## Autonomic control of adrenal function

## A. V. EDWARDS AND C. T. JONES

The Physiological Laboratory, University of Cambridge and Laboratory of Cellular and Developmental Physiology, Institute for Molecular Medicine, Oxford, UK

(Accepted 22 February 1993)

## ABSTRACT

Recent studies of adrenal function in conscious calves are reviewed. These have involved collecting the whole of the adrenal effluent blood from the right adrenal gland at intervals and, where necessary, prior functional hypophysectomy by destruction of the pituitary stalk under general halothane anaesthesia 3 d previously. The adrenal medulla was found to release numerous neuropeptides, in addition to catecholamines, in response to stimulation of the peripheral end of the right splanchnic nerve, which was carried out below behavioural threshold. Many of these responses were enhanced by stimulating intermittently at a relatively high frequency. Intra-aortic infusions of a relatively low dose of acetylcholine (4.5 nmol min<sup>-1</sup> kg<sup>-1</sup>) elicited similar responses. In the adrenal cortex, agonists which either potentiated the steroidogenic response to ACTH or exerted a direct steroidogenic action included VIP, CGRP, CRF and ACh acting via muscarinic receptors. Stimulation of the peripheral end of the right splanchnic nerve strongly potentiated the steroidogenic response to ACTH and there is compelling evidence that the innervation normally plays an important part in cortisol secretion.

## INTRODUCTION

It has been widely accepted that the adrenal medulla secretes catecholamines in response to activity in exclusively preganglionic sympathetic fibres which fire tonically at low frequencies and release acetylcholine which activates the chromaffin cells via nicotinic receptors. It has also been accepted that the secretion of cortisol from the adrenal cortex is determined solely by the release of adrenocorticotrophin (ACTH) from the anterior pituitary. The object of this review is to assess the extent to which these views need to be reappraised in the light of recent work, and to focus principally on studies carried out on conscious animals (mainly calves) in the Physiological Laboratory at Cambridge.

Briefly, the adrenal medulla releases numerous peptides, in addition to catecholamines, in response to stimulation of the splanchnic sympathetic innervation, and these responses can be enhanced by stimulating intermittently at frequencies well above those at which sympathetic fibres have generally been supposed to fire naturally. Postganglionic neurons have been identified in both the medulla and cortex and intraaortic infusions of small amounts of acetylcholine have been found to elicit adrenal medullary responses principally by activating muscarinic receptors, in marked contrast to many results of in vitro studies. Vasoactive intestinal peptide (VIP) is also present in nerve terminals in the gland and evokes release of catecholamines when administered intra-arterially. For the adrenal cortex agonists which either potentiate the steroidogenic response to ACTH or exert a direct steroidogenic action include VIP, calcitonin gene related peptide (CGRP), corticotrophin releasing factor (CRF) and acetylcholine, acting via muscarinic receptors. Finally, activation of the splanchnic sympathetic innervation strongly potentiates the steroidogenic action of ACTH and there is compelling evidence that the innervation normally plays an important part in the control of cortisol secretion.

## ADRENAL MEDULLARY FUNCTION

The 'adrenal clamp' technique involves removal of the right kidney, cannulation of the right renal vein and insertion of a specially designed clamp along the posterior vena cava in such a way that the jaws divert

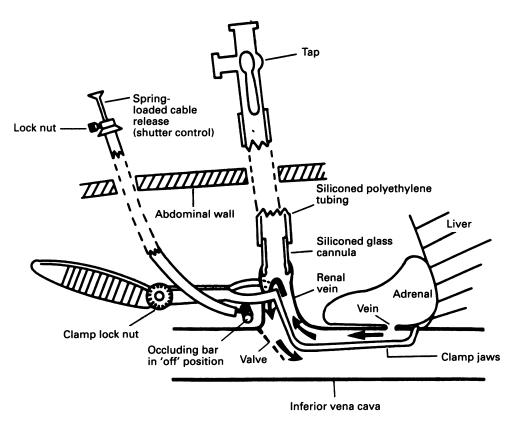


Fig. 1. Diagrammatic representation of the positioning of an 'adrenal clamp' in relation to the adrenal gland and renal vein. (From Edwards et al. 1974, by permission of the Physiological Society.)

the venous effluent adrenal blood back to enter the vena cava via the renal vein (Edwards et al. 1974) (Fig. 1). The operation is carried out under full general anaesthesia, during which appropriate intravascular catheters can be inserted and a fluid electrode attached to the peripheral end of the right splanchnic nerve. Following recovery from anaesthesia, the whole of the adrenal venous effluent can be collected at intervals simply by occluding the channel through the renal vein. Adrenal output can then be quantified precisely in the conscious animal, the bloodflow being estimated gravimetrically.

## Adrenal medullary sensitivity

The adrenal medulla was found to be remarkably resistant to a variety of stresses which readily produced an increase in the output of glucocorticoids from the adrenal cortex. Thus hypoxia failed to elicit a significant increase in catecholamine output unless the arterial  $P_{0_2}$  fell below 15 mmHg (Bloom et al. 1976, 1977), hypercapnia failed to produce an adrenal medullary response unless the arterial  $P_{CO_2}$  rose above 100 mmHg (Bloom et al. 1977) and insulin hypoglycaemia was only effective in this regard when the

arterial plasma glucose concentration had fallen below 2 mmol  $l^{-1}$  for a prolonged period (Bloom et al. 1975).

### Responses to splanchnic nerve stimulation

Dopamine- $\beta$ -monooxygenase. The gland responds very readily to stimulation of the peripheral end of the splanchnic nerve, which can be accomplished below any discernible behavioural threshold in conscious animals by employing a fluid electrode that minimises spread of activity. The outputs of both adrenaline and noradrenaline are linearly related to stimulus frequency over the range 2–10 Hz, with peak outputs occurring at 15 Hz. Continuous stimulation at either 4 or 10 Hz produced a small but significant rise in the output of dopamine-\beta-monooxygenase activity (Edwards et al. 1980). The amounts released could not be related either to the frequency of stimulation or to the output of adrenaline or noradrenaline and so proved not to be a reliable index of exocytosis under these conditions, as had previously been suggested (Geffen et al. 1969).

*Enkephalins*. Opioid peptides are present in the adrenal medullae in many species in the form of a proenkephalin precursor possessing 6 copies of metenkephalin and 1 copy of leuenkephalin, the pro-

cessing of which is under neural control (Schultzberg et al. 1978; Costa et al. 1979; Stern et al. 1979; Viveros et al. 1979). They are localised both in the chromaffin granules of adrenergic, but not noradrenergic, cells (Livett et al. 1982; Pelto-Huikko et al. 1982; Roisin et al. 1983), where they are synthesised (Stern et al. 1979; Chang et al. 1982) and within nerve terminals (Schultzberg et al. 1978). Electrical stimulation of the peripheral end of the splanchnic nerve causes the release of enkephalins from the adrenal gland (Hexum et al. 1980; Govoni et al. 1981). In the conscious dog 10-20% haemorrhage results in the release of enkephalins (Engeland et al. 1986) and in the conscious calf enkephalins are released from the gland in response to splanchnic nerve stimulation almost entirely in the form of high molecular weight precursors (Edwards et al. 1986; Bloom et al. 1988). Scrutiny of the profiles of chromatograms of met<sup>5</sup>enkephalin-like immunoreactivity from extracts of the gland showed it was virtually all material with M<sub>4</sub> values above 4000. In contrast, about 30% of the total in adrenal effluent plasma was accounted for by molecules with  $M_{4}$  values of about 4000 (Rossier et al. 1988). This peak might correspond to peptides I, E, B, F, BAM 22 or BAM 18, all of which have been isolated from bovine adrenal (Metters & Rossier, 1984).

The adrenal medulla contains numerous opioid binding sites (Castanas et al. 1985a, b), at least some of which are located postsynaptically on adrenal chromaffin cells (Kumakura et al. 1980), but their physiological functions and those of adrenal opioids have yet to be elucidated. It seems unlikely that endogenous adrenal opioids could act on adrenal chromaffin cells to modify their responses to prostaglandins (Marley et al. 1988) or the inositol phosphate second-messenger system (Bunn et al. 1988), or to affect their responses to histamine or angiotensin (Marley & Livett, 1986; Marley & Bunn, 1988). They have, however, been found to inhibit the adrenal catecholamine response to nicotine in isolated chromaffin cells (Marley et al. 1986; Marley & Livett, 1987) and the isolated, perfused adrenal gland of the rat (Chen & Dixon, 1990). In the conscious calf model, there is evidence that enkephalins released in response to stimulation of the peripheral end of the splanchnic nerve are autoinhibitory as their release was significantly enhanced by pretreatment with naloxone and the proportion of high molecular weight enkephalin molecules preferentially increased (Edwards & Jones, 1989a). Splanchnic nerve stimulation resulted in an abrupt rise in the output of both free and total met<sup>5</sup>-enkephalin-like immunoreactivity from

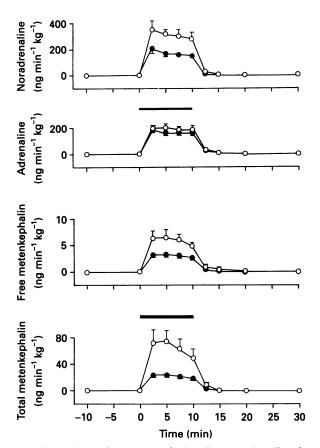


Fig. 2. Comparison of the output of adrenaline, noradrenaline, free and total met<sup>5</sup>-enkephalin-like immunoreactivity from the right adrenal medulla in response to stimulation of the peripheral end of the right splanchnic nerve in 7 conscious calves at either 4 Hz continuously ( $\bigcirc$ ), or at 40 Hz for 1 s at 10 s intervals ( $\bigcirc$ ), for 10 min. Vertical bars, s.E. of each mean value where these exceed the size of the symbol. Horizontal bar, duration of stimulation. (Modified from Bloom et al. 1988, by permission of the Physiological Society.)

the adrenal gland in conscious calves which was substantially potentiated by stimulating in bursts (Bloom et al. 1988) (Fig. 2). This pattern of stimulation also increased the proportion released in high molecular weight form.

Catecholamines. Adrenal medullary responses to splanchnic nerve stimulation are critically dependent on the pattern of stimulation that is applied. Stimulation in bursts at relatively high frequencies significantly potentiates the output of catecholamines, and enkephalins (Bloom et al. 1988) (Fig. 2). Thus stimulation at 40 Hz for 1 s at 10 s intervals elicited a substantial and significant increase in the output of noradrenaline, above that which occurred in response to stimulation at 4 Hz continuously, in conscious calves. This effect far exceeded the rise in adrenaline output and so produced a significant shift in the noradrenaline: adrenaline ratio, in favour of noradrenaline.

