

Pathophysiology of Acute Exercise-Induced Muscular Injury: Clinical Implications

Phillip Page, MS, ATC, PT, LAT

Abstract: Acute muscular injury is the most common injury affecting athletes and those participating in exercise. Nearly everyone has experienced soreness after unaccustomed or intense exercise. Clinically, acute strains and delayed-onset muscle soreness are very similar. The purpose of this paper is to review the predisposing factors, mechanisms of injury, structural changes, and biochemical changes associated with these injuries. Laboratory and clinical findings are discussed to help athletic trainers differentiate between the two conditions and to provide a background knowledge for evaluation, prevention, and treatment of exercise-induced muscular injury.

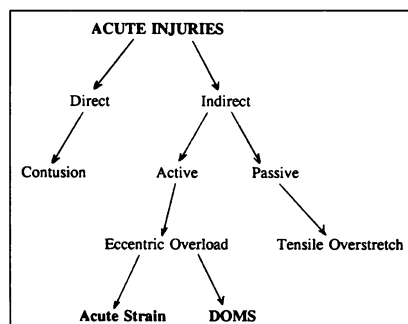
The musculotendinous unit is the force-generating component of functional movement. However, it is one of the least understood links in human movement and one of the most frequently injured tissues during exercise. Athletic trainers evaluate and treat musculoskeletal injuries daily. This paper provides a review of contemporary literature on the pathophysiology of acute strains and delayed-onset muscle soreness. Predisposing factors and mechanisms of injury are presented to aid the reader in the prevention of muscular injury, while structural and biochemical changes are reviewed for the purpose of evaluation and treatment. By understanding the pathophysiology of exercise-induced muscle injury, we may be better

able to prevent, evaluate, and treat these injuries.

Muscle Injury Classification

Musculoskeletal injuries are very common in both athletes and nonathletes. Although both acute and chronic injuries are seen, we will limit this review to acute muscular injuries. Acute injuries result from: 1) direct trauma, causing a contusion at the point of contact, or 2) indirect trauma, causing a disruption in the myofibers without contact. Indirect injuries may be passive or active. Passive injuries result from a tensile overstretch of the muscle without contraction, whereas active injuries usually result from eccentric overload of the muscles.^{28,32,46} These active injuries can be classified as acute strains or exercise-induced muscle soreness (see Figure).

Because most exercise-induced muscular injuries result from active contraction of muscles, our discussion will focus on injuries resulting from eccentric overload: acute strains and exercise-induced soreness. Exercise-induced muscular soreness or delayed-onset muscle soreness (DOMS) may be considered symptoms of injury. DOMS, however, limits performance resulting from decreased motion, decreased force production, and



Types of acute muscular injury.

pain^{2,14,15,26,48,51,58}; therefore, it is relevant to our discussion of acute exercise-induced muscular injury.

Both acute strains and DOMS usually occur as a result of eccentric overload and structural alteration^{26,54} and result in an inflammatory reaction.⁵⁶ Although each presents similar clinical symptoms and functional limitations, they should be differentiated to determine proper treatment. For example, acute strains may require rest, whereas DOMS may respond better to stretching and activity.

Clinically, acute muscle strain injuries can be distinguished from DOMS based on the history of the injury. A strain results from a specific episode, is painful, and is easily recognized by the patient as an injury.²⁸ DOMS, however, usually begins 12 to 48 hours postexercise, with no specific event causing the injury^{2,11,63} other than repeated eccentric muscle contraction. In both injuries, palpation, passive stretching, and active movement will cause pain.^{28,56} Classification and clinical findings of acute exercise-induced muscular injury are summarized in Table 1.

Acute Strain

Acute muscular strains result from macrotrauma with immediate and direct signs and symptoms.^{17,30} Most experimental studies, however, focus on acute muscular strain resulting from passive stretch in animal models. Taylor et al⁶¹ noted that both clinical and experimental strains result from large tensile forces that cause damage to the musculotendinous structure. In a review by Garret,²⁸ many laboratory studies were identified that were consistent with clinical findings of muscular injury.

Predisposing Factors

Garret²⁸ suggested three types of muscle that are at possible risk for injury:

Two-joint Muscles. Motion at one joint can place two-joint muscles in a position of increased passive tension. The increased passive tension may lead to an overstretch injury.

Muscles Contracting Eccentrically. Functional activities in exercise require both concentric and eccentric contractions. Muscular strains are considered to result most often from eccentric contrac-

Phillip Page is a staff physical therapist/athletic trainer at HealthSouth Sportsmedicine & Rehabilitation in Metairie, LA 70006.

