#### **CURRENT REVIEW • ACTUALITÉS**

# Would decreased aluminum ingestion reduce the incidence of Alzheimer's disease?

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Although the cause of Alzheimer's disease (AD) remains unknown there is mounting evidence that implicates aluminum as a toxic environmental factor of considerable importance. Four independent lines of evidence — laboratory studies of the effects of intracerebral aluminum on the cognitive and memory performance of animals, biochemical studies, epidemiologic studies and the slowing of the progress of the disease with the use of an agent that removes aluminum from the body — now support the concept that aluminum is one of the pathogenic factors in AD. The evidence warrants serious consideration of reducing human exposure to aluminum. We hypothesize that a public health effort to restrict human ingestion of aluminum would reduce the incidence of this common chronic illness in the elderly.

Même si l'on ne connaît toujours pas la cause de la maladie d'Alzheimer (MA), de plus en plus d'indices semblent démontrer que l'aluminium est un facteur environnemental toxique très important. Quatre niveaux de preuve indépendants — études de laboratoire au sujet des effets de l'aluminium intracérébral sur la cognition et la mémoire chez les animaux, études biochimiques, études épidémiologiques et ralentissement du progrès de la maladie lorsqu'on utilise un agent qui élimine l'aluminium du corps — appuient maintenant le concept selon lequel l'aluminium est un des facteurs pathogènes de la MA. Ces preuves justifient d'envisager sérieusement de réduire l'exposition des êtres humains à l'aluminium. Nous posons l'hypothèse suivante: une campagne d'hygiène publique visant à limiter l'ingestion d'aluminium par les êtres humains réduirait l'incidence de cette maladie chronique répandue chez les personnes âgées.

he cause of Alzheimer's disease (AD) remains unknown, but both genetic and environmental factors appear to be important. Mutation on chromosome  $21^{1,2}$  may account for a small number of families with familial AD; recently, two families of patients with AD were found to have a point mutation in the amyloid precursor protein, a gene on chromosome 21.3 This mutation causes an amino acid substitution of isoleucine for valine in the transmembrane domain of the  $\beta$ -amyloid precursor protein at a site just two amino acids from the

 $\beta$ -amyloid peptide. It is still uncertain whether this point mutation "causes" AD or whether the mutation is closely linked to the yet-to-be-discovered gene responsible for AD on chromosome 21. For most families, however, linkage to chromosome 21 has not been established, and the genetic form of the disease appears to involve mutations on more than one chromosome.

The much more common, sporadic form of AD appears to be related to unidentified environmental factors. Even in genetically identical twins nongenet-

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ic factors appear to be important; in one study the disease developed in only 40% of twin pairs.<sup>4</sup> Although a virus or scrapie-like infectious agent has been proposed as a pathogen AD is not transmissible to other humans or laboratory animals, and neither viral DNA nor viral antigens have been reproducibly found in AD-affected tissues. Immunologic mechanisms have also been suggested as factors, but whether the observed changes are cause or effect is uncertain.<sup>5</sup>

Aluminum, a widely recognized neurotoxin, has been found in increased concentrations in all AD-affected tissues examined by means of methods sufficiently sensitive and appropriately applied.<sup>6</sup>

However satisfying the idea, we may not be justified in searching for a single cause. A direct experimental approach to measuring the importance of aluminum, or any other postulated etiologic or pathogenic factor, is not possible because AD is found only in humans and cannot be replicated exactly in laboratory preparations. Aluminum induces neither the paired helical filament configuration found in Alzheimer-type neurofibrillary tangles nor the formation of senile plaques with amyloid cores. Nevertheless, it has been found in at least four sites in AD-affected brains and induces at least three cellular derangements also found in AD. However, no single observation or experimental result is, in itself, conclusive.

Four independent lines of evidence implicate aluminum's role in AD: (a) toxicologic studies and laboratory observations of the learning and memory performance of animals, (b) a large number of documented biochemical changes at concentrations of aluminum similar to those found in various subcellular compartments in AD-affected human brains, (c) epidemiologic evidence of the increased incidence of AD in relation to exposure to aluminum in drinking water and (d) the slowing of the clinical progression of AD by a drug that selectively removes aluminum from the body.

# Effect of aluminum on learning and memory performance of animals

The direct intracranial injection of small but lethal amounts of soluble aluminum salts produces delayed memory and learning impairment in sensitive species such as cats and rabbits. 7-9 A dose of intracranially injected aluminum sufficient to raise the content in grey matter about fourfold (from 1.5  $\mu$ g/g dry weight to the 50% lethal concentration of about 5.5  $\mu$ g/g) had no immediate effect on memory and motor tasks in trained cats; however, 7 to 10 days after injection the cats began to exhibit progressive impairment in precise motor control during jumping 10 and alterations in the performance of

learning and memory tasks.<sup>7</sup> Initially the learning and memory deficits were selective: the performance of visual discrimination tasks was not affected and the speed of motor response not altered. The selective effect of aluminum on the learning and memory system is element specific, because nine other trivalent toxic metals (boron, chromium, gallium, indium, lanthanum, scandium, thallium, vanadium and yttrium) have been shown not to elicit the sequence of clinical signs that follow aluminum administration.<sup>11</sup>

Ten days after aluminum injection rabbits have exhibited deficits in learning (acquisition of an active avoidance task) and failed to retain the task when tested 3 days later.<sup>12</sup> Impaired learning in a water maze,<sup>13</sup> defective classic<sup>14</sup> and eye-blink conditioning<sup>15,16</sup> and impaired long-term potentiation in hippocampus slices<sup>17</sup> (considered to be an electrophysiologic model of a learning response) have also been reported in rabbits given aluminum.

Shortly after the memory defects appear, cats and rabbits exhibit progressive deterioration in motor control, with difficulty executing body righting, increased muscle tone, ataxia of gait, tremors, myoclonic jerks and seizures. If the seizures are not treated with anticonvulsant medications the animals may die in convulsions; treated animals may survive, but with persistent severe neurologic and behavioural defects.

The unique, progressive clinical course after a single intracerebral lethal threshold dose of aluminum in susceptible mammals is marked first by disturbance of learning and memory and then by altered motor control, increased muscle tone, myoclonic jerks and seizures. It resembles the clinical course of AD in humans, although much shorter in duration.

The intracerebral aluminum levels (5 to 6  $\mu$ g/g dry weight<sup>18</sup>) that produce the toxic effects in animals are sometimes found in the brains of AD patients. Approximately 23% of randomly selected samples of neocortical grey matter from patients with AD have been found to contain aluminum in the amount of 5  $\mu$ g/g dry weight or more.<sup>19</sup>

## Aluminum-induced neurochemical changes in the brain

Aluminum affects many biochemical and neurochemical processes of the brain (Table 1). 12,17,20-105 Of particular importance is the observation that human neuroblastoma cells in culture treated with low doses of aluminum produce antigens that react with an antibody to an abnormally phosphorylated microtubule-associated protein known as Tau found in AD. 56 The antibody reacts specifically with AD neurofibrillary tangles. Considerable evidence now

#### Table 1: Actions of aluminum at the cellular level

#### **Nuclear effects**

Binds to DNA phosphate and bases<sup>20,21</sup>

Increases histone-DNA binding<sup>22</sup> Decreases RNA in neuroblastoma<sup>23</sup>

Blocks initiation sites for RNA polymerase<sup>24</sup>

Blocks RNA polymerase activity in vitro<sup>25</sup>

Blocks ribosylation of adenosine diphosphate<sup>26</sup>

Alters sister chromatid exchange<sup>27</sup>

Alters steroid-induced chromosome puffing<sup>28</sup>

Inhibits corticosterone binding to DNA<sup>29</sup>

Alters poly(A) RNA content in the forebrain of rabbits<sup>30</sup>

Reduces messenger RNA coding for neurofilaments in anterior horn cells of rabbits<sup>31</sup>

Decreases cell division and alters DNA synthesis<sup>32,33</sup>

Induces considerable changes in chromatin<sup>34</sup>

Alters the development of rat offspring<sup>35</sup>

#### Cytoplasmic effects

Induces conformational changes in calmodulin; blocks calmodulin-dependent calcium-magnesium adenosine triphosphatase (ATPase), which is important in extrusion of calcium ion from cells<sup>36,37</sup>

Increases intracellular calcium content<sup>17,38</sup>

Reduces sugar phosphorylation<sup>39</sup> Decreases respiration<sup>40</sup> Inhibits hexokinase activity<sup>41–46</sup>

