

Report

Children with Idiopathic Hemihypertrophy and Beckwith-Wiedemann Syndrome Have Different Constitutional Epigenotypes Associated with Wilms Tumor

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Idiopathic hemihypertrophy (IH) is a congenital overgrowth syndrome associated with an increased risk of embryonal cancers in childhood. A related developmental disorder is Beckwith-Wiedemann syndrome (BWS), which increases risk for embryonal cancers, including Wilms tumor. Constitutional epigenetic alterations associated with BWS have been well characterized and include epigenetic alterations of imprinted genes on 11p15. The frequency of hypermethylation of *H19* in children with IH and Wilms tumor, 20% (3/15), was significantly lower than the frequency in children with BWS and Wilms tumor, 79% (11/14; $P = .0028$). These results indicate that children with IH and Wilms tumor have different constitutional epigenotypes from those of children with BWS and Wilms tumor.

Idiopathic hemihypertrophy (IH [MIM 235000]) is a congenital overgrowth syndrome associated with an increased risk of embryonal cancers of childhood, including Wilms tumor (Hoyme et al. 1998). A related congenital overgrowth and cancer-predisposition syndrome is Beckwith-Wiedemann syndrome (BWS [MIM #130650]). BWS is also associated with hemihyperplasia and embryonal cancers of childhood, including Wilms tumor (DeBaun and Tucker 1998); however, there are many other manifestations, such as macroglossia, abdominal-wall defects (omphalocele, diastasis recti, or umbilical hernia), and neonatal hypoglycemia. In children with BWS, Wilms tumor is primarily associated with constitutive hypermethylation of the *H19* promoter and loss of imprinting of *IGF2*; in a case-cohort study of children with BWS, constitutional hypermethylation of *H19* was associated with a fourfold greater risk of embryonal cancers than that in children with other meth-

ylation abnormalities (DeBaun et al. 2002). In contrast to BWS, only uniparental disomy (UPD) of 11p15 has been associated with cancer in children with IH. Despite the similarities between these two syndromes, limited molecular data exist as to whether BWS and IH represent phenotype variations of the same genotype or, in fact, are two separate syndromes with different genotypes. To test the hypothesis that children with BWS and IH have the same genotype with variable expression, we performed genotype-phenotype studies in children identified as having IH or BWS and Wilms tumor from the National Wilms Tumor Study (NWTs).

Patients in the study were registered in NWTs 3 and 4; the design of NWTs has been reported elsewhere (Breslow et al. 1988). At the time of enrollment, registering physicians were requested to indicate the presence or absence of specific conditions, including BWS and IH. No systematically collected details about the clinical features of IH and BWS in the patients were available. A pediatric oncologist with expertise in evaluating children with overgrowth syndromes reviewed all records. All classifications of either IH or BWS were done prior to and independent of genetic analyses.

Patients with either BWS or IH and fresh frozen tissues submitted to the NWTs Wilms tumor bank were iden-

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tified and represent the sampling frame. Frozen aliquots of all these samples were then selected for analysis. Control samples of Wilms tumors from nonsyndromic patients were similarly obtained.

Genomic DNA was prepared from snap-frozen normal kidney tissue, tumor tissue, and peripheral blood lymphocytes by standard proteinase K digestion and phenol extraction (Cui et al. 1998). *H19* and *LIT1* methylation and UPD analysis by microsatellite marker typing were done as described elsewhere (DeBaun et al. 2002). Loss of heterozygosity (LOH) analysis was performed similar to UPD analysis by use of FAM-labeled primer pairs, detection on a model 377 automated fluorescent DNA sequencer (Applied Biosystems), and analysis with Genescan and Genotyper software (Applied Biosystems). Examination of the chromatograms from samples with UPD revealed a small peak (<20% of the expected size), corresponding to the lost allele and thereby indicating mosaicism for UPD. Patients in whose DNA an allele was reduced in the tumor tissue compared with the normal tissue were considered to have LOH. The threshold for LOH by visual chromatogram inspection was 50%.

