

Published in final edited form as:

J Am Chem Soc. 2005 August 3; 127(30): 10510–10511.

DNA Interstrand Cross-Link Formation Initiated by Reaction between Singlet Oxygen and a Modified Nucleotide

In Seok Hong and Marc M. Greenberg*

Department of Chemistry, Johns Hopkins University, 3400 North Charles Street, Baltimore, Maryland 21218

DNA is often the target of anti-cancer agents, which alkylate or oxidatively damage the biopolymer. Hydroxyl radical, metal–oxo complexes, such as that produced by bleomycin, and singlet oxygen are examples of agents that oxidatively damage DNA.^{1–4} Singlet oxygen, which selectively oxidizes deoxyguanosine, is an important reactive oxygen species in photodynamic therapy.^{3,5,6} The reactivity of a subset of DNA alkylating agents is distinguished from reactive oxygen species by their formation of interstrand cross-links (ISC's). For instance, interstrand cross-links are believed to be the source of the cytotoxicity of the anti-cancer agents, mitomycin C and chlorambucil.⁷ Herein, we describe a process involving a modified nucleotide (**2**) that potentiates the effects of singlet oxygen by reacting with this reagent to form ISC's.

We recently described a mechanism in which 5-(2'-deoxyuridinyl)methyl radical (**1**) forms an ISC with the opposing deoxyadenosine when it is photochemically generated from **2** in DNA (Scheme 1).^{8,9} In the course of investigating the mechanism for this process, we examined the effect of singlet oxygen on DNA containing **2**. Filtered ($\lambda \geq 400$ nm) aerobic photolysis (30 min) of 5'-³²P-**3** in the presence of 1–50 μ M of the singlet oxygen sensitizer, Rose Bengal, produced ISC's in as high as 48% yield (Figure 1B). Anoxic photolysis of a mixture of 5'-³²P-**3** and Rose Bengal (50 μ M) produces ~3% ISC.¹⁶ These observations suggest that direct photolysis of the phenyl selenide (**2**), which generates ISC's via **1** independent of O₂, and in lower yield, is not the source of cross-links under these conditions. Furthermore, the anoxic results suggest that ISC's do not result from a direct photoreaction between the sensitizer and DNA. Instead, the dependence of the rate of disappearance of monomeric **2** on D₂O content suggests that singlet oxygen, whose lifetime is enhanced 10-fold in the deuterated solvent, is responsible for phenyl selenide consumption (Figure 2).¹⁰

Photolysis of an otherwise identical duplex containing dT in place of **2** produces ~2% ISC, indicating that the phenyl selenide (**2**) plays an integral role in their formation in **3**.¹⁶ In contemplating a mechanism for this process, we recognized that phenyl selenides are oxidized to selenoxides by singlet oxygen in good yield, and allylic selenoxides undergo [2,3] sigmatropic rearrangements.^{11,12} Execution of these reactions in **2** would produce an electrophilic methide-type intermediate (**5**), akin to other molecules that alkylate DNA (Scheme 2).^{13,14} Evidence for this mechanism was gleaned from NMR analysis of the reaction of monomeric **2** with NaIO₄ (Figure 3) or its photosensitization by Rose Bengal in deuterated phosphate buffer.¹⁶ Periodate is shown because it rapidly and completely oxidizes **2**. The phenyl selenide (**2**) is completely consumed within 10 min and replaced by a diastereomeric

E-mail: mgreenberg@jhu.edu.

Supporting Information Available: Experimental procedures for carrying out the experiments on **2** and **3**. Autoradiograms showing ISC formation from **3** by NaIO₄ and hydroxyl radical footprinting of these ISC's, effect of glutathione on ISC formation, decomposition of singlet oxygen induced ISC's by piperidine treatment, and control experiments. ¹H NMR showing formation of **5** by Rose Bengal sensitized photolysis of **2**. This material is available free of charge via the Internet at <http://pubs.acs.org>.

mixture of the rearrangement product (**5**). Selenoxide **4** is not detected. Compound **5** reacts with the weak nucleophilic H₂O over the course of 24 h to produce 5-hydroxymethyl-2'-deoxyuridine (**6**), which is identical to independently prepared material.¹⁵ Furthermore, ISC's are efficiently formed with the opposing 2'-deoxyadenosine upon treatment of **3** with NaIO₄, indicating that the methide (**5**) produces cross-links.¹⁶