Stabilizes terminal phosphoryl group on ATP<sup>41–46</sup>

Forms long-lived complex with ATP41-46

Inhibits ATP41-46

Inhibits brain glycolysis, depression of yeast and rat brain cytosolic and mitochondrial hexokinase activity<sup>41–46</sup>

Stimulates brain pyruvate kinase<sup>46</sup>

Enhances adenylate cyclase stimulation by fluoride but inhibits activation by serotonin and guanine nucleotides in *Fasciola hepatica*, a requirement for activation of the regulatory component of adenylate cyclase by fluoride<sup>47,48</sup>

Increases the number of lysosomes; reduces thiamine pyrophosphatase and nucleotide diphosphatase in the Golgi apparatus<sup>49</sup>

Inhibits the synthesis of tetrahydrobiopterin<sup>50</sup>

Elevates cyclic AMP and GMP\* levels<sup>51</sup>

Increases ubiquitin response in neurites of cultured nervous tissue<sup>52</sup>

Binds to ferritin and is partially sequestered by this mechanism; may alter iron storage<sup>53</sup>

Proliferates peroxisomes (aluminum clofibrate)54

Inactivates phosphofructokinase and inhibits hepatic glycolysis<sup>55</sup>

Induces an alteration in Tau that is recognized by antibodies to Alz 50<sup>56</sup>

Perturbs elongation factor Tu<sup>57</sup> Inhibits protein kinase-C activation<sup>58</sup>

Inhibits proton translocating
ATPases in Streptococcus and
Lactobacillus<sup>59</sup>

#### Cytoskeletal effects

Induces neurofibrillary degeneration composed of 10-nm fibres identical in chemical composition to normal neurofilaments<sup>60–64</sup>

Alters slow axonal transport, although disputed; has no effect on anterograde transport<sup>65–67</sup>

Alters phosphorylation of cytoskeletal proteins, microtubule-associated protein 2 and the 200KD component of neurofilaments<sup>68</sup>

May have secondary effect on cyclic AMP-dependent protein kinase<sup>68</sup>

Promotes assembly of microtubules, which are more slowly depolymerized than magnesium-assembled tubules<sup>69,70</sup>

Induces chronic myelopathy in rabbits<sup>71</sup>

Induces neuronal cytoskeletal lesions (through intravenous and intrathecal injections)<sup>72</sup>

Alters neurofilament conformation in vitro<sup>73</sup>

Inhibits calpain-mediated proteolysis; induces human neurofilament proteins to form high-molecular-weight complexes<sup>74</sup>

### Effects on membranes and membrane-bound enzymes

Alters physical properties of membrane lipids<sup>75</sup>

Binds to positive-charged and negative-charged sites in membranes in vitro<sup>76</sup>

Alters membrane structure<sup>77,78</sup>
Alters adenylate cyclase activity required for activation of regulatory component of adenylate cyclase in vitro by fluoride<sup>47,58</sup>

Inhibits sodium—
potassium-activated ATP
activity at relatively high
concentrations<sup>79</sup>

Enhances brain-specific lipid peroxidation<sup>80,81</sup>

Accelerates peroxidation of membrane lipids stimulated by iron salts<sup>80,81</sup>

Decreases activity of superoxide dismutase in rat brain<sup>80,81</sup>

Alters blood-brain barrier<sup>82-85</sup> Inhibits saturatable transport system for *N*-tyrosinated peptides and encephalin from brain<sup>82-85</sup>

Increases permeability of blood-brain barrier to neuropeptides<sup>82-85</sup>

### Synaptic and neurotransmitter effects

Alters dendritic shape and synaptic density in chronic preparations<sup>12,86</sup>

Blocks high-affinity uptake of γ-aminobutyric acid (GABA) and glutamate from synaptosomes<sup>87,88</sup>

Blocks synaptosome uptake of neurotransmitter amines choline, dopamine and noradrenaline<sup>79,89</sup>

Inhibits acetylcholinesterase<sup>23,90,91</sup>
Blocks uptake of calcium and binding of acetylcholine<sup>92</sup>

Reduces glucose uptake by synaptosomes extracted from rat cortex<sup>93</sup>

Depresses norepinephrine and dopamine levels in cortex and activity of enzymes dopamine-β-hydroxylase and phenylethanolamine-N-methyltransferase when fed to rats with diets deficient in copper, zinc and iron<sup>94,95</sup>

Reduces choline acetyltransferase in hypoglossal nucleus and spinal cord grey matter in rabbits<sup>96–98</sup>

Inhibits fast phase of voltage-dependent calcium influx into synaptosomes<sup>99</sup>

Inhibits protein phosphatase (in synaptosomal cytosol fractions)<sup>100</sup>

Is toxic to key synaptosomal enzymes (dependent on sodium-potassium, magnesium and calcium ions)<sup>101</sup>

Stimulates sodium chloride-dependent release of taurine and GABA in rat cortical astrocytes<sup>102</sup>

#### Effects on blood

Alters activity of cholinesterase<sup>103,104</sup> Interacts with transferrin<sup>105</sup>

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<sup>\*</sup>AMP = adenosine 3',5'-cyclic monophosphate, GMP = guanosine monophosphate.

indicates that these tangles are composed of a polymer of normal Tau that has undergone abnormal phosphorylation<sup>106</sup> and that aluminum in healthy neurons induces hyperphosphorylation of Tau.

The determination of which of the other toxic effects listed in Table 1 are important in the pathogenesis of AD will require a more advanced understanding of the cellular and molecular characteristics of AD and aluminum neurotoxic effects than now possible. The key biochemical events responsible for neuronal dysfunction and neuron death may involve energy metabolism, calcium homeostasis, membrane receptor and channel functions or gene expression. Aluminum, or any other postulated pathogenic factor, must be shown to induce changes in model systems similar to those observed in the human disease and must be found at the putative site within brain cells in sufficient concentrations to induce the changes observed in AD-affected tissues.<sup>107</sup>

Abnormal accumulation of aluminum has now been found in at least four sites in the AD-affected brain.

#### Neurofibrillary tangles

Different analytic methods capable of precise tissue localization  $^{108-110}$  have shown remarkably high concentrations of aluminum (up to  $300~\mu g/g$  dry weight) in the bundles of paired helical filaments that make up AD neurofibrillary tangles. The observation that aluminum induces abnormal phosphorylation of Tau,  $^{56}$  the principal subunit of AD neurofibrillary tangles,  $^{106}$  supports the idea that aluminum accumulation may occur early in the disease process and result in the formation of tangles, not late in the process in a terminally damaged neuron.

#### Amyloid cores of senile plaques

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Focal deposits of aluminum and silicon, as aluminosilicates, are also a consistent and specific feature of the central core of senile plaques. 111,112 Aluminosilicates, in vitro, can seed the formation of polymeric fibrillary aggregates of model peptides, including the amyloid peptide, which resemble the amyloid core of senile plaques. The amyloid precursor protein is considered to be the major protein found in senile plaques. Increased amounts of the protein have been found in some patients with elevated brain aluminum concentrations due to chronic renal failure. In some cases of prolonged dialysis precocious development of immature senile plaques has been observed.111,112 These observations, together with the finding that aluminosilicates seed the formation of fibrillary aggregates, support the possibility that elevated amounts of aluminum in

serum and brain can induce some of the cellular responses contributing to amyloid deposition.

#### Ferritin

Ferritin is an ubiquitous intracellular iron storage protein capable of scavenging other metals, including zinc and beryllium. The amount of aluminum found in ferritin extracted from AD-affected brains was 5.6 times higher than in ferritin from matched control preparations.<sup>113</sup> The researchers considered that the increase may have been due to a general increase in the availability of aluminum to the brain of patients with AD and raised the possibility that aluminum may release iron as Fe<sup>+++</sup>. This could facilitate the production of highly toxic free radicals that can denature proteins and destroy membranes, thereby contributing to neuron death in AD.

#### Chromatin fractions

AD is associated with a change in the structure of the DNA-protein complex that constitutes the physical matrix within which genes are expressed.<sup>114</sup> This change results in the reduced transcription of certain neuron-specific genes, including the lowmolecular-weight messenger RNA for neurofilament protein. 115,116 Aluminum has been shown to accumulate on DNA-containing components of the cell nucleus in the cerebral cortex.<sup>117</sup> A ninefold increase in aluminum content is associated with the DNAprotein fraction containing repressed neuronal genes.<sup>114</sup> Although aluminum increases the affinity of binding of certain repressor proteins to DNA and contributes to the gene repression, some other event likely occurs first to allow the protein to dock at a particular DNA site.