In the dog several groups have reported an increase in the proportion of noradrenaline released from the adrenal gland in response to continuous splanchnic nerve stimulation as the stimulus frequency was increased (Rapela & Covian, 1954; Klepping, 1956; Rapela, 1956). However, the changes were comparatively small and interpretation of the results is complicated by the fact that experiments were carried out under anaesthesia. In certain species, notably the cat, the proportions of adrenaline and noradrenaline released from the adrenal have been found to vary with the particular type of stimulus employed (von Euler, 1956). Thus adrenaline is released preferentially in response to hypoglycaemia in cats (Dunér, 1954) and sheep (Crone, 1965). In the cat, von Euler & Folkow (1953) originally showed that carotid occlusion favours the release of noradrenaline whereas stimulation of either the brachial or sciatic plexus favours the release of adrenaline. The effect of carotid occlusion is in accord with more recent findings that in cats, but not dogs, reduction of the pressure in the carotid sinus preferentially releases noradrenaline whereas stimulation of the carotid chemoreceptors preferentially releases adrenaline (Anichkov et al. 1960; Critchley et al. 1980), as does asphyxia (Redgate & Gellhorn, 1953). This difference between the adrenal medullary responses of (anaesthetised) cats and dogs is further exemplified by the finding that the relative proportions of the 2 amines that are secreted in response to electrical stimulation of the hypothalamus vary in accordance with the position of the stimulating electrode in cats but not dogs (Redgate & Gellhorn, 1953; Folkow & von Euler, 1954; Goldfien & Ganong, 1962; Malmejac, 1964). Differential release of one or other of these catecholamines in response to such stimuli as hypoglycaemia or asphyxia could well be due to differential sensitivity of the chromaffin cells themselves, to a lack of glucose or oxygen, or to an excess of carbon dioxide. It is more difficult to understand how differential release in response to electrical stimulation of the hypothalamus could be mediated peripherally. However, the results obtained in the conscious calf show that this could be achieved simply by altering the pattern of efferent splanchnic nerve activity.

Corticotrophin releasing factor. The adrenal gland has also been reported to contain corticotrophinreleasing factor-like immunoreactivity (CRF) (Suda et al. 1984; Hashimoto et al. 1984) which is released from the gland in quite substantial amounts in response to haemorrhage in conscious dogs (Bruhn et al. 1987). In conscious calves, stimulation of the peripheral end of the right splanchnic nerve for 10 min increased the mean output of CRF progressively, so that it had risen about 20-fold, to a peak incremental value of  $24 \pm 4$  pmol min<sup>-1</sup> kg<sup>-1</sup> at 10 min (Edwards & Jones, 1988). This response was significantly increased by stimulating in bursts at 40 Hz for 1 s at 10 s intervals, which raised the mean adrenal CRF output by  $44 \pm 7 \text{ pmol min}^{-1} \text{ kg}^{-1}$  at 10 min (P < 0.05). In hypophysectomised calves, administration of synthetic adrenocorticotrophic hormone (ACTH<sub>1-24</sub>) at a dose of 5 ng min<sup>-1</sup> kg<sup>-1</sup> reduced the output of adrenal CRF in response to splanchnic nerve stimulation by about 50%. Release of this peptide from the adrenal gland thus appears to be susceptible to inhibition via the ACTH-adrenal cortical axis, just like its pituitary counterpart, to which it is also structurally closely related (Edwards & Jones, 1988).

## Responses to acetylcholine

Muscarinic stimulation of catecholamine release was first described by Feldberg et al. (1934) in the cat and has since been amply confirmed both in this (Douglas & Poisner, 1965; Lee & Trendelenberg, 1967; Kirpekar et al. 1982) and other species, including the gerbil (Douglas et al. 1967), rat (Yoshikazi, 1975; Wakade & Wakade, 1983) and guinea pig (Role & Perlman, 1983). In each of these, however, it has generally been supposed to have a trivial effect by comparison with the nicotinic mechanism. In other species release of catecholamines has been ascribed to activation of just one type of cholinergic receptor, supposedly exclusively muscarinic in the chick adrenal (Ledbetter & Kirschner, 1975; Knight & Baker, 1986) and nicotinic in the bovine adrenal (see e.g. Ballesta et al. 1989). In the conscious calf, this question has been examined employing an infusion of acetylcholine at a dose of 4.5 nmol min<sup>-1</sup> kg<sup>-1</sup> into the aorta above the origin of the adrenal arterial supply, in functionally hypophysectomised animals to prevent fluctuations in endogenous ACTH. Under these conditions acetylcholine produced an abrupt rise in the outputs of adrenaline and noradrenaline which reached a peak at 2.5 min and declined steadily thereafter for the duration of the 10 min infusion (Jones et al. 1991) (Fig. 3). Unlike the response to splanchnic nerve stimulation, the output of adrenaline was substantially greater than that of noradrenaline. Both responses are greatly reduced following the administration of atropine (roughly in the same proportion) but were by no means abolished thereby (Fig. 3). There was an abrupt rise in the output of enkephalin peptides from the right adrenal gland in response to acetylcholine. Like the response to nerve stimulation, large mol-

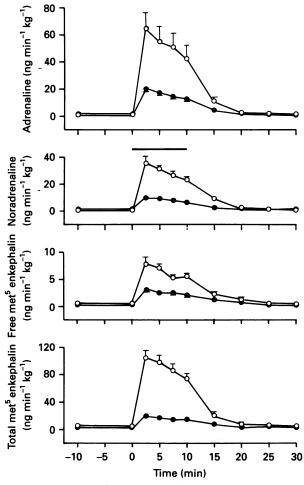


Fig. 3. Comparison of the outputs of adrenaline, noradrenaline, free and total enkephalin-like immunoreactivity from the right adrenal medulla in response to an intra-aortic infusion of acetylcholine (4.5 nmol min<sup>-1</sup> kg<sup>-1</sup> for 10 min), during an i.v. infusion of exogenous ACTH (2 ng min<sup>-1</sup> kg<sup>-1</sup>) in 4 conscious functionally hypophysectomised calves, in the presence ( $\bigcirc$ ) and absence ( $\bigcirc$ ) of atropine (0.2 mg kg<sup>-1</sup>). Vertical bars, s.E. of each mean value where these exceed the size of the symbol. Horizontal bar, duration of infusion. (Modified from Jones et al. 1991, by permission of the Physiological Society.)

ecular forms predominated and exogenous ACTH was found to inhibit the release of free, but not total, met<sup>5</sup>-enkephalin in response to acetylcholine (Jones et al. 1991) just as it does during splanchnic nerve stimulation (Edwards et al. 1986; Edwards & Jones, 1987*a*). The output of both forms of met<sup>5</sup>-enkephalin from the adrenal gland was substantially reduced, but not completely suppressed, by pretreatment with atropine (0.2 mg kg<sup>-1</sup>), both in the presence (Fig. 3) and the absence of ACTH.

Acetylcholine also produced an abrupt increase in the output of CRF from the right adrenal gland, which rose from  $0.8 \pm 0.1$  to a peak mean value of  $15.2 \pm 1.4$  pg min<sup>-1</sup> kg<sup>-1</sup> at 2.5 min. This response was significantly potentiated in the presence of exogenous

Table Average mean right adrenal outputs of catecholamines, enkephalins and of CRF in response to an intra-aortic infusion of acetylcholine (4.5 nmol min<sup>-1</sup> kg<sup>-1</sup> for 10 min) in conscious calves receiving exogenous ACTH (2 ng min<sup>-1</sup> kg<sup>-1</sup> i.v.)

	Α	В	С	D
Adrenaline (ng min <sup>-1</sup> kg <sup>-1</sup> )	$51.3 \pm 3.2$	15.7±1.7	40.6±2.9	56.3
Noradrenaline ng min <sup>-1</sup> kg <sup>-1</sup> )	30.0±0.2	8.0±0.9	$25.4 \pm 2.0$	33.4
Free met <sup>5</sup> -enkephalin (ng min <sup>-1</sup> kg <sup>-1</sup> )	6.9±0.8	$2.6\pm0.2$	$6.9\pm0.8$	9.5
Total met <sup>3</sup> -enkephalin (ng min <sup>-1</sup> kg <sup>-1</sup> )	87.6±7.1	16.6±1.3	76.3±6.9	92.9
CRF (pg min <sup>-1</sup> kg <sup>-1</sup> )	22.0±1.5	4.8±0.4	$22.2 \pm 0.5$	27.0

A, without blockade; B, atropine  $(0.2 \text{ mg kg}^{-1})$ ; C, hexamethonium (> 10 mg kg<sup>-1</sup>); D, B+C.

(From Jones & Edwards, 1992, by permission of the Physiological Society.)

ACTH 2 (ng min<sup>-1</sup> kg<sup>-1</sup>, i.v.) and the values during the infusion were consistently higher under these latter conditions. It was also substantially reduced, but not entirely abolished, following the administration of atropine (0.2 mg kg<sup>-1</sup>, i.v.).

These results, which strongly implicate muscarinic receptors in the release of catecholamines, among other agonists, from the adrenal medulla in the conscious calf conflict with the results of numerous in vitro studies, both on freshly isolated (Schneider et al. 1979; Oka et al. 1982) and on cultured bovine chromaffin cells (Yanagihara et al. 1979; Trifaro & Lee, 1980; Fisher et al. 1981). Presumably, the muscarinic responses to acetylcholine in the present experiments were particularly pronounced simply because the dose employed sufficed to activate muscarinic receptors without affecting the nicotinic receptors significantly. Most studies of the mechanism of catecholamine release from the adrenal medulla have involved splanchnic nerve stimulation at intensities far above anything which could conceivably occur naturally, or very high doses of acetylcholine, or cholinergic agonists, likely to favour activation of nicotinic receptors. In vivo studies have also almost invariably been carried out under anaesthesia, which substantially modifies catecholamine release and may well have led to preferential activation of nicotinic receptors. In the conscious calf there is a substantial hypertensive response to splanchnic nerve stimulation, following complete nicotinic blockade with hexamethonium, which is completely abolished by atropine and so clearly mediated by muscarinic receptors (A. V. Edwards & C. T. Jones, unpublished observations). Furthermore, even in isolated bovine chromaffin cell preparations muscarinic receptors are known to be present (Ballesta et al. 1989) and their activation leads to biochemical responses other than secretion of catecholamines, which include increasing levels of cGMP (Yanagihara et al. 1979), phospholipid turnover (Fisher et al. 1981) and intracellular inositol triphosphate (Forsberg et al. 1986). The finding that the adrenal medullary responses to intra-aortic infusions of acetylcholine, at a dose of 4.5 nmol min<sup>-1</sup> kg<sup>-1</sup>, are due mainly to activation of muscarinic receptors has now been vindicated by the further finding that they are not greatly reduced following full nicotinic blockade with hexamethonium (Jones & Edwards, 1992*a*) (Table).

