

Lopinavir Measurement in Pleural Effusion in a Human Immunodeficiency Virus Type 1-Infected Patient with Kaposi's Sarcoma

Human immunodeficiency virus type 1 (HIV-1) infection is highly compartmentalized within various organs. Tissue dissemination is evident in the lung (1), the male (3) and female (8) genital tracts, the lymph nodes (12), and especially in the brain (5, 12). This process may contribute to the increase in systemic HIV-1 heterogeneity.

Although multiple antiretroviral drug regimens have led to a dramatic decline in HIV-related morbidity and mortality (6), viral eradication is not yet achievable with existing regimens, a failure likely to be attributable to poor drug penetration into critical tissues. Despite adequate plasma drug concentrations and associated systemic virological efficacy (7), poor drug diffusion into particular body sites is thought to be responsible for the selection of drug-resistant HIV quasispecies which keep on seeding the systemic compartment, thus eventually leading to emergence of HIV-1 variants resistant to antiretroviral drugs.

Limited data available on drug concentrations of highly bound protease inhibitors show that penetration of these drugs into semen (10) and brain (4) is poor. In addition to considerations of protein binding, passage into body compartments is likely related to limited lipid solubility, high molecular weight, extensive binding to plasma proteins, or affinity for ATP-dependent efflux membrane transporters (11). Recently, high viral load values were found in the pleural space, and these were shown to be responsible for HIV quasispecies migration from the pleural space to the blood (2).

As no data exist on the penetration of antiretroviral drugs in pleural fluid, we measured concentrations of lopinavir, a highly specific HIV-1 protease inhibitor, in plasma and pleural fluid of an HIV-1-infected male admitted to our ward for cutaneous and pulmonary Kaposi's sarcoma associated with bilateral pleural effusion. The subject had a good response to antiretroviral therapy (consisting of stavudine, 40 mg twice a day [BID]; lamivudine, 150 mg BID; and lopinavir-ritonavir, 400 and 100 mg BID) with a CD4 cell count increase from 70 to 352 cells mm^{-3} and a plasma viral load decrease of 2.7 \log_{10} after 3 months of treatment. When blood and pleural fluid were collected for drug analysis, the patient had been on lopinavir-ritonavir for 19 days and was about to initiate a specific treatment for Kaposi's sarcoma.

Lopinavir concentrations were assessed 12 h after a dose of lopinavir-ritonavir by high-performance liquid chromatography–dual mass spectrometry (9) and were seen to be similar in pleural fluid and plasma (12,439 and 12,691 $\text{ng} \cdot \text{ml}^{-1}$, respectively). As lopinavir is coformulated with low-dose ritonavir to enhance plasma lopinavir concentrations, ritonavir concentrations in pleural fluid and plasma were also assessed and were 352 and 214 $\text{ng} \cdot \text{ml}^{-1}$, respectively.

In normal conditions, lower concentrations of lopinavir may be expected in pleural fluid, since passage of highly bound drug, such as lopinavir, through parietal and visceral pleura is limited. However, as suggested by laboratory findings (pleural protein content, 3.7 $\text{g} \cdot \text{dl}^{-1}$; pleural fluid protein/serum protein ratio, 0.6; pleural fluid lactate dehydrogenase [LDH], 544 $\text{IU} \cdot \text{liter}^{-1}$; pleural fluid LDH/serum LDH ratio, 2), the presence of acute inflammation in the pleural space with significant exudative effusion can account for the comparable concentra-

tions of lopinavir and ritonavir that we found in pleural fluid and plasma. However, drug concentrations were here measured at a single time point which may not reflect the pleural fluid/plasma ratio over the 12-h dosing schedule since the penetration of a drug like lopinavir-ritonavir into a closed space such as the pleura may be relatively slow.

To what extent the rate of penetration of antiretrovirals into pleural fluid may be of therapeutic concern is presently unknown. However, since enhanced HIV replication may result (both systemically and locally) from immune activation by opportunistic pathogens, the presence of suppressive concentrations of antiretrovirals in the site where the concomitant process is ongoing does appear to be important.

Further studies are clearly needed to investigate the relevance of HIV replication and antiretroviral drug penetration into pleural effusions and other tissue compartments.