Abnormal methylation of *H19* in kidney was defined as a methylation index >0.74 (mean + 2 SD of six kidney samples from control Wilms tumors with intralobar nephrogenic rests, the histological subtype of Wilms tumor not associated with epigenetic alterations of *H19*

Table 1

Clinical Features of Patients with BWS

Patient	Birth Weight ^{a,b} (kg)	Congenital Anomalies Noted in Chart ^b
BWS1	4.5	BWS
BWS2	4.7	BWS, somatic overgrowth
BWS3	3.9	BWS
BWS4	4.3	BWS, macroglossia, hemihypertrophy, adrenal adenoma
BWS5	4.7	BWS
BWS6	3.2	BWS, hemihypertrophy
BWS7	2.4	BWS, omphalocele
BWS8	3.5	BWS, macroglossia, hemihypertrophy, umbilical hernia, cryptorchidism
BWS9	NA	BWS, hemihypertrophy
BWS10	NA	Accessory renal artery
BWS11	NA	None
BWS12	NA	Accessory renal artery
BWS13	NA	Accessory renal artery, 4th and 5th anterior rib anomalies
BWS14	NA	Hemihypertrophy
BWS15	NA	NA
BWS16	NA	NA

^a Gestational ages were not available; however, at 40 wk gestational age, the 90th percentile for birth weight is ~3.9 kg, and the 50th percentile is ~3.4 kg (Oken et al. 2003).

^b NA = not available.

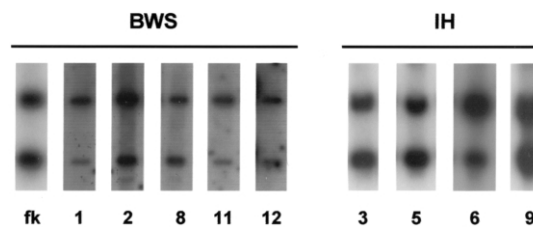


Figure 1 Analysis of *H19* methylation in patients with BWS and IH. A 1.0-kb fragment of the *H19* gene was hybridized to genomic DNA digested with *Pst*I and the methylcytosine-sensitive restriction enzyme *Sma*I. The upper band represents the 1.8-kb methylated fragment, and the lower band represents the 1.0-kb unmethylated digested fragment. Patient numbers corresponding to those in the tables are shown below the lanes. All DNA specimens were from the normal kidney. fk = fetal kidney.

and *IGF2* [Ravenel et al. 2001]). Abnormal methylation of *H19* in lymphocytes was defined as a methylation index >0.63 (mean + 2 SD of 15 normal individuals). Abnormal methylation of *LIT1* in kidney and lymphocytes was defined as a methylation index <0.39 (mean – 2 SD of six kidney samples from control Wilms tumors and of 15 normal individuals, respectively). Fisher's exact test was used to determine whether hypermethylation of *H19* was associated with IH and BWS. A *P* value <.05 was considered statistically significant.

We received samples from 24 patients with IH and Wilms tumor. On the basis of chart review, we excluded the diagnosis of IH for a total of 7 of the 24 patients. Six excluded patients had been initially classified as having IH but also had one or two of the cardinal features of BWS: macroglossia and macrosomia (*n* = 1), macroglossia and umbilical hernia (*n* = 1), umbilical hernia alone (*n* = 1), or macrosomia alone (*n* = 3). These patients were excluded because of the ambiguity of the clinical diagnosis and because they had not previously received a diagnosis of BWS. One patient was excluded for suspected neurofibromatosis because of café au lait pigmentation. The remaining 17 patients received the diagnosis of IH.

We received samples from 16 patients with BWS and Wilms tumor. When available, records of congenital anomalies and birth weight were assessed to confirm the diagnosis (table 1). Gestational ages were not available to determine whether the patient had macrosomia, defined as birth weight ≥90th percentile.

