Early Dissemination of CTX-M-Derived Enzymes in South America

In a recent publication by Bonnet et al. regarding a new CTX-M-derived enzyme, a short history of this group of enzymes is provided, stating that "initially found in Europe, microorganisms producing these enzymes have now been observed over a wide geographic area..." (3). We want to point out that although the description of the first members of enzymes was achieved in Europe, they had been widely distributed in South America at least since 1989. Three different groups of investigators reported, in a national meeting, the explosive dissemination of extended-spectrum cephalosporinresistant salmonella, initially in neonatology units in La Plata and Buenos Aires city hospitals (A. Picandet, S. Giugno, M. I. Caffer, and G. Schembri, Abstr. II Congreso Internacional de SADEBAC, Antimicrobianos '90, abstr. A-16, 1990; H. Lopardo, M. I. Caffer, N. Fernandez, et al., Abstr. II Congreso Internacional de SADEBAC, Antimicrobianos '90, abstr. A-15, 1990; E. Maiorini, A. Procopio, S. Furmanski, et al., Abstr. II Congreso Internacional de SADEBAC, Antimicrobianos '90, abstr. A-17, 1990). An outbreak in a single hospital was presented at the 17th Congress of Chemotherapy, Berlin, 1991 (H. Lopardo, N. Fernandez, M. Fernandez Cobo, et al., abstr. 2088, 1991).

The presence of a new enzyme in the implicated microorganisms was pointed out in a communication to the 1992 ASM General Meeting (A. Rossi, M. Woloj, G. Gutkind, et al., Abstr. 92nd Gen. Meet., abstr. A-135, 1992), and a brief historic perspective of their dissemination was reported in a paper that finally appeared in 1995 (7). In that paper, information relevant to the initial isolates, dissemination of this resistance marker within different salmonella serovars, and some biochemical characteristics of the enzymes present was displayed. In brief, they made their appearance in a really explosive way, initially in a single hospital in La Plata, from there to neonatology units in pediatric hospitals in Buenos Aires, and from them it was disseminated to Paraguay, Misiones (in northern Argentina), and Uruguay. However, formal sequencing of the structural gene of the enzyme responsible for this outbreak (and naming of the enzyme) was achieved by a more efficient collaboration of Bauernfeind and his group with Casellas and his group (1-2).

Since then, its presence has been suggested or demonstrated in different microorganisms, such as Escherichia coli (M. Radice, A. Rossi, M. Venuta, H. Lopardo, and G. Gutkind, XVI Congreso Chileno de Microbiologia, p. 31, 1994; (5a), Shigella sonnei (5), and Proteus mirabilis (M. Quinteros, M. Mollerach, M. Radice, et al., Abstr. 39th Intersci. Conf. Antimicrob. Agents Chemother., abstr. 893, 1999; M. Radice, M. Quinteros, M. Matteo, et al., Abstr. 9th Int. Congr. Infect. Dis., abstr. 16028, 2000) (as typical non-AmpC producers), Morganella morganii (reference 44 and P. Power, M. Radice, C. Barberis, et al., Abstr. 98th Gen. Meet. Am. Soc. Microbiol. 1998, abstr. V-125, 1998), Citrobacter freundii, Serratia marcescens, and Enterobacter aerogenes, among other "AmpC-inducible" enterobacteria (M. Quinteros, M. Radice, P. Power, et al., Abstr. II Int. Congr. β-Lactamases, abstr. A-27, 1999), Vibrio cholerae (reference 6 and M. Galas, A. Petroni, R. Melano, et al., Abstr. 38th Intersci. Conf. Antimicrob. Agents Chemother., abstr. C-174, 1998), and *Aeromonas hydrophyla* (M. Quinteros, M. Radice, P. Power, et al., Abstr. 9th Int. Congr. Infect. Dis., abstr. 15884, 2000).

A discussion on the ability of different susceptibility testing systems has been presented also (M. Quinteros, M. Matteo, P. Power, et al., Abstr. VIII Congreso Argentino de Microbiologia, abstr. A-44, 1998; M. Quinteros, M. Mollerach, M. Radice, et al., Abstr. 39th Intersci. Conf. Antimicrob. Agents Chemother., abstr. 893, 1999; S. Kauffman, M. Quinteros, M. Radice, et al., Abstr. 9th Int. Congr. Infect. Dis., abstr. 15883, 16028, and 16031, 2000). A more straightforward recommendation was presented almost a decade ago by the Subcomisión de Antimicrobianos, Sociedad Argentina de Bacteriologia Clinica, Asociación Argentina de Microbiología, for changing on a national scale the interpretation of NCCLS breakpoints (while keeping all the other NCCLS method recommendations), stating that any enterobacteria (except for those in which AmpC is typically inducible) with inhibition zones lower than 26 mm around 30-µg cefotaxime disks or with a MIC higher than 1 mg/liter should be considered as potentially resistant to all extended-spectrum cephalosporins. It would be interesting to learn if this new CTX-M-derived enzymes explosion would have been so easily detected if NCCLS recommendations had not moved the breakpoint to 27 mm.