Table 1.—Clinical Manifestations of Exercise-Induced Muscular Injury

Evaluation	Acute Strain	DOMS
History	One specific episode	Unaccustomed/intense exercise
Structures involved	Musculotendinous junction	Muscle and connective tissue
Muscle contractions	Eccentric	Eccentric
Pain	Immediate, localized	Delayed, diffuse, dull ache
Palpation	Point tenderness, \pm defect	Point tenderness, no defect
Swelling	\pm	\pm
Stretching	Painful	Painful
Weakness	Yes	Yes
Active movement	Painful	Painful

tions^{28,32,57,66} because higher specific tensions produced in eccentric exercise lead to myofiber overload injury.^{2,58,61} Eccentric contractions are common in the deceleration phase of activity.

Muscles With a Higher Percentage of Type II Fibers. These fast-twitch muscles create greater speed of contraction which may predispose a muscle to injury. Because most of the muscle action involved with running and sprinting is eccentric, muscle strains most often occur in sprinters or “speed athletes.”

Arnheim⁴ noted that the muscles with the highest incidence of strains in sports are the hamstrings, gastrocnemius, quadriceps, hip flexors, hip adductors, erector spinae, deltoid, and rotator cuff. In addition, he stated that a fault in the reciprocal coordination of the agonist and antagonist muscles may cause muscular strain. Knapić et al⁴³ reported that flexibility imbalances between agonists and antagonists may predispose athletes to injury. Interestingly, however, the flexible sides were most likely to be injured.

Previously injured muscle may also be more vulnerable to reinjury.³⁰ Jonhagen et al⁴¹ reported that sprinters with recent hamstring injuries had tighter and weaker hamstrings than uninjured sprinters, although it was not known if this finding was a cause or a result of injury. Inadequate rehabilitation may not restore full strength, flexibility, and endurance to the involved tissues before return to activity. Subsequently, muscle weakness, tightness, and fatigue may predispose muscle to injury.^{24,28}

Stretching and warmup before exercise have been advocated to prevent muscular injury^{6,60,64} on the assumption

that cold or tight muscles²³ might predispose one to muscular strain. Few experimental studies exist supporting these claims. However, inadequate warm-up exercises have been shown to be associated with muscle strains.³⁷ Increased tissue temperatures of 1°C⁵³ and 4°C⁶⁰ allowed greater elongation of the muscle before failure. Similarly, Noonan et al⁵⁰ reported that warm muscles (40°C) were less stiff than cold (25°C) muscles and required greater length to fail, offering experimental data to support warm-up as an aid in injury prevention and enhanced performance.

Mechanism of Injury

Acute muscular strains usually result from a specific event of macrotrauma.^{17,30} A strain may involve the muscle itself or adjacent tissue, such as fascia or tendon.⁴ The severity of the injury is in direct relation to the forces placed upon the muscle.⁴⁵ Mechanically, an acute strain may be caused by an abnormal muscular contraction,⁴ a response to high specific tensions,² or forcible stretching of a muscle while it is active.²⁸ Each of these mechanisms is related to eccentric contraction, in which sufficient tensile forces develop to cause irreversible changes in the structure of the musculotendinous unit.⁶¹

Prevention of Acute Strains

Prevention of acute muscular strains should begin with adequate preseason screening of flexibility and strength balances in major joints such as the knee, shoulder, and ankle. Evaluation of previous muscle injuries should be performed to assess flexibility,

strength, endurance, and proprioception. Preseason and in-season conditioning of muscle groups is also vital for prevention. Adequate agonist/antagonist ratios for strength and flexibility should be attained for major muscle groups, including quadriceps/hamstrings, shoulder internal rotators/external rotators, ankle dorsiflexors/plantarflexors, and abdominals/paraspinals. Muscles must be strengthened in the mode in which they are used functionally; ie, eccentric muscles should be strengthened eccentrically.

Warmup and stretching before activity are thought to prevent muscular strains. Smith⁵⁵ offers an excellent review of the literature supporting the efficacy of this practice. Active warmup such as jogging or biking should be performed before specific muscle stretching with emphasis on muscles at risk for strain, including two-joint muscles and those with high percentages of fast-twitch fibers (hamstrings, gastrocnemius, quadriceps, biceps), and muscles with high incidence of strain (hip flexors, hip adductors, erector spinae, rotator cuff). Muscles which contract eccentrically or decelerate in functional high-speed activities, such as the posterior rotator cuff in throwing athletes or the hamstrings in sprinters should be stretched for 15 to 20 seconds^{6,8,62} and repeated four times.⁶² Static stretching in combination with passive heating may be more effective than passive heating alone.³⁸

