Strong opioids for cancer pain

Matthew K Makin MA MRCP

J R Soc Med 2001;94:17-21

SECTION OF ANAESTHESIA, 27 APRIL 2000

The past decade has seen much research on the mechanisms by which opioids promote analgesia. The challenge for clinicians involved in the management of chronic cancer pain is now to translate the knowledge that has emerged from basic science and animal models into safe and effective treatment strategies, not only for patients in hospitals and hospices but also for patients at home. This is all the more important because of the new directions being taken in the use of strong-opioid alternatives to morphine, where much practice is based upon personal experience rather than rigorous clinical evidence¹. In this paper I outline the role of morphine in the management of cancer pain and discuss the role of strong-opioid alternatives.

As yet, no drug has shown sufficient advantages to supersede oral morphine as the strong opioid of first choice in chronic cancer pain. Three elements are critical to achieving an acceptable balance between side-effects and analgesia—thorough clinical assessment, careful dose titration and the use of laxatives. Most patients can be kept on a stable dose of morphine for weeks or months². If requirements increase, this is usually because nociceptive input has increased with progression of the disease, and this calls for a rigorous reassessment of the pain syndrome³.

SAFETY AND TOLERABILITY OF CHRONIC THERAPY WITH MORPHINE

The side-effects of chronic morphine therapy range from nausea and constipation to mild cognitive impairment, somnolence, myoclonus and hallucinations. Respiratory depression is an effect to which tolerance develops rapidly, allowing use in chronic cancer pain without serious respiratory risk. Constipation is mediated via enteric as well as spinal and possibly supraspinal opioid receptors⁴ and tolerance to this side-effect does not seem to develophence the almost universal need for laxatives. In most cases the cognitive impairment and dizziness that occurs within a few days of beginning therapy or increasing the dose is selflimiting. O'Neill et al.5, examining the effects of repeated oral doses of morphine, found that patients receiving small doses actually showed enhanced performance in some measures of cognitive function, and in cancer patients receiving long-term morphine treatment Vainio et al.6

detected only a slight and selective effect on functions related to driving.

Somnolence and a constellation of signs and symptoms referred to by some workers as opioid-induced neurotoxicity (delirium, hallucinations, multifocal myoclonus)⁷ can limit the dose during chronic therapy. The collective symptoms of opioid-induced neurotoxicity, which are not reversed by naloxone⁸, seem to be mediated by at present unidentified non-opioid receptors⁹.

Risk factors associated with the development of opioid-induced neurotoxicity include inappropriately rapid escalation of dose, advanced age of the patient, renal impairment, poor hydration and the concomitant use of other psychoactive drugs¹⁰. There are also case reports describing the rare phenomenon of morphine or diamorphine induced hyperalgesia and allodynia, which has occurred when the opioid dose was increased despite signs of opioid-induced neurotoxicity¹¹.

MORPHINE METABOLITES

Morphine is metabolized predominantly in the liver, by glucuronidation. The major metabolites are morphine-3-glucuronide (M3G) and morphine-6-glucuronide (M6G). Because the glucuronides are water-soluble and excreted in urine, there is an increased risk of side-effects in patients with renal impairment.

In laboratory animals M3G has no analgesic action; but, despite lacking affinity for opioid receptors, it is many times more potent than morphine in eliciting opioid-induced neurotoxicity¹². M6G, on the other hand, displays high affinity for the μ opioid receptor and its antinociceptive activity is greater than that of morphine in laboratory animals¹³. Recently, genetically modified mice lacking one or other opioid receptor have been produced. Morphine had no analgesic activity in mice lacking the μ opioid receptor, whereas M6G and diamorphine were potentially antinociceptive¹⁴. These findings suggest that, in addition to working via the μ opioid receptor, M6G acts at a receptor separate from that which mediates morphine analgesia. This is unlikely to be a δ or κ opioid receptor since the antinociception is not blocked by the relevant selective antagonists¹⁵.