## ADRENAL CORTICAL FUNCTION

Control of adrenal glucocorticoid secretion in the short term has generally been supposed to be largely, if not entirely, determined by adrenocorticotrophin (ACTH) released from the anterior pituitary. However, there have been an increasing number of reports of situations in which adrenal glucocorticoid output and plasma ACTH concentration have been poorly correlated and in which adrenal cortical function must be influenced by other factors (Krieger, 1979). Searches for such other mechanisms have produced a body of, mainly indirect, evidence to implicate the splanchnic sympathetic adrenal innervation (Engeland et al. 1981; Wood et al. 1982a, b; Dempsher & Gann, 1983) which is entirely consistent with the concept that the innervation to the gland exerts long-term effects on adrenal cortical growth (Engeland & Dallman, 1976; Dallman et al. 1976; Kleitman & Holzwarth, 1985a). Diurnal rhythms in adrenal cortical sensitivity have been demonstrated (Srivasta & Meier, 1972; Ottenweller et al. 1978; Kaneko et al. 1981; Ottenweller & Meier, 1982) and presumably normally potentiate the effect of diurnal fluctuations in plasma ACTH concentration. Furthermore, rats maintained on a restricted water schedule exhibit a rapid and pronounced fall in plasma corticosterone concentration immediately after drinking, which cannot be ascribed to a fall in plasma ACTH concentration, or to an increase in the rate of steroid catabolism (Wilkinson et al. 1982).

The contention that adrenal cortical function is influenced by the autonomic innervation is strongly supported by morphological evidence of various types of nerve terminal that are present within the adrenal cortex in a number of different species (Holzwarth et al. 1987; Kesse et al. 1987; Charlton, 1990; Hinson, 1990). In man, it has recently been reported that nerve terminals come into close contact with endocrine cells in the zona fasciculata (Dorovini-Zis & Zis, 1991) and seem likely to be cholinergic (Charlton et al. 1991).

# Physiological evidence of neural control of glucocorticoid secretion

Stimulation of the peripheral end of the splanchnic nerve in the conscious, functionally hypophysectomised, calf at a moderate frequency (4 Hz) produces no significant increase in adrenal glucocorticoid output (Edwards & Jones, 1987b) (Fig. 4), in the absence of exogenous ACTH, although it produces the expected abrupt increase in the adrenal output of catecholamines and enkephalins. In contrast, splanchnic nerve stimulation at the same frequency, under precisely similar conditions, except that the animals were receiving an i.v. infusion of ACTH<sub>1-24</sub>, resulted in a substantial increase in adrenal cortisol output, without affecting the concentration of ACTH in the arterial plasma (Edwards & Jones, 1987a). In these experiments the dose of exogenous ACTH raised cortisol output by about 50% of maximal (500 ng  $min^{-1} kg^{-1}$ ) and this was roughly doubled (increasing by a further 400 ng min<sup>-1</sup> kg<sup>-1</sup>) during splanchnic nerve stimulation (Fig. 4). This finding has since been confirmed in hypophysectomised dogs, in which splanchnic nerve stimulation was shown to increase the conversion of 11-deoxycortisol to cortisol, suggesting a change in  $11\beta$ -hydroxylase activity (Engeland & Gann, 1989) by a mechanism which has yet to be established.

Splanchnic nerve stimulation in the conscious, functionally hypophysectomised calf model also causes an increase in adrenal bloodflow. Accordingly, it might be supposed that the rise in cortisol output, in the presence of exogenous ACTH, was secondary to a rise in the rate at which ACTH was presented to the gland, even though there was no change in the concentration of the peptide in the arterial plasma, as Urquhart (1965) originally suggested. However, if it is assumed that changes in total adrenal bloodflow represent a reasonably accurate index of changes in cortical bloodflow, it is a simple matter to estimate ACTH presentation rate during these experiments. It can then be seen that there is a closely linear relation between adrenal ACTH presentation rate and adrenal cortisol output for each of the samples collected before and after splanchnic nerve stimulation. In contrast, cortisol output during splanchnic nerve stimulation was significantly greater than that which could be accounted for by the existing ACTH presentation rate (Fig. 5). The assumption that the

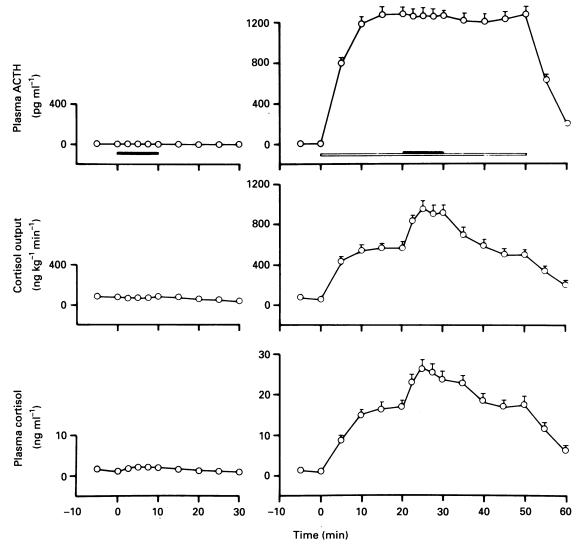


Fig. 4. Comparison of the changes in mean arterial plasma ACTH and cortisol concentration, and right adrenal cortisol output, in response to stimulation of the peripheral end of the right splanchnic nerve (4 Hz for 10 min) in 11 conscious functionally hypophysectomised calves in the presence and absence of exogenous ACTH<sub>1-24</sub> (5 ng min<sup>-1</sup> kg<sup>-1</sup>). Open horizontal bar, duration of ACTH infusion. Filled horizontal bars: duration of splanchnic nerve stimulation. Vertical bars, s.E. of each mean value where these exceed the size of the symbol. (From Edwards & Jones 1987*a*, by permission of the Physiological Society.)

total adrenal bloodflow represents a reliable index of cortical bloodflow is supported by the finding that medullary bloodflow amounts to a mere 7% of the total (Sparrow & Coupland, 1987), at least in the rat. In the dog, Breslow and colleagues (Breslow et al. 1987) have shown that the rise in adrenal bloodflow which occurs during haemorrhage, and is mediated via the innervation to the gland, is largely confined to the medulla. Accordingly, it is even less plausible that the rise in adrenal cortisol output which occurs during splanchnic nerve stimulation, in the presence of exogenous ACTH, could be due solely to an increase in ACTH presentation. This is not to say that an increase in ACTH presentation, secondary to an increase in cortical bloodflow, could not have contributed to the increase in adrenal cortisol output.

Changes in ACTH presentation in this particular model quite clearly result in appropriate changes in cortisol output as evidenced by the fall in adrenal cortisol which occurs when adrenal bloodflow is deliberately reduced, as with intra-aortic infusions of endothelin (Bloom et al. 1990). The conclusion is simply that the increase in cortisol output which occurs in response to splanchnic nerve stimulation, in the presence of exogenous ACTH, in the conscious functionally hypophysectomised calf cannot be accounted for by that mechanism.

## Cortical responses to peptidergic agonists

Vasoactive intestinal peptide (VIP). VIP-containing neurons are known to be present in the adrenal

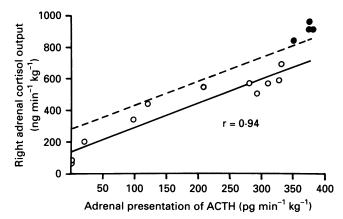


Fig. 5. Relation between mean right adrenal presentation rate of ACTH and mean right adrenal cortisol output in 11 conscious, functionally hypophysectomised, calves in which the peripheral end of the right splanchnic nerve was stimulated (4 Hz for 10 min) during an infusion of exogenous  $ACTH_{1-24}$  (5 ng min<sup>-1</sup> kg<sup>-1</sup>).  $\bigcirc$ , before and after splanchnic nerve stimulation.  $\bigcirc$ , during stimulation. Regression line calculated by the method of least squares applied to the values obtained before and after stimulation. Hatched line, regression line +2 s.D. (From Edwards & Jones 1989*b*, by permission of International Universities Press.)

cortex (Hökfelt et al. 1981; Holzwarth, 1984; Cheung & Holzwarth, 1986; Cunningham & Holzwarth, 1989) and VIP has been shown to stimulate steroid secretion by adrenal cortical cells, albeit at a higher concentration than ACTH (Kowal et al. 1977; Morera et al. 1979; Leboulanger et al. 1983). Furthermore VIP is released from the gland in response to stimulation of the peripheral end of the splanchnic nerve in the conscious calf (Bloom et al. 1988). Accordingly, the possibility that the potentiation of the steroidogenic response to ACTH, which occurs during splanchnic nerve stimulation, might be mimicked by intra-aortic infusions of this peptide was investigated in the conscious hypophysectomised calf model. This involved monitoring the adrenal cortical responses to intra-aortic infusions of VIP at a dose of 50 pmol  $\min^{-1} kg^{-1}$  in the presence and absence of exogenous ACTH. As with splanchnic nerve stimulation, VIP had no discernible effect on cortisol output in the absence of ACTH but produced quite a substantial rise in its presence, without affecting the concentration of ACTH in the arterial plasma (Bloom et al. 1987). The output of cortisol from the gland was consistently greater than that which could be accounted for by the presentation rate of ACTH alone throughout the VIP infusion. Furthermore, VIP faithfully mimicked the effect of splanchnic nerve stimulation on the adrenal vasculature causing an increase in adrenal bloodflow in the absence, but not in the presence, of exogenous ACTH. It was therefore concluded that release of this peptide from splanchnic nerve terminals in the adrenal cortex most probably accounts, at least in part, for the powerful adrenal cortical steroidogenic response to splanchnic nerve stimulation that occurs in the presence of submaximal doses of ACTH.

These results have recently been confirmed by Holst and his colleagues in the isolated, innervated, perfused adrenal gland of the pig (Erhardt-Bornstein et al. 1991), except that both VIP and splanchnic nerve stimulation were found to exert a steroidogenic action in both the presence and the absence of ACTH. This might be due to the fact that they employed higher stimulus frequencies and larger doses of VIP, possibly outside the physiological range. The amount of VIP it is practicable to infuse intra-aortically is limited by its hypotensive action in the conscious calf model whereas no such restraint is necessary in the perfused gland preparation.

