

# Of yeast and men

## The evolution of PtdIns(3,4,5)P<sub>3</sub> synthesis

Since the identification of the lipid second messenger phosphatidylinositol (3,4,5)-trisphosphate (PtdIns(3,4,5)P<sub>3</sub>; Traynor-Kaplan *et al*, 1988; Auger *et al*, 1989), this molecule has become a paradigm for how phosphoinositides (PIs) mediate complex signalling events in cells. PtdIns(3,4,5)P<sub>3</sub> can regulate chemotaxis, cell survival, translation and cytoskeletal rearrangement (Cantley, 2002), the deregulation of which contributes to the pathology of diseases such as diabetes, and breast and prostate cancer.

PtdIns(3,4,5)P<sub>3</sub> is a phosphorylated form of the PI phosphatidylinositol (PtdIns). All PIs have two fatty-acid chains that anchor them to membranes and a hydrophilic inositol head group that is exposed to the aqueous cytosol (Fig 1A). This head group can be phosphorylated at the 3-, 4- and 5-positions and so far, a total of seven phosphorylated derivatives of PtdIns have been reported (Fig 1A).

After the treatment of cells with certain agonists, the transient synthesis of PtdIns(3,4,5)P<sub>3</sub> is initiated by class I PI 3-kinases. These lipid kinases are normally inactive; however, agonist stimulation increases their ability to phosphorylate the 3-position of PtdIns(4,5)P<sub>2</sub> to generate PtdIns(3,4,5)P<sub>3</sub> (Hawkins *et al*, 1992). The receptor-driven activation of class I PI 3-kinases can give rise to greater than a tenfold increase in the cellular levels of PtdIns(3,4,5)P<sub>3</sub> (which rarely approach 0.1% of total PI levels, making its detection difficult—even for the more hardened lipid biochemists among us). In turn, subsets of proteins that specifically recognize and bind to PtdIns(3,4,5)P<sub>3</sub> (mediated by specialist lipid-binding modules such as pleckstrin homology (PH) domains) are then recruited to the membrane, which often leads to their activation and further transduction of the agonist signal (Fig 1B; Lemmon, 2003).

Recently, PtdIns(3,4,5)P<sub>3</sub> has stepped into the scientific limelight once again, this time not because of its biological function but intriguingly because of the way it is synthesized in fission yeast. A recent article in the *Journal of Cell Biology* (Mitra *et al*, 2004) has revealed for the first time that *Schizosaccharomyces pombe* can generate the 3-phosphorylated lipid PtdIns(3,4,5)P<sub>3</sub>. This is exciting because researchers have assumed that yeast are incapable of producing PtdIns(3,4,5)P<sub>3</sub>. This dogma has arisen as PtdIns(3,4,5)P<sub>3</sub> has never been found in yeast, and yeast do not have a class I PI 3-kinase, which suggests that they lack one of the essential reaction steps necessary to generate PtdIns(3,4,5)P<sub>3</sub>.

The paper by Mitra and co-workers begins with the identification of *ptn1*, a homologue of the mammalian phosphatase and tensin homologue (PTEN) gene. PTEN is a lipid phosphatase (Maehama *et al*, 1998), which dephosphorylates the 3-position of PtdIns(3,4,5)P<sub>3</sub> to generate PtdIns(4,5)P<sub>2</sub>. When the authors tested the lipid phosphatase

