

Clozapine disposition covaries with CYP1A2 activity determined by a caffeine test

LEIF BERTILSSON¹, JUAN ANTONIO CARRILLO², MARJA-LIISA DAHL¹, ADRIAN LLERENA^{1,2}, CHRISTINA ALM¹, ULF BONDESSON³, LEIF LINDSTRÖM³, INMACULADA RODRIGUEZ DE LA RUBIA², SARA RAMOS² & JULIO BENITEZ²

¹Department of Medical Laboratory Sciences and Technology, Division of Clinical Pharmacology at the Karolinska Institute, Huddinge Hospital, S-141 86 Huddinge, Sweden, ²Department of Pharmacology, Medical School, University of Extremadura, E-06071 Badajoz, Spain, and ³Psychiatric Research Center, University of Uppsala, S-750 17 Uppsala, Sweden

In a previous study we showed that the disposition of clozapine after a single oral dose is unrelated to either debrisoquine or S-mephenytoin hydroxylation polymorphism. The same 14 healthy subjects studied in that investigation were given 150 mg of caffeine. The reciprocal of plasma clozapine AUC (0,24), was correlated with an index of the *N*3-demethylation of caffeine ($r_s = 0.84$; $P = 0.0024$), used as a measure of cytochrome P4501A2 (CYP1A2) activity. *N*1- and *N*7-demethylation indices of caffeine also reflect CYP1A2 activity and were also correlated with clozapine clearance ($r_s = 0.89$ and 0.85 ; $P = 0.0013$ and 0.0023 ; respectively). No significant relationships with xanthine oxidase and *N*-acetyl transferase activity, also assessed by a caffeine test, were found. This study suggests that clozapine is metabolised by CYP1A2 to a major extent.

Keywords clozapine caffeine CYP1A2

Introduction

Clozapine is an atypical neuroleptic with potent anti-psychotic efficacy and few parkinsonian side-effects, but is associated with a high incidence of agranulocytosis [1]. Classical neuroleptics such as perphenazine [2], zuclopenthixol [3], thioridazine [4] and haloperidol [5] are metabolised by the polymorphic debrisoquine hydroxylase, CYP2D6 [6]. However, clozapine, which is also eliminated by oxidative metabolism, shows similar disposition in extensive (EM) and poor (PM) metabolisers of debrisoquine [7].

Recently, Jerling *et al.* [8] found that the plasma concentration of clozapine increased markedly in patients who also received fluvoxamine. A similar interaction between tricyclic antidepressants and fluvoxamine has been demonstrated [9, 10]. Spina *et al.* [11] showed that fluvoxamine markedly inhibited the *N*-demethylation of imipramine without affecting the CYP2D6 mediated hydroxylation of desipramine [12]. Brøsen *et al.* [13] have shown *in vitro* that fluvoxamine is a potent inhibitor of phenacetin *O*-deethylation, a reaction catalysed by CYP1A2.

The major metabolic pathway of caffeine is

N-3-demethylation, which is mainly mediated by CYP1A2 [14, 15]. The *N*-1 and *N*-7-demethylations are catalysed to a small extent by CYP2E1 in addition to CYP1A2 [14, 16]. The polymorphic *N*-acetyl transferase and xanthine oxidase are involved in the further metabolism of caffeine [14, 15]. Caffeine may thus be used as a probe drug for several drug metabolising enzymes. In the present study we performed a caffeine test in subjects previously investigated with respect to clozapine disposition [7].

Methods

In a previous study [7] a single oral 10 mg dose of clozapine was given to five PM of debrisoquine, five PM of S-mephenytoin and five individuals, who were EM of both probe drugs. The pharmacokinetic parameters were similar in the three groups. From that study we have taken reciprocal values of plasma clozapine AUC (0, 24) as an index of oral clearance.

Fourteen of the 15 healthy Swedish subjects studied [7] took part in the present study, which was conducted 1 year later. Only one subject was a smoker (10–20 cigarettes per day). All were drug free for at least 2 weeks before the study. Informed consent was obtained from all subjects and the study was approved by the Ethics Committee of the Huddinge Hospital.

The subjects abstained from methylxanthine containing food and beverages for 48 h before and during the study. Each subject emptied their bladder and collected a urine sample before administration of caffeine to detect compounds in the urine that might interfere with the analysis.

At 08.00 h 150 mg caffeine was taken in tablet form (Koffein, ACO) as a single oral dose with water. All urine voided during the next 24 h was collected in plastic vessels. The pH was adjusted to 3.5 by 5M hydrochloric acid and 10 ml aliquots were frozen at -20°C until analysis.