Calcitonin gene-related peptide. CGRP is also present in the adrenal gland (Kuramoto et al. 1987; Kong et al. 1989) and was tested in functionally hypophysectomised calves in the same way as VIP (Bloom et al. 1989). The peptide was infused into the aorta above the origin of the adrenal arterial supply at a dose of 33 pmol min<sup>-1</sup> kg<sup>-1</sup> and, like VIP, caused a pronounced increase in adrenal blood flow in the absence of exogenous ACTH; this effect was reduced by not abolished in the presence of ACTH. Unlike VIP, CGRP stimulated the release of cortisol from the adrenal gland of these animals in the absence of exogenous ACTH. This effect was reduced during an i.v. infusion of exogenous ACTH at a low dose  $(2 \text{ ng min}^{-1} \text{ kg}^{-1})$  and completely suppressed by ACTH at a higher dose (5 ng min<sup>-1</sup> kg<sup>-1</sup>). It exerted a direct steroidogenic action on the gland which in no way resembled the response to splanchnic nerve stimulation. CGRP has also been found to stimulate secretion of cortisol in the isolated, perfused in situ rat adrenal preparation (Hinson & Vinson, 1990). These authors attributed this latter effect to an increase in

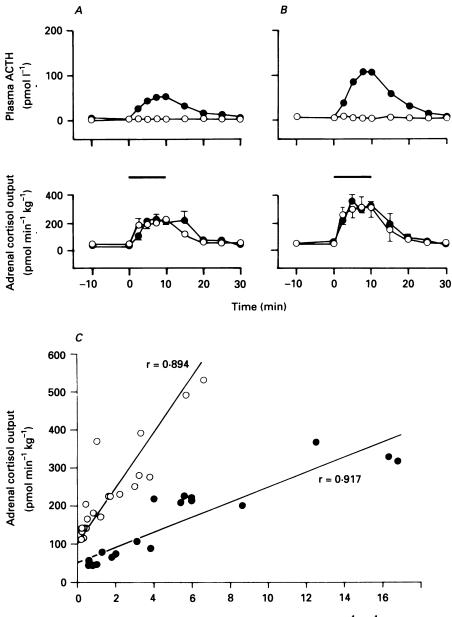




Fig. 6. (A) Comparison of the changes in mean plasma ACTH concentration and mean right adrenal cortisol output in conscious functionally hypophysectomised calves in response to intra-aortic infusion of CRF ( $\bigcirc$ ; n = 5) or ACTH<sup>1-24</sup> ( $\bigoplus$ ; n = 5) at 4 pmol min<sup>-1</sup> kg<sup>-1</sup>; (B) 8 pmol min<sup>-1</sup> kg<sup>-1</sup>. Vertical bars, S.E. of each mean value where these exceed the size of the symbol. Horizontal bars, duration of infusions. (C) Relation between mean estimated CRF ( $\bigcirc$ ) and ACTH ( $\bigoplus$ ) presentation rate to the right adrenal gland and mean right adrenal cortisol output in conscious functionally hypophysectomised calves in response to intra-aortic infusions of CRF and ACTH as in A and B. Regression lines calculated by the method of least squares applied to all mean values. (Modified from Jones & Edwards 1990*a*, by permission of the Physiological Society.)

adrenal bloodflow but it has yet to be established that an increase in bloodflow can cause an increase in cortisol output in vivo, above and beyond that attributable to the increase in ACTH presentation.

Corticotrophin releasing factor (CRF). Since CRF is also released from the adrenal gland in response to splanchnic nerve stimulation (see above) and it was suggested that it might have a direct effect on adrenal steroidogenesis, at least in the case of aldosterone production in the zona glomerulosa of the rat (Mazzochi et al. 1989), the possibility arose that this peptide might also be implicated in the steroidogenic response to splanchnic nerve stimulation. This was tested by infusing it intra-aortically in the functionally hypophysectomised calf model, as described for VIP and CGRP except that much lower doses were employed. CRF caused an increase in the output of cortisol from the adrenal gland, which was doserelated over the range 1.3-2.6 ng min<sup>-1</sup> kg<sup>-1</sup> and maximal at the higher of these doses (Jones & Edwards, 1990 *a*) (Fig. 6). This response was observed at a dose below that which caused a significant fall in adrenal vascular resistance (5.2 ng min<sup>-1</sup> kg<sup>-1</sup>). Cortisol output was also linearly related to the rate at which CRF was presented to the gland during these infusions. In addition, infusions of CRF produced the release of small but readily detectable amounts of ACTH-like peptides from the gland, mainly in the form of ACTH<sup>1-39</sup> but with some proopiomelanocortin being released. These findings have recently been confirmed in the rat by Andreis et al. (1991), who further suggested that the hypothalamopituitary CRF/ACTH system may be replicated in the adrenal gland and operate to control glucocorticoid secretion.

In the functionally hypophysectomised calf model this would appear to be unlikely as roughly equimolar amounts of CRF and ACTH are released from the gland in response to splanchnic nerve stimulation (about 10 fmol min<sup>-1</sup> kg<sup>-1</sup> during splanchnic nerve stimulation at 40 Hz for 1 s at 10 s intervals; Edwards & Jones, 1988; Jones & Edwards, 1990a). This is more suggestive of corelease than of a cascade phenomenon. Furthermore, CRF has significantly greater steroidogenic potency than ACTH (Jones & Edwards, 1990b) (Fig. 6). It is difficult to visualise a biological justification for any hypothetical causal relation between the release of the 2 peptides in view of the fact that they are released in about the same amounts and that CRF is actually the more potent agonist. If both peptides are released together, either from chromaffin cells or nerve terminals within the adrenal medulla, there is reason to doubt that they could reach the adrenal cortex before entering the peripheral circulation; the cortex and medulla are now thought to be supplied by largely separate capillary beds, at least in the rat (Sparrow & Coupland, 1987) and, to the extent that it is not, flows from cortex to medulla (Vinson et al. 1985). Both peptides exert other actions within the gland, such as vasodilatation, and it is entirely possible that, when released within the gland, they normally act to modulate the bloodflow without affecting steroidogenesis at all.

It was also concluded that it was unlikely that CRF, released in response to splanchnic nerve stimulation, contributes to the potentiation of ACTH-induced steroidogenesis which then occurs. Splanchnic nerve stimulation exerts a potent steroidogenic effect in the presence of exogenous ACTH but has no detectable effect in the absence of ACTH (Fig. 4) whereas CRF clearly does (Fig. 6). Furthermore, the release of CRF from the adrenal gland which occurs in response to splanchnic nerve stimulation is significantly reduced during infusions of exogenous ACTH in these animals (Edwards & Jones, 1988). This conclusion has been corroborated more recently by the finding that a dose of a specific CRF-antagonist, which completely blocks the adrenal steroidogenic response to intra-aortic infusions of CRF, in the presence of exogenous ACTH, in functionally hypophysectomised calves, has absolutely no effect on the steroidogenic response to splanchnic nerve stimulation in calves given i.v. infusions of exogenous ACTH (Jones & Edwards, 1992*b*).

## Cortical responses to nonpeptidergic agonists

Acetylcholine. Acetylcholine was first shown to stimulate steroidogenesis by a direct action on adrenal cortical cells in isolated perfused calf adrenal glands by Rosenfeld (1955). This has since been confirmed in isolated bovine adrenal cortical cells, in which the effect was found to depend on activation of muscarinic receptors (Hadjian et al. 1982), the adrenal gland of the frog (Benyamina et al. 1987) and rat (Porter et al. 1988). The presence of cholinergic fibres in the cortex has been reported in a number of species (see Robinson et al. 1977, for references); in the human adrenal, acetylcholinesterase positive nerve plexuses traverse the zona fasciculata in radial trunks and ramify throughout the cortical parenchyma (Charlton et al. 1991).

In the functionally hypophysectomised calf model intra-aortic infusions of a low dose of acetylcholine  $(4.5 \text{ nmol min}^{-1} \text{ kg}^{-1} \text{ for } 10 \text{ min})$  were tested in the presence and absence of exogenous ACTH (2 ng  $\min^{-1} kg^{-1}$ , i.v.). Very much like splanchnic nerve stimulation ACh elicited a significant increase in adrenal cortisol output, associated with an increase in peripheral plasma cortisol concentration, in the presence of ACTH, but not otherwise. However, unlike splanchnic nerve stimulation, the increase in cortisol output did not exceed that which would be predicted from the increase in ACTH presentation which occurred, secondary to acetylcholine-induced adrenal vasodilatation. However, when the study was extended to investigate the responses of animals pretreated with hexamethonium a direct steroidogenic response was revealed (Jones & Edwards, 1992a). In the absence of hexamethonium this effect is obscured by the greater change in ACTH presentation rate which occurs at the higher perfusion pressure which then obtains.

*Catecholamines*. Several groups have reported the fact that catecholamines are capable of exerting a steroidogenic effect on adrenocortical cells in vitro, leading to aldosterone production by cells of zona

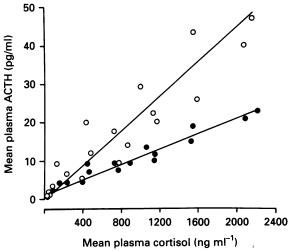


Fig. 7. Relations between mean plasma ACTH and cortisol concentration in conscious 2–6 wk calves given i.v. infusion of ACTH at a dose of either 5 or 10 ng min<sup>-1</sup> kg<sup>-1</sup>.  $\bigcirc$ , Control animals (n = 7, r = 0.93).  $\textcircledline$ , Cut splanchnic nerves (n = 7, r = 0.98. Regression lines calculated by the method of least squares. (From Edwards & Jones 1987b by permission of the Physiological Society.)

glomerulosa origin (DeLéan et al. 1984; Pratt et al. 1985) and cortisol production by cells derived from the zona reticularis (Water et al. 1988). In each case the effect has been found to be mediated by  $\beta$ adrenoceptors and, for glucocorticoid production, by receptors of the  $\beta_1$  subclass (Lightly et al. 1990). Activation of  $\beta$ -adrenoceptors apparently stimulates cleavage of the side chain of cholesterol, by promoting accumulation of mRNA for the appropriate cytochrome (Erhardt-Bornstein et al. 1991), which happens to be the rate-limiting step in steroid synthesis (Stone & Hechter, 1954). Holst and colleagues have recently confirmed the finding that catecholamines stimulate the secretion of both aldosterone and cortisol in the intact gland, employing the isolated, perfused adrenal gland of the pig (Bornstein et al. 1990). The existence of catecholaminergic neurons in the cortex (Robinson et al. 1977; Kleitman & Holzwarth, 1985b), together with reports of chromaffin cells scattered within the cortex and in close association with cortical cells (Gallo-Payet et al. 1987; Bornstein et al. 1990, 1991) rather suggests that catecholamines act via neural or paracrine pathways within the gland (Bornstein et al. 1990; Hinson, 1990).

### Physiological significance

The evidence reviewed above shows that stimulation of the splanchnic sympathetic innervation, at a frequency likely to fall within the physiological range, substantially potentiates the secretion of the adrenal cortex in response to ACTH in normal conscious animals, without detectable steroidogenic effect in the absence of ACTH. The question remains as to whether control of adrenal steroidogenesis via the sympathetic innervation is important under resting conditions in normal animals. It was addressed by comparing the steroidogenic responses of normal calves and lambs to ACTH with those of animals in which the splanchnic nerves, and consequently the sympathetic innervation to the adrenal gland, had been cut at least 7 d previously. In both species prior adrenal denervation significantly reduced adrenal steroidogenesis, assessed by the rise in arterial plasma cortisol concentration, in response to ACTH (Edwards et al. 1986; Edwards & Jones, 1987a). Mean plasma ACTH was linearly related to mean plasma cortisol concentration throughout each experiment and the slopes of the regression lines therefore provide an index of cortical sensitivity. The data from the calves are illustrated in Figure 7 which shows that adrenal cortical sensitivity to ACTH was reduced about twofold.