We postulate that aluminum replaces magnesium at a key DNA-protein binding site. Because the aluminum atom has a small ionic radius and high charge it is nearly a million times slower than the magnesium atom in dissociating from the DNA-protein complex. By replacing magnesium, aluminum locks the repressor protein in place.

#### **Environmental aluminum**

Although aluminum is a common constituent of the environment it has no recognized biologic function. Aluminum is absorbed primarily through the gastrointestinal tract but probably also through the respiratory epithelium and skin. Preliminary data using accelerator mass spectrometry with aluminum-26 ligated to citrate indicate that as much as 1% of aluminum ingested orally is absorbed into the blood stream (James Barker and J. Philip Day,

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Department of Chemistry, University of Manchester, Manchester: personal communication, 1991). Aluminum and fluoride, and probably silicon, are mutually antagonistic in competing for absorption in the gut; the more fluoride or silicon in the diet, the less aluminum absorbed.

In 1980 Shore and associates<sup>118</sup> did not show increased aluminum concentrations in AD patients. However, later studies<sup>119-121</sup> showed significantly elevated aluminum levels in the serum or whole blood of AD patients, as compared with the levels in carefully matched control patients. Aluminum appears to be transported in serum and carried into the brain by several proteins that may have different, genetically determined affinities for aluminum. One speculation is that individual variation in susceptibility to elevated aluminum concentrations may be related to the proteins responsible for transportation in serum. A portion of aluminum in serum is carried by high-molecular-weight proteins, including  $\alpha_2$ -macroglobulin, immunoglobulin, hepatoglobin, transferrin and albumin. In addition, a low-molecularweight protein (of about 18 kilodaltons) that transports aluminum has recently been discovered. 122

In cerebrospinal fluid most of the aluminum is bound to these newly identified low-molecular-weight proteins. Aluminum uptake by the brain is currently attributed to transferrin, and the highest densities of transferrin receptors are in regions of the brain selectively vulnerable to AD.<sup>123</sup> Recent evidence has indicated that the binding of gallium, an analogue of aluminum, by transferrin is defective in AD.<sup>124</sup> Thus, a serum transport defect could contribute to the accumulation of aluminum in the AD-affected brain.

Although it has been recognized for several years that the markedly elevated serum aluminum levels (200 µg/L) that occur in kidney failure may result in dialysis dementia, much lower concentrations are now recognized to be associated with impaired cognitive function. The psychomotor performance of 27 patients receiving long-term hemodialysis who had only a mildly raised serum aluminum level (mean 59 [normally less than 10]  $\mu$ g/L) was impaired, as compared with the performance of matched control subjects. 125 In another study 126 signs of neurologic dysfunction and impaired memory occurred in dialysis patients who had labile aluminum released by desferrioxamine, an aluminum chelating agent, in challenge tests. These observations further indicate that even moderate elevations in the serum aluminum level pose a risk of cognitive impairment.

#### Sequestration of aluminum

Aluminum may be sequestered in the human

body by several mechanisms. It is injected intramuscularly or subcutaneously into most of the world's population as an adjuvant in vaccines for diphtheria, pertussis, tetanus and hepatitis and in allergenic extracts. Alum-precipitated allergenic extracts contain up to 850  $\mu$ g of aluminum per dose, and injections of ragweed pollen every 2 weeks for 2 years results in a calculated dose of 44 mg of aluminum. The injected aluminum may persist in the tissues for weeks or years as subcutaneous nodules. 127 The granulomas comprise aluminum-laden histiocytes and dense lymphocytic infiltrates with germinal centres.127 The transportation and distribution of injected aluminum in the body are unknown, but tissue depots of aluminum are likely to release the metal for many years. Immunologic mechanisms may play a role in AD,5 and histiocytes that penetrate brain tissue may serve as an additional transport system for aluminum, which bypasses the blood-brain barrier.

Considerable concern has been expressed about the aluminum content of infant formulas and parenteral nutrients. Bishop and collaborators reported that a premature infant who had received an estimated total of 645  $\mu$ g of aluminum through parenteral feeding had seizures and died. At autopsy the temporal grey matter contained an average of 40.1  $\mu$ g/g wet weight of aluminum, as compared with a mean of 2.4  $\mu$ g/g wet weight in 12 infants dying unexpectedly within the first year of life. This 17-fold increase closely approaches that mentioned in the first reported fatal case of encephalopathy associated with respirable aluminum. 129

X-ray energy spectroscopy has detected aluminum in 9% of oral apical granulomas. <sup>130</sup> Possible sources of aluminum in the mouth include toothpaste, impression materials and amalgam.

Aluminum is also found in pigmented macrophages in Peyer's patches in all patients over 6 years of age<sup>131</sup> and is presumed to be of dietary origin. Whether aluminum is sequestered in these cells from potential neurotoxic actions is unknown. The recent development of accelerator mass spectroscopy for the long-life isotope <sup>26</sup>Al to be used in tracer studies will greatly help answer the many questions concerning aluminum metabolism.

#### **Epidemiologic findings**

Seven studies have related elevated aluminum concentrations in drinking water to an increased incidence of AD. 132-139

In a recent study<sup>137</sup> 2792 randomly selected subjects aged 65 years or more were screened for AD by psychologists using dementia screening tests.<sup>138</sup> They were then examined by senior neurologists using criteria established by the National Institute of

Neurological and Communicative Disorders and Stroke and the Alzheimer's Disease and Related Disorders Association.<sup>140</sup> The aluminum level in drinking water varied from 10 to 160 µg/L for the sample. The relative risk for probable AD at 100  $\mu$ g/L was 4.53 times greater than at 10  $\mu$ g/L. Martyn and colleagues<sup>136</sup> reported similar results in an incidence study of presenile Alzheimer's disease (in people 65 years of age or less) in 88 counties in England. By examining the records of computerized tomography (CT) units that served the districts they identified patients with probable AD (445 people) or possible AD (221). The control group comprised 519 people with dementia and multiple minor strokes visible on the CT scans and 2920 with adult-onset epilepsy without clinical dementia; the incidence of each of these diseases in the catchment areas was correlated with the aluminum content in the drinking water. No statistically significant relative risk between aluminum in the drinking water and dementia caused by multiple strokes or adult-onset epilepsy was found. However, the prevalence of AD in areas with an aluminum level of 111 µg/L of drinking water was 1.7 times higher than in areas with a level of less than 10 µg/L (confidence limits [CLs] 1.1 and 2.7).

Two epidemiologic studies in Norway used an ecologic approach. Vogt<sup>133</sup> divided the population of Norway into five zones according to the aluminum concentration in the lakes, which corresponded well with the aluminum levels found in drinking water. The lowest concentration was 20 µg/L and the highest 200 µg/L. The risk of death from senile dementia in the zone with the highest concentration was 1.48 times higher than in the zone with the lowest. Correlations between the rates of death from AD and the aluminum concentrations were statistically significant; multiple-stroke dementia and the risk of death from paralysis agitans were not related to the aluminum levels. Flaten<sup>134</sup> found a highly significant correlation between aluminum in processed drinking water and AD.

Table 2 lists the studies in order of increasing diagnostic accuracy. The relative risk of AD in relation to aluminum content of drinking water

appears to increase as the diagnostic accuracy increases in each of the studies.

In the United States Still and Kelly<sup>132</sup> examined first admissions to the state mental hospitals between 1971 and 1979 in three counties of South Carolina. Index cases were patients aged 55 years or more classified as having primary degenerative dementia (considered by the authors to be predominantly AD), vascular dementia, alcoholic dementia or other forms of dementia. Fluoride levels in water were measured instead of aluminum levels, because fluoride reduces aluminum uptake by the gut. In addition, the ingestion of aluminum compounds counteracts dental fluorosis and results in reduced fluoride stores in teeth and bones.<sup>141</sup> One county had 4.18 mg of fluoride per litre of drinking water; the levels in the other two counties were 0.49 and 0.61 mg/L. The incidence of primary degenerative dementia in the county with the highest fluoride level was about 20% of that in the other two; there were no significant differences between the counties in the number of first admissions because of the other types of dementia.

