# CCLXVI. THE CATALYTIC EFFECT OF BUFFERS ON THE REACTION $CO_2+H_2O \rightleftharpoons H_2CO_3$

BY F. J. W. ROUGHTON AND V. H. BOOTH<sup>1</sup>

From the Physiological Laboratory, Cambridge

(Received 1 October 1938)

FAURHOLT [1925, 1] has shown that in aqueous solution  $CO_2$  takes part in two independent reactions

$$CO_2 + H_2O \rightleftharpoons H_2CO_3 \rightleftharpoons H^+ + HCO_3^-, \qquad \dots \dots (1)$$
  
$$CO_2 + OH^- \rightleftharpoons HCO_3^-. \qquad \dots \dots (2)$$

The velocity constant of the reaction  $CO_2 + OH^- \rightarrow HCO_3^-$  is more than a million times greater than the velocity constant of the reaction  $CO_2 + H_2O \rightarrow H_2CO_3$ , the loss of the proton from the  $H_2O$  molecule apparently causing, as Faurholt has emphasized, an enormous increase in the affinity for  $CO_2$ . In consequence of this, it is only below pH 8 that the rate of (2) becomes negligible in comparison with the rate of (1): in the range pH 9-10 the two rates are of the same order, whilst above pH 10 the rate of (2) becomes predominant.

Similar studies on the rate of combination of  $CO_2$  with very weakly acidic organic hydroxides, such as methyl alcohol, ethyl alcohol and glucose, have shown [Faurholt, 1927] that  $CO_2$  reacts slowly and in an analogous way with the organic hydroxide molecule:

$$\operatorname{CO}_2 + \operatorname{HO}_3 X$$
. .....(3)

The inhibitory effect of the proton is again shown by the fact that the reaction with the organic anion—

$$\operatorname{CO}_2 + OX^- \rightleftharpoons \operatorname{CO}_3 X^- \qquad \dots \dots (4)$$

In the case of stronger oxy-acids the affinity of the anion for H<sup>+</sup> is correspondingly lower, and Faurholt tacitly assumed that the affinity of the anion for CO<sub>2</sub> would fall *pari passu* and hence that acids with pK of the order of 7 (e.g. cacodylic acid) could be safely used as buffers in the measurements of the velocity constants of the reactions  $CO_2 + H_2O \rightleftharpoons H_2CO_3$  and  $CO_2 + OH^- \rightleftharpoons HCO_3^-$ , without fear of the buffer taking any part other than the "instantaneous" supply or removal of the H<sup>+</sup> ions involved in the ionization of carbonic acid, i.e.  $H_2CO_3 \rightleftharpoons H^+ + HCO_3^-$ . The possibility of any direct reaction of the buffer with  $CO_2$ ,  $H_2CO_3$  or  $HCO_3^-$  (except in the case of the borate ion, which Faurholt suggested might form some special complex with  $CO_2$ ) was thus implicitly excluded, not only by Faurholt but by all others who have worked in this field.

Preliminary experiments on the activity of carbonic anhydrase in phosphate solutions of varying concentration and pH have, however, prompted us to examine more critically the role of the buffer in reactions (1) and (2). To this end we have measured manometrically both the rate of CO<sub>2</sub> uptake by buffer solutions shaken violently with CO<sub>2</sub>, and the rate of CO<sub>2</sub> output from bicarbonate buffer mixtures, in the presence of a much wider range of buffer concentration

<sup>1</sup> George Henry Lewes Student.

(2049)

and species than heretofore. Our results, especially those in which the buffer concentration is varied 10-fold or more, lead us to conclude that almost all common oxy-acid-salt mixtures which buffer above pH 6, not only promote the CO<sub>2</sub> reactions by supplying or removing H<sup>+</sup> ions, but also through their more negative constituents catalyse directly both phases of the reaction  $CO_2 + H_2O \rightleftharpoons H_2CO_3$ . A preliminary account of these results has already appeared [Booth & Roughton, 1938]. Faurholt's discovery of the activity of the borate ion is thus only a special, though pronounced, example of a general principle.

It was not until nearly the end of the research that we investigated the effect of nitrogen bases as buffers. We were deterred from doing so by the fact that straight-chain nitrogenous bases, such as  $NH_3$  or  $CH_2NH_2COO^-$ , combine rapidly and reversibly with  $CO_2$  to form carbamino compounds, according to the following scheme [Faurholt, 1925, 2]:

$RNH_2 + CO_2 \rightleftharpoons RNH.COOH,$	(5)
$RNH.COOH \rightleftharpoons H^+ + RNH.COO^-$ ,	(6)
$H^+ + RNH_2 \rightleftharpoons RNH_3^+$ .	(7)

(The inhibitory effect of the proton is shown in this case also by the absence of reaction between  $CO_2$  and  $RNH_3^+$ .) The simultaneous occurrence of this carbamino reaction might make it difficult to detect and measure accurately any catalytic effect on the  $CO_2 + H_2O \rightarrow H_2CO_3$  reaction.

Cyclic nitrogenous bases have not, so far as we know, been investigated as regards their carbamino-forming power, and it was therefore with special interest that we tested some members of the new range of glyoxaline buffers, recently described by Kirby & Neuberger [1938], to whom we are greatly indebted for samples. Both the cations and the molecules of these substances were found to be incapable of any carbamino reaction with  $CO_2$ , but on the other hand the molecules proved to be markedly catalytic towards the reaction  $CO_2 + H_2O \rightarrow H_2CO_3$  especially if their pK were greater than 7.0.  $(pK = -\log_{10}K)$ where  $K = [H^+] \times [$ molecule of base]/[cation of base].) This led us to test such other cyclic nitrogenous bases as were both readily available and sufficiently soluble in water. Similar catalytic activity was always found if the pK were greater than 7, whereas tests of a more extended range of straight-chain N bases showed that these were without catalytic effect, their only reaction with  $CO_2$ , if any, being the carbamino formation. These rather remarkable results are summarized in Section II, together with some preliminary data on sulphhydryl and other compounds.

A possible mechanism of these catalyses is given in the discussion, together with some further points of interest which arise in regard to the enzyme carbonic anhydrase.

# Methods

Rates of  $CO_2$  uptake were determined by an improvement of Meldrum & Roughton's [1933, 2] manometric boat method. The apparatus (Fig. 1) consists of a lagged tank, a stationary manometer containing toluene coloured with sudan III as gauge liquid, a compensating bottle E (about 200 ml.), a  $CO_2$ reservoir D (about 200 ml.) which can be connected through fine thermometer tubing R to a 60 ml. long-necked glass "boat" and a mechanism, not shown, for shaking the boat about AR as axis at an adjustable speed. The shaking must be rapid enough not to limit the rate of  $CO_2$  uptake, but not so violent as to splash solution up into the tube connecting with the manometer. At 315 oscillations per min. the limiting speed is exceeded without the readings becoming erratic. This speed is kept constant by a synchronous motor and is indicated by a neon lamp stroboscope.

The procedure in an experiment is as follows. The tank is emptied, and dried air is drawn through the connecting tubes to remove condensed moisture. In the meantime the test solution is measured into the "boat", which is stoppered and pushed into a holder in the shaker which holds it firmly by means of two rubber sleeves encircling the boat, all clips being open except  $C_4$ . The tank is then filled with water and the stirrer turned on. D is repeatedly evacuated and filled with CO<sub>2</sub> and left with a CO<sub>2</sub> pressure of 30-40 cm. Hg, the clips being closed and water-sealed. The boat, compensating bottle and manometer are evacuated to  $\frac{1}{7}$  atm. through  $C_6$  which is closed and placed below the water surface. After 9 min. the boat is shaken for 2 min. to bring the dissolved gases



Fig. 1. Simplified diagram of apparatus arranged for  $CO_2$  uptake. Not to scale. The boat is oscillated through 50° at right angles to the plane of the paper. Tube F is only used for anaerobic experiments.

in the test solution into equilibrium with the gas phase, and 1 min. is allowed for the solution to drain to the bottom of the boat. The short circuit  $C_5$  between the two limbs of the manometer is closed and the zero reading taken.  $CO_2$  is slowly let into the boat through the resistance R, by manipulation of  $C_4$ , up to the desired pressure. The time taken by this should be about 30 sec. A further period is allowed for the disturbance caused by the admission of the  $CO_2$  to settle down, and then, exactly 2 min. after  $CO_2$  begins to enter, shaking is started and manometer readings taken at 0, 5, 10, 15 sec. etc., up to 5 min. or more. The tank temperature is read. At the end of the experiment the clip  $C_5$  is opened, and air allowed to enter slowly through a capillary inserted at  $C_6$  until all parts of the apparatus are at atmospheric pressure. (If air enters suddenly into the apparatus, when at low pressure, the gauge liquid will shoot over.) Most of the water is meanwhile emptied out of the tank, and the boat then removed, washed and dried.