Being able to detect it does not prevent its dissemination or allows its control: CTX-M-2 is present in almost 75% of the extended-spectrum β -lactamase (ESBL)-producing enterobacteria submitted to our lab, coincident to an epidemiological study in Buenos Aires hospitals through a network of microbiological laboratories (Quinteros et al., unpublished data) being PER-2 (another nonclassical enzyme), the second most prevalent ESBL. Why are these enzymes and not the classical TEM- or SHV-derived enzymes (or any other classical family) are the most prevalent ESBLs in our region, as A. Medeiros has been asking Argentinean microbiologists in each of his visits? There is no formal answer to date.

REFERENCES

- Bauernfeind, A., J. M. Casellas, M. Goldberg, M. Holley, R. Jungwirth, P. Mangold, T. Röhnisch, S. Schweigart, and R. Wilhelm. 1992. A new plasmidic cefotaximase from patients infected with *Salmonella typhimurium*. Infection 20:168–173.
- Bauernfeind, A., I. Stemplinger, R. Jungwirth, S. Ernst, and J. M. Casellas. 1996. Sequences of β-lactamase genes encoding CTX-M-1 (MEN-1) and CTX-M-2 and relationship of their amino acid sequences with those of other β-lactamases. Antimicrob. Agents Chemother. 40:509–513.
- Bonnet, R., C. Dutour, J. L. M. Sampaio, C. Chanal, D. Sirot, R. Labia, C. De Champs, and J. Sirot. 2001. Novel cefotaximase (CTX-M-16) with increased catalytic efficiency due to substitution Asp-240→Gly. Antimicrob. Agents Chemother. 45:2269–2275.
- Power, P., M. Radice, C. Barberis, C. de Mier, M. Mollerach, M. Maltagliatti, C. Vay, A. Famiglietti, and G. Gutkind. 1999. β-lactamases in *Morganella morganii*: characterization of extended spectrum β-lactamases. Eur. J. Clin. Microbiol. Infect. Dis. 18:43–47.
- Radice, M., C. Gonzalez, P. Power, M. Vidal, and G. Gutkind. 2001. Third generation cephalosporins resistance in *Shigella sonnei*. Emerg. Infect. Dis. 7:442–443.

- 5a.Radice, M. 1999. Ph.D. dissertation. Universidad de Buenos Aires, Buenos Aires, Argentina.
- Rossi, A., M. Galas, A. Corso, M. Radice, M. Rivas, M. Caffer, N. Binztein, and G. Gutkind. 1993. Unussual multiresistant *Vibrio cholerae* O1 var. El Tor in Argentina. Lancet 342:1172–1173.
- Rossi, A., H. Lopardo, M. Woloj, A. M. Picandet, M. Mariño, M. Galas, M. Radice, and G. Gutkind. 1995. Non-typhoid Salmonella spp. resistant to cefotaxime. J. Antimicrob. Chemother. 36:697–702.

Marcela Radice Pablo Power Jose Di Conza Gabriel Gutkind* Cátedra de Microbiología Facultad de Farmacia y Bioquímica Universidad de Buenos Aires Junín 954 1113-Buenos Aires, Argentina

*Phone: 11 4964 8285 Fax: 11 4508 3645 E-mail: ggutkind@ffvb.uba.ar

Authors' Reply

We recently published a paper in this journal on the structure and properties of a novel extended-spectrum β -lactamase, CTX-M-16 (7). The enzyme differs from CTX-M-9 by a single amino acid substitution, Asp-240 \rightarrow Gly. The most interesting feature was the increased hydrolytic activity of the novel enzyme for ceftazidime. This finding shows the evolutionary potential of the spectrum of CTX-M activity.

In our article (7), the aim was not to trace the history of the dissemination of CTX-M enzymes; however, in the introduction we did mention that "initially found in Europe, CTX-M-producing strains have now been observed over a wide geographic area." G. Gutkind and coworkers have contested the phrase "initially found in Europe" and state that CTX-M β -lactamases "had been widely distributed in South America at least since 1989" and possibly before appearing in Europe.