A study in Newfoundland revealed clusters of high rates of death from dementia, recorded on death certificates as an immediate, antecedent, underlying or contributing cause of death.<sup>139</sup> Index cases were grouped according to birth place. In 1985 and 1986 there was a significant excess of deaths in a small area of Bonavista Bay that could not be explained by differences in age, sex, ethnic origin, family origin or mobility patterns. The area was reported to have a high aluminum concentration in the drinking water (165  $\mu$ g/L), the lowest pH (5.2) and the highest colour (turbidity) index in the region. The last two conditions would increase the probability of the formation of both polynuclear inorganic aluminum and organic aluminum ligands, which, as argued later, have greater aluminum neurotoxic effects than mononuclear aluminum.

A recent case-control epidemiologic survey of 130 matched pairs examined the association between AD and the lifetime exposure to aluminum in antiperspirants and antacids. <sup>142</sup> For aluminum-containing antiperspirants the overall adjusted odds

Study	Diagnostic criteria	No. of subjects	Aluminum level, μg/L drinking water	Relative risk*
Vogt <sup>133</sup>	Death certificates	18 664	20–200	1.48
Martyn et al <sup>136</sup> Michel et al <sup>137</sup>	Computerized tomograms DSM-III <sup>138</sup> and	307	0-> 110	1.7
	NINCDS-ADRDA140	2 792	10-160	4.53

ratio for AD was 1.6 (CL 1.04 and 2.4), the risk increasing with increased frequency of use (p =0.03), and the odds ratio for the 33% of those who used the highest amount was 3.2. For antacids the overall adjusted odds ratio, regardless of aluminum content, was 3.1, with a dose-response gradient (p =0.009), and the odds ratio for the highest tertile was 11.7. When only aluminum-containing antacids were analysed no significant risk or dose-response trend emerged. However, in another study the brain aluminum concentration was higher after ingestion of antacids with a high level of aluminum. Dollinger and coworkers<sup>143</sup> examined brain tissue specimens from 20 subjects scheduled for brain surgery who were given antacids for 10 days for stress prophylaxis. Half of them received 70 mL of a high-aluminumcontent antacid daily, the others an equal dose of a low-aluminum-content antacid. After the 10 days of antacid treatment the low-dose group had a mean aluminum concentration of 0.412 µg/g wet weight (an estimated 2.60  $\mu$ g/g dry weight) and the highdose group 1.05  $\mu$ g/g wet weight (an estimated 5.25  $\mu g/g$  dry weight). The mean aluminum level in brain tissue from 20 control subjects was 0.583  $\mu$ g/g wet weight.

The new tracer techniques demonstrate that uptake of aluminum between the bowel and the blood and between the blood and the brain occurs at a considerably higher rate than previously suspected. Accelerator mass spectrometry with the use of <sup>26</sup>Al ligated to citrate in rats revealed that approximately 1/55 000 of a single intraperitoneal injection of <sup>26</sup>Al is incorporated into the cerebrum. <sup>144</sup>

Epidemiologic studies demonstrate association but do not establish cause and effect. Nevertheless, each study reviewed reached the same conclusion: aluminum in drinking water is associated with an increased risk of dementia of the Alzheimer type.

The most satisfactory studies would include autopsy findings confirming the diagnosis and ruling out clusters of familial AD as confounding variables. The aluminum ligands in food, water and all substances ingested or injected into people should be measured and correlated with the incidence of AD. Such studies will be extremely expensive to conduct and will require several years to execute.

#### Organic and inorganic forms of aluminum

Are all forms of aluminum equally toxic? It is well known that organic mercury, methyl mercury, is much more toxic than metallic mercury. The same is true for aluminum. Some forms of aluminum, like aluminum hydroxide, are poorly absorbed from the gastrointestinal tract, whereas certain organic ligands of aluminum, such as aluminum citrate, pass very rapidly from the food chain into the blood.

The forms of aluminum in drinking water are of considerable significance for epidemiologic studies. They are incompletely studied, and those that could be risk factors for AD have not been identified. Driscoll and Letterman<sup>145</sup> studied the chemistry and fate of aluminum in water pumped from Lake Ontario and treated with alum for drinking in Syracuse, NY. They found that 52% of the aluminum was in the form of monomeric alumino-hydroxide complexes, 29% was associated with organic matter, 19% was in a fluoride complex, and a small amount was particulate. The relative intestinal absorption by mammals of these organic and inorganic forms has not been studied.

Recently, aluminum-27 magnetic resonance spectra of soil samples demonstrated the presence of inorganic polynuclear aluminum forms, which may account for up to 30% of aluminum in water. 146 Since these inorganic polynuclear forms appear to be 10 times more toxic than mononuclear forms in certain aquatic plants and fish their toxic effects in mammals and humans need to be investigated. Considerable seasonal variation has been observed in the distribution of aluminum forms in drinking water. Therefore, it is very important that the various organic and inorganic types of aluminum be correlated with the incidence of AD in each of the epidemiologic studies reviewed in this article and be considered in future studies.

#### Respiratory aluminum

Rifat and associates<sup>147</sup> examined prolonged exposure to respirable aluminum to determine whether it was associated with serious cognitive deficit. Between 1944 and 1979 finely ground alumina (particle diameter less than 2  $\mu$ m) was dispensed prophylactically for silicosis to groups of gold miners in northern Ontario. The alumina powder was dispersed in the miners' change rooms for 10 to 20 minutes at a concentration of 35 mg/m³ before each underground shift. These miners had significantly poorer performance on cognitive tests than an agematched group of miners who had not been exposed. The differences persisted after adjusting for potential confounding factors such as head injury, education and alcohol abuse. The relative risk of severe cognitive deficit was 4.5 times greater among miners with more than 20 years' exposure, 3.1 among those with 10 to 20 years' exposure and 2.4 among those with 1 to 10 years' exposure. Clinical and histopathologic examinations were not done, and thus no conclusion can be reached as to whether the cognitive deficit was more closely related to dialysis dementia or to AD. Although long-term follow-up examinations were not performed and brain tissue was not examined, the authors have concluded that prolonged

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exposure to respirable aluminum is associated with serious cognitive deficit.

### Clinical progression of AD after removal of brain aluminum

A fourth line of independent evidence has tested the hypothesis that if aluminum is an important pathogenic factor in AD, its removal by an ionspecific binding agent should slow the progress of the disease. A 2-year prospective, single-blind clinical trial was conducted to determine whether the sustained use of desferrioxamine, a trivalent metal ion binding agent, would slow the progression of the dementia.148 A total of 48 people living at home with probable AD were randomly assigned to three treatment groups: desferrioxamine, lecithin (in oral homeopathic doses of 1 g/d) and no treatment. A structured performance test measuring daily living skills was videotaped in the home and was the outcome measure over the 2-year period. The tapes were analysed at random by trained behaviour raters blind to the purpose and protocol of the study. There was no statistical difference in the average rate of decline in performance between the lecithin group and the no-treatment group. However, when data from these two groups were combined the average 2-year decline in the desferrioxamine group was 25% of the maximum score, as compared with 57% in the no-treatment group.

A double-blind placebo-controlled multicentre trial must now be conducted to confirm these results. However, on the basis of current evidence desferrioxamine appears to slow the progression of AD. These results support the hypothesis that aluminum is a significant toxic environmental factor in the pathogenesis of the disease.

#### **Conclusions**

Four independent lines of evidence support the conclusion that aluminum is an important risk factor in AD: (a) prolonged exposure to trace amounts of aluminum induces cognitive deficits in experimental animals and humans, (b) aluminum accumulates in at least four sites in AD-affected brain tissue at concentrations known to affect several biochemical reactions, (c) seven epidemiologic studies have demonstrated an association between AD and aluminum exposure in drinking water and antiperspirants (high levels of aluminum in the air have been found to increase significantly the risk of cognitive defects. but an association between this type of aluminum exposure and AD has not been investigated) and (d) treatment with a trivalent metal ion binding agent slows, but does not arrest, the clinical progression of AD.