Biochem. 1938 xxxII

The preliminary equilibration period is 12 min.; if duplicate boats are used and one is prepared during the equilibration of the other, an observer and an assistant can perform serial experiments at the rate of one every 25 min.

With axial shaking, only a slight vibration is transmitted to the manometer and volume variations from distortion of the thick rubber tubing are negligible. Furthermore all the rubber joints can be water-sealed, the dead space between the boat and the liquid meniscus in the manometer cut down to 1 ml., and the volume of the system exposed to room temperature reduced to about 0.2 ml. These improvements, together with the substitution of toluene for water in the manometer gauge, the modified boat shape and the constant speed motor, are responsible for the increase in accuracy and reliability of the method. When the apparatus is working properly, readings should duplicate to 0.2 mm. At temperatures other than that of the room stirring of the water in the tank must be vigorously maintained during equilibration; otherwise the temperature, and therefore the pressure, in the compensating bottle or boat may vary from one another.

From time to time the whole apparatus must be washed out with water and acetone and dried by flushing with dried air. The manometer is then refilled with de-aerated coloured toluene through the rubber tube H. This tube is then filled with de-aerated water to form a seal and immersed in water. A little water is always left in E to saturate the gas with vapour.

When autoxidation, leaks or other complications which would give spurious results by manometric methods were suspected, readings were taken in absence of  $CO_2$  during a prolonged first shaking period. For autoxidizable substances experiments were done under anaerobic conditions, by applying a slight modification of the method used by Meldrum & Roughton [1933, 2] for studying the rate of uptake of  $CO_2$  by reduced blood to the present technique. If necessary one component of the required solution could be weighed solid directly into a dry boat, the oxygen removed by washing out with nitrogen and the remaining de-aerated solution added from a tonometer through F. In this case the substance makes no contact with oxygen whilst in solution.

Unless otherwise stated all experiments by the  $CO_2$  uptake method were done at 0° with 4.2 ml. solution and an initial  $CO_2$  pressure of 12.8 mm. Hg in a total gas space of 60 ml.

In order to avoid the effect of complications due to the  $CO_2 + OH^- \rightarrow HCO_3^-$  reaction, most experiments were done below pH 8.0.

Rates of  $CO_2$  output were determined in the same apparatus but with the double compartment boat method of Meldrum & Roughton [1933, 1]. After equilibration of the boat in the tank the solutions were suddenly mixed (by starting the shaker) and the  $CO_2$  evolution followed manometrically. Most of the experiments were done at  $0^{\circ}$  and at a total gas pressure of  $\frac{1}{5}$  atm.

The majority of the experiments were done with the illustrated apparatus which has however now been simplified. The outlet clip, water-seal etc., at H have been eliminated and the gauge fluid is introduced into the manometer tube through M.

Notes on solutions. Carbonic anhydrase was prepared in semi-purified state by Meldrum & Roughton's [1933, 1] chloroform method.

Buffer solutions were made up with a known acid/ion ratio by adding the calculated quantity of HCl or alkali, and checking the pH or titrating to an unbuffered end-point. The pH of solutions was determined in most cases colorimetrically but for special cases it was measured by the hydrogen electrode.

Na and K salts were used indiscriminately, except when high concentrations were required and one salt (usually K) is markedly more soluble than the other.

#### I. OXY-ACID BUFFERS

The two buffers which have been most used hitherto in work on  $CO_2$  kinetics are phosphate ( $H_2PO_4^- + HPO_4^-$ ) and cacodylate (HCac+Cac<sup>-</sup>), where Cac<sup>-</sup> is

$$CH_{3} \rightarrow O.$$

$$CH_{3} \rightarrow O.$$

We have accordingly investigated these with special care, and since the results and inferences therefrom seem qualitatively the same as for the general class of oxy-acid buffers which function above pH 6, we shall first describe the results with phosphate and cacodylate in detail.

# CO2 uptake experiments

Curve A, Fig. 2, shows the rate of  $CO_2$  uptake by water, recorded manometrically. The gaseous  $CO_2$  pressure drops to a steady value in about 20 sec.: this rapid uptake is simply due to physical solution of  $CO_2$ , the subsequent  $H_2CO_3$  and  $HCO_3^-$  formation being negligible.

Curves B, C, D, Fig. 2, show the effect of increasing concentration of total phosphate, the ratio of  $[HPO_4^{-}]$  to  $[H_2PO_4^{-}]$  being kept constant. In each case there is a similar rapid drop of CO<sub>2</sub> pressure in the first 20 sec. (though the drop may be less owing to the solubility of CO<sub>2</sub> in the buffer being lower than in water), and then a prolonged slow phase in which the dissolved CO<sub>2</sub> enters into chemical combination. The slope of this slow phase is seen to be steeper



Fig. 2. Curves showing the observed course of CO<sub>2</sub> uptake by water and by phosphate at 0°. A, water; B, 0.0095 M phosphate; C, 0.14 M; D, 0.76 M. The pH was 7.4 at 0.2 M.

Fig. 3. Curves showing the expected course of  $CO_2$  uptake by various concentrations of a given buffer, if the buffer itself has no direct effect on the  $CO_2$  reaction. Corrected for effect of changing buffer concentration on physical solubility of  $CO_2$ .

throughout its whole course, as the phosphate concentration is increased; whereas if the phosphate did no more than just "instantaneously" remove the H<sup>+</sup> ions formed by the reaction  $H_2CO_3 \rightarrow H^+ + HCO_3^-$ , the slopes of curves B, C, D should be the same in their early stages, and the increased buffer concentration should exercise no effect until the latter part of the uptake, when the

velocity of the back reactions becomes significant, i.e. a family of curves like those shown in Fig. 3 should have been obtained.

Clearly there is, in addition, some direct effect of the phosphate upon the reaction, as is indeed shown quantitatively by equation (8) which fits the curves plotted in Fig. 2, i.e.

$$-\frac{d [CO_2]}{dt} = v_u [CO_2] = k_u [CO_2] \{1 + l_u [HPO_4^{=}]\}, \qquad \dots \dots (8)$$

where  $v_u$  is the apparent overall velocity constant for CO<sub>2</sub> uptake,

 $k_u$  is the true velocity constant of the reaction  $\rm CO_2 + H_2O \rightarrow H_2CO_3$ ,

 $l_u$  is a catalytic coefficient.

The numerical values of  $v_u$  in Tables I and III have been calculated by the method of Brinkman *et al.* [1933], but a correction has also been inserted for the rate of the back reaction, which in the case of phosphate amounts to 2–20 % and in the case of cacodylate to even higher figures. The allowance for the back reaction was made on usual principles and since it requires a knowledge of the end point of the reaction, this was determined either by continuing the experiment for 30 min. or preferably by a separate experiment in which a suitable amount of carbonic anhydrase was added to the mixture, so that the end point was reached in 5 min.

The time intervals over which  $v_u$  has been calculated have usually been 2 min. each, and in any given experiment at least four such intervals have been chosen between 1.5 and 8.0 min.,  $v_u$  as given in Tables I and III thus being the average of four or more values. The individual figures in any one experiment usually agree with the mean to within 10%.

A rougher, but less laborious, mode of calculation was used for the other data, which are mostly of a preliminary nature. This consisted in measuring the time t for the CO<sub>2</sub> pressure to drop through one or more small specified ranges in the early part of the slow phase of CO<sub>2</sub> uptake. Clearly  $v_u = 1/t \times a$  constant, which is obtained from the curve of a solution of known  $v_u$  (determined as in Table I).



Fig. 4. Effect of [HPO<sub>4</sub><sup>=</sup>] on velocity of CO<sub>2</sub> uptake. ⊙ our experiments; × calculated from data of Brinkman *et al.* [1933].

Fig. 4 shows that there is a linear relation between  $v_u$  and [HPO<sub>4</sub>=], at a ratio of [HPO<sub>4</sub>=]: [H<sub>2</sub>PO<sub>4</sub>-] of 2:1, as equation (8) requires. The corresponding

value of  $l_u$ , namely 8, together with  $l_u$  values for ratios ranging from 8:1 to 1:1 are given in Table I. The  $l_u$  values agree with one another within experimental error, thus demonstrating the validity of equation (8).