In a study of 109 cancer patients no correlation was shown between myoclonus and cognitive dysfunction and plasma concentrations of morphine and M6G¹⁶. On the

other hand, three case series have suggested that high concentrations of M3G in plasma and a high ratio of M3G to M6G in cerebrospinal fluid are important factors in the development of opioid-induced neurotoxicity in patients receiving chronic morphine therapy^{16–18}. Some workers have speculated that M3G might antagonize the action of morphine, but this seems unlikely. An electrophysiological study in rats demonstrated that M3G did not possess any significant antinociceptive effect¹⁹ and in a series of 39 cancer patients no correlation was found between high M3G concentration and poor analgesia. None of the patients in this series, however, displayed signs or symptoms of opioid-induced neurotoxicity²⁰.

MORPHINE 'POORLY RESPONSIVE' PAIN

The World Health Organization, in its approach to cancer pain management, advocates the use of strong opioids not as single agents but in combination with non-opioids, usually non-steroidal anti-inflammatory drugs and appropriate adjuvant analgesics. This strategy should offer good or moderate relief of pain in about 80% of cancer patients. Morphine has no ceiling dose and requirements vary considerably between individual patients. A 'poor response' to morphine—i.e. inadequate analgesia—is best viewed as a consequence of dose-limiting side-effects. Neuropathic pain and movement-related pain are disproportionately represented amongst patients whose pain responds poorly to morphine²¹.

Neuropathic pain

Neuropathic pain arises from dysfunction of the peripheral or central nervous system and is typically experienced in an area of altered sensation. The examining clinician may be able to elicit pain by means of a stimulus that does not normally cause pain (allodynia), or identify an exaggerated response to a normally painful stimulus (hyperalgesia). In a series of 595 cancer patients referred to a pain service based in an anaesthesia department, 213 (36%) had pain with a neuropathic component²².

Neuropathic pain states are maintained by complex mechanisms involving altered peripheral activity, central excitatory or inhibitory activity and sympathetic nervous system activity²³. Another factor is disruption of nerve fibres, which can reduce opioid sensitivity through loss of opioid receptors²⁴.

In animal models that simulate peripheral nerve damage, a wide variety of neuropeptides and their receptors are affected. Most notable is cholecystokinin (CCK), which may have a critical role in modulating response to opioid therapy²⁵. CCK receptors are intimately linked to μ opioid receptors and, when CCK binds to its receptor, changes are induced that lessen the response to μ opioid agonists such as

morphine²⁶. High levels of CCK are associated with a decline in opioid effectiveness; conversely, opioid sensitivity is increased by the use of CCK antagonists²⁷. In animal models, CCK receptors appear to be upregulated in neuropathic pain, and this could further contribute to the poor response to morphine²⁸.

There is increasing evidence that neuropathic pain states involve prolonged activation of the *N*-methyl-D-aspartate (NMDA) receptor, leading to increased neuronal activity or 'wind-up'²³. In this state of hypersensitization, mechanical and thermal stimuli are amplified, with development of hyperalgesia. Activation of the NMDA receptor has also been implicated in the development of tolerance to morphine, and a likely site of action for both hyperalgesia and morphine tolerance is the dorsal horn of the spinal cord²⁹. Serotonin and noradrenaline dampen incoming pain impulses in descending inhibitory pathways³⁰ but become deactivated following reuptake at nerve terminals; drugs that inhibit this reuptake, such as amitriptyline, have proved effective in the management of neuropathic pain.

Some commentators have suggested that neuropathic pain should be regarded as inherently insensitive to opioids³¹. Others argue that opioid responsiveness in neuropathic pain should be regarded as relative, as a consequence of a shift in the dose response curve to the right, though recognizing that in practice, dose-limiting side-effects prohibit escalation to a level sufficient to produce effective analgesia²¹.