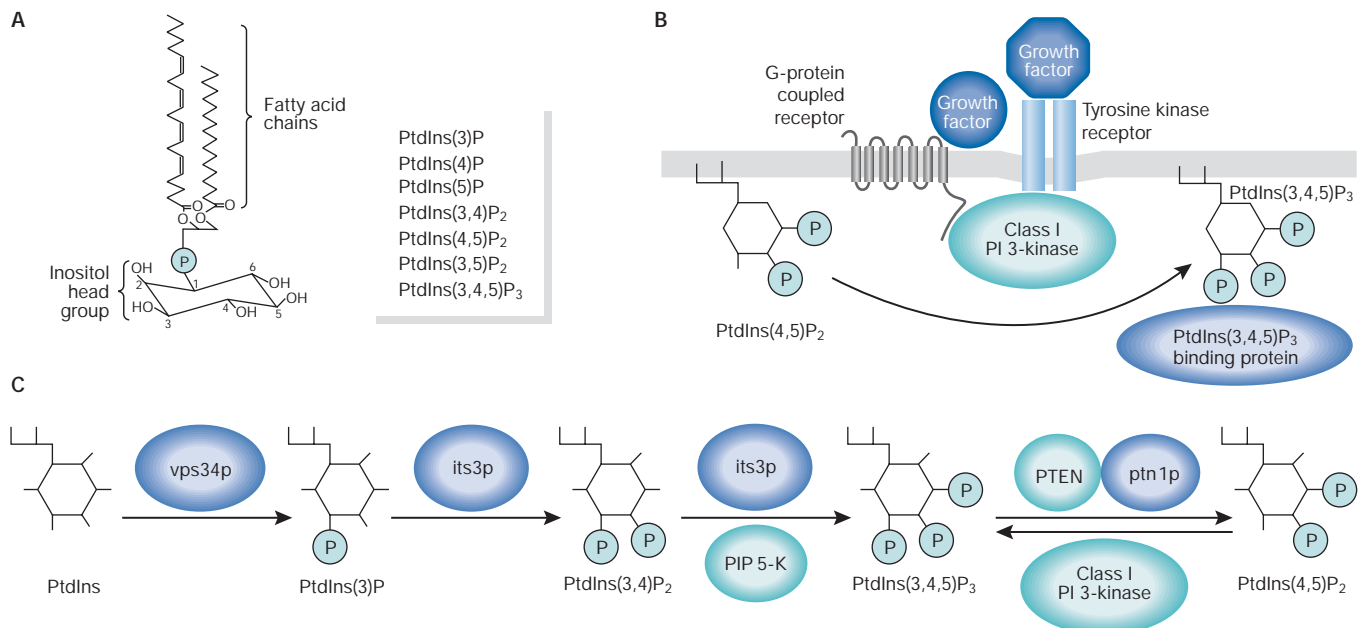
activity of Ptn1 *in vitro*, they found that it also dephosphorylated PtdIns(3,4,5)P<sub>3</sub>. This led Mitra and colleagues to question why *S. pombe* would have maintained a functional PtdIns(3,4,5)P<sub>3</sub> phosphatase in its genome if not to regulate PtdIns(3,4,5)P<sub>3</sub> levels.

In mammalian cells, disruption of PTEN activity leads to elevated PtdIns(3,4,5)P<sub>3</sub> levels and surprisingly, examination of the phosphoinositide levels in an *S. pombe* strain lacking *ptn1* (*ptn1Δ*) revealed a dramatic increase in PtdIns(3,4)P<sub>2</sub> and PtdIns(3,4,5)P<sub>3</sub>. This finding is noteworthy in itself; however, the authors went on to postulate that *S. pombe* synthesizes PtdIns(3,4,5)P<sub>3</sub> through a pathway that is different to the one used by mammalian cells after growth factor stimulation.