When **5** is produced in duplex DNA, rotation about the N-glycosidic bond into the *syn*-conformation positions the exocyclic methylene to react with N1 of the opposing deoxyadenosine, which is the same position that **1** is believed to cross-link with. However, the cross-link products formed in the presence of singlet oxygen migrate more slowly than those produced via **1** (Figure 1A). Exposure of 5'-³²P-**3** to singlet oxygen (Figure 1A, lane 2) previously cross-linked via formation of **1** (Figure 1A, lane 1) indicates that the cross-links produced under singlet oxygen conditions contain additional damage. Oxidized deoxyguanosines within the ISC products were deemed to be the most likely lesions formed in addition to cross-links. The damaged purines were revealed by treatment of the ISC's with piperidine and IrCl₆ (Figure 1A, lanes 4 and 5).^{17–20} Piperidine treatment following oxidation with IrCl₆ converts many of the ISC products into shorter, faster migrating fragments, indicating that not all of the cross-linked products also contain additional damaged nucleotides.¹⁶

The reaction of **2** with singlet oxygen is also distinguished from the radical pathway (**1**) by the effect of glutathione on ISC formation. ISC formation is unaffected by physiologically relevant glutathione concentrations (5 mM) when **3** is subjected to singlet oxygen.¹⁶ This observation reinforces those reported above, which suggest that the combination of the phenyl selenide derivative of thymidine (**2**) and singlet oxygen offers a novel and potentially important means for producing interstrand cross-links in DNA. The physiological importance of interstrand cross-links suggests that the incorporation of phenyl selenide **2** in DNA could be an effective adjuvant in photodynamic therapy.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgements

We are grateful for support from the National Institute of General Medical Sciences (GM-054996).

References

1. Chen J, Stubbe J. *Nat Rev Cancer* 2005;5:102–112. [PubMed: 15685195]
2. von Sonntag, C. *The Chemical Basis of Radiation Biology*; Taylor & Francis: London, 1987.
3. Lee PCC, Rodgers MAJ. *Photochem Photobiol* 1987;45:79–86. [PubMed: 3031708]
4. McCallum JEB, Kaniyoshi CY, Foote CS. *J Am Chem Soc* 2004;126:16777–16782. [PubMed: 15612716]
5. Dolmans DE, Fukumura D, Jain R. *Nat Rev Cancer* 2003;3:380–387. [PubMed: 12724736]
6. Prat F, Hou CC, Foote CS. *J Am Chem Soc* 1997;119:5051–5052.
7. Schärer OD. *ChemBioChem* 2005;6:27–32. [PubMed: 15637664]
8. Hong IS, Greenberg MM. *J Am Chem Soc* 2005;127:3692–3693. [PubMed: 15771492]
9. Monomeric compounds are referred to by the same number as when in **3**.
10. Turro, N. J. *Modern Molecular Photochemistry*; Benjamin Cummings: Menlo Park, CA, 1978.
11. Krief A, Lonz F. *Tetrahedron Lett* 2002;43:6255–6257.
12. Reich HJ, Yelm KE, Wollowitz S. *J Am Chem Soc* 1983;105:2503–2504.
13. Douki T, Vadesne-Bauer G, Cadet J. *J Org Chem* 2003;68:478–482. [PubMed: 12530874]

14. Veldhuyzen WF, Shallop AJ, Jones RA, Rokita SE. *J Am Chem Soc* 2001;123:11126–11132. [PubMed: 11697955]
15. Sowers LC, Beardsley GP. *J Org Chem* 1993;58:1664–1665.
16. See Supporting Information.
17. Muller JG, Duarte V, Hickerson RP, Burrows CJ. *Nucleic Acids Res* 1998;26:2247–2249. [PubMed: 9547288]
18. Ye Y, Muller JG, Luo W, Mayne CL, Shallop AJ, Jones RA, Burrows CJ. *J Am Chem Soc* 2003;125:13926–13927. [PubMed: 14611206]
19. Duarte VGD, Yamaguchi LF, Ravanat JL, Martinez GR, Medeiros MHG, DiMascio PD, Cadet J. *J Am Chem Soc* 2000;122:12622–12628.
20. Ravanat JL, Di Mascio P, Martinez GR, Medeiros MHG, Cadet J. *J Biol Chem* 2000;275:40601–40604. [PubMed: 11007783]

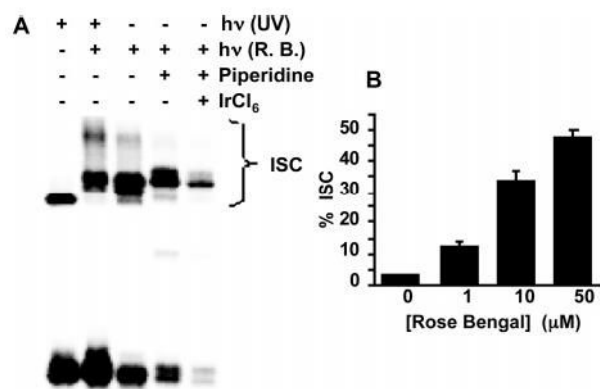


Figure 1. Formation of DNA interstrand cross-links (**3**, 10 nM) via UV or Rose Bengal sensitized aerobic photolysis. (A) Autoradiogram comparing ISC's produced upon UV or Rose Bengal (50 μM) sensitized photolysis. (B) Effect of Rose Bengal concentration on ISC formation (30 min photolysis).