## Adrenal bloodflow

The significance of changes in adrenal bloodflow which are associated with changes in adrenal cortical or medullary activity is controversial. For the medulla, Breslow and colleagues have shown that increased activity in response to hypoxia, haemorrhagic hypotension and splanchnic nerve stimulation, in anaesthetised dogs, is associated with 3-4 fold rises in bloodflow, which are restricted to the medulla (Breslow et al. 1987, 1989). However, prevention of the vascular response, by reducing the perfusion pressure, did not significantly influence the output of adrenaline or noradrenaline in response to supramaximal splanchnic nerve stimulation for a short period (20 Hz for 5 min) (Breslow et al. 1991 a). This finding is in accord with expectation, in view of the large amounts of catecholamine stored in the chromaffin cells and the importance of this release mechanism during life-threatening episodes of hypotension. The further observation that glucocorticoid output was unaffected by reducing cortical bloodflow in these animals has been criticised on the grounds that cortical responsiveness could have been blunted by pretreatment with dexamethasone for 48 h; also that cortisol output was so low to start with that a further reduction might be below detection limits (Engeland, 1991).

The suggestion that adrenal cortical blood flow contributes to the control of glucocorticoid secretion dates from the mid 1960s when corticosterone output in anaesthetised hypophysectomised rats, given a steady infusion of exogenous ACTH, rose with (whole gland) bloodflow for 30-40 min until a steady state was established (Porter & Klaiber, 1965). At about the same time the estimated rate of adrenal presentation of ACTH was found to correlate better with steadystate cortisol secretion than the concentration of ACTH in blood perfusing canine adrenal in situ (Urquhart, 1965). A little later it was reported that changes in adrenal bloodflow in conscious dogs pretreated with dexamethasone correlated well with adrenal cortisol output, so long as an exogenous ACTH infusion was below maximal levels (L'Age et al. 1970). Unfortunately this study can be criticised on the grounds that dexamethasone suppression of endogenous ACTH release is not absolute and the agents employed to vary adrenal bloodflow (histamine and methacholine) could well have had direct effects on the adrenal cortical cells.

Adrenal cortical bloodflow is inexorably linked to glucocorticoid secretion by virtue of the fact that ACTH exerts a specific vasodilator effect on the adrenal vasculature (above a certain concentration) which has been demonstrated in anaesthetised calves (Balfour, 1953), dogs (Hartman et al. 1955; Stark et al. 1965), rats (Holzbauer & Vogt, 1957; Sapirstein & Goldman, 1959) and human patients (Grant et al. 1957), conscious sheep (Wright, 1963), dogs (Stark et al. 1965; L'Age et al. 1970) and calves (Edwards et al. 1974, 1975) and the isolated perfused adrenal gland of the rat (Hinson et al. 1986). The vasodilatation is not secondary to increased steroidogenesis as it persists in the presence of cycloheximide at a dose which completely blocks steroidogenesis (Maier & Staehlin, 1968; Edwards et al. 1975) and the suggestion that it might be due to production of prostaglandins (Maier & Staehlin, 1968) has not been vindicated by subsequent studies. ACTH is thought to act mainly on capsular and subcapsular arterioles (Vinson et al. 1985), around which mast cells are congregated (Hinson et al. 1991a). Recent evidence, obtained in the in situ isolated perfused rat adrenal gland shows that degranulation of these cells with compound 48/80 mimics the vascular response to ACTH, while sodium cromoglycate, which prevents mast cell degranulation, inhibits adrenal vasodilatation in response to ACTH (Hinson et al. 1989, 1991b). This strongly suggests that mast cells are implicated in the adrenal vasodilator response to ACTH but this has yet to be confirmed in a whole animal model.

To the extent that adrenal vasodilatation increases the rate of ACTH presentation to adrenal cortical cells, it could be expected to lead to an increase in corticosteroidogenesis, although doubts have recently been expressed as to whether increased binding of ACTH to its receptor(s) can be achieved without increasing ACTH concentration (Breslow et al. 1991 b). Of course it would be the concentration of ACTH in the adrenal extracellular fluid which would be determinative and this could well rise with an increase in plasma flow without any change in plasma ACTH concentration, depending upon the kinetics. The consensus is certainly that glucocorticoid output varies, up to a maximum, with ACTH presentation and the close linearity of the relations between these 2 variables illustrated in Figures 5 and 6B testifies to this. The question has been addressed directly in the conscious hypophysectomised calf model, in which the potent vasoconstrictor agonist endothelin was employed to vary adrenal blood flow during the course of ongoing i.v. infusions of exogenous ACTH (Jones et al. 1990). Under these conditions endothelin produced a fall in adrenal cortisol output which correlated precisely with that predicted from the fall in ACTH presentation. The possibility that this might be attributable to some other, nonvascular, action of endothelin was eliminated by adjusting the dose of exogenous ACTH during the infusion of endothelin, so as just to compensate for the fall in adrenal plasma flow and maintain a constant ACTH presentation rate. Cortisol output could still be related to ACTH presentation with precision. It is also important to note that these infusions of endothelin, which raised the concentration in the plasma by no more than 10 pmol l<sup>-1</sup>, had no detectable effect of the output of catecholamines, enkephalins, aldosterone or CRF from the gland.

The adrenal vasculature is less sensitive to ACTH than the cells of the zona fasciculata and it is generally agreed that corticosteroidogenic responses occur at ACTH levels below those necessary to produce vasodilatation in the gland (Wright, 1963; L'Age et al. 1970; Edwards et al. 1975). Just recently, Breslow and colleagues have reported that in dexamethasonepretreated, pentobarbital-fentanyl-anaesthetised ventilated dogs increased adrenal cortisol output, in response to exogenous ACTH, was associated with a significant increase in oxygen extraction from the blood with no significant increase in flow (Sakima et al. 1991). Our contention is merely that, when the flow does increase, it will necessarily produce an increase in glucocorticoid output by increasing ACTH presentation.

In the in situ isolated perfused rat adrenal gland it is reported that increases in flow which are not associated with any increase in ACTH presentation also lead to increased corticosteroidogenesis (Hinson et al. 1989). It has further been suggested that the effect is mediated by release of endothelins within the gland since ACTH stimulates the release of detectable (statistically significant) amounts of endothelin from this preparation (Hinson et al. 1991c) and endothelins elicit a dose-dependent and highly sensitive release of both glucocorticoids and aldosterone from dispersed preparations of adrenal cortical cells (Hinson et al. 1991 a, c). One difficulty in interpreting these data arises from the fact that the increase in steroidogenesis in the in situ perfused rat adrenal preparation in response to an increase in flow (with no increase in ACTH presentation) is limited to the glucocorticoids, whereas an increase in association with aldosterone production would be expected. Also it does not appear to apply in the conscious hypophysectomised calf model. This might be due to a species difference as it has yet to be established whether or not endothelins are released from the bovine adrenal gland in response to ACTH.

## CONCLUSIONS

Studies in conscious calves have revealed the fact that medullary responses to stimulation of the splanchnic sympathetic innervation are substantially potentiated by employing an intermittent pattern of stimulation. It has yet to be established what mechanisms underlie this phenomenon. It might be due to enhanced synaptic transmission due to increased mobilisation of Ca<sup>2+</sup>, or of some other intracellular messenger. Alternatively, the probability of vesicle release might be increased by modulation of K<sup>+</sup> efflux and Ca<sup>2+</sup> influx across the chromaffin cell membrane, as apparently occurs in postganglionic sympathetic nerve terminals supplying the tail artery of the rat and determines the probability of vesicle release therefrom (Stjärne et al. 1991). Yet another possibility is suggested by the finding by Ip & Zigmond (1984) that, in the superior cervical ganglion of the rat, acute activation of tyrosine hydroxylase (which catalyses the rate-limiting step in noradrenaline synthesis) is mediated in part by acetylcholine, but also in part by a noncholinergic agonist. Release of the noncholinergic transmitter is potentiated by high frequency stimulation. In the bovine adrenal medulla, VIP is released in response to splanchnic nerve stimulation. The amounts appearing in the venous effluent plasma were too small to indicate whether release was potentiated by high-frequency stimulation, but this has been clearly established in other tissues such as the submandibular gland of the cat (Andersson et al. 1982). Furthermore the peptide has been shown to release adrenal catecholamines when infused in small amounts intra-aortically (Edwards & Jones 1993).

Secretion of cortisol from the zona fasciculata in response to ACTH is potentiated by stimulation of the splanchnic sympathetic innervation. It has yet to be established whether the effect can be potentiated by stimulating in bursts, but it occurs at a frequency (4 Hz) likely to fall within the physiological range. The contention that the effect is operative under normal resting conditions has been confirmed by the finding that the sensitivity of the adrenal cortex to ACTH is substantially reduced, within 7 d, following adrenal denervation. To the extent that activity of the innervation produces adrenal cortical vasodilatation it is likely to contribute to enhancement of corticosteroidogenesis by increasing the rate at which ACTH is presented to the cortical cells. It also appears to be due, at least in part, to VIP and possibly to acetylcholine or catecholamines. It does not appear to be due to release of CGRP, enkephalins or CRF.

## REFERENCES

- ANDERSSON P-O, BLOOM SR, EDWARDS AV, JÄRHULT J (1982) Effects of stimulation of the chorda tympani in bursts on submaxillary responses in the cat. *Journal of Physiology* 322, 469–483.
- ANDREIS PG, NERI G, NUSSDORFER GG (1991) Corticotrophinreleasing hormone (CRH) directly stimulates corticosterone secretion by the rat adrenal gland. *Endocrinology* 128, 1198–1200.
- ANICHKOV SV, MALYGHINA EI, POSKALENKO AN, RYZHENKOV V (1960) Reflexes from carotid bodies upon the adrenals. Archives Internationales de Pharmacodynamie et de Thérapie 129, 156–165.
- BALFOUR WE (1953) Changes in the hormone output of the adrenal cortex of the young calf. Journal of Physiology 122, 59P-60P.
- BALLESTA JJ, BORGES R, GARCIA AG, HIDALGO MJ (1989) Secretory and radioligand binding studies on muscarinic receptors in bovine and feline chromaffin cells. *Journal of Physiology* **418**, 411–426.
- BENYAMINA M, LEBOULENGER F, LIHRMANN I, DELARUE C, FEUILLOLEY M et al. (1987) Acetylcholine stimulates steroidogenesis in isolated frog adrenal gland though muscarinic receptors: evidence for a desensitisation mechanism. Journal of Endocrinology 113, 339–348.
- BLOOM SR, EDWARDS AV, HARDY RN, MALINOWSKA KW, SILVER M (1975) Endocrine responses to insulin hypoglycaemia in the young calf. *Journal of Physiology* 244, 783–803.
- BLOOM SR, EDWARDS AV, HARDY RN, SILVER M (1976) Adrenal and pancreatic endocrine responses to hypoxia in the conscious calf. *Journal of Physiology* **261**, 271–283.
- BLOOM SR, EDWARDS AV, HARDY RN (1977) Adrenal and pancreatic endocrine responses to hypoxia and hypercapnia in the calf. *Journal of Physiology* **269**, 131–154.
- BLOOM SR, EDWARDS AV, JONES CT (1987) Adrenal cortical responses to vasoactive intestinal peptide in conscious hypophysectomized calves. *Journal of Physiology* **391**, 441–450.
- BLOOM SR, EDWARDS AV, JONES CT (1988) The adrenal contribution to the neuroendocrine responses to splanchnic nerve stimulation in conscious calves. *Journal of Physiology* 397, 513-526.