#### Recommendations

- Human exposure to aluminum should be limited. After considering the available evidence a prudent person may wish to limit daily exposure to aluminum. In addition to reducing head trauma, which is considered to be a risk factor for AD,149 reduction in aluminum exposure may be the only change in lifestyle that offers hope of reducing the incidence of AD. Unfortunately, the sources of aluminum are largely unknown to the public. As well as occurring naturally in food and water, aluminum is added to drinking water, many processed foods, cosmetics, toothpaste, antiperspirants and adjuvants in various parenteral preparations and other pharmaceutical agents. Current evidence supports the hypothesis that a major reduction in the ingestion of aluminum would significantly reduce the incidence of AD.
- Public policy action is required. No risk factor other than aluminum that might be controlled by public action to reduce the incidence of AD has yet been identified. Sources of public health information have an obligation to inform the public about the current state of knowledge concerning the relation between aluminum and AD so that each individual may make an informed decision. Additional important pathogenic factors in AD will become recognized as knowledge grows about this disease.
- The aluminum content should be listed on the packages of all substances marketed for human contact and ingestion including processed foods, potable water, cosmetics, toothpaste and pharmaceutical products.
- Municipal processed water should be regulated so that the aluminum concentration is less than  $50 \mu g/L$ ; the long-term goal should be a concentration of less than  $10 \mu g/L$ .
- A goal for the daily intake of aluminum from all sources by adults should be 3 mg or less.
- Further research should be conducted to understand fully the health risks of aluminum.

This work was supported by grants from the Ontario Mental Health Foundation, the Medical Research Council of Canada, the Department of National Health and Welfare, the Scottish Rite Charitable Foundation and the Alzheimer Association of Ontario.

#### References

- St George-Hyslop PH, Tanzi RE, Polinsky RJ et al: The genetic defect causing familial Alzheimer's disease maps on chromosome 21. Science 1987; 235: 885-890
- St George-Hyslop PH, Haines JL, Farrer LA et al: Genetic linkage studies suggest that Alzheimer's disease is not a single homogeneous disorder. Nature 1990; 347: 194-197

- Goate A, Chartier-Harlin MC, Mullan M et al: Segregation of a missense mutation in the amyloid precursor protein gene with familial Alzheimer's disease. Nature 1991; 349: 704-706
- Nee LE, Elvidge R, Sunduland T et al: Dementia of the Alzheimer type: clinical and family study of 22 twin pairs. Neurology 1987; 37: 359-363
- McGeer PL, McGeer E, Rogers J et al: Anti-inflammatory drugs and Alzheimer disease [C]. Lancet 1990; 335: 1037
- Krishnan SS, McLachlan DR, Krishnan B et al: Aluminum toxicity to brain. Sci Total Environ 1988; 71: 59-64
- Crapper DR, Dalton AJ: Alterations in short term retention, conditioned avoidance response acquisition and motivation following aluminum induced neurofibrillary degeneration. *Physiol Behav* 1973; 10: 925-933
- Idem: Aluminum induced neurofibrillary degeneration, brain electrical activity and alterations in acquisition and retention. Ibid: 935-945
- Crapper DR: Experimental neurofibrillary degeneration and altered electrical activity. Electroencephalogr Clin Neurophysiol 1973; 35: 575-588
- Idem: Functional consequences of neurofibrillary degeneration. In Gershon S, Terry RD (eds): The Neurobiology of Aging, Raven, New York, 1976: 405-432
- Idem: Dementia: recent observations on Alzheimer's disease and experimental aluminum encephalopathy. In Seeman P, Brown GM (eds): Frontiers in Neurology and Neuroscience (Neuroscience Institute, University of Toronto, symposium 1), U of Toronto Pr, Toronto, 1974: 97-111
- Petit TL, Biederman GB, McMullen PA: Neurofibrillary degeneration, dendritic dying back and learning-memory deficits after aluminum administration: implications for brain aging. Exp Neurol 1980; 67: 152-162
- Rabe A, Lei M, Shek J et al: Learning deficit in immature rabbits with aluminum-induced neurofibrillary degeneration. Exp Neurol 1982; 76: 441-446
- Yokel RA: Repeated systemic aluminum exposure effects on classical conditioning of the rabbit. Neurobehav Toxicol Teratol 1983; 5: 41-46
- Pendlebury WW, Perl DP, Schwentker A et al: Aluminuminduced neurofibrillary degeneration disrupts acquisition of the rabbit's classically conditioned nictitating membrane response. *Behav Neurosci* 1988; 102: 615-620
- Solomon PR, Pingree TM, Baldwin D et al: Disrupted retention of the classically conditioned nictitating membrane response in rabbits with aluminum-induced neurofibrillary degeneration. Neurotoxicology 1988; 2: 209-222
- Farnell BJ, Crapper McLachlan DR, Baimbridge K et al: Calcium metabolism in aluminum encephalopathy. Exp Neurol 1985; 88: 68-83
- Crapper DR, Tomko GJ: Neuronal correlates of an encephalopathy associated with aluminum neurofibrillary degeneration. *Brain Res* 1975; 97: 253-264
- Crapper DR, Krishnan SS, Quittkat S: Aluminum, neurofibrillary degeneration and Alzheimer disease. *Brain* 1976; 99: 67-79
- Karlik SJ, Eichhorn GL, McLachlan DRC: Molecular interactions of aluminum with DNA. Neurotoxicology 1980; 1: 83-88
- Karlik SJ, Eichhorn GL, Crapper DR: Interactions of aluminum species with deoxyribonucleic acid. *Biochemistry* 1980; 19: 5991-5998
- Lukiw WJ, Kruck TPA, McLachlan DR: Alterations in human linker histone-DNA binding in the presence of aluminum salts in vitro and in Alzheimer's disease. Neurotoxicology 1987; 8: 291-301
- 23. Miller CA, Levine EM: Affects of aluminum salts on cultured neuroblastoma cells. *J Neurochem* 1974; 22: 751-758
- 24. Sarkander HI, Balb G, Schlosser H et al: Blockade of neuronal brain RNA initiation sites by aluminum: a primary

- molecular mechanism of aluminum-induced neurofibrillary changes. In Cervos-Navarro J, Sarkander HI (eds): *Brain Aging: Neuropathology and Neuropharmacology*, Raven, New York, 1983: 645-647
- 25. Crapper McLachlan DR, Dam TV, Farnell BJ et al: Aluminum inhibition of ADP-ribosylation in vivo and in vitro. *Neurobehav Toxicol Teratol* 1983; 5: 645-647
- Crapper McLachlan DR, Farnell B, Galin H et al: Aluminum in human brain disease. In Sarkar B (ed): Biological Aspects of Metals and Metal Related Diseases, Raven, New York, 1983: 209-218
- 27. De Boni U, Seger M, Crapper McLachlan DR: Functional consequences of chromatin bound aluminum in cultured human cells. *Neurotoxicology* 1980; 1: 65-81
- 28. Sanderson CL, Crapper McLachlan DR, De Boni U: Altered steroid induced puffing by chromatin bound aluminum in a polytene chromosome of the black fly Simulium vittatum. Can J Genet Cytol 1982; 24: 27-36
- Idem: Inhibition of corticosterone binding in vitro, in rabbit hippocampus, by chromatin bound aluminum. Acta Neuropathol (Berl) 1982; 57: 249-254
- 30. Van Berkum MFA, Wong L, Lewis PN et al: Total and poly(A) RNA yields during an aluminum encephalopathy in rabbit brains. *Neurochem Res* 1986; 11: 1347-1359
- 31. Muma NA, Troncoso JC, Hoffman P et al: Aluminum neurotoxicity altered expression of cytoskeletal genes.