REFERENCES

1. Ait-Khaled, M., J. E. McLaughlin, M. A. Johnson, and V. C. Emery. 1995. Distinct HIV-1 long terminal repeat quasispecies present in nervous tissues compared to that in lung, blood and lymphoid tissues of an AIDS patient. *AIDS* 9:675–683.
2. Collins, K. R., M. E. Quinones-Mateu, M. Wu, H. Luzze, J. L. Johnson, C. Hirsch, Z. Toossi, and E. J. Arts. 2002. Human immunodeficiency virus type 1 (HIV-1) quasispecies at the sites of *Mycobacterium tuberculosis* infection contribute to systemic HIV-1 heterogeneity. *J. Virol.* 76:1697–1706.
3. Coombs, R. W., C. E. Speck, J. P. Hughes, W. Lee, R. Sampoleo, S. O. Ross, J. Dragavon, G. Peterson, T. M. Hooton, A. C. Collier, L. Corey, L. Koutsky, and J. N. Krieger. 1998. Association between culturable human immunodeficiency virus type 1 (HIV-1) in semen and HIV-1 RNA levels in semen and blood: evidence for compartmentalization of HIV-1 between semen and blood. *J. Infect. Dis.* 177:320–330.
4. Enting, R. H., R. M. Hoetelmans, J. M. Lange, D. M. Burger, J. H. Beijnen, and P. Portegies. 1998. Antiretroviral drugs and the central nervous system. *AIDS* 12:1941–1955.
5. Korber, B. T., K. J. Kunstman, B. K. Patterson, M. Furtado, M. M. McEvilly, R. Levy, and S. M. Wolinsky. 1994. Genetic differences between blood- and brain-derived viral sequences from human immunodeficiency virus type 1-infected patients: evidence of conserved elements in the V3 region of the envelope protein of brain-derived sequences. *J. Virol.* 68:7467–7481.
6. Palella, F. J., Jr., K. M. Delaney, A. C. Moorman, M. O. Loveless, J. Fuhrer, G. A. Satten, D. J. Aschman, S. D. Holmberg, et al. 1998. Declining morbidity and mortality among patients with advanced human immunodeficiency virus infection. *N. Engl. J. Med.* 338:853–860.
7. Pialoux, G., S. Fournier, A. Moulignier, J. D. Poveda, F. Clavel, and B. Dupont. 1997. Central nervous system as a sanctuary for HIV-1 infection despite treatment with zidovudine, lamivudine and indinavir. *AIDS* 11:1302–1303.
8. Poss, M., A. G. Rodrigo, J. J. Gosink, G. H. Learn, D. de Vange Panteleeff, D. J. White, Jr., J. Bwayo, J. K. Kreiss, and J. Overbaugh. 1998. Evolution of envelope sequences from the genital tract and peripheral blood of women infected with clade A human immunodeficiency virus type 1. *J. Virol.* 72:8240–8251.
9. Reynolds, H. E., J. F. Tjia, S. E. Gibbons, S. H. Khoo, and D. J. Back. 2001. Simultaneous determination of four HIV protease inhibitors by HPLC-MS/MS for use in a therapeutic drug monitoring service. *Br. J. Clin. Pharmacol.* 52:481P.
10. Taylor, S., D. J. Back, S. M. Drake, J. Workman, H. Reynolds, S. E. Gibbons, D. J. White, and D. Pillay. 2001. Antiretroviral drug concentrations in semen of HIV-infected men: differential penetration of indinavir, ritonavir and saquinavir. *J. Antimicrob. Chemother.* 48:351–354.
11. van Praag, R. M., G. J. Weverling, P. Portegies, S. Jurriaans, X. J. Zhou,

M. L. Turner-Foisy, J. P. Sommadossi, D. M. Burger, J. M. Lange, R. M. Hoetelmans, and J. M. Prins. 2000. Enhanced penetration of indinavir in cerebrospinal fluid and semen after the addition of low-dose ritonavir. *AIDS* **14**:1187–1194.

12. **Wong, J. K., C. C. Ignacio, F. Torriani, D. Havlir, N. J. Fitch, and D. D. Richman.** 1997. In vivo compartmentalization of human immunodeficiency virus: evidence from the examination of *pol* sequences from autopsy tissues. *J. Virol.* **71**:2059–2071.

Marta Boffito*
Patrick G. Hoggard
David J. Back
Pharmacology Research Laboratories
University of Liverpool
Block H, First Floor
70 Pembroke Place
Liverpool L69 3GF, United Kingdom

Stefano Bonora
Agostino Maiello
Anna Lucchini
Giovanni Di Perri
Department of Infectious Diseases
University of Turin
Turin, Italy

*Phone: 44 151 7945565
Fax: 44 151 7945656
E-mail: martalbb@hotmail.com