- BLOOM SR, EDWARDS AV, JONES CT (1989) Adrenal responses to calcitonin gene-related peptide in conscious hypophysectomized calves. *Journal of Physiology* 409, 29–41.
- BLOOM SR, EDWARDS AV, JONES CT (1990) The effect of changes in adrenal blood flow on adrenal cortical responses to adrenocorticotrophin in conscious calves. *Journal of Physiology* 429, 377–386.
- BORNSTEIN SR, ERHARDT-BORNSTEIN M, SCHERBAUM WA, PFEIFFER EF, HOLST JJ (1990) Effects of splanchnic nerve stimulation on the adrenal cortex may be mediated by chromaffin cells in a paracrine manner. *Endocrinology* 127, 900–906.
- BORNSTEIN SR, ERHART-BORNSTEIN M, USADEL H, BÖCKMANN M, SCHERBAUM WA (1991) Morphological evidence for a close interaction of chromaffin cells with cortical cells within the adrenal gland. *Cell and Tissue Research* **265**, 1–9.
- BRESLOW MJ, JORDAN DA, THELLMAN CT, TRAYSTMAN RJ (1987) Neural control of adrenal medullary and cortical bloodflow during hemorrhage. *American Journal of Physiology* 252, H521-528.
- BRESLOW MJ, BALL TD, MILLER CF, RAFF H, TRAYSTMAN RJ (1989) Adrenal blood flow and secretory relationships during hypoxia in anaesthetized dogs. *American Journal of Physiology* 257, H1458-1465.
- BRESLOW MJ, TOBIN JR, KUBOS KL, RAFF H, TRAYSTMAN RJ (1991 a) Effect of adrenal hypotension on elicited secretory activity in anesthetized dogs. *American Journal of Physiology* 260, H21-26.
- BRESLOW MJ, TOBIN JR, KUBOS KL, TRAYSTMAN RJ, RAFF H (1991b) Coupling of adrenal blood flow and secretion reply. *American Journal of Physiology* 261, H1351.
- BRUHN TO, ENGELAND WC, ANTHONY ELP, GANN DS, JACKSON IMD (1987) Corticotrophin-releasing factor in the dog adrenal medulla is secreted in response to hemorrhage. *Endocrinology* 120, 25–33.
- BUNN SJ, MARLEY PD, LIVETT BG (1988) Effects of opioid compounds on basal and muscarinic induced accumulation of inositol phosphates in cultured bovine chromaffin cells. *Biochemical Pharmacology* 37, 395–399.
- CASTANAS E, BOURHIM N, GIRAUD P, BOURDOURESQUE F, CANTOU P et al. (1985*a*) Interactions of opiates with opioid binding sites in the bovine adrenal medulla. I. Interactions with  $\delta$  and  $\mu$  sites. *Journal of Neurochemistry* **45**, 688–699.
- CASTANAS G, BOURHIM N, GIRAUD P, BOURDOURESQUE F, CANTOU P et al. (1985b) Interactions of opiates with opiate binding sites in the bovine adrenal medulla. II. Interactions with K sites. *Journal of Neurochemistry* **45**, 688–699.
- CHANG KJ, WILSON SP, VIVEROS OH (1982) Co-storage and cosecretion of opioid peptides and catecholamines. *Advances in Biosciences* 36, 243–248.
- CHARLTON BG (1990) Adrenal cortical innervation and glucocorticoid secretion. Journal of Endocrinology 126, 5–8.
- CHARLTON BG, NKOMAZANA OF, MCGADEY J, NEAL DE (1991) A preliminary study of acetylcholinesterase-positive innervation in the human adrenal cortex. *Journal of Anatomy* 176, 99–104.
- CHEN YN, DIXON WR (1990) The effect of etorphine in nicotineand muscarine-induced catecholamine section from perfused rat adrenal glands. Life Science. 40, 1167-1173.
- CHEUNG CH, HOLZWARTH MA (1986) Fetal adrenal VIP: distribution and effect on medullary catecholamine secretion. *Peptides* 7, 413–418.
- COSTA E, DIGIULIO A, FRATTA W, HONG J, YANG HY-YT (1979) Interactions of enkephalinergic and catecholaminergic neurones in CNS and periphery. In *Catecholamines: Basic and Clinical Frontiers* (ed. E. Usdin, I. J. Kopin & J. Barchas), pp. 1020–1025. Oxford: Pergamon Press.
- CRITCHLEY JAJH, ELLIS P, UNGAR A (1980) The reflex release of adrenaline and noradrenaline from the adrenal glands of cats and dogs. *Journal of Physiology* 298, 71–78.

- CRONE C (1965) The secretion of adrenal medullary hormones during hypoglycaemia in intact, decerebrate and spinal sheep. *Acta Physiologica Scandinavica* 63, 213–224.
- CUNNINGHAM LA, HOLZWARTH MA (1989) Autoradiographic distribution of <sup>125</sup>I-VIP binding in the rat adrenal cortex. *Peptides* 10, 1105–1108.
- DALLMAN MF, ENGELAND WC, SHINSAKO J (1976) Compensatory adrenal growth, a neurally mediated reflex. American Journal of Physiology 231, 408–414.
- DELÉAN A, RACZ K, MCNICOLL N, DESROSIERS M-C (1984) Direct β-adrenergic stimulation of aldosterone secretion in cultured bovine adrenal subcapsular cells. *Endocrinology* **115**, 485–492.
- DEMPSHER DS, GANN DS (1983) Increased cortisol secretion after a small hemorrhage is not attributable to ACTH. *Endocrinology* 113, 86–93.
- DOROVINI-ZIS K, ZIS AP (1991) Innervation of the zona fasciculata of the adult human adrenal cortex: a light and electron microscopic study. Journal of Neural Transmission 84, 75–84.
- DOUGLAS WW, POISNER AM (1965) Preferential release of adrenaline from the adrenal medulla by muscarine and pilocarpine. *Nature* 208, 1102–1103.
- DOUGLAS WW, KANNO T, SAMPSON SR (1967) Effects of acetylcholine and other medullary secretagogues and antagonists on the membrane potential of adrenal chromaffin cells: an analysis employing techniques of tissue culture. *Journal of Physiology* 188, 107–120.
- DUNÉR H (1954) The effect of insulin hypoglycaemia on the secretion of adrenaline and noradrenaline from the suprarenal gland of the cat. Acta Physiologica Scandinavica 32, 63-68.
- EDWARDS AV, HARDY RN, MALINOWSKA KW (1974) The effects of infusions of synthetic adrenocorticotrophin in the conscious calf. *Journal of Physiology* 239, 477–498.
- EDWARDS AV, HARDY RN, MALINOWSKA KM (1975) The sensitivity of adrenal responses to synthetic adrenocorticotrophin in the conscious unrestrained calf. *Journal of Physiology* 245, 639-653.
- EDWARDS AV, FURNESS PN, HELLE KB (1980) Adrenal medullary responses to stimulation of the splanchic nerve in the conscious calf. Journal of Physiology 308, 15–27.
- EDWARDS AV, HANSELL D, JONES CT (1986) Effects of synthetic adrenocorticotrophin on adrenal medullary responses to splanchnic nerve stimulation in conscious calves. *Journal of Physiology* **379**, 1–16.
- EDWARDS AV, JONES CT, BLOOM SR (1986) Reduced adrenal cortical sensitivity to ACTH in lambs with cut splanchnic nerves. *Journal of Endocrinology* 110, 81–85.
- EDWARDS AV, JONES CT (1987*a*) The effect of splanchnic nerve stimulation on adrenocortical activity conscious calves. *Journal* of *Physiology* **382**, 385–396.
- EDWARDS AV, JONES CT (1987b) The effect of splanchnic nerve section on the sensitivity of the adrenal cortex to adrenocorticotrophin in the calf. Journal of Physiology **390**, 23–31.
- EDWARDS AV, JONES CT (1988) Secretion of corticotrophin releasing factor from the adrenal during splanchnic nerve stimulation in conscious calves. *Journal of Physiology* **400**, 89–100.
- EDWARDS AV, JONES CT (1989*a*) Adrenal responses to splanchnic nerve stimulation in conscious calves given naloxone. *Journal of Physiology* **418**, 339–351.
- EDWARDS AV, JONES CT (1989b) The role of sympathetic nerves in the control of adrenal cortical function. In *The Control of the Hypothalamo-Pituitary-Adrenocortical Axis* (ed. F. Clifford Rose), pp. 275–295. Madison: International Universities Press.
- EDWARDS AV, JONES CT (1993) Adrenal cortical and medullary responses to acetylcholine and vasoactive intestinal peptide in conscious calves. *Journal of Physiology*, in press.
- ENGELAND WC, DALLMAN MF (1976) Neural mediation of compensatory adrenal growth. *Endocrinology* **99**, 1659–1677.