Structural Changes

The most vulnerable site for an indirect strain injury is just distal to the musculotendinous junction or the tendon-bone junction^{5,28-31,61}; therefore, knowledge of musculoskeletal surface anatomy and palpation skill is important. Pain, swelling, deformity, and point tenderness may be localized over the musculotendinous junction immediately after injury. Tendons themselves are very resilient to injury and are rarely injured acutely. Tendons will pull away from a bone, a bone will break, or a muscle will tear before tendons are injured.⁴ However, in contrast to the musculotendinous junction, tendons are more susceptible to microtrauma.³⁰

A muscle strain may be partial or complete, depending on the amount and degree of fiber disruption within the muscle.²⁸ Strains can be classified by three degrees: first degree, a minute separation of muscle fibers; second degree, partial tearing of some fibers; and third degree, a complete rupture or tendinous avulsion. While full-thickness tears do occur, partial or incomplete tears or strains are more common³⁰ and are clinically characterized by focal pain and swelling.²⁸ A palpable defect such as a bulge or gap in the muscle may be found in a complete or partial tear. Disruptions in the fibers cause biochemical changes both from direct injury to the fibers and from the inflammatory reaction.

Biochemical Changes

Serum creatine kinase (CK) and lactate dehydrogenase (LDH) enzyme levels are used to indirectly assess muscle damage following eccentric exercise.^{1,2,12,14-16,26,47,48,54,58} In addition to the release of these biochemical markers, all acute strains undergo similar inflammatory reactions. Acute inflammation is the fundamental reaction designed to protect, localize, and remove injurious agents from the body in preparation for healing and repair.⁴ Chemical mediators are present in acute muscular strain, as with any inflammation. These include: histamine, serotonin, bradykinin, and prostaglandin.⁴ These chemicals increase the capillary membrane permeability, change blood vessel diameter, and stimulate pain receptors. Edema results from an accumulation of proteins and transudate in the interstitial space. This accumulation is a result of increased capillary membrane permeability secondary to the chemical mediators. Therefore, the characteristic swelling, heat, redness, and pain of inflammation are due to biochemical changes⁴ caused by the chemical mediators. Nikolaou et al⁴⁹ noted an inflammatory reaction and edema at 1 to 2 days after a stretch-induced muscular injury. The acute phase of inflammation lasts up to 3 to 4 days after the initial injury⁴ unless the tissue continues to be traumatized, as is commonly seen when injured athletes return to activity too soon or are progressed too rapidly. Proliferation of fibroblasts, increased collagen production, and degradation of ma-

ture collagen have an overall weakening effect on the tissue; therefore, efforts to stretch the tissue perpetuate the irritation and progressive limitation,⁴² leading to chronic muscle strains.

As the inflammatory phase subsides, repair begins and lasts for 2 to 3 weeks. The repair phase is characterized by capillary growth and fibroblast activity to form immature collagen. This immature collagen is easily injured if overstressed; however, proper growth and alignment can be stimulated with appropriate tensile loading in the line of normal stresses.⁴² The final stage of healing is maturation and remodeling of collagen, occurring from 2 to 3 weeks after onset until there is pain-free functional use of the muscle.⁴² If the healing fibers are not properly stressed, the fibers adhere to surrounding tissue and form a restricting scar which is resilient to remodeling.

Treatment of Acute Strains

The key to returning any injured athlete to competition safely is to provide an optimal environment for healing and to progress the patient according to: 1) the severity of the injury, 2) the natural healing process of the body, and 3) the response of the tissue to new demands. Treatment goals for any soft-tissue injury must take the natural healing process into consideration. The overall goal is to assist the body with its natural healing process. The body must go through each stage with any soft tissue injury; therefore, the athletic trainer must not return the athlete to activity too soon. Two to three weeks of restricted activity may be a minimum to allow for collagen formation and prevent reinjury in all soft-tissue injuries.

Inflammatory Phase. Effective management of muscular strains begins at the time of injury. Care for acute injuries must be initiated as quickly as possible with rest, ice, compression, and elevation (RICE) for at least 48 hours. Cold slows the inflammatory process and decreases pain and muscle spasm; compression and elevation reduce edema. An ice bag wrapped with an elastic wrap, elevation, and crutches may be used. Rest protects the injured tissue; however, immobilization may be detrimental to healing and the uninjured tissue. As the inflammation subsides, pain-free, passive range of mo-

tion (ROM) and gentle joint mobilization should be initiated to maintain soft-tissue and joint integrity. Gentle, pain-free, submaximal isometric muscle sets may be used at multiple angles to maintain strength and keep the developing scar tissue mobile. Aggressive stretching and strengthening is contraindicated. Any increase in pain, swelling, warmth, or redness indicates a proliferation of the inflammatory phase which should be treated only with RICE. Modalities for pain and edema such as electrical stimulation and pulsed ultrasound should be used during both the inflammatory and repair phases.