  Mol Brain Res 1988; 3: 115-122
- 32. Gelfant S: Inhibition of cell division a critical experimental analysis. *Rev Cytol* 1963; 14: 249-254
- Sampson M, Clarkson D, Davies DD: DNA synthesis in aluminum treated roots of barley. Science 1965; 148: 1476– 1477
- 34. Walker PR, LeBlanc J, Sikorska M: Effects of aluminum and other cations on the structure of brain and liver chromatin. *Biochemistry* 1989; 28: 3911-3915
- 35. Muller G, Bernuzzi V, Desor D et al: Developmental alterations in offspring of female rats orally intoxicated by aluminum lactate at different gestation periods. *Teratology* 1990; 42: 253-261
- Siegel N, Haug A: Aluminum interaction with calmodulin: evidence for altered structure and function from optical and enzymatic studies. *Biochim Biophys Acta* 1983; 744: 36-45
- 37. Idem: Calmodulin dependent formation of membrane potential in barley root plasma membrane vescicles a biochemical model of aluminum toxicity in plants. *Physiol Plant* 1983; 59: 285-291
- 38. Burnatowska-Hledin MA, Mayor GH: The effect of aluminum (Al) loading on specific tissue calcium (Ca) and magnesium (Mg) concentrations in normal rats [abstr]. Clin Res 1982; 30: A741
- Rorison IH: The effect of aluminum on the uptake and incorporation of phosphate by excised sanfoin roots. New Physiol 1965; 64: 23-27
- Foy CD, Chaney RL, White MC: The physiology of metal toxicity in plants. Annu Rev Plant Physiol 1978; 29: 511-566
- 41. Trapp GA: Studies of aluminum interaction with enzymes and proteins: the inhibition of hexokinase. *Neurotoxicology* 1980; 1: 89-100
- 42. Bock JL, Ash AE: NMR and infrared spectroscopic investigations of the Al (III), Ga (III), and Be (II) complexes of ATP. J Inorg Biochem 1980; 13: 105-110
- Womack FC, Colowick SP: Proton-dependent inhibition of yeast and brain hexokinase by aluminum in ATP preparations. Proc Natl Acad Sci USA 1979; 76: 5080-5084
- Karlik SJ, Elgavish GA, Eichhorn GL: Multinuclear NMR studies on Al (III) complexes of ATP and related compounds. J Am Chem Soc 1983; 105: 602-609
- 45. Karlik SJ, Tariene, Elgavish GA et al: Aluminum-27 NMR study of Al (III) interactions with carboxylate ligands. *Inorg Chem* 1983; 22: 525-529
- 46. Lai JCK, Blass JP: Inhibition of brain glycolysis by alumi-

- num. J Neurochem 1984; 42: 438-445
- 47. Mansour JM, Ehrlich A, Mansour TE: The dual effects of aluminum as activator and inhibitor of adenylate cyclase in the liver fluke Fasciola hepatica. Biochem Biophys Res Commun 1983; 112: 911-918
- Sternweiss PC, Gilman AG: Aluminum a requirement for activation of the regulatory component of adenylate cyclase by fluoride. Proc Natl Acad Sci USA 1982; 79: 4888-4891
- Gruca S, Wisniewski HM: Cytochemical study on the effect of aluminum on neuronal Golgi apparatus and lysosomes. Acta Neuropathol (Berl) 1984; 63: 287-295
- Altmann P, Alsalihi F, Butter K et al: Serum aluminum levels and erythrocyte dihydropteridine reductase activity in patients on thermodialysis. N Engl J Med 1987; 317: 80-84
- Johnson GVW, Jope RS: Aluminum alters cyclic AMP and cyclic GMP levels but not presynaptic cholinergic markers in rat brain in vivo. *Brain Res* 1987; 403: 1-6
- 52. Morandi A, Los B, Osofsky L et al: Ubiquitin and heat shock proteins in cultured nervous tissue after different stress conditions. In Iqbal K, Wisniewski H, Winblad B (eds): Alzheimer's Disease and Related Disorders, AR Liss, New York, 1989: 819-827
- Fleming J, Joshi JG: Ferritin: isolation of aluminum-ferritin complex from brain. Proc Natl Acad Sci USA 1987; 84: 7866-7870
- 54. Takagi A, Sai K, Umemura T et al: Relationship between hepatic peroxisome proliferation and 8-hydroxydeoxyguanosine formation in liver DNA of rats following long-term exposure to three peroxisome proliferators: di (2-ethylhexyl) phthalate, aluminum clofibrate and simfibrate. Cancer Lett 1990; 35 (1): 33-38
- Xu ZX, Fox L, Melethil S et al: Mechanism of aluminuminduced inhibition of hepatic glycolysis: inactivation of phosphofructokinase. *J Pharmacol Exp Ther* 1990; 254: 301-305
- 56. Guy SP, Jones D, Mann DMA et al: Human neuroblastoma cells treated with aluminium express an epitope associated with Alzheimer's disease neurofibrillary tangles. *Neurosci* Lett 1991; 121: 166-168
- Hazlett TL, Higashijima T, Jameson DM: Examination of elongation factor Tu for aluminum fluoride binding sites using fluorescence and F-19-NMR methodologies. FEBS Lett 1991; 278: 225
- Cochran M, Elliott DC, Brennan P et al: Inhibition of protein kinase C activation by low concentrations of aluminium. Clin Chim Acta 1990; 194: 167-172
- 59. Sturr MG, Marquis RE: Inhibition of proton-translocating ATPases of *Streptococcus mutans* and *Lactobacillus casei* by fluoride and aluminum. *Arch Microbiol* 1990; 155: 22-27
- Klatzo I, Wisniewski H, Streicher E: Experimental production of neurofibrillary degeneration: 1. Light microscopic observations. J Neuropathol Exp Neurol 1965; 24: 187-199
- Terry RD, Peña C: Experimental production of neurofibrillary degeneration: 2. Electron microscopy, phosphatase histochemistry and electron probe analysis. Ibid: 200-210
- Dahl D, Bignami A: Immunochemical cross-reactivity of normal neuro-fibrils and aluminum-induced neurofibrillary tangles — immunofluorescence study and antineural filament serum. Exp Neurol 1978; 58: 79-80
- 63. Selkoe DJ, Liem RKH, Yen S et al: Biochemical and immunological characterization of neurofilaments in experimental neurofibrillary degeneration induced by aluminum. *Brain Res* 1979; 163: 235-252
- 64. Munoz-Garcia D, Pendlebury WW, Kessler JB et al: An immunocytochemical comparison of cytoskeletal proteins in aluminum-induced and Alzheimer-type neurofibrillary tangles. *Acta Neuropathol (Berl)* 1986; 70: 243-248
- 65. Bizzi A, Crane RC, Autilio-Gambetti L et al: Aluminum effect on slow axonal transport: a novel impairment of

- neurofilament transport. J Neurosci 1984; 4: 722-731
- 66. Kosik KS, McCluskey AH, Walsh FX et al: Axonal transport of cytoskeletal proteins in aluminum toxicity — aluminum toxicity in axonal transport. Neurochem Pathol 1985; 3: 99-108
- Liwnicz BH, Kristensson K, Wisniewski HM et al: Observations on axoplasmic transport in rabbits with aluminuminduced neurofibrillary tangles. *Brain Res* 1974; 80: 413– 420
- 68. Johnson GVM, Jope RS: Cytoskeletal protein phosphorylation is altered in the brains of aluminum-treated rats [abstr]. Presented at the 17th Annual Meeting of the Society for Neurosciences, New Orleans, 1987: A366.10 (vol 13)
- Macdonald TL, Humphries WG, Martin RB: Promotion of tubulin assembly by aluminum ion in vitro. Science 1987; 236: 183-186
- 70. Macdonald TL, Martin RB: Aluminum ion in biological systems. *Trends Biochem Sci* 1988; 13: 15-19
- Strong MJ, Wolff AV, Wakayama I et al: Aluminuminduced chronic myelopathy in rabbits. Neurotoxicology 1991: 12: 9-21
- Katsetos CD, Savory J, Herman MM et al: Neuronal cytoskeletal lesions induced in the CNS by intraventricular and intravenous aluminium maltol in rabbits. *Neuropathol* Appl Neurobiol 1990; 16: 511
- 73. Troncoso JC, March JL, Haner M et al: Effect of aluminum and other multivalent cations on neurofilaments in vitro: an electron microscopic study. *J Struct Biol* 1990; 103: 2-12
- Nixon RA, Clarke JF, Logvinenko KB et al: Aluminum inhibits calpain-mediated proteolysis and induces human neurofilament proteins to form protease-resistant high molecular weight complexes. J Neurochem 1990; 55: 1950-1959
- 75. Viestra R, Haug A: The effect of aluminum 3+ on the physical properties of membranes in *Thermoplasma acidophilum. Biochem Biophys Res Commun* 1978; 84: 134-144
- Deleers M: Cationic atmosphere and cation competition binding at negatively charged membranes: pathological implication of aluminum. Res Commun Chem Pathol Pharmacol 1985; 49: 277-294
- 77. Deleers M, Servais JP, Wülfert E: Micromolar concentrations of Al<sup>3+</sup> induce phase separation, aggregation and dye release in phosphatidylserine-containing lipid vesicles. *Biochim Biophys Acta* 1985; 813: 195-200
- 78. Idem: Neurotoxic cations induce membrane rigidification and membrane fusion at micromolar concentrations. *Biochim Biophys Acta* 1986; 855: 271-276
- 79. Lai JCK, Guest JF, Leung TKC et al: The effect of cadmium, manganese and aluminum on sodium-potassium-activated and magnesium-activated adenosine triphosphate activity and choline uptake in rat brain synaptosomes. *Biochem Pharmacol* 1980; 29: 141-146
- Ohtawa M, Seko M, Takayama F: Effect of aluminum on lipid peroxidation in rats. Chem Pharm Bull (Tokyo) 1983; 31: 1415-1418
- 81. Gutteridge JMC, Quinlan GJ, Clark I et al: Aluminum salts accelerate peroxidation of membrane lipids stimulated by iron salts. *Biochim Biophys Acta* 1985; 835: 441-447
- 82. Banks WA, Kastin AJ, Fasold MB: Differential effect of aluminum on the blood-brain barrier, transport of peptides, technetium and albumin. *J Pharmacol Exp Ther* 1988; 244: 579-585
- 83. Banks WA, Kastin AJ: Aluminium increases permeability of the blood-brain barrier to labelled DSIP and β-endorphin: possible implications for senile and dialysis dementia. Lancet 1983; 2: 1227-1229
- Idem: Aluminum alters the permeability of the blood-brain barrier to some non-peptides. *Neuropharmacology* 1985; 24: 407-412
- 85. Idem: The aluminum induced increase in blood-brain barrier permeability to delta-sleep-inducing peptide occurs throughout the brain and is independent of phosphorus and