The CTX-M-1 (2) and MEN-1 (1) β -lactamases, which were the first fully characterized CTX-M reported, were initially detected from strains isolated in Europe. The first came from Germany and the second was isolated in France from a patient who was an Italian national.

At an International Congress on Infectious Diseases meeting held in Montreal in 1990, one of us (R. Labia) met A. Bauernfeind, who was presenting a poster about an enzyme which was then named CTX-M-1 (A. Bauernfeind, S. Schweighart, and H. Grimm, Int. Congr. Infect. Dis., abstr. 570, p. 17, 1990), and they discussed the similarity of their enzymes, CTX-M-1 and MEN-1, which they were studing. They exchanged strains and observed that the enzymes were identical. One sequence was published in 1992 (1) and the other was published in 1996 (4), but both strains had been isolated at the beginning of 1989 (5). In 1992, Bauernfeind et al. published a preliminary paper on a CTX-M-2-producing Salmonella enterica serovar Typhimurium strain (pI 7.9) isolated "in the beginning of August 1990" in Argentina without further details of its origin (3). In 1995, Rossi and coworkers described cefotaxime-resistant Salmonella spp. strains isolated in Argentina in 1991 (17). They described unidentified β -lactamases of alkaline pI (7.4, 8.1) which hydrolyze cefotaxime. These enzymes could be CTX-M extended-spectrum β -lactamases (ESBLs), and it would be interesting to characterize their β-lactamaseencoding genes.

In Japan, Matsumoto et al. had already described an FEC-1

enzyme in 1988, which induced higher resistance levels to cefotaxime than to ceftazidime (15). The enzyme had a pI and kinetic constants similar to those of CTX-M ESBLs. In our opinion, this work may have been the first report of a CTX-M-type enzyme. Currently, Japan is a large reservoir of CTX-M enzymes (13, 14, 19), as are South America 6; M. Galas, F. Pasteran, R. Melano, A. Petroni, G. Lopez, A. Corso, A. Rossi, and WHONET collaborative group, Abstr. 38th Intersci. Conf. Antimicrob. Agents Chemother., abstr. E-109, p. 201, 1998) and Eastern Europe (8–12, 16, 18).

A considerable delay is usually observed between the isolation of a strain and the publication of its characteristics. Thus, we agree that CTX-M β -lactamases may well have been present in Argentina in 1989, or even before, and were quite likely present in other countries as well.