[HPO4=]:[H2PO4=]	Conc. range $(total phosphate, M)$	l <sub>u</sub>
8.0	0.01-0.20	7.7
$3 \cdot 2$	0.01-0.20	8.0
2.0	0.01-0.10	7.7
1.0	0.01-0.10	8.5
	Mean	8.0
[Cac-]:[HCac]	Conc. range $(total cacodylate, M)$	l <sub>u</sub>
7.0	0.02-0.20	8.5
4.0	0.02-0.20	<b>9·4</b>
	Mean	9.0

Table I. Effects of phosphate and cacodylate on  $CO_2$  uptake rate

A  $[HPO_4^{-}]: [H_2PO_4^{-}]$  ratio of 1:1 was the lowest that could be used, for below this the CO<sub>2</sub> taken up becomes so small and the back reaction increases so fast that accurate determinations are not feasible. For the same reasons it was impossible to reduce the  $[Cac^{-}]: [HCac]$  ratio below 4:1, for the pK of cacodylic acid is about 0.8 pH below the  $pK_2$  of phosphoric acid, and even at 4:1 the back reaction corrections amount to as much as 30%, though in spite of this the  $v_u$  values over different intervals in any one experiment tally satisfactorily.

Above a total buffer concentration of 0.2 M, the physical solubility of  $CO_2$  is appreciably depressed and the interpretation of the results thus becomes uncertain: in rough experiments, however, up to M it was found that  $v_u$  continued to increase nearly linearly with  $[HPO_4^{-}]$ . In the case of cacodylate  $v_u$ increased rather faster than  $[Cac^{-}]$ : this may be just an ordinary deviation from the law of mass action in strong solution, but it might also be due to some change in the state of the cacodylate at higher concentrations, such as the formation of a polymeride which is catalytically more active. A similar deviation occurs with chromate buffer solutions, in which there is known to be an equilibrium between the hydrochromate ion,  $HCrO_4^-$ , and the dichromate ion,  $Cr_2O_7^-$ , the amount of the latter increasing rapidly as the total chromate concentration is raised.

It was of interest to see whether equation (8) could also be applied to results published by earlier authors before the effect of the buffer was suspected. Adequate data are available in two such papers, and  $l_u$  values calculated therefrom are given in Table II. The excellent agreement with the present findings is a strong confirmation of the validity of our conclusions. Table II further shows that equation (8) is valid over a wide range of CO<sub>2</sub> concentration, namely 0.001 M(present paper) to 0.03 M (Faurholt).

Table II. Values of  $l_{\mu}$  calculated from data in the literature

Source	Buffer	CO <sub>2</sub> , M	l <sub>u</sub>	$l_u$ from Table I
Faurholt [1925, 1, Table IV]	Cacodylate	0.03	9.5	9.0
Faurholt [1925, 1, Table VI]	Phosphate	0.018	8.4	8.0
Brinkman et al. [1933, Table III]	Phosphate	0.002	8.3	8.0

Extrapolation of the data, summarized in Table I (or in Table III [Brinkman et al. 1933]) to zero [HPO<sub>4</sub>=] gives us for the first time an accurate value for  $k_u$ ,

131-3

the true velocity constant of the reaction  $CO_2 + H_2O \rightarrow H_2CO_3$ . The mean value of  $k_u$  is 0.0021, i.e. about 25 % lower than the hitherto accepted value of 0.0027 [Table I, Roughton, 1935, 1], which is, of course, wrong, for in the calculations on which it was based no account was taken of the direct catalytic effect of the buffer.

Arguments against the effect being due to neutral salt action. It will be convenient now to exclude the possibility that the effect of the buffer may be accounted for by some kind of "neutral salt action" or ionic activity factor. This we do on the following grounds.

(a) The observed effects are larger than expected, and indeed are appreciable at concentrations, e.g. 0.01 M, much lower than those at which typical neutral salt action manifests itself. This holds not only for phosphate and cacodylate, but more so for the very active substances, such as selenite, tellurate and sulphite, to be dealt with later.

(b) Table III shows that addition of high concentrations of typical neutral salts, such as KCl and NaNO<sub>3</sub>, to 0.04 M phosphate buffer (1:1), only increases the CO<sub>2</sub> uptake rate to a relatively slight extent, the  $l_u$  values for these salts being at most only one-sixteenth that of phosphate.

Table III. Effect of neutral salts on the rate of  $CO_2$  uptake by 0.04 M phosphate

Salt	$l_u$
$(NH_4)_2SO_4$	0.2
NaNO <sub>a</sub>	<0.05
NaNO,	0
KCl	0.3
NH <sub>4</sub> Cl	<0.05
KI T	0.07
KCNS	0.2
K <sub>3</sub> Fe(CN) <sub>6</sub>	0.4

# $CO_2$ output experiments

If the action of the buffer (or some accompanying impurity) is purely catalytic, then the reverse reaction  $H_2CO_3 \rightarrow CO_2 + H_2O$  should be equally affected. On the face of it, this should be easy to test by measuring the rate of  $CO_2$  output from mixtures of bicarbonate with varying strengths of buffer. Unfortunately, however, variation of the buffer strength will also affect the activities of the ions in solution, and may thus independently alter the rate of  $CO_2$  output, which is directly proportional to the product of the H<sup>+</sup> and HCO<sub>3</sub><sup>-</sup> activities. The exact nature of the possible "activity effects" is shown by the following treatment.

The rate of CO<sub>2</sub> output = 
$$\frac{d [CO_2]}{dt} = v_o [H_2CO_3] = \frac{v_o a_H f_{HCO_3} [HCO_3^{-}]}{K_1}$$
, .....(9)

where

r

Now

 $v_o = \text{overall output velocity constant},$ 

 $a_{\rm H} =$  hydrogen ion activity,

 $f_{\rm HCO_3}$  = activity coefficient of HCO<sub>3</sub><sup>-</sup>,

 $K_1 =$ true first ionization constant of  $H_2CO_3$ .

$$a_{\rm H} = \frac{K_2 f_{\rm H_2PO_4} [\rm H_2PO_4^{--}]}{f_{\rm HPO_4} [\rm HPO_4^{--}]}, \qquad \dots \dots (10)$$

where  $f_{\text{H}_2\text{PO}_4}$ ,  $f_{\text{HPO}_4}$  = activity coefficients of  $\text{H}_2\text{PO}_4^-$  and  $\text{HPO}_4^-$  respectively,  $K_2$  = second ionization constant of  $\text{H}_3\text{PO}_4$ .

Therefore 
$$\frac{d[\text{CO}_2]}{dt} = \frac{v_0 K_2 f_{\text{HCO}_3} f_{\text{H}_2\text{PO}_4}[\text{H}_2\text{PO}_4^{--}]}{K_1 f_{\text{HPO}_4}[\text{HPO}_4^{---}]} [\text{HCO}_3^{---}]. \qquad \dots \dots (11)$$

The activity coefficients of divalent ions, such as  $\text{HPO}_4^{=}$ , are much more affected, and in a more specific manner, than those of univalent ions, by changes in ionic strength, and there is no reason to suppose that the fraction  $f_{\text{HCO}_3} \times f_{\text{H}_2\text{PO}_4}/f_{\text{HPO}_4}$  should remain constant as the phosphate concentration is reduced by dilution with water. But if the buffer is, instead, diluted with a mixture of K<sub>2</sub>SO<sub>4</sub> and KCl, of the same total molarity as the highest concentration of buffer and with the same molar proportions of SO<sub>4</sub><sup>=</sup> to Cl<sup>-</sup> as of HPO<sub>4</sub><sup>=</sup> to H<sub>2</sub>PO<sub>4</sub><sup>-</sup> in the buffer, this difficulty should be practically overcome: for Landolt & Bornstein's Tables show that the activity coefficients of K<sub>2</sub>SO<sub>4</sub> and Na<sub>2</sub>SO<sub>4</sub> agree to within 0.01 with the activity coefficient of Na<sub>2</sub>HPO<sub>4</sub> in pure solutions of the respective salts at the same concentration (up to 0.1 *M*), and it is therefore reasonable to suppose that in phosphate + sulphate mixtures the activity coefficients would be the same as in pure phosphate buffers of the same total molarity. A similar argument is applied to the Cl<sup>-</sup> and H<sub>2</sub>PO<sub>4</sub><sup>-</sup> ions.

All solutions, except bicarbonate, were freed from  $CO_2$  by repeated evacuation and shaking. To compensate for the carbonate content of the bicarbonate solution, an equivalent amount of HCl was placed in the other solution used in the double compartment boat.



Fig. 5. Effect of  $[HPO_4^{=}]$  on velocity of  $CO_2$  output.