Repair Phase. The inflammatory and repair phases overlap during the first week after injury. As the inflammation subsides, the athlete may attempt too much activity too soon. This prolongs the inflammatory phase and leads to chronic muscle strain. However, as collagen is laid down, it must be appropriately stressed in the normal lines of tension. This is a critical "turning point" in the treatment of muscle injuries and may be the most important stage for the trainer or therapist in any rehabilitation. Signs of inflammation (pain, swelling, redness, warmth) are used to determine whether the tissues are being overstressed with activity. The rehabilitation program must be constantly evaluated. Frequency, intensity, and duration of exercises are altered to allow for healing and to prevent inflammation for the next 1 to 2 weeks. Cold in the form of cryostretch or cryokinetics (see Ref. 44) may be beneficial initially to allow for pain-free exercise to aid in the formation of the scar tissue. Heat in the form of warm whirlpools, moist heat, or ultrasound is used to promote capillary growth and increase ROM. Contrast baths may be most beneficial during this period. Gentle, pain-free stretching and pain-free submaximal isometrics can be incorporated into contract-relax techniques to help align collagen fibers. These exercises are progressed to active ROM for the agonist. Active or resistive motion of the antagonist or contralateral extremity may also be incorporated. Finally, a cardiovascular conditioning program should be incorporated for any athlete not capable of full athletic participation.

Maturation and Remodeling Phase.

As collagen matures, it requires tension in the line of normal stresses to remodel properly. Clinically, this stage presents at about 2 to 3 weeks after injury and is characterized by: 1) the absence of inflammation, 2) full, pain-free ROM, and 3) pain after tissue resistance, which is felt with passive ROM.⁴² The athlete is progressed as tolerated with limited participation in his/her sport. Rehabilitation includes more vigorous stretching, closed- and open-chain strengthening, cardiovascular training, and sport-specific activities. It is vital to remember that muscles must be stressed and overloaded in the manner in which they are used functionally, following the principle of specificity. Type of contraction (eccentric vs concentric), metabolism (aerobic vs anaerobic), and functional pattern (diagonal vs cardinal plane) should be specific to the activity in which they are used. Eccentric exercise is functional in most athletic activities, develops greater tension than concentric exercise, and may be more comfortable in the early stages of rehabilitation.⁷ Eccentric contraction may be performed against manual, isotonic, isokinetic, or elastic resistance. Massage, aquatic therapy, and plyometrics are beneficial to any soft-tissue treatment. Proprioceptive and endurance training are used in the advanced stages of rehabilitation. Modalities before and after activity may be beneficial as well. An elastic or neoprene wrap with a felt pad directly over the injury site provides warmth and compression. After the athlete has regained full, pain-free active ROM and over 90% strength bilaterally, full participation is allowed. Maintenance programs should be independent and individualized to avoid any dysfunctional adaptation or compensation.

Delayed-Onset Muscle Soreness

DOMS is commonly seen in patients performing new exercises or in athletes involved in weight-lifting or other eccentric activities. DOMS results from muscle damage^{2,11,15,26,58} following eccentric exercise. The onset of DOMS is characterized as a dull, aching pain usually beginning 12 to 48 hours after exer-

cise.^{2,11,63} Clarkson et al¹⁵ found that soreness peaks 2 to 3 days following eccentric exercise and subsides linearly within 10 days. In addition to pain, other symptoms include decreased motion and decreased force production.^{2,14,15,26,48,51,58,65} Newham and associates,⁴⁷ however, reported a decreased force production with electrical stimulation of these muscles, indicating that the soreness itself does not inhibit force production.

Muscles adapt to a single bout of eccentric exercise. This is evidenced by less damage to the muscle after the same exercise months later.¹⁵ The muscle is repaired without any residual dysfunction or scarring and the muscle is often able to resist even greater forces.^{12,16,61}

Predisposing Factors

Armstrong¹ suggested two possible causative factors during the initial events of DOMS^{2,58}: high tensions and metabolic changes.

High Tensions. High tensions produced during eccentric exercise are more apt to produce myofiber injury than isometric or concentric contractions.

Metabolic Changes. Increased temperature, decreased aerobic capacity, and decreased pH of the muscle may have a role in causing DOMS.

Mechanism of Injury

Exercise that results in the development of soreness is associated with the rapid destruction of muscle tissue.^{11,26} The soreness usually results from muscle damage following repetitive eccentric exercise or after the first or second session of a new training program.^{2,11,15,22,58} DOMS is also associated with muscle spasm, as evidenced by increased EMG activity.^{20,21,46,52} DOMS results primarily from structural muscle damage and microtrauma and may be related to the resultant biochemical changes of the inflammatory process.