Yeast have only one PI 3-kinase homologue, Vps34, which regulates vesicle transport to the vacuole (Takegawa *et al*, 1995). When a strain lacking *vps34* was crossed with *ptn1Δ*, the authors found that levels of PtdIns(3,4,5)P<sub>3</sub> and PtdIns(3,4)P<sub>2</sub> decreased. Unlike class I PI 3-kinases, Vps34 can only phosphorylate PtdIns to generate PtdIns(3)P *in vitro*, indicating that it is not the 3-phosphorylation of PtdIns(4,5)P<sub>2</sub>, but the synthesis of PtdIns(3)P that is the important step in *S. pombe* PtdIns(3,4,5)P<sub>3</sub> synthesis. Mitra and colleagues then turned their attention to the kinases that might phosphorylate the 4- and 5-positions of the inositol head group. The lipid kinase Fab1 can phosphorylate PtdIns(3)P at the 5-position; however, its deletion did not affect the PtdIns(3,4)P<sub>2</sub> or PtdIns(3,4,5)P<sub>3</sub> production in *ptn1Δ* cells, indicating that there was no role for Fab1 in PtdIns(3,4,5)P<sub>3</sub> synthesis. Next, they examined Its3, the *S. pombe* homologue of the mammalian phosphatidylinositol-4-phosphate 5-kinase (PIP 5-K) family, which are key enzymes in the regulation of PtdIns(4,5)P<sub>2</sub> levels. When a strain retaining only 10% of wild-type PIP 5-K activity was crossed with *ptn1Δ*, levels of PtdIns(4,5)P<sub>2</sub> were lowered as expected. Surprisingly, PtdIns(3,4)P<sub>2</sub> and PtdIns(3,4,5)P<sub>3</sub> levels were also dramatically attenuated. As PtdIns(4,5)P<sub>2</sub> is not a substrate for Vps34, this suggests a novel role for Its3 in PtdIns(3,4,5)P<sub>3</sub> synthesis. (It is possible that yeast have a PtdIns(4,5)P<sub>2</sub> 3-kinase that is yet to be identified or that PtdIns(4,5)P<sub>2</sub> can allosterically regulate PtdIns(3,4,5)P<sub>3</sub> synthesis.) *In vitro*, mammalian PIP 5-Ks can sequentially phosphorylate PtdIns(3)P at the 4- and 5-positions to generate PtdIns(3,4,5)P<sub>3</sub> (Zhang *et al*, 1997), and this led Mitra and colleagues to propose that PtdIns(3)P generated by Vps34 is converted to PtdIns(3,4,5)P<sub>3</sub> by Its3 (Fig 1C).

What is the function of PtdIns(3,4,5)P<sub>3</sub> in yeast? Although the *ptn1Δ* strain displayed vacuolar defects and was more susceptible to osmotic stress than wild-type yeast, neither of these observations were directly linked to elevated PtdIns(3,4)P<sub>2</sub> and PtdIns(3,4,5)P<sub>3</sub> levels. Importantly, the authors demonstrated that at least two *S. pombe* proteins can bind PtdIns(3,4,5)P<sub>3</sub> *in vitro*, with the intracellular localization of one of them showing some dependency on the presence of Ptn1. This raises the possibility that PtdIns(3,4,5)P<sub>3</sub>-dependent signalling networks exist in yeast, analogous to those described in metazoan systems.

So, what are the consequences of these findings? The fact that the PIP 5-K-driven PtdIns(3,4,5)P<sub>3</sub> synthesis pathway is present in yeast indicates that, historically, this was the original pathway that organisms evolved to generate PtdIns(3,4,5)P<sub>3</sub>. Class I PI-3-kinases



**Fig 1** | Pathways of PtdIns(3,4,5)P<sub>3</sub> synthesis. **(A)** Phosphatidylinositol (PtdIns) is the basic building block of phosphorylated phosphoinositides (PIs). The inositol head group can be phosphorylated in the 3-, 4- and 5-positions to generate derivatives of PtdIns, which are listed on the right. **(B)** Schematic for receptor-driven PtdIns (3,4,5)-trisphosphate (PtdIns(3,4,5)P<sub>3</sub>) production. Agonist stimulation (tyrosine kinase or G-protein-coupled receptors) activates class I PI 3-kinase to induce phosphorylation of PtdIns(4,5)P<sub>2</sub> on the 3-position of the inositol group. **(C)** The proposed new pathway for PtdIns(3,4,5)P<sub>3</sub> production in *Schizosaccharomyces pombe*. Fission yeast enzymes are shown in blue. Their mammalian counterparts are depicted in turquoise. PIP 5-K, phosphatidylinositol-4-phosphate 5-kinase; PTEN, phosphatase and tensin homologue.

evolved much later, which suggests that the growth-factor-driven pathway found in *Caenorhabditis elegans* and mammalian cells is a 'new kid on the block'. If so, then what became of the PIP 5-K-driven PtdIns(3,4,5)P<sub>3</sub> pathway in mammalian cells? Intriguingly, whereas growth-factor-stimulated PtdIns(3,4,5)P<sub>3</sub> synthesis is dependent on class I PI 3-kinases in mammalian cells, we recently showed that certain stress agonists, such as H<sub>2</sub>O<sub>2</sub>, increase PtdIns(3,4,5)P<sub>3</sub> levels *in vivo* by the PIP 5-K-mediated 5-phosphorylation of PtdIns(3,4)P<sub>2</sub> (Halstead *et al*, 2001).

