Plasmid-Mediated Carbapenem-Hydrolyzing β-Lactamase KPC in a *Klebsiella pneumoniae* Isolate from France

Carbapenem-hydrolyzing β -lactamases can be metallo- β -lactamases (7), expanded-spectrum oxacillinases (8), or Ambler class A enzymes (1, 2, 4, 6, 9–12). The class A KPC β -lactamases hydrolyze all β -lactams except for cephamycins and have been rarely reported in studies of enterobacterial species (*Klebsiella pneumoniae*, *Klebsiella oxytoca*, *Salmonella enterica*, *Enterobacter* sp.) found in New England and the mid-Atlantic region of the United States; when they are reported, they are found to come from New York City hospitals in particular (2, 4, 6, 9–12). We report here a *K. pneumoniae* isolate from Paris that produced β -lactamase KPC-2.

An 80-year-old man with a 5-year history of prostatic carcinoma and metastasis was admitted in February 2005 at the Cochin hospital in Paris for acute urine retention in the right nephrostomy. The bilateral nephrostomy was performed in December 2004 at a New York City hospital. No hospitalization or antibiotic treatment is known to have taken place during the period between the patient's stay in New York and his emergency hospitalization in Paris. *K. pneumoniae* YC was isolated from urine and blood cultures and, according to disk diffusion susceptibility testing, was resistant or of intermediate susceptibility to all antibiotics except colistin and fosfomycin (3). No other *K. pneumoniae* isolates with similar antibiotic resistance patterns were recovered from the Cochin hospital during this same period of time.

The MICs of β -lactams (3) for *K. pneumoniae* YC confirmed the disk diffusion results, with the MICs of expanded-spectrum cephalosporins and carbapenems being only slightly modified after the addition of clavulanic acid (Table 1). A crude β -lactamase extract of that isolate had significant imipenem hydrolysis activity (0.37 μ mol of imipenem/min/mg of total protein) (5, 8).

TABLE 1. MICs of β -lactams for K. pneumoniae YC, the transconjugant strain, and the reference strain

β-Lactam(s) ^a	MIC (μg/ml) for:		
	K. pneumoniae YC	Tc E. coli J53	E. coli J53
Amoxicillin	>256	>256	4
Amoxicillin + CLA	32	32	4
Ticarcillin	>256	>256	2
Ticarcillin + CLA	>256	256	2
Piperacillin	>256	>256	1
Piperacillin + TZB	256	64	1
Cephalothin	>256	>256	4
Cefoxitin	64	2	1
Cefotaxime	64	4	0.06
Cefotaxime + CLA	16	2	0.06
Ceftazidime	64	4	0.06
Aztreonam	>256	16	0.03
Aztreonam + CLA	>256	8	0.03
Imipenem	4	8	0.12
Imipenem + CLA	2	2	0.12
Meropenem	2	2	0.03
Meropenem + CLA	1	0.25	0.03

 $[^]a$ CLA, clavulanic acid at a fixed concentration of 2 μ g/ml; TZB, tazobactam at a fixed concentration of 4 μ g/ml.

Plasmid analysis detected two plasmids of >100 kb and ca. 75 kb in isolate YC. Using ampicillin (100 µg/ml) and sodium azide (100 µg/ml; Sigma-Aldrich, Saint-Quentin-Fallavier, France) as selective agents and sodium azide-resistant Escherichia coli J53 as the recipient strain, the ca.-75-kb plasmid was self-conjugative and conferred β-lactam resistance patterns for E. coli transconjugants similar with respect to the MICs obtained, except for those for cefoxitin, cefotaxime, ceftazidime, and aztreonam, which were lower than those for the parental strain (Table 1). A β-lactamase extract from a transconjugant culture subjected to analytical isoelectric focusing (8) identified two β-lactamases with pI values of 5.4 and 6.8. Using primers for the detection of Ambler class A and class B β-lactamase genes (7, 8), PCR experiments followed by sequencing identified a β-lactamase gene coding for the carbapenemase KPC-2 (pI 6.8) and the narrow-spectrum TEM-1 (pI 5.4), which were also identified from K. pneumoniae YC.

Then, by the use of a series of successive PCR primers, the 2.8-kb surrounding sequences of the $bla_{\rm KPC-2}$ gene were found to be identical to those surrounding the same $bla_{\rm KPC-2}$ gene from a *Salmonella* isolate from Maryland (6).

This report identified a carbapenem-hydrolyzing β -lactamase KPC in France that likely resulted from an intercontinental transfer of the KPC producer from the United States. Up to the time of this discovery, plasmid-mediated KPC enzymes had been a unique feature of the United States, whereas most other broad-spectrum antibiotic resistance genes have been reported in the United States after their initial discoveries elsewhere. The identification of KPC enzymes is worrisome, since they hydrolyze expanded-spectrum cephalosporins and carbapenems and thus could jeopardize therapy for serious infections caused by major nosocomial pathogens. In addition, their detection, based on careful interpretation of reduced carbapenem susceptibility in *Enterobacteriaceae*, remains difficult.