- ENGELAND WC, BYRNES CJ, PRESNELL K, GANN D (1981) Adrenocortical sensitivity to ACTH in awake dogs changes as a function of time of observation independently of changes in ACTH. *Endocrinology* **107**, 2149–2153.
- ENGELAND WC, BEREITER DF, GANN DS (1986) Sympathetic control of adrenal secretion of enkephalins after hemorrhage in awake dogs. *American Journal of Physiology* **251**, R341–348.
- ENGELAND WC, GANN DS (1989) Splanchnic nerve stimulation modulates steroid secretion in hypophysectomized dogs. *Neuro*endocrinology 50, 124–131.
- EHRHARDT-BORNSTEIN M, BORNSTEIN SR, SCHERBAUM WA, PFEIFFER EF, HOLST JJ (1991) Role of the vasoactive intestinal peptide in a neuroendocrine regulation of the adrenal cortex. *Neuroendocrinology* 54, 623–628.
- ERHARDT-BORNSTEIN M, BORNSTEIN SR, TREZCLAK WH, USADEL H, GÜSEBEHLING H et al. (1991) Adrenaline stimulates cholesterol side-chain cleavage cytochrome P450 mRNA accumulation in bovine adrenocortical cells. *Journal of Endocrinology* **131**, R5–8.
- EULER US VON (1956) Noradrenaline. Springfield, Illinois: Charles C. Thomas.
- EULER US VON, FOLKOW B (1953) Einfluss verschiedener afferenter Nervenreize auf die Zusammensetzung des Nebennierenmarkinkretes bei der Katze. Archiv für experimentell Pathologie und Pharmakologie 219, 242–247.
- FELDBERG W, MINZ B, TSUDZIMURA H (1934) The mechanism of the nervous discharge of adrenaline. *Journal of Physiology* 81, 286–304.
- FISHER SK, HOLZ RW, AGRANOFF BW (1981) Muscarinic receptors in chromaffin cell cultures mediate enhanced phospholipid labelling but not catecholamine secretion. *Journal of Neurochemistry* 37, 491–497.
- FOLKOW B, EULER US VON (1954) Selective activation of noradrenaline and adrenaline producing cells in the cat's adrenal gland by hypothalamic stimulation. *Circulation Research* 2, 191–195.
- FORSBERG EJ, ROJAS E, POLLARD HB (1986) Muscarinic receptor enhancement of nicotine-induced catecholamine secretion may be mediated by phophoinositide metabolism in bovine adrenal medullary chromaffin cells. *Journal of Biological Chemistry* **261**, 4915–4920.
- GALLO-PAYET N, POTHIER P, ISLER H (1987) On the presence of chromaffin cells in the adrenal cortex: their possible role in adrenocortical function. *Biochemistry & Cell Biology* 65, 588-000.
- GEFFEN LB, LIVETT BG, RUSH RA (1969) Immunohistochemical localisation of protein and chromogranins of sheep sympathetic neurones and their release by nerve impulses. *Journal of Physiology* **204**, 58–59P.
- GOLDFIEN A, GANONG WF (1962) Adrenal medullary and adrenal cortical response to stimulation of diencephalon. *American Journal of Physiology* **202**, 205–211.
- GOVONI S, HANBAUER I, HEXUM TD, YANG H-YT, KELLY GD et al. (1981) In vivo characterization of the mechanisms that secrete enkephalin-like peptides stored in dog adrenal medulla. Neuropharmacology 20, 639–645.
- GRANT JK, FORREST AMP, SYMINGTON T (1957) The secretion of cortisol and corticosterone by the human adrenal cortex. Acta Endocrinological (Copenhagen) 26, 195–207.
- HADJIAN AJ, GUIDICELLI C, CHAMBAZ EM (1982) Cholinergic muscarinic stimulation of steroidogenesis in bovine adrenal cortex fasciculata cell suspensions. *Biochimica et Biophysica Acta* 714, 157–163.
- HARTMAN FA, BROWNELL KA, LIU Y (1955) Bloodflow through the dog adrenal. American Journal of Physiology 180, 375–377.
- HASHIMOTO K, MURAKAMI K, HATTORE T, NIIMI M, FUJINO K et al. (1984) Corticotrophin releasing factor (CRF)-like immunoreactivity in the adrenal medulla. *Peptides* 5, 707–712.
- HEXUM TD, HANBAUER I, GOVONI S, YANG H-YT, COSTA E (1980) Secretion of enkephalin-like peptide from canine adrenal gland

following splanchnic nerve stimulation. Neuropeptides 1, 137–142.

- HINSON JP, VINSON GP, WHITEHOUSE BJ, PRICE GM (1986) Effects of stimulation on steroid output and perfusion medium flow rate in the isolated perfused rat adrenal gland *in vitro*. Journal of Endocrinology **109**, 279–285.
- HINSON JP, VINSON GP, PUDNEY J, WHITEHOUSE BJ (1989) Adrenal mast cells modulate vascular and secretory responses in the intact adrenal gland of the rat. *Journal of Endocrinology* 121, 253–260.
- HINSON JP (1990) Paracrine control of adrenocortical function: a new role for the medulla? *Journal of Endocrinology* **124**, 7–8.
- HINSON JP, VINSON GP (1990) Calcitonin gene-related peptide stimulates adrenocortical function in the isolated perfused rat adrenal gland in situ. *Neuropeptides* **16**, 129–133.
- HINSON JP, KAPAS S, TEJA R, VINSON GP (1991*a*) Effect of the endothelins on aldosterone secretion by rat zona glomerulosa cells *in vitro*. Journal of Steroid and Biochemical Molecular Biology **40**, 437–439.
- HINSON JP, VINSON GP, KAPAS S, TEJA R (1991 b) The relationship between adrenal vascular events and steroid secretion: the role of mast cells and endothelin. *Journal of Steroid and Biochemical Molecular Biology* **40**, 381–389.
- HINSON JP, VINSON GP, KAPAS S, TEJA R (1991c) The role of endothelin in the control of adrenocortical function: stimulation of endothelin release by ACTH and the effects of endothelin-1 and endothelin-3 on steroidogenesis in rat and human adrenocortical cells. *Journal of Endocrinology* **128**, 275–280.
- HÖKFELT T, LUNDBERG JM, SCHULTZBERG JM, FAHRENKRUG J (1981) Immunohistochemical evidence for a local VIP-ergic neuron system in the adrenal gland of the rat. Acta Physiologica Scandinavica 113, 575–576.
- HOLZBAUER M, VOGT M (1957) Functional changes produced in the adrenal cortex of the rat by administration of corticotrophin. *Journal of Physiology* **138**, 449–459.
- HOLZWARTH MA (1984) The distribution of vasoactive intestinal peptide in the rat adrenal cortex and medulla. *Journal of the Autonomic Nervous System* 11, 269–283.
- HOLZWARTH MA, CUNNINGHAM LA, KLEITMAN N (1987) The role of the adrenal nerves in the regulation of adrenal cortical functions. Annals of the New York Academy of Sciences 512, 449–464.
- IP NY, ZIGMOND RE (1984) Pattern of presynaptic activity can determine the type of neurotransmitter regulating a postsynaptic event. *Nature* **311**, 472–474.
- JONES CT, EDWARDS AV (1990*a*) Adrenal responses to corticotrophin-releasing factor in conscious hypophysectomized calves. *Journal of Physiology* **430**, 25–36.
- JONES CT, EDWARDS AV (1990*b*) Release of adrenocorticotrophin from the adrenal gland in the conscious calf. *Journal of Physiology* **426**, 397–407.
- JONES CT, EDWARDS AV, BLOOM SR (1990) The effect of change in adrenal blood flow on adrenal cortical responses to adrenocorticotropin in conscious calves. *Journal of Physiology* **429**, 377–386.
- JONES CT, EDWARDS AV (1991) Muscarinic adrenal responses to acetylcholine in conscious calves. *Journal of Physiology* 444, 605–614.
- JONES CT, EDWARDS AV, BLOOM SR (1991) Endocrine responses to intra-aortic infusions of acetylocholine in conscious calves. *Journal of Physiology* **439**, 481–499.
- KANEKO M, KANEKO K, SHINSAKO J, DALLMAN MF (1981) Adrenal sensitivity to adrenocorticotropin varies diurnally. *Endocrinology* 109, 70–75.
- KESSE WK, PARKER TL, COUPLAND RE (1987) The innervation of the adrenal gland. I. The source of pre- and postganglionic nerve fibres to the rat adrenal gland. *Journal of Anatomy* **157**, 33-41.
- KIRPEKAR SM, PRAT JC, SCHIAVONE MT (1982) Effect of muscarine

on release of catecholamines from the perfused adrenal gland of the cat. *British Journal of Pharmacology* 77, 455–460.

- KLEITMAN N, HOLZWARTH MA (1985*a*) Compensatory adrenal growth is inhibited by sympathectomy. *American Journal of Physiology* 248, E261–263.
- KLEITMAN N, HOLZWARTH MA (1985b) Catecholaminergic innervation of the rat adrenal cortex. Cell and Tissue Research 241, 139–147.
- KLEPPING J (1956) Modalités sécrétoires de la médullosurrénale en fonction de la fréquence de stimulation du nerf splanchnique. Comptes Rendus de la Société de Biologie 150, 705–708.
- KNIGHT DE, BAKER PF (1986) Observations on the muscarinic activation of catecholamine secretion in the chick adrenal. *Neuroscience* 19, 357–366.
- KONG JY, THYRESAN-KLEIN A, KLEIN RL (1989) Differential distribution of neuropeptides and serotonin in pig adrenal glands. *Neuroscience* 28, 765–775.
- KOWAL J, HORST I, PENSKY J, ALFONZO M (1977) A comparison of the effects of ACTH, vasoactive intestinal peptide and cholera toxin on adrenal cAMP and steroid synthesis. *Annals of the New York Academy of Sciences* 297, 314–328.
- KREIGER DP (1979) Plasma ACTH and corticosteroids. In *Endocrinology* vol. 2 (ed. L. J. Groot, G. F. Cahill, L. Martin, D. H. Nelson, W. D. Odell et al.), pp. 1139–1156. New York: Grune & Stratton.
- KUMAKURA K, KAROUM F, GUIDOTTI A, COSTA E (1980) Modulation of nicotinic receptors by opiate receptor agonists in cultured adrenal chromaffin cells. *Nature* 283, 489–492.
- KURAMOTO K, KONDO H, FUJITA T (1987) Calcitonin gene-related peptide (CGRP)-like immunoreactivity in scattered chromaffin cells and nerve fibres in the adrenal gland of rats. *Cell and Tissue Research* 247, 309–315.
- L'AGE M, GONZALEZ-LUQUE A, YATES EF (1970) Adrenal blood flow dependence of cortisol secretion rate in unanesthetized dogs. *American Journal of Physiology* **219**, 281–287.
- LEBOULANGER F, LEROUX P, DELARUE C, TONON MC, CHARNAY Y et al. (1983) Co-localisation of vasoactive intestinal peptide (VIP) and enkephalin in chromaffin cells of the adrenal gland of amphibia. Stimulation of corticosteroid secretion of VIP. *Life Sciences* 32, 375–383.
- LEDBETTER FH, KIRSCHNER N (1975) Studies of chick adrenal medulla in organ culture. *Biochemical Pharmacology* 24, 967–974.
- LEE FL, TRENDELENBERG U (1967) Muscarinic transmission of preganglionic impulses to the adrenal medulla in the cat. Journal of Pharmacology and Experimental Therapeutics 158, 73-79.
- LIGHTLY ERT, WALKER SW, BIRD IM, WILLIAMS BC (1990) Subclassification of  $\beta$ -adrenoceptors responsible for steroidogenesis in primary cultures of bovine adrenocortical zona fasciculata/reticularis cells. *British Journal of Pharmacology* **99**, 709-712.
- LIVETT BG, DAY R, ELDE RP, HOWE PRC (1982) Co-storage of enkephalins and adrenaline in the bovine adrenal medulla. *Neuroscience* 7, 1323–1332.
- MAIER R, STAEHLIN M (1968) Adrenal responses to corticotrophin in the presence of an inhibitor of protein synthesis. Acta Endocrinologica (Copenhagen) 58, 619–629.
- MALMEJAC J (1964) Activity of the adrenal medulla and its regulation. *Physiological Reviews* 44, 186–218.
- MARLEY PD, MITCHELHILL KI, LIVETT BG (1986*a*) Effects of opioid peptides containing the sequence of met<sup>5</sup>-enkephalin or Leu<sup>5</sup>-enkephalin on nicotine-induced secretion from bovine adrenal chromaffin cells. *Journal of Neurochemistry* **46**, 1–11.
- MARLEY PD, MITCHELHILL KI, LIVETT BG (1986b) Metorphamide, a novel endogenous adrenal opioid peptide, inhibits nicotineinduced secretion from bovine adrenal chromaffin cells. Brain Research 363, 10–17.
- MARLEY PD, LIVETT BG (1987) Effects of opioid compounds on desensitization of the nicotinic response of isolated bovine

adrenal chromaffin cells. *Biochemical Pharmacology* 36, 2937–2944.