We tested normal kidney tissue for methylation status at *H19* and *LIT1* and typed the normal and tumor tissues for microsatellite markers at 11p15 to identify UPD or LOH (figs. 1 and 2 and tables 2 and 3). If no normal tissue was available, we analyzed the methylation and microsatellites of the tumor tissue. We accepted the tumor data as representative of the state of the normal

tissue if the tumor had the half methylation pattern that occurs in normal tissues and showed retention of heterozygosity by microsatellite analysis (patients IH10, IH13, IH14, and IH15). The assumption is that if tumor tissue from a particular sample shows normal methylation of *H19* or heterozygosity, the corresponding normal tissue will not show hypermethylation or homozygosity, respectively. This assumption was reasonable and necessary because, for four samples, only tumor tissue was available. We tested this assumption in six patients (patients BWS1, BWS2, BWS3, BWS10, BWS11, and BWS16) by examining the methylation status of paired tumors and normal kidneys, which all showed abnormal methylation. In all cases, both the tumor tissue and the normal kidney tissue showed abnormal methylation. These data support the concept that normal methylation of *H19* in the tumor does not occur when there is abnormal *H19* methylation in the normal tissue. There is no mechanism for acquisition of heterozygosity at multiple microsatellite markers, other than microsatellite instability, which is rare in Wilms tumor (Mason et al. 2000), and the pattern of heterozygosity observed was not that seen in microsatellite instability—that is, with narrowly spaced alleles. We excluded patients in whom the tumor was the only available sample and had abnormal methylation (patients IH16 and BWS13), because we could not determine whether the normal tissue had normal or abnormal methylation. When available, we also tested patient blood genomic DNA to provide convincing evidence of UPD and LOH and tested pa-

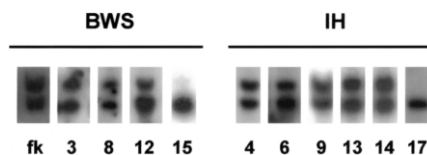


Figure 2 Analysis of *LIT1* methylation in patients with BWS and IH. An EST probe from the *LIT1* gene was hybridized to genomic DNA digested with *Bam*H1 and the methylcytosine-sensitive restriction enzyme *Not*I. The upper band represents the 6.0-kb unmethylated fragment, and the lower band represents the 4.2-kb unmethylated digested fragment. Patient numbers corresponding to those in the tables are shown below the lanes. All DNA specimens were from the normal kidney except samples for IH13, IH14, and IH17, which were from the Wilms tumor. fk = fetal kidney.

rental blood genomic DNA to identify the parental origin of microsatellite alleles. For one patient, although kidney tissue was not available, patient and parental blood genomic DNA was available for analysis (patient BWS9). The finding of UPD in the patient’s blood sample made the testing of other tissues unnecessary.

The majority of children with IH had a normal hemimethylated pattern of *H19* and *LIT1*. Molecular analyses of the 15 evaluable patients with IH (table 2) indicated that 12 patients had normal methylation of *H19* and *LIT1*. Of these 12, 4 had LOH of 11p15 (patients IH3, IH4, IH5, and IH11), 6 did not have LOH (patients IH1, IH2, IH10, IH13, IH14, IH15), and 2 did not have tumor tissue available to test for LOH (patients IH8 and

Table 2

Molecular Analysis of Kidney Tissue from Patients with IH and Wilms Tumor

PATIENT	NORMAL KIDNEY TISSUE			TUMOR KIDNEY TISSUE			NOTE
	<i>H19</i> Methylation	<i>LIT1</i> Methylation	Microsatellite Analysis	<i>H19</i> Methylation	<i>LIT1</i> Methylation	Microsatellite Analysis	
IH1	Normal	Normal	Normal	Normal	...
IH2	Normal	Normal	Normal	Normal	...
IH3	Normal	Normal	Normal	Normal	...
IH4	Normal	Normal	Normal	LOH	...
IH5	Normal	Normal	Normal	LOH	...
IH6	Abnormal	Normal	Normal	Normal	...
IH7	Abnormal	Abnormal	UPD
IH8	Normal	Normal	Normal
IH9	Normal	Normal	Normal
IH10	Normal	Normal	Normal	...
IH11	Normal	Normal	Normal	LOH	...
IH12	Abnormal	Abnormal	UPD
IH13	Normal	Normal	Normal	...
IH14	Normal	Normal	Normal	...
IH15	Normal	Normal	Normal	...
IH16	Abnormal	Normal	Normal	Excluded ^a
IH17	Abnormal	Abnormal	LOH or UPD?	Excluded ^b

^a Patient IH16 was excluded because it was not possible to determine whether the hypermethylation of *H19* in the tumor was constitutive or tumor specific.