Concentrations of caffeine and metabolites in urine were measured by h.p.l.c. according to Carrillo & Benitez [15], and the analyst was blind to the clozapine data. The following molar ratios were calculated to indicate various enzyme activities [15]: Caffeine N1-demethylation: $7\text{X} + 37\text{U} + 37\text{X}/137\text{X}$; Caffeine N3-demethylation: $\text{AAMU} + 1\text{U} + 1\text{X} + 17\text{U} + 17\text{X}/137\text{X}$; Caffeine N7-demethylation: $3\text{X} + 13\text{U} + 13\text{X}/137\text{X}$; Xanthine oxidase: $1\text{U}/1\text{X} + 1\text{U}$; N-acetylation: $\text{AAMU}/1\text{X}$.

AAMU: 5-acetylamino-6-amino-3-methyluracil; 7X: 7-methylxanthine; 1U: 1-methyl uric acid; 3X: 3-methylxanthine; 37U: 3,7-dimethyl uric acid; 1X: 1-methylxanthine; 13U: 1,3-dimethyl uric acid; 37X: 3,7-dimethylxanthine (theobromine); 17U: 1,7-dimethyl uric acid; 17X: 1,7-dimethylxanthine (paraxanthine); 13X: 1,3-dimethylxanthine (theophylline); 137X: 1,3,7-trimethylxanthine (caffeine).

Results

All subjects tolerated the 48 h abstinence from caffeine containing food and beverage, although some subjects had headache. The reciprocal of plasma AUC (0,24) of clozapine [7] correlated with measures of caffeine N1-, 3- and 7-demethylations ($r_s = 0.89, 0.84$ and 0.85 ; all $P < 0.01$) (Table 1). The relationship with N3-demethylation is shown in Figure 1. The three N-demethylations were closely

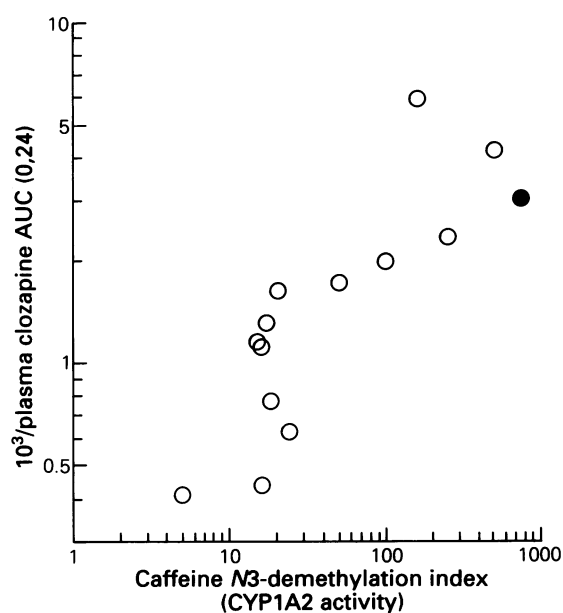


Figure 1 Relationship between reciprocal plasma clozapine AUC (0,24) and a caffeine N3-demethylation index of CYP 1A2 activity ($r_s = 0.84$; $n = 14$; $P = 0.0024$). Data for the only smoker are indicated as a filled circle.

intercorrelated ($r_s = 0.88$ – 0.92) (Table 1). There was no correlation between clozapine AUC (0,24) and caffeine indices of xanthine oxidase ($r_s = -0.32$) or N-acetyl transferase ($r_s = -0.33$) activity.

The only smoker participating in this study had the highest N3-demethylation index and the third highest reciprocal clozapine AUC (0,24) value.

Discussion

The pharmacokinetic interaction observed in patients between fluvoxamine and clozapine [8] suggests that clozapine might be metabolised by CYP1A2, since fluvoxamine was shown to be a potent inhibitor of this isoform of cytochrome P450 *in vitro* [13]. In the present study about 70% of the variance in the oral clearance of clozapine was accounted for by an index of caffeine N3-demethylation, used as an *in vivo* marker of CYP1A2 activity. This suggests that clozapine is to a major extent metabolized by CYP1A2, and is consistent with the knowledge that both clozapine disposition [17] and CYP1A2 activity [14, 15] are inducible by smoking. The only smoker

Table 1 Spearman rank correlation coefficients for relationships between the reciprocal of plasma clozapine AUC (0,24) and various enzyme activities measured by a caffeine test

	N1	N3	N7	XO	NAT
Reciprocal clozapine AUC (0,24)	0.89**	0.84**	0.85**	-0.32	-0.33
Caffeine N1-demethylation (N1)		0.88**	0.92***	-0.08	-0.43
Caffeine N3-demethylation (N3)			0.88**	-0.02	-0.31
Caffeine N7-demethylation (N7)				-0.28	-0.36
Xanthine oxidase (XO)					0.05

** $P < 0.01$; *** $P < 0.001$; NAT = N-acetyl transferase.

in this study had the highest index of CYP1A2 activity and a high oral clearance of clozapine. Jerling *et al.* [8] showed that carbamazepine decreases plasma concentrations of clozapine in patients given both drugs. Carbamazepine is an inducer of CYP3A4 and this enzyme may metabolise clozapine in addition to CYP1A2. Carbamazepine is not an inducer of caffeine metabolism [18] and, therefore, not an inducer of CYP1A2.