Prevention of DOMS

Few studies exist on the prevention of DOMS. Because it is known that eccentric contractions cause muscle damage, DOMS may be prevented with warmup and stretching before and after eccentric or novel exercise. Ideally, DOMS is prevented by avoiding eccentric or unaccus-

tomized exercise. Clinically, this is not possible. Athletes in competition, weight lifters, and patients undergoing early rehabilitation commonly perform eccentric exercise. This eccentric training develops strength to resist further damage.¹⁵ Therefore, specific eccentric training is necessary for any sport activity to help prevent further damage or injury. Eccentric training sessions should be limited to two per week to allow adequate rest and recovery between sessions. Patients should be educated about the importance and need for eccentric exercises, as well as about the possibility of DOMS.

Effective prevention of DOMS may begin in the acute stages of treatment before symptoms begin. Prophylactic ibuprofen administered before or immediately after heavy eccentric exercise may decrease the pain associated with DOMS.³⁴ Yackzan et al⁶⁵ found that subjects who received ice massage immediately after eccentric exercise had more ROM 24 hours later than those who did not receive it. Further research is needed on the prevention of DOMS, including the roles of warmup, stretching, and immediate treatment after intense eccentric exercise.

Structural Changes

Structural damage to subcellular components following eccentric exercise has been found by microscopic evaluation.^{1,3,11,26,40} High specific tensions seen in eccentric contractions could mechanically disrupt the connective tissue, myofilaments, sarcomere, sarcolemma, or sarcoplasmic reticulum.^{1,2,9,10} Friden et al^{25,26} found alteration of Z-bands in type II fibers both immediately and 3 days after eccentric exercise.

Damage to the extracellular matrix (ECM, the interface between the myofiber and fascia) following eccentric exercise has been evaluated by Stauber and associates.^{58,59} Myofiber and ECM damage result directly from eccentric contractions.²⁷ Stauber et al⁵⁹ reported that eccentric muscle action is related to mechanical shearing at the ECM. These structural changes then cause biochemical changes within the injured tissue.

Biochemical Changes

Damage to the sarcolemma and ECM creates an altered chemical environment

Table 2.—Treatment of DOMS

Authors	Treatment	Efficacy
Ciccone et al ¹³	Trolamine salicylate phonophoresis	Effective
Denegar et al ¹⁸	TENS	Effective
DeVries ^{20,21}	Static stretching	Effective
Hasson et al ³³	High speed, voluntary muscle contraction	Effective
Hasson et al ³⁴	Ibuprofen	Effective
Haynes & Perrin ³⁶	Topical counterirritant	Effective
Hill & Richardson ³⁹	Topical trolamine salicylate cream	Effective
Prentice ⁵²	Static or PNF stretching + cold or heat	Effective
McGlynn et al ⁴⁶	Stretching and biofeedback	Reduced EMG, but not pain
Prentice ⁵²	Heat	Not effective alone
Yaczan et al ⁶⁵	Ice massage	Not effective alone
	Continuous ultrasound	Increases pain
Denegar et al ¹⁹	Microcurrent	Not effective, + analgesic effect
Hasson et al ³⁵	Dexamethasone Iontophoresis	Questionable

within the muscle. The release of proteins and ions into the plasma as a result of inflammation is similar to that found in acute strains.^{1,2,59} Increases in these levels indicate damage to the sarcolemma. Elevations of CK, LDH, protein metabolites, and myoglobin have been found in plasma up to 48 hours following eccentric exercise.^{2,3,14,54,58,63} These biochemical events occur within the muscle cells themselves and begin approximately 24 hours postexercise,¹⁵ before phagocytic cells enter the injury site.¹ Time-specific clinical events (such as peak soreness at 2 to 3 days) may correspond to the time of increased enzyme levels (such as CK increase at 2 days). While Tiidus⁶³ reported such a correlation between soreness and enzyme levels, Clarkson et al¹⁵ cautioned against claiming a cause-and-effect relationship based on limited research.

The structural disruption leads to the normal inflammatory response: an increase in chemical mediators such as histamine, bradykinin, prostaglandin, and serotonin,⁴ causing pain and swelling. The products of the inflammatory response sensitize free nerve endings in muscle,^{11,56} thus increasing soreness. Stauber et al⁵⁹ concluded that the DOMS after repeated eccentric muscle action is not because of actual myofiber damage, but more likely results from inflammation.