The methods of calculation were similar to those used in the CO<sub>2</sub> uptake work. Fig. 5 shows that there is again a linear relation between  $[HPO_4^{=}]$  and  $v_o/K_1$  and we may therefore write

$$\frac{v_o}{K_1} = \frac{k_o}{K_1} \{ 1 + l_o \ [\text{HPO}_4^{-1}] \}, \qquad \dots \dots (12)$$

where  $k_o$  is the true velocity constant of the reaction  $H_2CO_3 \rightarrow CO_2 + H_2O$ , and  $l_o$  is the catalytic coefficient for  $CO_2$  output corresponding to  $l_u$  for  $CO_2$  uptake. Table IV gives the values of  $l_o$  for phosphate + sulphate + chloride mixtures and also for cacodylate + chloride  $\pm$  acetate mixtures. It will be noted that in the case of both buffers  $l_o$  is independent of wide variations in pH and that the  $l_o$  values agree, within experimental error, not only with the  $l_u$  values of Table I but also with the  $l_u$  values given in Table IV, which were specially determined in buffer salt mixtures of the same composition as used in the output experiments. This last result proves that phosphate and cacodylate are both true catalysts of the reaction  $CO_2 + H_2O \rightleftharpoons H_2CO_3$ .

The assumption that the ionic dissociation and reformation of  $H_2CO_3$  are too fast to limit the rates of uptake or output of  $CO_2$ , even when the buffer

Table IV. Effects of phosphate and cacodylate on  $CO_2$  output rate

	$[HPO_4^{=}]: [H_2PO_4^{-}]$	lo	$l_u$
Phosphate $+ SO_4 = + Cl^-$	3:1	8.5	8.9
$Phosphate + SO_4^{-} + Cl^{-}$	1:3	8.6	
	[Cac-] : [HCac]	lo	$l_u$
$Cacodylate + Cl^{-}$	7:1	_	8.5
Cacodylate + Cl-	2:1	9.2	
$Cacodylate + Cl^- + acetate$	1:2	8.9	

concentration is below M/100, perhaps requires some further words. Ionic reactions of this kind are generally assumed to be "instantaneous". A minimum value for their rate is given by rapid reaction velocity experiments such as those of Roughton [1930] which have shown that in presence of M/10 buffer the ionization of weak acids similar to  $H_2CO_3$  has a half-period of less than 0.0003 sec. -probably far less. Whether decrease of buffer concentration would increase the half-period appreciably is very doubtful, since the ionization in complete absence of buffer, i.e.  $H_2CO_3 + H_2O \rightleftharpoons HCO_3^- + H_3O^+$ , is probably per se extremely fast. But, even if the rapid ionization of  $H_2CO_3$  in buffer solutions were entirely due to a direct reaction with the buffer anion, i.e.  $H_2CO_3 + A^- \rightleftharpoons HCO_3^- + AH$ , a decrease of buffer concentration from M/10 to M/100 could not raise the upper limit for the half-period to more than  $10 \times 0.0003$  sec. = 0.003 sec. Now calculation shows that, for the overall rate of CO<sub>2</sub> uptake in M/100 buffer solution to be retarded appreciably by the ionization of  $H_2CO_3$ , the half-period for the latter must be >0.01 sec. The action of the buffers cannot therefore be due to any effect on the ionic reaction  $H_2CO_3 \rightleftharpoons H^+ + HCO_3^-$ , but must be due entirely to their effect on the molecular reactions of CO<sub>2</sub> with water. Of several further points in favour of this we need only mention that the numerical values of  $k_{\mu}$ and  $k_o/K_1$ , as found by extrapolating to zero buffer concentration the respective rates of CO<sub>2</sub> uptake and output, check satisfactorily in two independent ways with the requirements of the law of mass action when applied to the molecular reaction  $CO_2 + H_2O \rightleftharpoons H_2CO_3$ . (Details will be given in a later paper by one of us.) If the rate of ionization of  $H_2CO_3$  were of any limiting influence at low buffer concentrations no such checks would be obtained.

Additive test. An experimental comparison of the rates of  $CO_2$  output from a mixture containing

(a) 0.017 M NaHCO<sub>3</sub>+0.075 M K<sub>2</sub>HPO<sub>4</sub>+0.025 M KH<sub>2</sub>PO<sub>4</sub>+0.1 M KCac, with

(b) 0.017 M NaHCO<sub>3</sub>+0.075 M K<sub>2</sub>HPO<sub>4</sub>+0.025 M KH<sub>2</sub>PO<sub>4</sub>+0.1 M Na accetate,

showed that the rate of (a) was 1.52 times that of (b). This demonstrates that the catalytic effects of the phosphate and cacodylate ions are additive, for on this basis we should have

 $\frac{\text{rate of } (a)}{\text{rate of } (b)} = \frac{1 + (8.55) (0.075) + (9.05) (0.1)}{1 + (8.55) (0.075) + (0.6) (0.1)} = 1.49.$ 

In this fraction 8.55, 9.05 and 0.6 are the respective  $l_o$  values of HPO<sub>4</sub><sup>=</sup>, Cac<sup>-</sup> and CH<sub>3</sub>COO<sup>-</sup> (Table IV).

The observed and calculated ratios agree much better than might have been expected from the size of the experimental errors.

Evidence that the catalytic action of the buffers is not due to an accompanying impurity. An impurity present in the salts and/or alkalis used in making the buffer solutions might be responsible for the catalytic effects observed. The alkalis should first be considered, for they are used both by the manufacturer in preparing salts, such as  $\rm KH_2PO_4$ , and by ourselves in making up the buffers. Two tests eliminate this possibility. (a) The effects are the same whether NaOH or KOH is used. (b) No increased effect is found when the buffer is treated with additional alkali if the latter is afterwards neutralized with an equivalent amount of HCl. It will be recalled that Cl<sup>-</sup> itself does not inhibit—see Table III.

As regards the phosphate, we find exactly the same amount of catalysis whether all the phosphate solution is made up from  $KH_2PO_4$  (Kahlbaum puriss. or AnalaR) or from Na<sub>2</sub>HPO<sub>4</sub>, 12H<sub>2</sub>O (Kahlbaum puriss. or AnalaR). It seems unlikely that all these reagents would contain just the same amount of catalytic impurity. Furthermore since the effect is always proportional to the  $[HPO_4^{-}]$ or [Cac<sup>-</sup>], the impurity would have to have the same *p*H-activity curve as both phosphate and cacodylate, the *pK* of which differ by 0.8. This is impossible if the impurities in the two cases are the same, whereas if there are two different impurities it would be a remarkable coincidence that their respective *p*H-activity curves should both happen to be identical with the ionization curves of the corresponding buffers.

Addition of sufficient carbonic anhydrase to double the rate of  $CO_2$  uptake by M/50 phosphate, is also found to double the rate of  $CO_2$  uptake by M/5phosphate which, in absence of enzyme, it will be remembered, is nearly twice the rate of uptake by M/50 phosphate. The catalytic actions of carbonic anhydrase and phosphate thus appear to multiply one another instead of being merely additive, as would be expected if the phosphate catalysis were due to traces of an impurity. A possible mechanism for the multiplicative effect is put forward later.

That the effect can be due to metallic impurities is very unlikely because:

(a) Certain metals, notably Fe, are precipitated by phosphate at pH 8.0, yet the supernatant fluid from phosphate solutions which had been kept a long time showed the same effect as freshly prepared solutions.

(b) No increased effect was observed on adding to 0.04 M phosphate, pH 7.1, various possible impurities in traces, including many cations, e.g. Li<sup>+</sup>, Cu<sup>+</sup>, Cu<sup>++</sup>, Ca<sup>++</sup>, Sr<sup>++</sup>, Ba<sup>++</sup>, Mg<sup>++</sup>, Hg<sup>++</sup>, Pb<sup>++</sup>, Fe<sup>++</sup>, Fe<sup>+++</sup>, Sb, La, U, Sn. The following cations had no (or very slight) additive effect on the rate of CO<sub>2</sub> uptake by cacodylate buffer, pH 6.8: Ca<sup>++</sup> (0.2 M), Ba<sup>++</sup> (0.2 M), Sr<sup>++</sup> (0.2 M), Th<sup>++++</sup> (0.001 M), La<sup>+++</sup> (0.0005 M). At 0.0006 M, however, La<sup>+++</sup> had a marked accelerating effect, but at this concentration a precipitate developed in the boat. 0.012 M Cu<sup>++</sup> and 0.1 M Rb<sup>+</sup> in  $\beta$ -glycerophosphate, pH 6.8, had no effect.

(c) The rate of CO<sub>2</sub> uptake by phosphate, pH 7·1, is unaffected by adding 0.08 M cyanide.

This varied evidence, taken as a whole, makes it safe for the present to attribute the catalytic action in the main, if not entirely, to the more electronegative constituent of the buffer itself.