STRONG-OPIOID ALTERNATIVES TO MORPHINE

When cancer pain responds poorly to morphine despite appropriate adjuvant analgesia, the patient and his or her pain syndrome must be carefully reassessed. One strategy is to use lower doses of morphine (or diamorphine) via the epidural or intrathecal route. If this is impracticable or undesirable an alternative is to use an alternative strong opioid, to provide a better balance between side-effects and analgesia. In the UK the number of alternative strong opioids licensed for management of chronic cancer pain has expanded rapidly. These drugs are now available in immediate-release and sustained-release preparations as well as novel delivery systems.

Oxycodone, used widely in the US, is now licensed in the UK. In controlled trials including patients with cancer, oxycodone was at least as effective an analgesic as controlled-release morphine³¹. There have been reports of a lower incidence of delirium with oxycodone³². Vomiting is less frequent than with morphine, constipation more so³³.

Transdermal fentanyl is a popular alternative to morphine and is particularly useful if the oral route is not possible. An open randomized cross-over study compared transdermal fentanyl with sustained-release oral morphine in 202 cancer patients. In this study fentanyl was associated with less constipation and daytime drowsiness than morphine but greater sleep disturbance and shorter sleep duration³⁴. Oral transmucosal fentanyl citrate, a novel preparation resembling a lollipop, may have a place in the treatment of breakthrough and incident pain in cancer patients. The onset of action is rapid. Dose titration is required, however, since there is no clear relation between the total daily dose of the long-acting opioid the patient is taking and the dose of transmucosal fentanyl needed to manage breakthrough pain³⁵.

Galer³⁶ reported success, in a series of patients with morphine 'poorly responsive' pain, after switches from morphine to hydromorphone, levorphanol or methadone. The benefit of such switches would depend on incomplete cross-tolerance between the drugs, with cross-tolerance to the analgesic effect less than cross-tolerance to the sideeffects. Bruera et al.37 examined this phenomenon in cancer patients by studying 48 switches between morphine and hydromorphone and 65 switches from hydromorphone to methadone. The dose ratio for morphine/hydromorphone did not change over a wide range of doses, and they saw this as evidence for complete cross-tolerance between these two opioids. But the hydromorphone/ methadone ratios were higher in those patients receiving large hydromorphone doses, and this they took to be evidence of incomplete cross-tolerance between hydromorphone and methadone.

When a patient has toxic effects from morphine, should an alternative strong opioid then be tried? The rationale for this controversial proposal is that opioid toxicity associated with the accumulation of active metabolites should be relieved by a switch to an opioid free of such metabolites e.g. oxycodone, fentanyl or methadone³⁸. Some centres suggest that as many as 41% of patients require a switch to an alternative opioid, whilst others quote a figure close to 2%. Consensus amongst UK palliative care physicians is that a diagnosis of opioid toxicity necessitates careful evaluation, being one of the many potential causes of similar symptoms and signs in patients with advanced cancer. In a retrospective review of 138 referrals to a hospital-based palliative care team, opioid toxicity was identified as being a major contributing factor in 13 out of 57 patients loosely categorized as 'confused'. Symptoms resolved in 11 of these 13 patients with a reduction in opioid dose rather than a switch to an alternative strong opioid³⁹.

OPIOID ACTIVITY AT NON-OPIOID RECEPTORS

Opioids act by binding to and activating μ , δ and κ opioid receptors. Morphine has a high affinity for the μ receptor with at least 50 times less affinity for the δ receptor and

negligible affinity for the κ receptor⁴⁰. Some opioids are distinct from morphine in having additional activity, acting as non-competitive NMDA receptor antagonists and inhibitors of monoamine reuptake. Dextromethorphan, methadone and levorphanol seem the most promiscuous of these opioids with affinity for the NMDA receptor in the low μ mol range (similar to that of ketamine) and an ability to inhibit the reuptake of noradrenaline and serotonin in some cases at nmol concentrations. NMDA receptor antagonists can block the hypersensitivity seen in neuropathic pain, potentiating the analgesic action of morphine⁴¹ and attenuating the development of morphine tolerance. Serotonin and noradrenaline reuptake inhibitors act in synergy with morphine to promote analgesia⁴² and, interestingly, most of the opioids that display activity at the NMDA receptor also inhibit the reuptake of these monoamines.