Treatment of DOMS

Because DOMS results from microtrauma, structural damage in DOMS is

not as severe as in acute strains resulting from macrotrauma. The symptoms of DOMS resolve relatively quickly without any residual dysfunction; therefore, DOMS can be treated symptomatically. In any exercise-induced muscular injury, RICE is the ideal immediate treatment to decrease inflammation and pain. However, because DOMS begins at 24 to 48 hours after exercise and peaks at 2 to 3 days after exercise, treatment may not begin immediately after injury.

The goal of treatment of DOMS is to reduce the pain, swelling, inflammation, and muscle spasm. These goals are similar to those in the acute stage of any soft-tissue injury. Several authors have studied the efficacy of these treatments (Table 2). Static or proprioceptive neuromuscular facilitation (PNF) stretching,^{20,21,52} high-speed muscular concentrics,³³ nonsteroidal anti-inflammatory drugs (NSAIDs),³⁴ and topical counterirritants^{36,39} have been shown to be effective in reducing pain associated with DOMS. Phonophoresis¹³ and transcutaneous electrical nerve stimulation (TENS)¹⁸ may be effective, but the benefits of iontophoresis³⁵ and biofeedback⁴⁶ remain questionable.

Most studies report significant improvement in DOMS with combinations of exercise and modalities. DOMS can be treated symptomatically by reducing the pain, soreness, swelling, and muscle spasm. NSAIDs and topical counterirritants may decrease soreness. Static or PNF stretching in combination with cryotherapy (spray-and-stretch or ice

massage) also help to decrease the symptoms of DOMS. In addition, high-speed, rapid concentric muscular contractions may provide relief. Further research is needed on the use of massage, pulsed ultrasound, and electrical stimulation (including TENS, iontophoresis, and microcurrent) in the treatment of DOMS.

Conclusion

Both acute strains and DOMS present similar clinical signs; however, they can be differentiated by history of the injury. While many studies exist on the structural changes and biochemical changes of exercise-induced muscular injury, many questions remain unanswered. The exact changes in human muscle after an acute strain have not been determined. A cause-and-effect relationship for DOMS has not been firmly established. We have reviewed the literature on these acute injuries and provided clinical findings to aid in the care of musculoskeletal injuries. Further research is needed on the causes of these injuries, as well as on effective preventive and treatment techniques to return athletes and patients back to preinjury levels.

References

1. Armstrong RB. Initial events in exercise-induced muscular injury. *Med Sci Sports Exerc.* 1990;22:429-435.
2. Armstrong RB. Mechanisms of exercise-induced delayed onset muscular soreness: a brief review. *Med Sci Sports Exerc.* 1984;16:529-538.
3. Armstrong RB, Ogilvie RW, Schwane JA. Eccentric exercise-induced injury to rat skeletal muscle. *J Appl Physiol.* 1983;54:90-93.
4. Arnheim DD. *Modern Principles of Athletic Train-*