# Further experiments on the nature of the catalysis

The natural question next arises as to whether this effect is limited to the cacodylate, the secondary phosphate and the borate ions (Faurholt's work) or whether it is shown by the more negative constituent of oxy-acid buffers in general. We have tested the matter by  $CO_2$  uptake experiments on a wide range of buffers, either in pure solution or when mixed with phosphate buffer, in which case their additive effect, if any, on the rate of uptake by phosphate has been worked out. At pH > 9.0 results become less easy to interpret owing to the

appreciable intervention of the  $CO_2 + OH \rightarrow HCO_3$  reaction, and we have therefore, for the present, restricted ourselves in the main to buffers with acid constituents of pK below 9.0. Our object being to make a broad preliminary survey of the whole field, we have in most cases contented ourselves with the rougher method of calculating  $l_u$  described above, and we have neglected any complications which may arise from the possible effects upon the solubility of  $CO_2$  of certain of the substances which were used in high concentration: for these reasons we do not claim the same exactitude for these results as for those given in Tables I and IV, though we do believe that their order of magnitude is correct. The values of  $l_u$ , in terms of the more negative constituent of the buffer, are

	Buffer	<i>рК</i> а	t 0°	[ion] : [acid]	[ion] + [acid],* 1	lí l <sub>u</sub>
I.	Phosphate Phosphite	$pK_2$ $pK_3$	7·1 6·7	2.0	0.20	8·0 6
	Pyrophosphate		-		_	· 10–50
	α-Glycerophosphate		6·4	10.0	0.40	4.5
	β-Glycerophosphate		6.3	3.0	0.25	<b>3</b> ·0
	Phosphoglycerate		6.0	18.0	0.12	2.5
	Hexosediphosphate		6.3	9.0	0.23	8.5
	Cacodylate		6.3			9.0
	Arsenate	$pK_2$	<b>6</b> ∙8	$2 \cdot 0$	1.20	6.0
II.	Maleate	$pK_2$	6.1	8.0	0.95	2.0
					1.00	1.64
	Citrate	$pK_3$	$5 \cdot 4$	c. 20·0	2.00	1.5†
	Veronal		<b>8</b> ∙0	3∙0	0.18	<b>8</b> ∙0
III.	Chromate	$pK_{\bullet}$	6.4	3.0	0.1	c. 50
	Borate	· ·	9.5	0.016	1.45	c. 150†
IV.	Sulphite	$pK_2$	<b>7</b> ·0	3.0	0.1	900
	Selenite	$pK_2$	<b>8</b> ∙0	0.3	0.02	1700
					0.0036	$>2000^{+}$
	Tellurate	$pK_1$	7.8	1.0	0.05	600
v.	Formate		3.8	_	1.0	0.5†
	Acetate		<b>4</b> ·7		1.0	0.64
	Phthalate	$pK_2$	5.3	-	0.8	0·8†
	Oxalate	$pK_2$	<b>4</b> ·1		0.6	1.44

Table V. Effect of various oxy-acid buffers on rate of  $CO_2$  uptake

\* Highest total concentration used.

 $\dagger$  Measured in 0.04 M phosphate buffer.

given in Table V which shows clearly that the effect sought for is very widespread, being considerable with all oxy-acids of pK > 6, and in general increasing with pK. For convenience in discussion Table V has been divided into the following sections.

Group I consists of buffers closely related in structure to phosphate and cacodylate, and with pK in the range 6.0-7.0. Phosphite, with a  $pK_2$  rather lower than the  $pK_2$  of phosphate, has an  $l_u$  value also slightly less (i.e. 6.0 compared with 8.0). The organic phosphates with still lower pK have their  $l_u$  values correspondingly lower, except hexosediphosphate. But the molecule of this substance has two phosphate groups, each with pK at 6.3. Hence for calculating  $l_u$ , a M solution of hexosediphosphate should from the present point of view correspond to a 2.0 M solution of the ion. On this basis,  $l_u$  comes to be of the same order as for the other organic P compounds. Pyrophosphate shows considerable catalytic effect, but it is present both in the trebly and quadruply ionized forms under the pH conditions studied, and the respective catalytic contributions of these two forms have not yet been worked out.

Group II. Two organic acids, maleate and citrate, with pK near the lower limit at which activity is shown, have rather small  $l_u$  values. Veronal with a pK of 8.0 shows, as would be expected, a higher  $l_u$  value.

Group III. The two inorganic buffers chromate and borate have  $l_u$  values about 10 times greater than those in group I. In the case of chromate there is, as already mentioned, some uncertainty owing to the presence in solution of appreciable amounts of  $Cr_2O_7^{-}$ , besides the  $HCrO_4^{-}$  and  $CrO_4^{-}$  ions.

Faurholt [1925, 1] commented on the anomalous results found by him with borate even at pH 8.0, where complications arising from the  $CO_2 + OH^- \rightarrow HCO_3^$ reaction should only be slight. Inspection of his figures shows that the discrepancies to which he refers could be explained if we assign an  $l_u$  value to the borate ion of the order of 100–200—a reasonable figure in view of the pK of boric acid, i.e. 9.5. In borate-phosphate mixture we find an  $l_u$  value of about 150, thus confirming this suggestion.

Group IV. Sulphite, selenite and tellurate,<sup>1</sup> showed the greatest effects of any so far tested,  $l_u$  being in the neighbourhood of 1000. Careful controls showed that the results were not due to autoxidation. Their activity, being so much greater than that of other substances of similar pK, must be due to their special chemical constitution: in this connexion it is of interest to find this high activity associated with three elements of the same group (VI) of the Periodic Table. A further point of interest about the action of selenite will be discussed later.

The effects of chromate and selenite on  $CO_2$  output were also investigated. The  $l_0$  values in the two cases were found to be of the same order as the  $l_u$  values.

Group V contains some typical carboxylic acids with  $pK < 5 \cdot 0$ . Their  $l_u$  values, with the doubtful exception of oxalate, are all distinctly below 1.0. It thus appears that as the strength of the acid rises a range is reached ( $pK \ 6 \cdot 0 - 5 \cdot 0$ ) in which the catalytic effect of the acid anion tends to fade out.

### The effect of the bicarbonate ion

The  $l_u$  value for the HCO<sub>3</sub><sup>-</sup> ion is not included in Table V, since it cannot be obtained either by the methods so far used in this paper or from data given in previous papers. It is, however, important to measure it, both from the point of view of the physico-chemical mechanism of the  $CO_2 + H_2O \Rightarrow H_2CO_3$  reaction and also of practical applications to biological problems. Since the true first pK of  $H_2CO_3$  is about 3.7 the  $l_u$  value for  $HCO_3^-$  would not, from Table V, be expected to exceed 1.0. The following modification of the usual technique shows that, in point of fact, the  $l_u$  value must be less than 0.5 and may be zero.

A 0.2 M NaHCO<sub>3</sub> in 0.004 M Na<sub>2</sub>CO<sub>3</sub> mixture was prepared by dissolving the requisite weights of the two salts in recently boiled distilled water with minimal contact with air so as to reduce CO<sub>2</sub> exchange. 5 ml. of this solution were placed in the boat and shaken at 0° with the standard CO<sub>2</sub> pressure (in the usual way) for 90 min., i.e. until equilibrium was practically reached. The [CO<sub>3</sub>=] was found to have increased to 0.005 M, and the [HCO<sub>3</sub>-] to have dropped correspondingly. The pH of the solution was calculated to be 8.6. After 1 min. pause for drainage, a second vol. of CO<sub>2</sub> equal to the first was introduced into the boat in the usual way, and an ordinary experiment then carried out to completion. Still more CO<sub>3</sub><sup>=</sup> was formed and there was a further slight fall in pH. The overall velocity constant of CO<sub>2</sub> uptake was then calculated from the time course of the CO<sub>2</sub> uptake during the second shaking though far larger corrections were necessary than usual for (a) the velocity of the back reaction (50-60 %) and (b) the velocity

<sup>1</sup> Prepared from B.D.H. telluric acid. Two samples from other sources were unsatisfactory as they gave heavy precipitates when partially neutralized by alkali.

of the  $\text{CO}_2 + \text{OH}^- \rightarrow \text{HCO}_3^-$  reaction which at pH 8.6 is about 30% of the velocity of the  $\text{CO}_2 + \text{H}_2\text{O} \rightarrow \text{H}_2\text{CO}_3$  reaction. Fortunately, these two large corrections are opposite in sign, and in consequence the final value of the overall velocity constant should be accurate to about  $\pm 10\%$ . The average of four calculated values came out to 0.00208, in very close agreement with the value, 0.0021, given above for  $k_u$ , the true velocity constant of the reaction  $\text{CO}_2 + \text{H}_2\text{O} \rightarrow \text{H}_2\text{CO}_3$ . The joint catalytic effects of the  $\text{HCO}_3^-$  and  $\text{CO}_3^-$  ions in this experiment thus cannot at most have exceeded 10%, which means that  $l_u$  for  $\text{HCO}_3^-$  ion alone  $\geq 0.5$ —a negligible value for practical purposes in work at  $[\text{HCO}_3^-] < 0.05 M$ .

