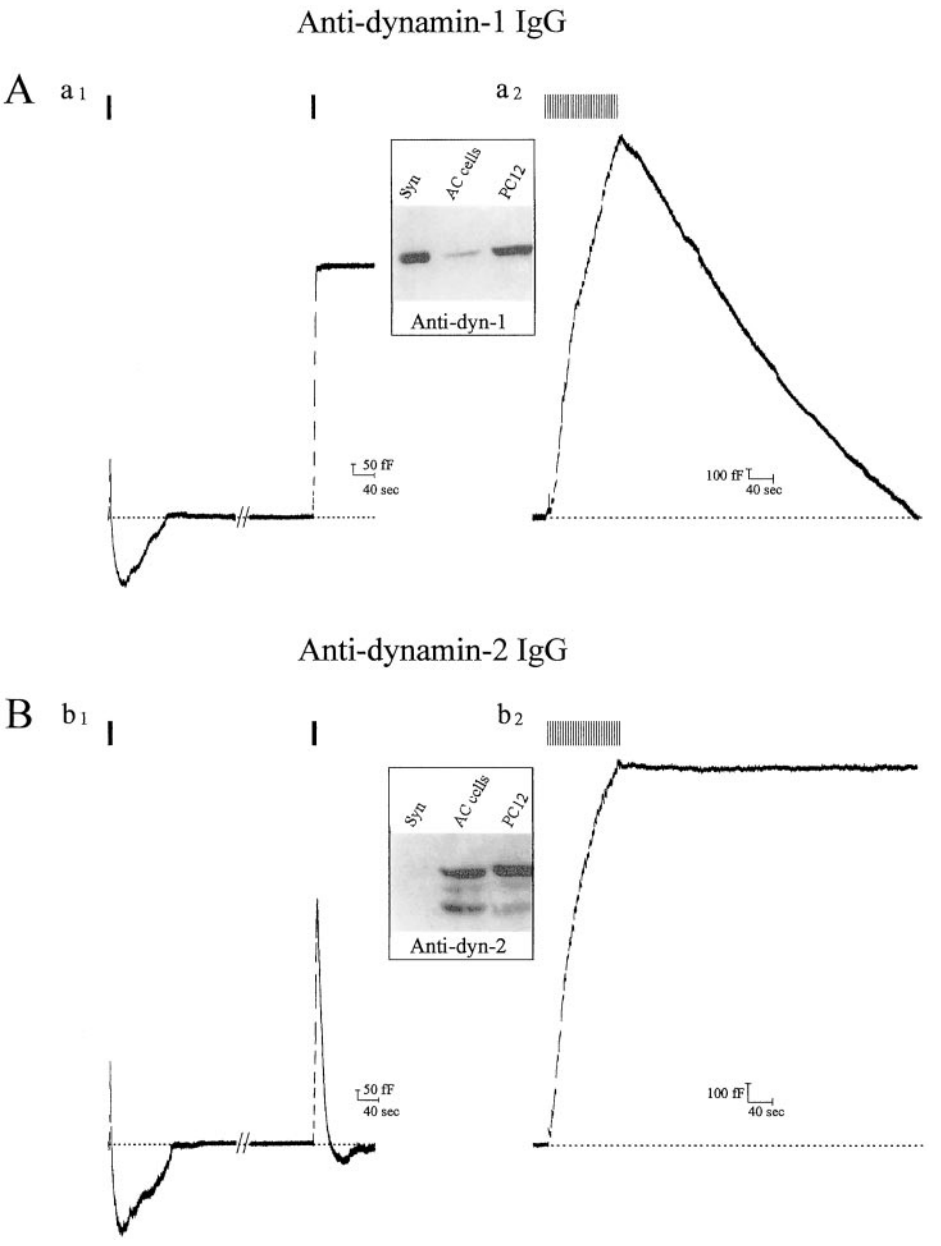

Fig. 3. Dynamin-1 mediates RE, whereas dynamin-2 mediates SE. C_m records from cells in which either affinity-purified (*A*) antidynamin-1-specific IgG or (*B*) antidynamin-2-specific IgG, both at 1 mg/ml, were introduced into calf chromaffin cells followed by transient or sustained stimulation. In *A*, note that antidynamin-1 IgG inhibits RE (*a*1) but has no effect on SE (*a*2). In *B*, antidynamin-2 IgG has no effect on RE (*b*1) but blocks SE (*b*2). Note in *Bb*1 that two rounds of exocytosis/RE occur, whereas in *Aa*1 the second round of RE is blocked after the antibody has diffused into the cell (in the first round, RE is normal because insufficient antibody has diffused into the cell; the extent of the first exocytosis in *Aa*1 and *Bb*1 appears smaller because of simultaneous endocytosis that is largely absent in the second round). (*Insets*) Reactivity of antidynamin-1 (*c*1)- and -2 (*c*2)-specific antibodies with lysates from rat brain synaptosomes (Syn; 10 μg of protein); calf chromaffin (AC) cells (100 μg), and PC12 cells (100 μg); immunoblots were performed as described (9) and developed by using enhanced chemiluminescence.

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