^b Patient IH17 was excluded because it was not possible to determine whether multiple homozygous microsatellite markers were due to LOH or UPD.

Table 3**Molecular Analysis of Kidney Tissue from Patients with BWS and Wilms Tumor**

PATIENT	NORMAL KIDNEY TISSUE			TUMOR KIDNEY TISSUE			NOTE
	<i>H19</i> Methylation	<i>LIT1</i> Methylation	Microsatellite Analysis	<i>H19</i> Methylation	<i>LIT1</i> Methylation	Microsatellite Analysis	
BWS1	Abnormal	Normal	Normal	Abnormal	Normal	Normal	...
BWS2	Abnormal	Normal	Normal	Abnormal	Normal	Normal	...
BWS3	Abnormal	Normal	Normal	Abnormal	Normal	Normal	...
BWS4	Abnormal	Abnormal	UPD
BWS5	Abnormal	Abnormal	UPD
BWS6	Abnormal	Abnormal	UPD
BWS7	Abnormal	Abnormal	UPD
BWS8	Normal	Abnormal	Normal	LOH	...
BWS9	UPD in blood
BWS10	Abnormal	Normal	Normal	Abnormal	Normal	Normal	...
BWS11	Abnormal	Normal	Normal	Abnormal	Normal	Normal	...
BWS12	Normal	Abnormal	Normal	LOH	...
BWS13	Abnormal	Abnormal	Normal	Excluded ^a
BWS14	Abnormal	Abnormal	LOH or UPD?	Excluded ^b
BWS15	Normal	Normal	Normal	LOH	...
BWS16	Abnormal	Normal	Normal	Abnormal	Normal	Normal	...

^a Patient BWS13 was excluded because it was not possible to determine whether the hypermethylation of *H19* and *LIT1* in the tumor was constitutive or tumor specific.

^b Patient BWS14 was excluded because it was not possible to determine whether multiple homozygous microsatellite markers were due to LOH or UPD.

IH9). Three patients had an abnormal methylation pattern in *H19*, one patient had hypermethylation of *H19* in the normal kidney tissue and retention of heterozygosity in the tumor (patient IH6), and two patients had constitutive UPD (patients IH7 and IH12).

Unlike the children with IH and Wilms tumor, the majority of children with BWS and Wilms tumor had abnormal methylation patterns of *H19* or *LIT1*. Molecular analyses of 14 evaluable patients with BWS (table 3) indicated that 13 of them had abnormal methylation of either *H19* or *LIT1*. Six children had hypermethylation of *H19* alone, with retention of heterozygosity in the tumor (patients BWS1, BWS2, BWS3, BWS10, BWS11, and BWS16). Five patients had constitutive UPD (patients BWS4, BWS5, BWS6, BWS7, and BWS9) that included abnormal methylation in *H19* and *LIT1*, and two patients had hypomethylation of *LIT1* and LOH in the tumor (patients BWS8 and BWS12). One patient had normal methylation of *H19* and *LIT1* and LOH in the tumor (patient BWS15).

The frequency of hypermethylation of *H19* in children with IH and Wilms tumor, 20% (3/15), was significantly lower than the frequency in children with BWS and Wilms tumor, 79% (11/14; $P = .0028$) (table 4). In addition, this difference between IH and BWS remained significant when patients with UPD for 11p15, which encompasses *H19* along with many other genes, were excluded: hypermethylation frequencies of 67% (6/9) and 8% (1/13) for IH and BWS, respectively ($P = .0066$) (table 4).

The higher proportion of abnormal methylation of *H19* in patients with BWS and Wilms tumor corroborated our previous results in the BWS Registry and was expected (DeBaun et al. 2002). Given the significant overlap between BWS and IH, we did not expect the low frequency of *H19* methylation abnormalities in the patients with IH and Wilms tumor.

Several possibilities exist as to why the children with IH and Wilms tumor did not have the expected epigenetic abnormalities. Perhaps children with IH and Wilms tumor are genetically different from children with IH and without Wilms tumor. Alternatively, patients with IH may have a different genotype that acts in a common causal pathway and results in a similar phenotype to that of BWS. The rare cases of IH with UPD of 11p15.5, described here and by others (Grundy et al. 1991), suggest that there might be another gene on 11p15.5 included in the disomic region that is responsible for the phenotype.