In animal models, methadone reduces pain behaviour provoked by stimuli mimicking neuropathic pain, and attenuates the development of tolerance—although electrophysiological studies have suggested that this is primarily an effect mediated via opioid receptors⁴³. As yet, reports of the efficacy of methadone in the management of cancerrelated neuropathic pain are anecdotal, but clinical trials are in progress.

Dextromethorphan, however, in combination with morphine (Morphidex) enhanced analgesia in a double-blind study⁴⁴, although when used as a single agent it did not show any analgesic activity⁴⁵.

This striking synergy between NMDA receptor antagonists and μ opioid agonists is seen with the subcutaneous coadministration of ketamine (a potent NMDA receptor antagonist) and diamorphine—a strategy that has been reported highly effective in many cases of otherwise intractable cancer pain⁴⁶. There is also a good case for assessing more closely the utility of opioids such as methadone or levorphanol, particularly when cancer pain is associated with hyperalgesia, allodynia or the rapid development of tolerance to morphine.

Levorphanol, obtainable in the USA and Scandinavia, is no longer available in the UK. Unfortunately methadone, with its long and variable half-life, requires careful dose titration during the initial switch to avoid toxicity. Long-term follow-up has also shown an appreciable incidence of side-effects, with nearly half of 54 cancer patients reporting excessive drowsiness⁴⁷.

CONCLUSION

Whilst the under-use of opioid drugs undoubtedly leads to unnecessary suffering⁴⁸, clinicians are faced with a bewildering number of strong opioids to choose from. Carefully designed studies are needed to clarify the role of

strong-opioid alternatives to morphine. Meanwhile, oral morphine remains the strong opioid of first choice for management of chronic cancer pain.

Note

Conflicts of interest: I have received educational grants from Parke-Davies, Boehringer Ingelheim, Janssen Cilag and Napp Pharmaceuticals.

REFERENCES

- 1 McQuay H, Moore A. Need for rigorous assessment of palliative care. BMJ 1994;309:1315–16
- 2 Brescia FJ, Portenoy RK, Ryan M, Krasnoff L, Gray G. Pain, opioid use, and survival in hospitalized patients with advanced cancer. J Clin Oncol 1992;10:49–55
- 3 Collin E, Poulain P, Gauvain-Piquard A, Petit G, Pichard-Leandri E. Is disease progression the major factor in morphine 'tolerance' in cancer pain treatment? *Pain* 1993;55:319–26
- 4 Culpepper-Morgan JA. Treatment of opioid-induced constipation with oral naloxone: a pilot study. Clin Pharmacol Ther 1992;52:90–5
- 5 O'Neill WM, Hanks GW, Simpson P, Fallon MT, Jenkins E, Wesnes K. The cognitive and psychomotor effects of morphine in healthy subjects: a randomized controlled trial of repeated (four) oral doses of dextropropoxyphene, morphine, lorazepam and placebo. *Pain* 2000:85:209–15
- 6 Vainio A, Ollila J, Matikainen E, Rosenberg P, Kalso E. Driving ability in cancer patients receiving long-term morphine analgesia. *Lancet* 1995;345:667–70
- 7 Daeninck PJ, Bruera E. Opioid use in cancer pain. Is a more liberal approach enhancing toxicity? Acta Anaesthesiol Scand 1999;43:924–38
- 8 Mercadante S. The role of morphine glucuronides in cancer pain. Palliative Med 1999;13:95–104
- 9 Bartlett SE, Dodd PR, Smith MT. Pharmacology of morphine and morphine-3-glucuronide at opioid, excitatory amino acid, GABA and glycine binding sites. *Pharmacol Toxicol* 1994;75:73–81
- 10 Potter JM, Reid DB, Shaw RJ, Hackett P, Hickman PE. Myoclonus associated with treatment with high doses of morphine: the role of supplemental drugs. BMJ 1989;299:150–3
- 11 Heger S, Maier C, Otter K, Helwig U, Suttorp M. Morphine induced allodynia in a child with brain tumour. *BMJ* 1999;319:627–9
- 12 Halliday AJ, Bartlett SE, Colditz P, Smith MT. Brain region-specific studies of the excitatory behavioral effects of morphine-3-glucuronide. *Life Sci* 1999;**65**:225–36
- 13 Sullivan AF, McQuay HJ, Bailey D, Dickenson AH. The spinal antinociceptive actions of morphine metabolites morphine-6glucuronide and morphine in the rat. Brain Res 1989;482:219–24
- 14 Schuller AG, King M, Zhang J, et al. Retention of heroin and morphine-6 beta-glucuronide analgesia in a new line of mice lacking exon 1 of MOR-1. Nat Neurosci 1999;2:151–6
- 15 Rossi GC, Brown GP, Leventhal L, Yang K, Pasternak GW. Novel receptor mechanisms for heroin and morphine-6-glucuronide induced analgesia. *Neurosci Lett* 1996;216:1–4
- 16 Tiseo PJ, Thaler HT, Lapin J, Inturrisci CE, Portenoy RK, Foley KM. Morphine-6-glucuronide concentrations and opioid-related side effects: a survey in cancer patients. *Pain* 1995;61:47–54
- Morley JS, Watt J, Wells JC, McKibbin PE, Miles JB. Paradoxical and other pains uncontrolled by morphine. In: Gebhart GK, Hammond DL, Jensen TS, eds. Proceedings of the Seventh World Congress on Pain (Progress in Pain Research and Management Vol 2). Seattle, WA: IASP Press. 1994:621–30
- 18 Smith MT, Wright A, Williams B, Stuart G, Gramond T. Cerebrospinal fluid and plasma concentrations of morphine,