This work was funded by grants from the Ministère de l'Education Nationale et de la Recherche (UPRES-EA3539), Université Paris XI, France; from the European Community (6th PCRD, LSHM-CT-2003-503335); and from the Fondation pour La Recherche Médicale and Université Paris V, Paris, France.

REFERENCES

- Aubron, C., L. Poirel, R. J. Ash, and P. Nordmann. 2005. Carbapenemaseproducing *Enterobacteriaceae*, U.S. rivers. Emerg. Infect. Dis. 11:260–264.
- Bradford, P. A., S. Bratu, C. Urban, M. Visalli, N. Mariano, D. Landman, J. J. Rahal, S. Brooks, S. Cebular, and J. Quale. 2004. Emergence of carbapenem-resistant Klebsiella species possessing the class A carbapenemhydrolyzing KPC-2 and inhibitor-resistant TEM-30 β-lactamases in New York City. Clin. Infect. Dis. 39:55-60.
- Clinical and Laboratory Standards Institute. 2005. Performance standards for antimicrobial susceptibility testing; 15th informational supplement. M100-S15. Clinical and Laboratory Standards Institute, Wayne, Pa.
- Hossain, A., M. J. Ferraro, R. M. Pino, R. B. Dew III, E. S. Moland, T. J. Lockhart, K. S. Thomson, R. V. Goering, and N. D. Hanson. 2004. Plasmidmediated carbapenem-hydrolyzing enzyme KPC-2 in an *Enterobacter* sp. Antimicrob. Agents Chemother. 48:4438–4440.
- Matagne, A., A. M. Misselyn-Baudouin, B. Joris, B. Erpicum, B. Granier, and J. M. Frere. 1990. The diversity of the catalytic properties of class A β-lactamases. Biochem. J. 265:131–146.
- Miriagou, V., L. S. Tzouvelekis, S. Rossiter, E. Tzelepi, F. J. Angulo, and J. Whichard. 2003. Imipenem resistance in a Salmonella clinical strain due to

- 4424
 - plasmid-mediated class A carbapenemase KPC-2. Antimicrob. Agents Chemother. 47:1297–1300.
- Nordmann, P., and L. Poirel. 2002. Emerging carbapenemases in Gramnegative aerobes. Clin. Microbiol. Infect. 8:321–331.
- Poirel, L., C. Héritier, V. Tolun, and P. Nordmann. 2004. Emergence of oxacillinase-mediated resistance to imipenem in *Klebsiella pneumoniae*. Antimicrob. Agents Chemother. 48:15–22.
- Smith Moland, E., N. D. Hanson, V. L. Herrera, J. A. Black, T. J. Lockhart, A. Hossain, J. A. Johnson, R. V. Goering, and K. S. Thomson. 2003. Plasmidmediated, carbapenem-hydrolysing β-lactamase, KPC-2, in *Klebsiella pneumoniae*. J. Antimicrob. Chemother. 51:711–714.
- 10. Woodford, N., P. M. Tierno, Jr., K. Young, L. Tysall, M. F. I. Papelou, E. Ward, R. E. Painter, D. F. Suber, D. Shungu, L. L. Silver, K. Inglima, J. Kornblum, and D. M. Livermore. 2004. Outbreak of Klebsiella pneumoniae producing a new carbapenem-hydrolyzing class A β-lactamase, KPC-3, in a New York medical center. Antimicrob. Agents Chemother. 48:4793–4799.
- 11. Yigit, H., A. M. Queenan, G. J. Anderson, A. Domenech-Sanchez, J. W. Biddle, C. D. Steward, S. Ablerti, K. Bush, and F. C. Tenover. 2001. Novel carbapenem-hydrolyzing β-lactamase KPC-1 from a carbapenem-resistant strain of Klebsiella pneumoniae. Antimicrob. Agents Chemother. 45:1151–1161.
- 12. Yigit, H., A. M. Queenan, J. K. Rasheed, J. W. Biddle, A. Domenech-Sanchez, S. Ablerti, K. Bush, and F. C. Tenover. 2003. Carbapenem-resistant strains of *Klebsiella oxytoca* harboring carbapenem-hydrolyzing β-lactamase KPC-2. Antimicrob. Agents Chemother. 47:3881–3889.

Thierry Naas Patrice Nordmann*

Service de Bactériologie-Virologie Hôpital de Bicêtre Assistance Publique/Hôpitaux de Paris Faculté de Médecine Paris-Sud Université Paris XI 94275 Le Kremlin-Bicêtre, France

Gérard Vedel Claire Poyart

Service de Bactériologie Hôpital Cochin Assistance Publique/Hôpitaux de Paris Faculté de Médecine René Descartes Université Paris V 75014 Paris, France

*Phone: 33-1-45-21-36-32 Fax: 33-1-45-21-63-40

E-mail: nordmann.patrice@bct.ap-hop-paris.fr