Finally, on a clinical note, PTEN (a tumour suppressor) deletion in human cancers leads to the activation of cell survival pathways through the upregulation of PtdIns(3,4,5)P<sub>3</sub> levels. In light of these new findings that connect PTEN with PIP 5-K-driven PtdIns(3,4,5)P<sub>3</sub> synthesis, it would be interesting to dissect how PtdIns(3,4,5)P<sub>3</sub> is generated in these cancer cells.

At the moment, this new, but ancient, biochemical pathway for PtdIns(3,4,5)P<sub>3</sub> synthesis seems to be an 'outsider' in the field of cell biology. Hopefully, with further study, it too can find a place of its own.

REFERENCES

Auger KR, Serunian LA, Soltoff SP, Libby P, Cantley LC (1989) PDGF-dependent tyrosine phosphorylation stimulates production of novel polyphosphoinositides in intact cells. *Cell* **57**: 167–175  
 Cantley LC (2002) The phosphoinositide 3-kinase pathway. *Science* **296**: 1655–1657  
 Halstead JR, Roefs M, Ellson CD, D'Andrea S, Chen C, D'Santos CS, Divecha N (2001) A novel pathway of cellular phosphatidylinositol(3,4,5)-trisphosphate synthesis is regulated by oxidative stress. *Curr Biol* **11**: 386–395  
 Hawkins PT, Jackson TR, Stephens LR (1992) Platelet-derived growth factor stimulates synthesis of PtdIns(3,4,5)P<sub>3</sub> by activating a PtdIns(4,5)P<sub>2</sub> 3-OH kinase. *Nature* **358**: 157–159  
 Lemmon MA (2003) Phosphoinositide recognition domains. *Traffic* **4**: 201–213

Maehama T, Dixon JE (1998) The tumor suppressor, PTEN/MMAC1, dephosphorylates the lipid second messenger, phosphatidylinositol 3,4,5-trisphosphate. *J Biol Chem* **273**: 13375–13378  
 Mitra P *et al* (2004) A novel phosphatidylinositol(3,4,5)P<sub>3</sub> pathway in fission yeast. *J Cell Biol* **166**: 205–211  
 Takegawa K, DeWald DB, Emr SD (1995) *Schizosaccharomyces pombe* Vps34p, a phosphatidylinositol-specific PI 3-kinase essential for normal cell growth and vacuole morphology. *J Cell Sci* **108**: 3745–3756  
 Traynor-Kaplan AE, Harris AL, Thompson BL, Taylor P, Sklar LA (1988) An inositol tetrakisphosphate-containing phospholipid in activated neutrophils. *Nature* **334**: 353–356  
 Zhang X, Loijens JC, Boronenkov IV, Parker GJ, Norris FA, Chan J, Thum O, Prestwich GD, Majerus PW, Anderson RA (1997) Phosphatidylinositol-4-phosphate 5-kinase isozymes catalyze the synthesis of 3-phosphate-containing phosphatidylinositol signaling molecules. *J Biol Chem* **272**: 17756–17761

**Nullin Divecha & Jonathan R. Halstead<sup>+</sup> are at the Netherlands Cancer Institute, Antoni van Leeuwenhoek ziekenhuis, Plesmanlaan 121, 1066 CX Amsterdam, the Netherlands**  
<sup>+</sup>Corresponding author.  
 Tel: +31 20 512 2009;  
 Fax: +31 20 512 1989;  
 E-mail: j.halstead@nki.nl



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