- morphine-3-glucuronide, and morphine-6-glucuronide in patients before and after initiation of intracerebroventricular morphine for cancer pain management. *Anesth Analg* 1999;88:109–16
- 19 Hewett K, Dickenson AH, McQuay H. Lack of effect of morphine-3-glucuronide on the spinal antinociceptive actions of morphine in the rat: an electrophysiological study. *Pain* 1993;52:659–63
- 20 Faura CC, Moore A, Horga JF, Hand C, McQuay H. Morphine and M-6-G plasma concentrations and effect in cancer pain. J Pain Symptom Management 1996;11:95–102
- 21 Portenoy RK, Foley KM, Inturrisi CE. The nature of opioid responsiveness and its implications for neuropathic pain: new hypotheses derived from studies of opioid infusions. *Pain* 1990;43:273–86
- 22 Grond S, Radbruch L, Meuser T, Sabatowski R, Loick G, Lehmann A. Assessment and treatment of neuropathic cancer pain following WHO guidelines. *Pain* 1999;79:15–20
- 23 Dickenson AH. NMDA receptor antagonists: interactions with opioids. Acta Anaesthesiol Scand 1997;41:112–15
- 24 Besse D, Lombard MC, Zajac JM. Pre- and postsynaptic distribution of mu, delta and kappa opioid receptors in the superficial layers of the cervical dorsal horn of the rat spinal cord: a quantitative autoradiographic study. Brain Res 1990;521:15–22
- 25 Zhang X, Dangerlind Å, Elde RP, et al. Marked increase in cholecystokinin B receptor mesenger RNA levels in rat dorsal root ganglia after peripheral axotomy. Neuroscience 1993;57:227–33
- 26 Dertwinkel R, Zenz M, Strumpf M, Donner B. Clinical status of opioid tolerance in long-term therapy of chronic noncancer pain. In: Kalso E, McQuay H, Wiesenfeld-Hallin Z, eds. Opioid Sensitivity of Chronic Noncancer Pain. Seattle: IASP Press, 1999:129–41
- 27 Dickenson AH. Neurophysiology of opioid poorly responsive pain. Cancer Surv 1994;21:5–16
- 28 Antunes Bras JM, Benoliel JJ, Bourgoin S, et al. Effects of peripheral axotomy on cholecystokinin neurotransmission in the rat spinal cord. J Neurochem 1999;72:858–67
- 29 Mao J, Price DD, Mayer DJ. Mechanisms of hyperalgesia and morphine tolerance: a current view on their possible interactions. *Pain* 1995;62:259–74
- 30 Budd K, Monoamine function and analgesia. Pain Rev 1994;1:3-8
- 31 Mucci-LoRusso P, Berman BS, Silberstein PT, et al. Controlled-release oxycodone compared with controlled-release morphine in the treatment of cancer pain: a randomized, double-blind, parallel-group study. Eur J Pain 1998;2:239–49
- 32 Maddocks I, Somogyi A, Abbott F, Hayball P, Parker D. Attenuation of morphine-induced delirium in palliative care by substitution with infusion of oxycodone. J Pain Symptom Management 1996;12:182–9
- 33 Heiskanen T, Kalso E. Controlled-release oxycodone and morphine in cancer related pain. Pain 1997;73:37–45
- 34 Ahmedzai S, Brooks D. Transdermal fentanyl versus sustained-release oral morphine in cancer pain: preference, efficacy, and quality of life. The TTS-Fentanyl Comparative Trial Group. J Pain Symptom Management 1997;13:254–61
- 35 Portenoy RK, Payne R, Coluzzi P, et al. Oral transmucosal fentanyl citrate (OTFC) for the treatment of breakthrough pain in cancer patients: a controlled dose titration study. Pain 1999;79:303–12
- 36 Galer BS, Coyle N, Pasternak GW, Portenoy RK. Individual variability in the response to different opioids: report of five cases. *Pain* 1992;49:87–91
- 37 Bruera E, Pereira J, Watanabe S, Belzile M, Kuehn N, Hanson J. Opioid rotation in patients with cancer pain. A retrospective comparison of dose ratios between methadone, hydromorphone and morphine. *Cancer* 1996;78:852–7
- 38 de Stoutz ND, Bruera E, Suarez-Almazor M. Opioid rotation for toxicity reduction in terminal cancer patients. J Pain Symptom Management 19995;10:378–84