- ing. 7th ed. St Louis, MO: Times Mirror/Mosby; 1989:198-231.
5. Bach BR, Warren RF, Wickiewicz TL. Triceps rupture: a case report and literature review. *Am J Sports Med.* 1987;15:285-289.
 6. Beaulieu JE. Developing a stretching program. *Phys Sportsmed.* Nov 1981;9:59-65.
 7. Bennet JG, Stauber WT. Evaluation and treatment of anterior knee pain using eccentric exercise. *Med Sci Sports Exerc.* 1986;18:526-530.
 8. Bohannon RW. Effect of repeated eight-minute loading on the angle of straight-leg raising. *Phys Ther.* 1984;64:491-497.
 9. Byrd SK. Alterations in the sarcoplasmic reticulum: a possible link to exercise-induced muscle damage. *Med Sci Sports Exerc.* 1992;24:531-536.
 10. Byrd SK, McCutcheon LJ, Hodgson DR, Gollnick PD. Altered sarcoplasmic reticulum function after high-intensity exercise. *J Appl Physiol.* 1989;67:2072-2077.
 11. Byrnes WC, Clarkson PM. Delayed onset muscle soreness and training. *Clin Sports Med.* 1986;5:605-614.
 12. Byrnes WC, Clarkson PM, White JS, Hsieh SS, Frykman PN, Maughan RJ. Delayed onset muscle soreness following repeated bouts of downhill running. *J Appl Physiol.* 1985;59:710-715.
 13. Ciccone CD, Leggin BG, Callamaro JJ. Effects of ultrasound and trolamine salicylate phonophoresis on delayed-onset muscle soreness. *Phys Ther.* 1991;71:666-678.
 14. Clarkson PM, Byrnes WC, McCormick KM, Turcotte LP, White JS. Muscle soreness and serum creatine kinase activity following isometric, eccentric, and concentric exercise. *Int J Sports Med.* 1986;7:152-155.
 15. Clarkson PM, Nosaka K, Braun B. Muscle function after exercise-induced muscle damage and rapid adaptation. *Med Sci Sports Exerc.* 1992;24:512-520.
 16. Clarkson PM, Tremblay I. Rapid adaptation to exercise induced muscle damage. *J Appl Physiol.* 1988;65:1-6.
 17. Davies GJ, Wallace LA, Malone TR. Mechanisms of selected knee injuries. *Phys Ther.* 1980;60:1590-1596.
 18. Denegar CR, Perrin DH, Rogol AD, Rutt R. Influence of transcutaneous electrical nerve stimulation on pain, range of motion, and serum cortisol concentration in females experiencing delayed onset muscle soreness. *J Orthop Sports Phys Ther.* 1989;11:100-103.
 19. Denegar CR, Yoho AP, Borowicz AJ, Bifulco N. The effects of low-volt microamperage stimulation on delayed onset muscle soreness. *J Sport Rehabil.* 1992;1:95-102.
 20. DeVries H. Quantitative electromyographic investigation of the spasm theory of muscle pain. *Am J Phys Med.* 1966;45:119-135.
 21. DeVries H. Prevention of muscle distress after exercise. *Res Q.* 1961;32:177-185.
 22. Ebbeling C, Clarkson PM. Exercise-induced muscle damage and adaptation. *Sports Med.* 1989;7:210-226.
 23. Ekstrand J, Gillquist J. The frequency of muscle tightness and injury in soccer players. *Am J Sports Med.* 1982;10:75-78.
 24. Evans WJ, Cannon JG. The metabolic effects of exercise-induced muscle damage. *Exerc Sports Sci Rev.* 1991;19:99-125.
 25. Friden J, Lieber RL. Structural and mechanical basis of exercise-induced muscle injury. *Med Sci Sports Exerc.* 1992;24:521-530.
 26. Friden J, Sjostrom J, Ekblom B. Myofibrillar damage following intense eccentric exercise in man. *Int J Sports Med.* 1983;4:170-176.
 27. Fritz VK, Stauber WT. Characterization of muscles injured by forced lengthening: II. Proteoglycans. *Med Sci Sports Exerc.* 1988;20:354-361.
 28. Garret WE. Muscle strain injuries: clinical and basic aspects. *Med Sci Sports Exerc.* 1990;22:436-443.
 29. Garret WE. Injuries to the muscle-tendon unit. *Instr Course Lect.* 1988;37:275-282.
 30. Garret WE, Duncan PW, Malone TR. Muscle injury and rehabilitation. *Sports Inj Manage.* 1988;1:1-42.
 31. Garret WE, Rich FR, Nikolaou PK, Vogler JB. Computed tomography of hamstring muscle strains. *Med Sci Sports Exerc.* 1989;21:506-514.
 32. Glick JM. Muscle strains. Prevention and treatment. *Phys Sportsmed.* Nov 1980;8:73-77.
 33. Hasson S, Barnes W, Hunter M, Williams J. Therapeutic effect of high speed voluntary muscle contractions on muscle soreness and muscle performance. *J Orthop Sports Phys Ther.* 1989;11:499-507.
 34. Hasson SM, Daniels JC, Divine JG, et al. Effect of ibuprofen use on muscle soreness, damage, and performance: a preliminary investigation. *Med Sci Sports Exerc.* 1993;25:9-17.
 35. Hasson SM, Wible CL, Reich M, Barnes WS, Williams JH. Dexamethasone iontophoresis: effect on delayed onset muscle soreness and muscle function. *Can J Sport Sci.* 1992;17:8-13.
 36. Haynes SC, Perrin DH. Effect of a counterirritant on pain and restricted range of motion associated with delayed onset muscle soreness. *J Sport Rehabil.* 1992;1:113-118.