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- acetylcholinesterase levels. Psychopharmacology (Berlin) 1985; 86: 84-89
- Uemura E, Ireland WP: Synaptic density in chronic animals with experimental neurofibrillary changes. Exp Neurol 1984; 85: 1-9
- 87. Wong PCL, Lai JCK, Lim L et al: Selective inhibition of L-glutamate and gamma-aminobutyrate transporter in nerve ending particles by aluminum, manganese and cadmium chloride. *J Inorg Biochem* 1981; 14: 253-260
- Sturman JA, Wisniewski HM, Shek JW: High affinity uptake of GABA and glycine by rabbits with aluminuminduced neurofibrillary degeneration. *Neurochem Res* 1983; 8: 1097-1109
- 89. Lai JCK, Lim L, Davison AN: Effects of Cd<sup>2+</sup>, Mn<sup>2+</sup>, and Al<sup>3+</sup> on rat brain synaptosomal uptake of noradrenaline and serotonin. *J Inorg Biochem* 1982; 17: 215-225
- Marquis JK: Aluminum neurotoxicity: an experimental perspective. Bull Environ Contam Toxicol 1982; 29: 43-49
- 91. Marquis JK, Lerrick AJ: Noncompetitive inhibition of bi-aluminum, scandium and yttrium of acetylcholinesterase from *Electrophorus electricus*. *Biochem Pharmacol* 1983; 31: 1437-1440
- 92. Hava M, Hurwitz A: The relaxing effect of aluminum and lanthanum on rat and human gastric smooth muscle in vitro. Eur J Pharmacol 1973; 22: 156-161
- Lipman JJ, Colowick SP, Lawrence PL et al: Aluminum induced encephalopathy in the rat. Life Sci 1988; 42: 863– 875
- 94. Wenk GL, Stemmer KL: The influence of ingested aluminum upon norepinephrine and dopamine levels in the rat brain. *Neurotoxicology* 1981; 2: 347-353
- 95. Idem: Activity of the enzymes dopamine-beta-hydroxylase and phenylethanolamine-N-methyltransferase in discrete brain regions of the copper-zinc deficient rat following aluminum ingestion. Neurotoxicology 1982; 3: 93-99
- Yates CM, Simpson J, Russell D et al: Cholinergic enzymes in neurofibrillary degeneration produced by aluminum. Brain Res 1980; 197: 269-274
- 97. Hetnarski D, Wisniewski HM, Iqbal K et al: Central cholinergic activity in aluminum-induced neurofibrillary degeneration. *Ann Neurol* 1989; 7: 489-490
- Simpson J, Yates CM, Whyler DK et al: Biochemical studies on rabbits with aluminum-induced neurofilament accumulations. Neurochem Res 1985; 10: 229-238
- Koenig ML, Jope RS: Aluminum inhibits the fast phase of voltage-dependent calcium influxes into synaptosomes. J Neurochem 1987; 49: 316-320
- 100. Yamamoto H, Saitoh Y, Yasugawa S et al: Dephosphorylation of Tau factor by protein phosphatase 2A in synaptosomal cytosol fractions, and inhibition by aluminum. J Neurochem 1990; 55: 683-690
- Rao KSJ: Effects of aluminum salts on synaptosomal enzymes: an in vitro kinetic study. Biochem Int 1990; 22: 725-734
- 102. Albrecht J, Norenberg MD: Aluminum chloride stimulates NaCl-dependent release of taurine and γ-aminobutyric acid in rat cortical astrocytes [abstr]. Neurochem Int 1991; 18: 125
- 103. Patocka J: The influence of Al3+ on cholinesterase and acetylcholinesterase activity. Acta Biol Med Ger 1971; 26: 845-846
- Marquis JK: Aluminum inhibition of human serum cholinesterase. Bull Environ Contam Toxicol 1983; 31: 164-169
- Trapp GA: Plasma aluminum is bound to transferrin. Life Sci 1983; 33: 311-316
- 106. Lee VMY, Balin BJ, Otvos L et al: A68: a major subunit of paired helical filaments and derivatized forms of normal tau. Science 1991; 251: 675-678
- 107. McLachlan DRC: Aluminum neurotoxicity: criteria for assigning a role in Alzheimer's disease. In Lewis TE (ed): Environmental Chemistry and Toxicology of Aluminum,

- Lewis Publs, Chelsea, Mich, 1989: 299-315
- Perl DP, Brody AR: Alzheimer's disease: x-ray spectrometric evidence of aluminum accumulation in neurofibrillary tangle-bearing neurons. Science 1980; 208: 297-299
- 109. Perl DP, Pendlebury WW: Aluminum accumulation in neurofibrillary tangle-bearing neurons of senile dementia, Alzheimer's type: detection by intraneuronal x-ray spectrometry studies of unstained tissue sections [abstr]. J Neuropathol Exp Neurol 1984; 43: 349
- 110. Perl DP: Aluminum and Alzheimer neurofibrillary degeneration [abstr]. Presented at the Aluminum and Health International Symposium, Orlando, Fla, Dec 12-14, 1989
- 111. Candy JM, Klinowski RH, Perry EK et al: Aluminosilicates and senile plaque formation in Alzheimer's disease. Lancet 1986; 1: 354-357
- 112. Edwardson JA, Candy JM: Aluminum and the aetiopathogenesis of Alzheimer's disease [abstr]. Neurobiol Aging 1990; 11: 314
- 113. Fleming J, Joshi JG: Ferritin: isolation of aluminum-ferritin complex from brain. Proc Natl Acad Sci USA 1987; 84: 7866-7870
- 114. McLachlan DRC, Lukiw WJ, Kruck TPA: New evidence for an active role of aluminum in Alzheimer's disease. Can J Neurol Sci 1989; 16: 490-497
- 115. McLachlan DRC, Lukiw WJ, Wong L et al: Selective messenger RNA reduction in Alzheimer's disease. Mol Brain Res 1988; 3: 255-262
- 116. Clark AW, Krekoski CA, Parhad IM et al: Altered expression of genes for amyloid and cytoskeletal proteins in Alzheimer cortex. Ann Neurol 1989; 25: 331-339
- 117. Crapper DR, Quittkat S, Krishnan SS et al: Intranuclear aluminum content in Alzheimer's disease: dialysis encephalopathy. Acta Neuropathol (Berl) 1980; 50: 19-24
- 118. Shore D, King SW, Kaye W et al: Serum and CSF aluminum and circulating parathyroid hormone in primary degenerative (senile) dementia. *Neurotoxicology* 1980; 1: 55-63
- Naylor GJ, Smith AHW, McHarg A et al: Raised serum aluminum concentration in Alzheimer's disease. Trace Elem Med 1989; 6: 93-95
- 120. Van Rhijn A, Corrigan FM, Ward NI: Serum aluminum in senile dementia of Alzheimer's type and in multi-infarct dementia. Ibid: 24-26
- 121. Kellet JM, Taylor A, Oram JJ: Aluminosilicates and Alzheimer's disease [C]. Lancet 1986; 1: 682
- 122. Favarato M, Mizzen C, Kruck TPA et al: Chromatographic resolution of aluminum binding components in human serum [abstr]. Neurobiol Aging 1990; 11: 315
- 123. Edwardson JA, Candy JM: Aluminium and the pathogenesis of senile plaques in Alzheimer's disease, Down's syndrome and chronic renal dialysis. *Ann Med* 1989; 21: 95-97
- 124. Farrar G, Altmann P, Welch S et al: Defective gallium-transferrin binding in Alzheimer disease and Down syndrome: possible mechanism for accumulation of aluminium in brain. *Lancet* 1990; 335: 747-750
- 125. Altmann P, Dhanesha U, Hamon C et al: Disturbance of cerebral function by aluminium in haemodialysis patients without overt aluminium toxicity. Lancet 1989; 1: 7-11
- 126. Sprague SM, Corwin HL, Tanner CM et al: Relationship of aluminum to neurocognitive dysfunction in chronic dialysis patients. Arch Intern Med 1988; 148: 2169-2172
- 127. Slater DN, Underwood JCE, Durrant TE et al: Aluminium hydroxide granulomas: light and electron microscopic studies and x-ray microanalysis. *Br J Dermatol* 1982; 107: 102, 108
- 128. Bishop NJ, Robinson MJ, Lendon M et al: Increased concentration of aluminium in the brain of a parenterally fed preterm infant. Arch Dis Child 1989; 64: 1316-1317
- 129. McLaughlin AIG, Kazantzis G, King E et al: Pulmonary fibrosis and encephalopathy associated with the inhalation of aluminium dust. *Br J Ind Med* 1962; 19: 253-263
- 130. Levison DA, Crocker PR, Lee G et al: Unexpected inorganic