The experiment also gives an upper limit of about 25 for the  $l_u$  value of the  $CO_3^{-}$  ion. Recent experiments at higher  $[CO_3^{-}]$  suggest that the limit is almost certainly much lower.

#### II. BASIC NITROGENOUS BUFFERS AND OTHER SUBSTANCES

#### **Basic nitrogenous buffers**

We have tested three classes of N-containing bases: (a) compounds with the basic N in a closed ring, e.g. glyoxaline, nicotine, (b) compounds with the basic N in a straight chain, e.g. aniline and (c) mixed compounds containing both cyclic basic N and straight chain basic N, e.g. histidine. The results with class (a) being the most straightforward will first be described.

Cyclic N bases. Four members of the glyoxaline series and three other compounds were tested as regards their effect on  $CO_2$  uptake by the methods described in Section I (see Table VI). Fig. 6 shows a typical pair of results for

Buffer	$pK$ at $0^\circ$	[base] : [ion]	$l_u$	lo
2:4(or 5)-Dimethylglyoxaline	8.80	0.33	12.5	
4(or 5)-Methylgloxaline	7.97	2·0 1·0 0·5 0·25	9·5 11 9·5 10	
Glyoxaline	7.40	1·0 2·0	1.5	c. 2
4(or 5)-Hydroxymethylglyoxaline	6.85	4.0	1.6	
Nicotine	8·5 (8·0 at 25°)	0•5	13	
Pilocarpine	7·30 (7·0 at 15°)	1·0 1·0	<u>4</u> ·6	 5·0
Pyridine (in phosphate)	5·7 (5·3 at 20°)	50.0	<0.6	

Table VI. Effect of cyclic nitrogenous bases

an equimolar buffer mixture of glyoxaline and glyoxaline hydrochloride at total concentrations of glyoxaline ranging from 0.036 to 0.38 M. It will be noted that the CO<sub>2</sub> uptake during the initial rapid phase is in both cases practically the same and is equal in amount to that expected from the solubility coefficient of CO<sub>2</sub>, thus showing that in this case there is no rapid reversible combination of the base with CO<sub>2</sub> to form a compound of a carbamino type. (Contrast the results obtained in Fig. 7 with a typical straight chain N compound.) Nor was there any such indication in the case of any of the other compounds listed in Table VI.

When  $v_u$ , the overall velocity constant for CO<sub>2</sub> uptake, is calculated as in Section I and plotted against the concentration of unionized base (i.e. the more regative constituent of the buffer) a straight line is obtained. The slope of the

line shows that the  $l_u$  value for glyoxaline is about 1.6. Extrapolation of the line back to zero base concentration gives a value for  $k_u$ , the true velocity constant of the reaction  $CO_2 + H_2O \rightarrow H_2CO_3$ , in close agreement with that already found in Section I.

The  $l_u$  values given in Table VI have all been calculated in terms of the more negative constituent: the validity of this procedure was tested thoroughly in the case of 4(or 5)-methylglyoxaline by experiments at four different [N base: [N cation]] ratios ranging from 2.0 to 0.25. It is seen that  $l_u$  is constant within experimental error. It may be reasonably assumed that the other compounds in the Table would, on test, yield the same result.

Table VI also shows that there is a correlation between  $l_u$  and pK similar to that already found for the oxy-acid buffers in Table V (though in the present case the available range is smaller): thus, for example, the two compounds with pK > 8.0, nicotine and 2:4(or 5)-dimethylglyoxaline, each have an  $l_u$  of about 12, whereas pyridine with a pK < 6 has a barely detectable  $l_u$  value. The compounds of intermediate pK have, broadly speaking, intermediate  $l_u$  values.



Fig. 6. Observed course of CO<sub>2</sub> uptake by glyoxaline. A, 0.018 M unionized glyoxaline + 0.018 M glyoxaline ion; B, 0.19 M unionized glyoxaline + 0.19 M glyoxaline ion.

Fig. 7. Observed course of CO<sub>2</sub> uptake by hydroxylamine buffer solutions. A, 0.036 M NH<sub>2</sub>OH +0.012 M NH<sub>3</sub> + OH; B, 0.085 M NH<sub>2</sub>OH +0.026 M NH<sub>3</sub>+OH/HPO<sub>4</sub>=+0.011 M; C, 0.033 M H<sub>2</sub>PO<sub>4</sub> for comparison.

Evidence for the catalytic nature of the effect was sought for as before, namely by comparing the  $l_0$  values, obtained from  $CO_2$  output experiments at different buffer concentrations, with the  $l_u$  values. Unfortunately, owing to experimental error and theoretical uncertainty, satisfactory accuracy can only be attained, if the  $l_0$  value is >5 and the pK < 7 (as is the case both for phosphate and cacodylate buffers). Of the compounds listed in Table VI pilocarpine approaches this ideal most nearly: the  $l_0$  value from experiments over a range of 0.05–0.20 *M* total pilocarpine concentrations is 5.0, in satisfactory agreement with the  $l_u$  value of 4.6. A similar, but much less accurate test showed that the  $l_o$  and  $l_u$  values of glyoxaline are of the same order of magnitude. The actual equation for the overall velocity constant of CO<sub>2</sub> output by N base buffers differs slightly from equation (11): it is

$$\frac{d[\text{CO}_2]}{dt} = \frac{v_o K_{\text{B}} f_{\text{HCO}_3} f_{\text{B}}[\text{B}^+]}{[\text{B}]} [\text{HCO}_3^-], \qquad \dots \dots (13)$$

where [B<sup>+</sup>], [B] are the concentrations of N cation and base respectively.

 $K_{\rm B}$  is the ionization constant of the base = [B] [H<sup>+</sup>]/[B<sup>+</sup>].

 $f_{\rm B}$  is the activity coefficient of the N cation.

The calculation of  $l_o$  in the case of N base buffers is thus much more sensitive to activity coefficient errors than in the case of the oxy-acid buffers, since a product of activity coefficients is involved instead of a quotient of activity coefficients as in equation (11).

Straight chain N bases. Faurholt [1925, 2] has measured the equilibrium constant  $K_{diss}$  for the carbamino reaction between CO<sub>2</sub> and the following straight chain amines: NH<sub>3</sub>, CH<sub>3</sub>NH<sub>2</sub>, (CH<sub>3</sub>)<sub>2</sub>NH and CHNH<sub>2</sub>COO<sup>-</sup> (glycinate). As a measure of affinity of the bases for CO<sub>2</sub> we may take

$$1/K_{\rm diss} = \frac{[{\rm CO}_2][{\rm B}]^2}{[{\rm B}^+][{\rm carb}]},$$
 .....(14)

where [carb] is the concentration of carbamino compound formed. The values of  $1/K_{\rm diss}$  at 0° ranged from 10<sup>3</sup> to 10<sup>6</sup> indicating a very large affinity of these bases for CO<sub>2</sub>. This, together with the fact that the pK value at 0° in all four cases is >10, makes it very difficult by the present methods to determine whether these bases have an appreciable  $l_u$  value, though Faurholt's work indirectly speaks against such a possibility.

We have, however, tested several straight chain bases of pK < 9.0, and, in accordance with Faurholt's views, have found a much lower carbamino affinity for CO<sub>2</sub> so that it has been possible to allow for it fairly well in calculation of  $l_u$ . Typical results for hydroxylamine at a [B]: [B<sup>+</sup>] ratio of 3:1 and total concentrations 0.048 M and 0.11 M, are shown in Fig. 7.

The  $CO_2$  uptake in the rapid phase is much greater than that taken up in physical solution (see control curve for  $CO_2$  uptake by phosphate, and also Fig. 2) the excess, due to carbamino formation, is, as is to be expected from equation (14), proportional (at constant [B]: [B+]) to the total hydroxylamine concentration. The data of Fig. 7 indicate that  $1/K_{diss}$  for hydroxylamine is about 4, i.e. about 1/500 the value for ammonia, thus showing the marked effect of substituting an —OH group both on pK and on the affinity for  $CO_2$ .

It will be seen that the rate of  $CO_2$  uptake during the slow phase is less in the 0.048 *M* hydroxylamine solution than in the phosphate control, and in the 0.11 *M* hydroxylamine solution less still. This is partly due to the lower pressure of  $CO_2$  remaining in the gas phase and partly due to the fact that the  $[HCO_3^{-}]$ in the hydroxylamine solution is formed not only from  $CO_2$  coming from the gas phase but also from  $CO_2$  dissociating from the carbamino compound, which is maximal at the end of the rapid phase and decreases as the solution becomes more acid [v. Roughton, 1935, 2]. Approximate corrections can be made for both of these effects: the former is obviously proportional to the extent by which  $p_{CO_2}$  is lower than the control, whilst the latter can be shown to be roughly equal to the [carb]/[ $CO_2$ ] in solution at the end of the rapid phase, provided that the [carb] is not too large. The carbamino corrections in the case of Fig. 7 amount to 7 and 20% respectively and when applied, together with the  $p_{CO_2}$ corrections, show that the true rate of  $HCO_3^{-}$  formation in the hydroxylamine

solution is, within experimental error, equal to the basal rate in absence of catalyst, thus showing that hydroxylamine has no appreciable  $l_u$  value. Similar results were obtained with the other compounds listed in Table VII.