- MARLEY PD, BUNN SJ (1988) Lack of effect of opioid compounds on angiotensin II responses of bovine adrenal medullary cells. *Neuroscience Letters* **90**, 343–348.
- MARLEY PD, BUNN SJ, LIVETT BG (1988) Prostanoid responses of bovine adrenal medullary cells: lack of effect of opioids. *European Journal of Physiology* **145**, 173–181.
- MAZZOCHI G, REBUFFAT P, MENEGHELI V, NUSSDORFER GG (1989) Effects of the infusion with ACTH or CRF on the secretory activity of rat adrenal cortex. *Journal of Steroid Biochemistry* 32, 841–843.
- METTERS KM, ROSSIER J (1984) Biosynthesis of the opioid peptides. In Les Colloques de l'INSERM, Spinal Opioids and the Relief of Pain (ed. J. M. Besson & Y. Lazorthes). INSERM 127, 31-40.
- MORERA AM, CATHIARD AM, LABURTHE M, SAEZ JM (1979) Interaction of vasoactive intestinal peptide (VIP) with a mouse adrenal cell line (Y-1): specific binding and biological effects. *Biochemical and Biophysical Research Communications* 90, 78-85.
- OKA M, ISOSAKI M, WANTANABE J (1982) Calcium influx and catecholamine release in isolated bovine adrenal medullary cells: effects of nicotinic and muscarinic stimulation. In Advances in the Biosciences, vol. 36 Synthesis, Storage and Secretion of Adrenal Catecholamines (ed. F. Izumi, M. Oka & K. Kumaakara), pp. 29-36. Oxford: Pergamon Press.
- OTTENWELLER JE, MEIER AH, FERRELL BR, HORSEMAN ND, PROCTOR A (1978) Extrapituitary regulation of the circadian rhythm of plasma corticosteroid concentration in rats. *Endo*crinology 103, 1875–1879.
- OTTENWELLER JE, MEIER AH (1982) Adrenal innervation may be an extrapituitary mechanism able to regulate adrenocortical rhythmicity in rats. *Endocrinology* 111, 1334–1338.
- PELTO-HUIKKO M, SALMINEN T, HERNOVEN A (1982) Enkephalinlike immunoreactivity is restricted to the adrenaline cells in the hamster adrenal medulla. *Histochemistry* **73**, 493–497.
- PORTER ID, WHITEHOUSE BJ, TAYLOR AH, NUSSEY SS (1988) Effect of arginine vasopresin and oxytocin on acetylcholine-stimulation of corticosteroid and catecholamine secretion from the rat adrenal gland perfused *in situ. Neuropeptides* **12**, 265–271.
- PORTER JC, KLAIBER MS (1965) Corticosterone secretion in rats as a function of ACTH input and adrenal blood flow. *Federation Proceedings* 24, 383.
- PRATT JH, TURNER DA, MCATEER JA, HENRY PD (1985)  $\beta$ adrenergic stimulation of aldosterone production by rat adrenal capsular explants. *Endocrinology* 117, 1189–1194.
- RAPELA CE (1956) Differential secretion of adrenaline and noradrenaline. Acta Physiologica Latinoamericana 6, 1-14.
- RAPELA CE, COVIAN MR (1954) Fréquence de stimulation des nerfs splanchnique et sécrétion d'adrénaline et de noradrénaline. *Comptes Rendus de la Société de Biologie* **48**, 1667–1669.
- REDGATE ES, GELLHORN E (1953) Nature of sympathetico-adrenal discharge under conditions of excitation of central autonomic structures. *American Journal of Physiology* **174**, 475–480.
- ROBINSON PM, PERRY RA, HARDY KJ, COGHLAN JP, SCOGGINS BA (1977) The innervation of the adrenal cortex of the sheep, *Ovis* ovis. Journal of Anatomy 124, 117–129.
- ROISIN MP, ARTOLA A, HENRY JP, ROSSIER J (1983) Enkephalins are associated adrenergic granules in bovine adrenal medulla. *Neuroscience* 10, 83–88.
- ROLE LW, PERLMAN RL (1983) Both nicotinic and muscarinic receptors mediate catecholamine secretion by isolated guinea pig chromaffin cells. *Neuroscience* 10, 979–985.
- ROSENFELD G (1955) Stimulative effect of acetylcholine on the adrenocortical function of isolated perfused calf adrenals. *American Journal of Physiology* 183, 272–278.
- Rossier J, Barres E, CUPO A, EDWARDS AV (1988) The release of enkephalin-containing peptides from the adrenal gland in conscious calves. In *Neurosecretion: Cellular Aspects of the*

Production and Release of Neuropeptides (ed. B. T. Pickering, J. B. Wakerley & A. J. S. Summerlee), pp. 53–59. New York: Plenum Publishing Corporation.

- SAKIMA NT, BRESLOW MJ, RAFF H, TRAYSTMAN RJ (1991) Lack of coupling between adrenal cortical metabolic activity and blood flow in anesthetized dogs. *American Journal of Physiology* 261, H410-415.
- SAPIRSTEIN LA, GOLDMAN HA (1959) Adrenal blood flow in the albino rat. American Journal of Physiology 196, 159-162.
- SCHNEIDER AS, CLINE HT, LEMAIRE S (1979) Rapid rise in cyclic AMP accompanies catecholamine secretion in suspensions of isolated adrenal chromaffin cells. *Life Sciences* 24, 1389–1394.
- SCHULTZBERG M, LUNDBERG JM, HÖKFELT T, TERENIUS L, BRANDT J et al. (1978) Enkephalin-like immunoreactivity in gland cells and nerve terminal of the adrenal medulla. *Neuroscience* 3, 1169–1186.
- SPARROW RA, COUPLAND RE (1987) Blood flow to the adrenal gland of the rat: its distribution between the cortex and medulla before and after haemorhage. *Journal of Anatomy* **155**, 51–56.
- SRIVASTA AK, MEIER AH (1972) Daily variation in concentration of cortisol in plasma in intact and hypophysectomized killifish. *Science* 177, 185–187.
- STARK E, VARGA B, ACS Z, PAPP M (1965) Adrenal bloodflow response to adrenocorticotrophic hormone and other stimuli in the dog. *Pflüger's Archives für gesamte Physiologie* 285, 296–301.
- STERN AS, LEWIS RV, KIMURA S, ROSSIER J, GERBER LD et al. (1979). Isolation of the opioid heptapeptide met-enkephalin [arg<sup>6</sup>, phe<sup>7</sup>] from bovine adrenal medullary granules and straitum. *Proceedings of the National Academy of Sciences of the USA* **76**, 6680–6683.
- STJÄRNE L, STJÄRNE E, MSGHINA M, BAO J-X (1991) K<sup>+</sup> and Ca<sup>+</sup> channel blockers may enhance or depress sympathethic transmitter release via a Ca<sup>2+</sup> dependent mechanism 'upstream' of the release site. Neuroscience 44, 673–692.
- STONE D, HECHTER O (1954) Studies on ACTH action in perfused bovine adrenal: the site of action of ACTH in corticosteroidogenesis. Archives of Biochemistry and Biophysics 51, 457-469.
- SUDA T, TOMORI N, TOZAWA F, MOURI, DEMURA H et al. (1984) Distribution and characterization of immunoreactive corticotrophin-releasing factor in human tissues. *Journal of Clinical Endocrinology and Metabolism* **55**, 861–867.

- TRIFARO JM, LEE RWH (1990) Morphological characteristics and stimulus-secretion coupling in bovine adrenal chromaffin cell cultures. *Neuroscience* 5, 1533–1546.
- URQUHART J (1965) Adrenal bloodflow and the adrenocortical response to corticotropin. *American Journal of Physiology* 209, 1162–1168.
- VINSON GP, PUDNEY JA, WHITEHOUSE BJ (1985) The mammalian adrenal circulation and the relationship between adrenal bloodflow and steroidogenesis. *Journal of Endocrinology* 105, 285–294.
- VIVEROS OH, DILIBERTO EJ JR, HAZUM E, CHANG K-J (1979) Opiate-like materials in the adrenal medulla: evidence for storage and secretion with catecholamines. *Molecular Pharmacology* 16, 1101–1108.
- WAKADE AR, WAKADE TD (1983) Contribution of nicotinic and muscarinic receptors in the secretion of catecholamines evoked by endogenous and exogenous acetylcholine. *Neuroscience* 10, 973–978.
- WALKER SW, LIGHTLY ERT, MILNER SW, WILLIAMS BC (1988) Catecholamine stimulation of cortisol secretion by 3-day primary cultures of purified zona fasciculata/reticularis cells isolated from bovine adrenal cortex. *Molecular and Cellular Endocrinology* 57, 139–147.
- WILKINSON CW, SHINSAKO J, DALLMAN MF (1982) Rapid decreases in adrenal and plasma corticosterone concentrations after drinking are not mediated by changes in plasma adrenocorticotropin concentration. *Endocrinology* **110**, 162–169.
- WOOD CE, SHINSAKO MJ, DALLMAN MF (1982a) Comparison of canine corticosteroid responses to mean and phasic increases in ACTH. American Journal of Physiology 242, E102–108.
- WOOD CE, SHINSAKO J, KEIL JC, RAMSAY DJ, DALLMAN MF (1982b) Apparent dissociation of ACTH and corticosteroid responses to 15 ml/kg haemorrhage in conscious dogs. *Endo*crinology 110, 1416–1421.
- WRIGHT RD (1963) Blood flow through the adrenal gland. Endocrinology 72, 418-428.
- YANAGIHARA N, ISOSAKI M, OHUCHI T, OKA M (1979) Muscarinic receptor-mediated increase in cyclic GMP levels in isolated bovine adrenal medullary cells. FEBS Letters 105, 296–298.
- YOSHIKAZI T (1975) Effects of cholinergic drugs and their blockers on adrenaline release from rat adrenal. *Biochemical Pharmacology* **12**, 1401–1405.