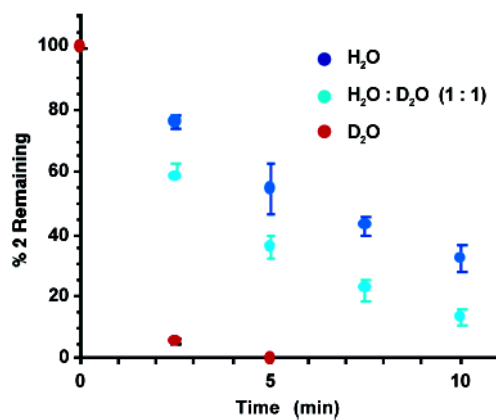


Figure 2. Effect of D₂O on the consumption of monomeric **2** (50 μM) upon irradiation of Rose Bengal (10 μM).

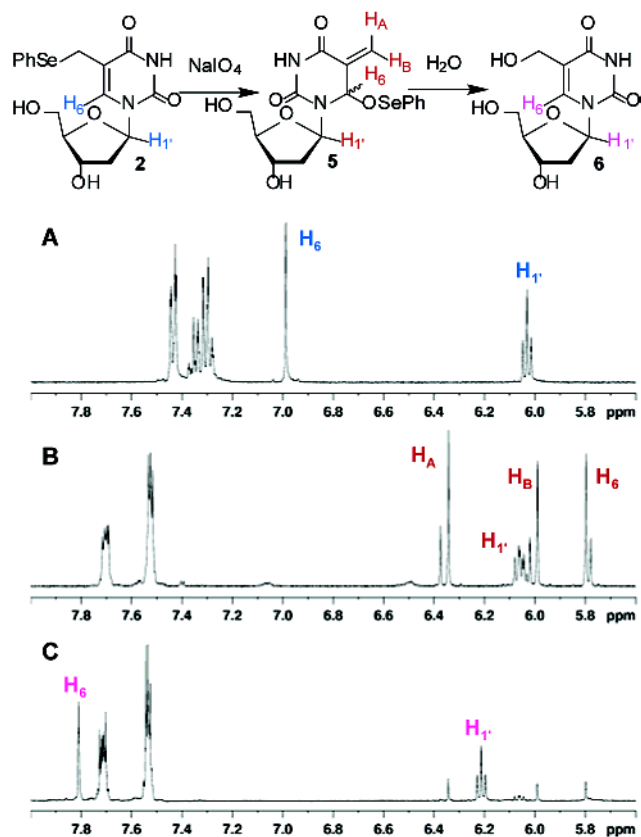
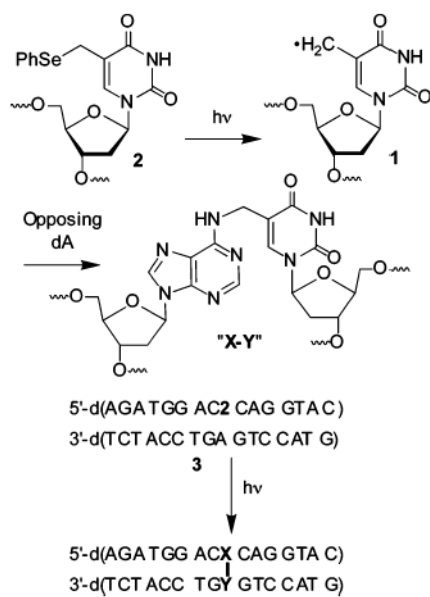
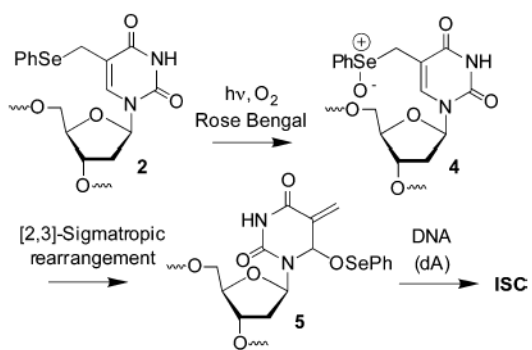


Figure 3. ¹H NMR analysis of the reaction of **2** (50 mM) with NaIO₄ (50 mM) in deuterated phosphate buffer (50 mM, pD 7.4). (A) Before NaIO₄ addition, (B) 10 min after NaIO₄ addition, and (C) 24 h after NaIO₄ addition.



Scheme 1.



Scheme 2.