# Table VII. Straight chain N bases

Compound	pK	Carbamino formation	l <sub>u</sub>
Hydrazine	8·4 at 20°	+ + +	Undetectable
Hydroxylamine	$6 \cdot 2$ at $20^{\circ}$	+ +	,,
Aniline	4·5 at 20°	+	,,
Acetamide	c. 0	0	,,
Urea	c. 0	0	,,
+ + + denotes that 1 + + denotes that 1	$/K_{\rm diss}$ is of same of $/K_{\rm diss}$ is of the order	rder as $1/K_{\rm diss}$ for ler of $10^{-2}$ times	r NH <sub>3</sub> . 1/K <sub>diss</sub> for NH <sub>3</sub> .

denotes that  $1/K_{diss}^{diss}$  is of the order of  $10^{-4}$  times  $1/K_{diss}^{diss}$  for NH<sub>2</sub>. denotes that  $1/K_{diss}$  is less than  $10^{-6}$  times  $1/K_{diss}$  for NH<sub>3</sub>.

 $\stackrel{+}{0}$ 

Mixed compounds. From the above results we should expect that a compound which possesses both a basic cyclic N and a basic straight chain N should act both as CO<sub>2</sub> carrier and CO<sub>2</sub> catalyst. We have verified this in preliminary experiments on histidine, and two of its derivatives of physiological interest, dissolved in phosphate buffer. The results are shown in Table VIII.

Table VIII. Effects of histidine and derivatives on  $CO_2$  uptake rate

$pK^*$ at 22°						
Compound	Glyoxa- line N	Side chain N	l <sub>u</sub>	Carbamino formation		
Histidine	6.15	9.3	c. 1·2	+ + +		
Carnosine	6.8	9.5	c. 6	+ + +		
Anserine	7.0	9.5	c. 3	+ + +		

\* Values and assignments from Deutsch & Eggleton [1938] who kindly supplied samples of anserine and carnosine.

Other buffers

We have also tested the catalytic effects of the anions of three non-oxy-acid weak acids, namely HCN (pK at 0° c. 10.0), H<sub>2</sub>S (pK at 0° c. 7.0) and HF.

Cyanide. The  $l_u$  of CN<sup>-</sup> was difficult to measure owing to the volatility and high pK of HCN. The following special technique was used. Scheele's HCN solution (4%, B.D.H.) was mixed with an equal volume of water in a burette over mercury, to avoid HCN loss by evaporation. Titration to pH 6.0 gave the mineral acid content of the solution (0.031 N), and further titration to pH c. 12 (Tropoeolin O as indicator) gave the HCN content (0.79 N). 4 ml. of this solution +0.8 ml. of M/2 KCN were placed in the boat, the latter stoppered at once and the experiment carried out as usual except that the gas pressure was left at 1 atm. instead of being reduced. The standard amount of CO<sub>2</sub> was introduced from a reservoir at 19 cm. Hg positive pressure. The rate of uptake of CO<sub>2</sub> by this cyanide buffer mixture, which contains 0.66 M HCN and 0.06 M $CN^-$ , was found to be about twice that of the basal rate below pH 7.5 in absence of catalyst. Fourfold dilution of the cyanide buffer in a second experiment showed a CO<sub>2</sub> uptake rate about 1.9 times the basal rate. The increase above the basal rate in the two experiments was mainly due to the rate of the  $CO_2 + OH^$ reaction, which in a 1:11 cyanide-HCN buffer mixture (pH c. 9.0) should be of the order of 80  $\%^1$  of the CO<sub>2</sub>+H<sub>2</sub>O  $\rightarrow$  H<sub>2</sub>CO<sub>3</sub> basal rate. If the residue of

<sup>1</sup> Exact allowance is difficult owing to uncertainty as to the pK of HCN at 0° and of the size of the appropriate activity corrections.

the increase is attributed to the catalytic effect of the  $CN^-$  ion, its  $l_u$  value comes out to be roughly 1.0 or less.  $CN^-$  therefore does not appear to belong to the catalytic cyclic N family, which at such a high pK should show an  $l_u > 10.0$ . Nor does it form any carbamino compound, according to the present experiment and the previous ones of Meldrum & Roughton [1933, 2].

Sulphide. The  $pK_1$  of  $H_2S$  at 0° is about 7.0 so that a catalytic effect of HSmight possibly be expected. Unfortunately the high volatility of  $H_2S$  leads to appreciable loss of  $H_2S$  into the gas phase of the boat from the liquid as the latter becomes more acid during the CO<sub>2</sub> uptake. To minimize this, the [HS-] was kept down to 0.012 *M* by dissolving the requisite amount of NaHS (obtained by half-neutralizing Na<sub>2</sub>S (A.R.) with HCl) in a buffer mixture containing 0.05 *M* 2:4(or 5)-dimethylglyoxaline base +0.05 *M* dimethylgloxaline cation (*p*H c. 9.0). Calculation showed that, even so, a correction of 15% must be inserted. The experiment was done in much the same way as the cyanide experiment and the uptake rate was found to be about 15% slower than the control uptake rate by the same buffer mixture without added NaHS. Thus when the correction for H<sub>2</sub>S volatility is applied the two rates agree within experimental error, showing that the catalytic effect of 0.012 *M* HS<sup>-</sup> is inappreciable. The  $l_u$  value of the HS<sup>-</sup> cannot therefore exceed 10.0 and may be zero.

The amount of  $CO_2$  taken up during the rapid phase was found to be the same whether HS<sup>-</sup> was present or not, thus showing no evidence of any appreciable rapid reversible combination between  $CO_2$  and HS<sup>-</sup> analogous to the carbamino reaction.

We have not yet tested any more complicated sulphhydryl acids, such as thiolacetic acid, owing to the higher pK of their -SH group (c. 10.0) and their instability.

Fluoride. The pK of HF is 4.45 at 25°, and the  $l_u$  value, calculated from an additive experiment with 0.6 M NaF + phosphate buffer, is 1.1. This value agrees closely with that of the oxy-acids of pK c. 4.0, and is distinctly higher than the range found for neutral salts of strong acids in Table III.

#### DISCUSSION

# The mechanism of the catalysis

In the CO<sub>2</sub> output experiments the additional rate due to the catalysis is proportional to the product of the  $[H_2CO_3]$  and the more negative constituent of the buffer, e.g.  $[HPO_4^-]$  or  $[Cac^-]$ , and is independent of the  $[HCO_3^-]$  except in so far as the latter conditions the  $[H_2CO_3]$ . This means that  $H_2CO_3$ , not  $HCO_3^-$ , is the substrate acted on by the catalyst, i.e. the reaction catalysed must be  $CO_2 + H_2O \rightleftharpoons H_2CO_3$  and not  $CO_2 + OH^- \rightleftharpoons HCO_3^-$ . Preliminary confirmation of this has been obtained by CO<sub>2</sub> uptake experiments between pH 9.0and 10.0 with two different  $[H_2BO_3^-]$ :  $[H_3BO_3]$  ratios and varying total borate concentration. It was found that the fraction of the overall uptake rate due to the  $CO_2 + OH^- \rightarrow HCO_3^-$  rate was practically unaffected by changes in  $[H_2BO_3^-]$  although the fraction due to the  $CO_2 + H_2O \rightarrow H_2CO_3$  was greatly affected by  $[H_2BO_3^-]$ , which showed its usual  $l_u$  value > 100.

Whilst not excluding the possibility of a chain mechanism we think it more likely that the catalysis can be explained by the intermediate compound formation between  $CO_2$  and  $H_2CO_3$  on the one hand, and the more negative constituent of the buffer on the other.