An unexpected finding was LOH of 11p15.5 in Wilms tumors from patients with BWS. Specifically, three tumors from patients with BWS showed LOH; two arose in patients with hypomethylation of *LIT1*. In addition, four tumors from patients with IH were determined to have LOH, all arising in patients without any known epigenetic alteration. These data are consistent with a model we have proposed in which epigenetic alterations (hypermethylation of *LIT1*, in this case) represent the first “hit” in Knudson’s two-hit model, and subsequent genetic changes represent the second hit (Feinberg and

Table 4
Significant Difference in Frequency of Hypermethylation of *H19* in BWS and IH

Sample Tested and Patient Group	Hypermethylation of <i>H19</i> (%)	Normal <i>H19</i> (%)
Entire sample ^a :		
Patients with BWS	11/14 (79)	3/14 (21)
Patients with IH	3/15 (20)	12/15 (80)
Sample excluding UPD for 11p15 ^b :		
Patients with BWS	6/9 (67)	3/9 (33)
Patients with IH	1/13 (8)	12/13 (92)

^a *P* = .0028 for hypermethylation values.

^b *P* = .0066 for hypermethylation values.

Tycko 2004). Knudson’s hypothesis does not require allelism for the two hits, and, by our modified Knudson model, epigenetic alterations can lead to an expanded population of precursor cells that are targets for subsequent oncogenetic activation, thereby increasing the risk of malignancy in the presence of constitutional epigenetic alterations in BWS.

In summary, children with IH and Wilms tumor do not have the constitutional hypermethylation of *H19* that is associated with cancer, unlike those with BWS, in which cancer is associated with constitutional hypermethylation of *H19*.

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Web Resources

The URL for data presented herein is as follows:

Online Mendelian Inheritance in Man (OMIM), <http://www.ncbi.nlm.gov/Omim/> (for BWS and IH)

References

Breslow N, Beckwith JB, Ciol M, Sharples K (1988) Age distribution of Wilms’ tumor: report from the National Wilms’ Tumor Study. *Cancer Res* 48:1653–1657

Cui H, Horon IL, Ohlsson R, Hamilton SR, Feinberg AP (1998) Loss of imprinting in normal tissue of colorectal cancer patients with microsatellite instability. *Nat Med* 4:1276–1280

DeBaun MR, Niemitz EL, McNeil DE, Brandenburg SA, Lee MP, Feinberg AP (2002) Epigenetic alterations of *H19* and *LIT1* distinguish patients with Beckwith-Wiedemann syndrome with cancer and birth defects. *Am J Hum Genet* 70:604–611

DeBaun MR, Tucker MA (1998) Risk of cancer during the first four years of life in children from The Beckwith-Wiedemann Syndrome Registry. *J Pediatr* 132:398–400

Feinberg AP, Tycko B (2004) The history of cancer epigenetics. *Nat Rev Cancer* 4:143–153

Grundy P, Telzerow P, Paterson MC, Haber D, Berman B, Li F, Garber J (1991) Chromosome 11 uniparental isodisomy predisposing to embryonal neoplasms. *Lancet* 338:1079–1080

Hoyme HE, Seaver LH, Jones KL, Procopio F, Crooks W, Feingold M (1998) Isolated hemihyperplasia (hemihypertrophy): report of a prospective multicenter study of the incidence of neoplasia and review. *Am J Med Genet* 79:274–278

Mason JE, Goodfellow PJ, Grundy PE, Skinner MA (2000) 16q Loss of heterozygosity and microsatellite instability in Wilms’ tumor. *J Pediatr Surg* 35:891–896

Oken K, Kleinman KP, Rich-Edwards J, Gillman MW (2003) A nearly continuous measure of birth weight for gestational age using a United States national reference. *BMC Pediatr* 3:6

Ravenel JD, Broman KW, Perlman EJ, Niemitz EL, Jayawardena TM, Bell DW, Haber DA, Uejima H, Feinberg AP (2001) Loss of imprinting of insulin-like growth factor-II (IGF2) gene in distinguishing specific biologic subtypes of Wilms tumor. *J Natl Cancer Inst* 93:1698–1703