- elements in oral lesions: results of x-ray energy spectroscopy (XES) on particulate matter in paraffin sections. *J Pathol* 1984; 144: 119-129
- 131. Shepherd NA, Crocker PR, Smith AP et al: Exogenous pigment in Peyer's patches. *Hum Pathol* 1987; 18: 50-54
- 132. Still CN, Kelly P: On the incidence of primary degenerative dementia vs. water fluoride content in South Carolina. *Neurotoxicology* 1980; 4: 125-131
- 133. Vogt T: Water quality and health study of a possible relationship between aluminum in drinking water and dementia [abstr] (Sosiale og okonomiske studier 61), Central Bureau of Statistics of Norway, Oslo, 1986
- 134. Flaten TP: Geographical associations between aluminum in drinking water and registered death rates with dementia (including Alzheimer's disease, Parkinson's disease and amyotrophic lateral sclerosis) in Norway. Environ Geochem Health 1990; 12: 152-167
- 135. Leventhal GH: Alzheimer's Disease and Environmental Aluminum in Maryville and Morristown, Tennessee, PhD thesis, University of Tennessee, Knoxville, Tenn, 1986
- 136. Martyn CN, Osmond C, Edwardson JA et al: Geographical relation between Alzheimer's disease and aluminium in drinking water. *Lancet* 1989; 1: 59-62
- 137. Michel P, Commenges D, Dartigues JF et al: Study of the relationship between Alzheimer's disease and aluminum in drinking water [abstr]. *Neurobiol Aging* 1990; 11: 264
- 138. Diagnostic and Statistical Manual of Mental Disorders, 3rd ed, rev, American Psychiatric Association, Washington, 1987
- 139. Frecker MF: Dementia in Newfoundland: Identification of a geographical isolate? J Epidemiol Community Health (in press)
- 140. McKhann G, Drachman D, Folstein M et al: Clinical diagnosis of Alzheimer's disease: report of the NINCDS-ADRDA Work Group under the auspices of Department of Health and Human Services Task Force on Alzheimer's Disease. Neurology 1984; 34: 939-944
- 141. Nania JM: Effect of minerals on dental caries. In Gould RF (ed): Dietary Chemicals Versus Dental Caries (Advances in Chemistry ser, 94), American Chemical Society, Washington, 1970: 123-160
- 142. Graves AB, White E, Koepsell TD et al: The association between aluminum-containing products and Alzheimer's disease. *J Clin Epidemiol* 1990; 43: 35-44
- 143. Dollinger HC, Zumkey H, Spieker C et al: Aluminum in antacids shown to accumulate in brain and bone tissue. Gastroenterol Obs 1986; 5: 478
- 144. Kobayashi K, Yumoto S, Nagai H et al: <sup>26</sup>Al tracer experiment by accelerator mass spectromety and its application to the studies for amyotrophic lateral sclerosis and Alzheimer's disease. *Proc Jpn Acad* 1990; 66B (10): 189-192
- 145. Driscoll CT, Letterman P: Chemistry and fate of Al(III) in treated drinking water. J Environ Eng 1988; 114: 21-37
- 146. Hunter D, Ross DS: Evidence for a phytotoxic hydroxyaluminum polymer in organic soil horizons. Science 1991; 251: 1056-1058
- 147. Rifat S, Eastwood MR, McLachlan DR et al: Evidence regarding the effect of prolonged aluminium exposure on cognitive function. *Lancet* 1990; 336: 1162-1165
- 148. Crapper McLachlan DR, Dalton AJ, Kruck TPA et al: Intramuscular desferrioxamine in patients with Alzheimer's disease. Lancet 1991; 337: 1034-1038
- 149. Mortimer JA: Genetic and environmental risk factors for Alzheimer's disease: key questions and new approaches. In Altman H (ed): Alzheimer's Disease and Dementia: Problems, Prospects and Perspectives, Plenum Pr, New York, 1989: 85-100

[This issue's Encore selection (starting on page 823) features a 1936 case report on Alzheimer's disease. — Ed.]

### Conferences continued from page 769

- Oct. 24-25, 1991: Gairdner Foundation Lectures University of Toronto
- Sally-Anne Hrica, Gairdner Foundation, 220-255 Yorkland Blvd., Willowdale, ON M2J 1S3; (416) 493-3101, fax (416) 493-8158
- Les 24 et 25 oct. 1991 : 9c Assemblée scientifique annuelle du Collège des médecins de famille du Canada (section Québec) Culture et santé : défis et perspectives Montréal
- Micheline Guilbault, secrétaire administrative, CP 146, Succ. Champlain, LaSalle, QC H8P 3J1; (514) 762-9889, fax (514) 762-9870
- Oct. 24-27, 1991: Canadian Association of Gerontology 20th Annual Scientific and Educational Meeting Regal Constellation Hotel, Toronto Canadian Association of Gerontology, 110-1565 Carling
- Oct. 25, 1991: Research Day in Family Medicine Radisson Hotel, London, Ont.

Ave., Ottawa, ON K1Z 3R1; (613) 728-9347

- Margot Meijer, Thames Valley Family Practice Research Unit, 1489 Richmond St., London, ON N6G 2M1; (519) 439-0121, fax (519) 439-0124
- Oct. 25-27, 1991: Canadian Medical Society on Alcohol and Other Drugs 3rd Annual Scientific Meeting
- Clarke Institute of Psychiatry and the Addiction Research Foundation, Toronto
- Canadian Medical Society on Alcohol and Other Drugs, 13-100 College St., Toronto, ON M5G 1L5; (416) 595-6000, ext. 7363, fax (416) 595-1214
- Oct. 30-Nov. 1, 1991: Catholic Health Association of Canada Administrators' Seminar Threats to the Mission of Catholic Health Care: a Strategic Response Radisson Hotel, Ottawa
- Freda Fraser, director of communications, Catholic Health Association of Canada, 1247 Kilborn Pl., Ottawa, ON K1H 6K9; (613) 731-7148
- Nov. 1-2, 1991: Pediatric AIDS Conference (sponsored by the Sunny Hill Hospital for Children, Vancouver, and the Division of Continuing Education in the Health Sciences, University of British Columbia)
- Coast Plaza Hotel at Stanley Park, Vancouver Pediatric AIDS Conference, Continuing Education in the Health Sciences, 105-2194 Health Sciences Mall, Vancouver, BC V6T 1Z3; (604) 822-2626, fax (604) 822-4835
- Nov. 1-2, 1991: Women, Food and Weight New Perspectives
- 519 Church Street Community Centre, Toronto National Eating Disorder Information Centre, 200 Elizabeth St., Ste. CW1-328, Toronto, ON M5G 2C4; (416) 340-4156

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