 37. Heiser TM, Wever J, Sullivan G, Clare P, Jacobs RR. Prophylaxis and management of hamstring injuries in intercollegiate football players. *Am J Sports Med.* 1984;12:368-370.
 38. Henricson AS, Fredriksson K, Persson I, Pereira R, Rostedt Y, Westlin NE. The effect of heat and stretching on range of hip motion. *J Orthop Sports Phys Ther.* 1984;6:110-115.
 39. Hill DW, Richardson JD. Effectiveness of 10% trolamine salicylate cream on muscular soreness induced by a reproducible program of weight training. *J Orthop Sports Phys Ther.* 1989;11:19-23.
 40. Hoppeler H. Exercise-induced ultrastructural changes in skeletal muscle. *Int J Sports Med.* 1986;7:76-92.
 41. Jonhagen S, Nemeth G, Eriksson E. Hamstring injuries in sprinters. The role of concentric and eccentric hamstring muscle strength and flexibility. *Am J Sports Med.* 1994;22:262-266.
 42. Kisner C, Colby LA. *Therapeutic Exercise. Foundations and Techniques.* 2nd ed. Philadelphia, PA: FA Davis; 1990:211-227.
 43. Knapic JJ, Bauman CL, Jones CL, Jones BH, Harris JM, Vaughan L. Preseason strength and flexibility imbalances associated with athletic injuries in female collegiate athletes. *Am J Sports Med.* 1991;19:76-81.
 44. Knight KL. *Cryotherapy: Theory, Technique, Physiology.* Chattanooga, TN: Human Kinetics; 1985.
 45. McCully KK, Faulkner JA. Characteristics of lengthening contractions associated with injury to skeletal muscle fibers. *J Appl Physiol.* 1986;61:293-299.
 46. McGlynn GH, Laughlin NT, Rowe V. The effect of electromyographic feedback and static stretching on artificially induced muscle soreness. *Am J Phys Med.* 1979;58:139-148.
 47. Newham DJ, Clarkson PM. Repeated high force eccentric exercise: effects on muscle pain and damage. *J Appl Physiol.* 1987;63:1381-1386.
 48. Newham DJ, McPhail G, Mills DR, Edwards RHT. Ultrastructural changes after concentric and eccentric contractions of human muscle. *J Neurol Sci.* 1983;61:109-122.
 49. Nikolaou PK, Macdonald BL, Glisson RR, Seaber AV, Garret WE. Biochemical and histological evaluation of muscle after controlled strain injury. *Am J Sports Med.* 1987;15:9-14.
 50. Noonan JT, Best TM, Seaber AV, Garret WE. Thermal effects on skeletal muscle tensile behavior. *Am J Sports Med.* 1993;21:517-522.
 51. Ogilvie RW, Hoppeler H, Armstrong RB. Decreased muscle function following eccentric exercise in the rat. *Med Sci Sports Exerc.* 1983;15:17-195.
 52. Prentice WE. An electromyographic analysis of the effectiveness of heat or cold and stretching for inducing relaxation in injured muscles. *J Orthop Sports Phys Ther.* 1982;3:133-140.
 53. Safran MR, Garret WE, Seaber AV, Glisson RR, Ribbeck BM. The role of warmup in muscular injury prevention. *Am J Sports Med.* 1988;16:123-129.
 54. Schwane JA, Johnson SR, Vandenalker CB, Armstrong RB. Delayed-onset muscular soreness and plasma CPK and LDH activities after downhill running. *Med Sci Sports Exerc.* 1983;15:51-56.
 55. Smith CA. The warm-up procedure: to stretch or not to stretch. A brief review. *J Orthop Sports Phys Ther.* 1994;19:12-17.
 56. Smith LL. Acute inflammation: the underlying mechanism in delayed onset muscle soreness? *Med Sci Sports Exerc.* 1991;23:542-551.
 57. Stanton P, Purdam C. Hamstring injuries in sprinting: the role of eccentric exercise. *J Orthop Sports Phys Ther.* 1989;3:343-349.
 58. Stauber WT. Eccentric action of muscles: physiology, injury, and adaptation. *Exerc Sports Sci Rev.* 1989;17:157-185.
 59. Stauber WT, Clarkson PM, Fritz VK, Evans WJ. Extracellular matrix disruption, stiffness and pain following eccentric muscle action. *J Appl Physiol.* 1991;69:868-874.
 60. Strickler T, Malone T, Garrett WE. The effects of passive warming on muscle injury. *Am J Sports Med.* 1990;18:141-145.
 61. Taylor DC, Dalton JD, Seaber AV, Garrett WE. Experimental muscle strain injury: early functional and structural deficits and the increased risk for reinjury. *Am J Sports Med.* 1993;21:190-194.
 62. Taylor DC, Dalton JD, Seaber AV, Garrett WE. Viscoelastic properties of muscle-tendon units: the biomechanics of stretching. *Am J Sports Med.* 1990;18:300-309.
 63. Tiidus PM, Januzzo CD. Effects of intensity and duration of muscular exercise on delayed soreness and serum enzyme activities. *Med Sci Sports Exerc.* 1983;15:461-465.
 64. Wiktorsson-Moller M, Oberg B, Ekstrand J, Gillquist J. Effects of warming up, massage, and stretching on range of motion and muscle strength in the lower extremity. *Am J Sports Med.* 1983;11:249-252.
 65. Yackzan L, Adams C Francis KT. The effects of ice massage on delayed onset soreness. *Am J Sports Med.* 1984;12:159-165.
 66. Zairs B, Ciullo JV. Acute muscle and tendon injuries in athletes. *Clin Sports Med.* 1983;2:167-182.