In the case of the oxy-acid buffers the compounds might be of a "carbonato"

type, analogous to the reversible compounds between CO<sub>2</sub> and organic hydroxides studied by Faurholt:

$$\bigcup_{0}^{0} C + 0X^{-} \rightleftharpoons \bigcup_{0}^{-0} C - 0X (+H_{2}0) \rightleftharpoons \bigcup_{H_{0}}^{-0} C - 0X \rightleftharpoons \bigcup_{H_{0}}^{0} C + 0X^{-}$$

We have to suppose that  $H_2O$  can combine with the  $CO_2$  bound to  $OX^-$  to form the corresponding  $H_2CO_3^-$ . OX intermediate compound which in turn decomposes to  $H_2CO_3 + OX^-$ . We then have the catalytic scheme formulated in the equation. This scheme explains why (a) the reaction  $CO_2 + OH^- \rightleftharpoons HCO_3^-$  is not catalysed; probably the electrostatic repulsion between  $HCO_3^-$  and  $OX^$ hinders the formation of the necessary  $HCO_3^-$ .  $OX^-$  intermediate compound, and (b) the catalytic activity of the oxy-acid ion tends to disappear when the pK of the acid is < 6.0. Table VII shows how the affinity of the straight chain N bases for  $CO_2$  declines *pari passu* with the decrease in pK, i.e. decreases in affinity for H<sup>+</sup>. Similarly we might suppose that the combination of  $CO_2$  with the  $-O^-$  in weak acid anions persists up to a certain point as the strength of the acid increases but finally tends to become negligible. On this view the lack of catalysis is due to failure to form the necessary intermediate compound.

In the case of the cyclic nitrogenous base buffers the intermediate compound with  $CO_2$  might be of a carbamino type:

$$\begin{array}{c} 0 \\ 0 \\ 0 \end{array} C + HNY \rightleftharpoons \begin{array}{c} 0 \\ HO \end{array} C - NY(+HO_2) \rightleftharpoons \begin{array}{c} HO \\ HO \end{array} CNY \rightleftharpoons \begin{array}{c} 0 \\ HO \end{array} C + HNY$$

With straight chain N compounds no reaction occurs between water and the carbamino compound of the N bases, but in the cyclic N compounds the postulated carbamino compound might, for some reason connected with the chemistry of the ring, tend to hydrate, forming the unstable intermediate which breaks down to  $H_2CO_3$  and the free N base. The catalytic scheme is thus complete. Although at present we have no direct evidence in favour of these hypotheses, we have found them very useful working guides, and therefore have felt justified in mentioning them.

# Additive effects

We have already alluded to the fact that the catalytic effects of  $HPO_4^{=}$  and Cac<sup>-</sup>, when jointly present, are additive and the same is also true of the following pairs—maleate + phosphate, chromate + phosphate and probably also borate + phosphate. Carbonic anhydrase and phosphate, on the other hand, tend to multiply the effects of each other. It was therefore of special interest to see whether our most active inorganic catalyst, selenite, is additive or multiplicative. It is difficult, owing to ionic activity uncertainties, to make such experiments fully satisfactory from a theoretical standpoint, but some preliminary results with selenite + phosphate, and selenite + chromate mixtures do suggest that in both cases selenite multiplies the effect of the weaker catalytic ion. Selenite may thus turn out to be a weak inorganic analogue of carbonic anhydrase, in which case a further detailed study of its catalytic mechanism would be well worth while.

As to the mechanism of the multiplicative effect it is possible that selenite and/or carbonic anhydrase may catalyse the formation of the intermediate compound between  $CO_2$  and the more weakly catalytic anions, which we have supposed to be a preliminary to the catalysis and which may not itself proceed very rapidly. Various other explanations are however possible.

#### The effect of pH on carbonic anhydrase

As already hinted in the introduction, the original aim of the present research was to determine the effect of pH upon the activity of carbonic anhydrase.

Two difficulties were already obvious at the start, namely (a) in the pH range above 8.0, where CO<sub>2</sub> reacts in two ways (see equations (1) and (2)) there is the problem of sorting out the respective effects of the enzyme on these two different processes, and (b) the Michaelis constant of the enzyme is very high, and hence it is impracticable to work at substrate concentrations sufficient to saturate the enzyme—a prerequisite in pH-activity work. The results described in this paper, though at first sight adding yet a further complication, may on further investigation prove to simplify the study of the effect of pH on carbonic anhydrase. Thus in regard to (a) preliminary experiments suggest that carbonic anhydrase, like phosphate and other buffers, only catalyses the  $CO_2 + H_2O \rightleftharpoons H_2CO_3$ , and does not affect the  $CO_2 + OH \rightarrow HCO_3$ . In regard to (b), it is already known from unpublished observations of the writers and others, that the activity of carbonic anhydrase increases markedly over the pH range 6.0–8.0. If it turns out that the activity of the enzyme in promoting CO<sub>2</sub> uptake at constant CO<sub>2</sub> pressure when plotted against pH gives a curve resembling the ionization curve of a weak acid, a result exactly analogous to the results with phosphate etc. would be obtained, and it might by further analogy be fair to infer therefrom the actual pK of the active group of the enzyme, even though the enzyme had not been saturated with substrate in the experiments. All this is perhaps too much to hope for, but at all events we have some valuable new pointers as to the investigation of the mode of action of carbonic anhydrase.

The catalytic effects herein described occur in many physiological buffers containing  $CO_2$ , and should therefore be borne in mind as possible factors in diverse physiological and biochemical processes and experiments. In particular, they may be of interest in bone equilibria, wherein carbonato-phosphate reactions of a type somewhat similar to that postulated in our catalytic schemes may take place, especially in regions of high local phosphate concentration.

#### SUMMARY

1. Detailed improvements have been made in the manometric technique for measuring the rate of  $CO_2$  uptake by solutions.

2. When  $CO_2$  is taken up by buffer solutions the buffer substance itself has some direct effect on the reaction besides "instantaneous" removal of H<sup>+</sup> concerned in the ionization of H<sub>2</sub>CO<sub>3</sub> to H<sup>+</sup>+HCO<sub>3</sub><sup>-</sup>. Experimental proof is offered that the effect cannot be due to impurities in the solutions or to "neutral salt" action, but is proportional to the concentration of the more negative constituent of the buffer. Thus in the case of phosphate buffer, the overall velocity constant,  $v_u$ , is given by

$$v_u = 0.0021 [CO_2] \{1 + l_u [HPO_4^-]\},\$$

 $l_u$ , the catalytic coefficient, being 8, and for cacodylate 9. The figure 0.0021 represents the true velocity constant of the reaction  $CO_2 + H_2O \rightarrow H_2CO_3$  and is about 25% lower than the erroneous value previously accepted.

3. All other oxy-acids, so far tested, which buffer in the pH range 6–9 show similar effects, whereas salts of stronger acids have much smaller effects ( $l_u < 1.5$ ). The effects with sulphite, selenite and tellurate are much larger ( $l_u$  of order of 1000).

4. Straight chain N bases, e.g.  $\rm NH_2OH$ , show no appreciable catalytic activity, though they combine readily with  $\rm CO_2$  to form carbamino compounds. Cyclic N bases, e.g. glyoxaline, on the other hand do not form carbamino compounds but do act catalytically. The effect is proportional again to the concentration of the more negative constituent—in this case the unionized N base. Values for  $l_u$  of the order of 10 were found for several cyclic bases of pK > 7.0. With weaker bases the effect tends to disappear. Mixed compounds, e.g. histidine, show both catalytic action and carbamino formation.

5. Similar effects are observed on the rate of *output* of  $CO_2$  from bicarbonate solution suddenly mixed with buffer,  $l_0$  (output) for the oxy-acid buffers phosphate, cacodylate, chromate, selenite, and for the N base buffers pilocarpine and glyoxaline being equal to  $l_u$ . The effect must therefore be a catalysis.

6. It is shown that the buffers only catalyse the  $CO_2 + H_2O \rightleftharpoons H_2CO_3$  reaction and not the  $CO_2 + OH^- \rightleftharpoons HCO_3^-$  reaction. A mechanism is suggested in which  $CO_2$  (or  $H_2CO_3$ ) combines reversibly with the more negative constituent of the buffer.

7. The effects of phosphate and cacodylate are additive, but of the enzyme and phosphate are probably multiplicative.

8. Some biochemical implications of these results are discussed. They must be allowed for in work involving velocities of  $CO_2$  reactions. Selenite is suggested as a possible inorganic model of carbonic anhydrase.

#### REFERENCES

Booth & Roughton (1938). J. Physiol. 92, 36 P.
Brinkman, Margaria & Roughton (1933). Philos. Trans. 232, 65.
Deutsch & Eggleton (1938). Biochem. J. 32, 209.
Faurholt (1925, 1). J. Chim. phys. 21, 400.
— (1925, 2). J. Chim. phys. 22, 1.
— (1927). Z. phys. Chem. 126, 72.

Kirby & Neuberger (1938). Biochem. J. 32, 1146.

Meldrum & Roughton (1933, 1). J. Physiol. 80, 113.

----- (1933, 2). J. Physiol. 80, 143. Roughton (1930). Proc. roy. Soc. A, 126, 470.

----- (1935, 2). J. Physiol. 85, 17 P.