- 39 Hawley P, Forbes K, Hanks GW. Opioids, confusion and opioid rotation. Palliative Med 1998;12:63–4
- 40 Dickenson AH. Pain and analgesia: new light on mechanism and therapy. CME Bulletin Palliative Med 1999;1(4):98–103
- 41 Chapman V, Dickenson AH. The combination of NMDA receptor antagonists and morphine produces profound antinociception in the rat dorsal horn. *Brain Res* 1992;573:321–3
- 42 Taiwo YO, Fabian A, Pazoles CJ, Fields HL. Potentiation of morphine antinociception by monoamine reuptake inhibitors in the rat spinal cord. *Pain* 1985;21:329–37
- 43 Carpenter K, Chapman V, Dickenson AH. Neuronal inhibitory effects of methadone are predominantly opioid receptor mediated in the rat spinal cord in vivo. Europ J Pain 2000;4:19–26

- 44 Katz N. MorphiDex (double-blind, multiple-dose studies in chronic pain patients). *J Pain Symptom Management* 2000;19(suppl):37–41
- 45 McQuay H, Carroll D, Jadad AR, et al. Dextromethorphan for the treatment of neuropathic pain: a double-blind randomised controlled crossover trial with integral n-of-1 design. Pain 1994;59:127–33
- 46 Finlay I. Ketamine and its role in cancer pain. Pain Rev 1999;6: 303–13
- 47 Makin MK, Coackley A, Duckitt H, Fleming J, Skinner J, Ellershaw JE. A report of adverse events occurring during methadone therapy for chronic cancer pain. *Proc Palliative Care Congress*, Warwick, 27–29 March, 2000
- 48 Zenz M, Willweber-Strumpf A. Opiophobia and cancer pain in Europe. *Lancet* 1993;341:1075–6