REFERENCES

- Barthélémy, M., J. Péduzzi, H. Bernard, C. Tancrède, and R. Labia. 1992. Close amino acid sequence relationship between the new plasmid-mediated extended-spectrum β-lactamase MEN-1 and chromosomally encoded enzymes of *Klebsiella oxytoca*. Biochim. Biophys. Acta 1122:15–22.
- Bauernfeind, A., J. M. Casellas, M. Goldberg, R. Holley, R. Jungwirth, P. Mangold, T. Röhnisch, S. Schweighart, and R. Wilhelm. 1992. A new plasmidic cefotaximase from patients infected with *Salmonella typhimurium*. Infection 20:158–163.
- Bauernfeind, A., H. Grimm, and S. Schweighart. 1990. A new plasmidic cefotaximase in a clinical isolate of *Escherichia coli*. Infection 18:294–298.
- Bauernfeind, A., I. Stemplinger, R. Jungwirth, S. Ernst, and J. M. Casellas. 1996. Sequences of β-lactamase genes encoding CTX-M-1 (MEN-1) and CTX-M-2 and relationship of their amino acid sequences with those of other β-lactamases. Antimicrob. Agents Chemother. 40:509–513.
- Bernard, H., C. Tancrède, V. Livrelli, A. Morand, M. Barthélémy, and R. Labia. 1992. A novel plasmid-mediated extended-spectrum β-lactamase not derived from TEM- or SHV-type enzymes. J. Antimicrob. Chemother 29: 590–592.
- Bonnet, R., J. L. M. Sampaio, R. Labia, C. De Champs, D. Sirot, C. Chanal, and J. Sirot. 2000. A novel CTX-M β-lactamase (CTX-M-8) in cefotaximeresistant *Enterobacteriaceae* isolated in Brazil. Antimicrob. Agents Chemother. 44:1936–1942.
- Bonnet, R., C. Dutour, J. L. M. Sampaio, C. Chanal, D. Sirot, R. Labia, C. De Champs, and J. Sirot. 2001. Novel cefotaximase (CTX-M-16) with increased catalytic efficiency due to substitution Asp-240→Gly. Antimicrob. Agents Chemother. 45:2269–2275.
- Bradford, P. A., Y. Yang, D. Sahm, I. Grope, D. Gardovska, and G. Storch. 1998. CTX-M-5, a novel cefotaxime-hydrolyzing β-lactamase from an outbreak of *Salmonella typhimurium* in Latvia. Antimicrob. Agents Chemother. 42:1980–1984.
- Gazouli, M., N. J. Legakis, and L. S. Tzouvelekis. 1998. Effect of substitution of Asn for Arg-276 in the cefotaxime-hydrolyzing class A β-lactamase CTX-M-4. FEMS Microbiol. Lett. 169:289–293.
- Gazouli, M., E. Tzelepi, A. Markogiannakis, N. J. Legakis, and L. S. Tzouvelekis. 1998. Two novel plasmid-mediated cefotaxime-hydrolyzing β-lactamases (CTX-M-5 and CTX-M-6) from *Salmonella typhimurium*. FEMS Microbiol. Lett. 165:289–293.
- Gazouli, M., E. Tzelepi, S. V. Sidorenko, and L. S. Tzouvelekis. 1998. Sequence of the gene encoding a plasmid-mediated cefotaxime-hydrolyzing class A β-lactamase (CTX-M-4): involvement of serine 237 in cephalosporin hydrolysis. Antimicrob. Agents Chemother. 42:1259–1262.
- Gniadkowski, M., I. Schneider, A. Palucha, R. Jungwirth, B. Mikiewicz, and A. Bauernfeind. 1998. Cefotaxime-resistant *Enterobacteriaceae* isolates from a hospital in Warsaw, Poland: identification of a new CTX-M-3 cefotaximehydrolyzingβ-lactamase that is closely related to the CTX-M-1/MEN-1 enzyme. Antimicrob. Agents Chemother. 42:827–832.
- Ishii, Y., A. Ohno, H. Taguchi, S. Imajo, M. Ishiguro, and H. Matsuzawa. 1995. Cloning and sequence of the gene encoding a cefotaxime-hydrolyzing class A β-lactamase isolated from *Escherichia coli*. Antimicrob. Agents Chemother. 39:2269–2275.
- Ma, L., Y. Ishii, M. Ishiguro, H. Matsuzawa, and K. Yamaguchi. 1998. Cloning and sequencing of the gene encoding Toho-2, a class A β-lactamase preferentially inhibited by tazobactam. Antimicrob. Agents Chemother. 42: 1181–1186.

604 LETTERS TO THE EDITOR

- Matsumoto, Y., F. Ikeda, T. Kamimura, Y. Yokota, and Y. Mine. 1988. Novel plasmid-mediated β-lactamase from *Escherichia coli* that inactivates oxyimino-cephalosporins. Antimicrob. Agents Chemother. 32:1243–1246.
- Palucha, A., B. Mikiewicz, W. Hryniewicz, and M. Gniadkowski. 1999. Concurrent outbreaks of the family *Enterobacteriaceae* in a Warsaw hospital. J. Antimicrob. Chemother. 44:489–499.
- Rossi, A., H. Iopardo, M. Woloj, A. M. Picandet, M. Marino, M. Galas, M. Radice, and G. Gutkind. 1995. Non-typhoid *Salmonella* spp. resistant to cefotaxime. J. Antimicrob. Chemother. 36:697–702.
- Tassios, P. T., M. Gazouli, E. Tzelepi, H. Milch, N. Kozlova, S. Sidorenko, N. J. Legakis, and L. S. Tzouvelekis. 1999. Spread of a *Salmonella typhi-murium* clone resistant to expanded-spectrum cephalosporins in three European countries. J. Clin. Microbiol. 37:3774–3777.
- Yagi, T., H. Kurokawa, K. Senda, S. Ichiyama, H. Ito, S. Ohsuka, K. Shibayama, K. Shimokata, N. Kato, M. Ohta, and Y. Arakawa. 1997. Nosocomial spread of cephem-resistant *Escherichia coli* strains carrying multiple Toho-1-like β-lactamases genes. Antimicrob. Agents Chemother. 41: 2606–2611.

Richard Bonnet* Danielle Sirot Jacques Sirot Laboratoire de Bacteriologie Faculte de Medecine 28 Place Henri-Dunant 63001 Clermont-Ferrand Cedex France

Roger Labia UMR 175, CNRS-MNHN 29000 Quimper France

Fax: 33 4 73 27 74 94 E-mail: richard.bonnet@u-clermont1.fr