Several neuroleptics are metabolised by CYP2D6 and pharmacokinetic interactions have been reported with drugs metabolised by this enzyme including neuroleptics and tricyclic antidepressants [6]. However, measurement of plasma concentrations of cloza-

pine in psychiatric patients indicated no interaction between clozapine and other neuroleptics or tricyclic antidepressants metabolised by CYP2D6 [8]. In theory, the metabolism of clozapine may be inhibited by several drugs metabolised by CYP1A2, such as imipramine, phenacetin, theophylline and possibly caffeine.

The study was supported by the Swedish Medical Research Council (3902), the Bank of Sweden Tercentenary Foundation, the Karolinska Institute, the Spanish Comisión Interministerial de Ciencia y Tecnología (SAF 92-0333) and Fondo de Investigaciones Sanitarias (93/0632), and was carried out in coordination with COST B1.

References

- Baldessarini RJ, Frankenburg FR. Clozapine. A novel antipsychotic agent. *New Engl J Med.* 1991; **324**: 746–754.
- Dahl-Puustinen M-L, Lidén A, Alm C, Nordin C, Bertilsson L. Disposition of perphenazine is related to polymorphic debrisoquine hydroxylation in human beings. *Clin Pharmac Ther* 1989; **46**: 78–81.
- Dahl M-L, Ekqvist B, Widén J, Bertilsson L. Disposition of the neuroleptic zuclopenthixol cosegregates with the polymorphic hydroxylation of debrisoquine in humans. *Acta Psychiat Scand* 1991; **84**: 99–102.
- von Bahr C, Movin G, Nordin C, Lidén A, Hammarlund-Udenaes M, Hedberg A, Ring H, Sjöqvist F. Plasma levels of thioridazine and metabolites are influenced by the debrisoquine hydroxylation phenotype. *Clin Pharmac Ther* 1991; **49**: 234–240.
- Llerena A, Alm C, Dahl M-L, Ekqvist B, Bertilsson L. Haloperidol disposition is dependent on debrisoquine hydroxylation phenotype. *Ther Drug Monit* 1992; **14**: 92–97.
- Dahl M-L, Bertilsson L. Genetically variable metabolism of antidepressants and neuroleptic drugs in man. *Pharmacogenetics* 1993; **3**: 61–70.
- Dahl M-L, Llerena A, Bondesson U, Lindström L, Bertilsson L. Disposition of clozapine in man: Lack of association with debrisoquine and S-mephenytoin hydroxylation polymorphisms. *Br J clin Pharmac* 1994; **37**: 71–74.
- Jerling M, Lindström L, Bondesson U, Bertilsson L. Fluvoxamine inhibition and carbamazepine induction of the metabolism of clozapine: evidence from a therapeutic drug monitoring service. *Ther Drug Monit* 1994; **16**: 368–374.
- Bertschy G, Vandel S, Vandel R, Allers G, Volmat R. Fluvoxaminetricyclic antidepressant interaction: an accidental finding. *Eur J clin Pharmac* 1991; **40**: 119–120.
- Spina E, Campo GM, Avenoso A, Pollicino MA, Caputi AP. Interaction between fluvoxamine and imipramine/desipramine in four patients. *Ther Drug Monit* 1992; **14**: 194–196.
- Spina E, Pollicino AM, Avenoso A, Campo GM, Perucca E, Caputi AP. Effect of fluvoxamine on the pharmacokinetics of imipramine and desipramine in healthy subjects. *Ther Drug Monit* 1993; **15**: 243–246.
- Bertilsson L, Åberg-Wistedt A. The debrisoquine hydroxylation test predicts steady-state plasma levels of desipramine. *Br J clin Pharmac* 1983; **15**: 388–390.
- Brøsen K, Skjelbo E, Rasmussen BB, Poulsen HE, Loft S. Fluvoxamine is a potent inhibitor of cytochrome P4501A2. *Biochem Pharmac* 1993; **45**: 1211–1214.
- Kalow W, Tang B-K. The use of caffeine for enzyme assays: A critical appraisal. *Clin Pharmac Ther* 1993; **53**: 503–514.
- Carrillo JA, Benitez J. Caffeine metabolism in a healthy Spanish population: *N*-acetylator phenotype and oxidation pathways. *Clin Pharmac Ther* 1994; **55**: 293–304.
- Gu L, Gonzalez FJ, Kalow W, Tang BK. Biotransformation of caffeine, paraxanthine, theobromine and theophylline by cDNA-expressed human CYP1A2 and CYP2E1. *Pharmacogenetics* 1992; **2**: 73–77.
- Haring C, Meise U, Humpel C, Saria A, Fleischhacker WW, Hinterhuber H. Dose-related plasma levels of clozapine: influence of smoking behaviour, sex and age. *Psychopharmacology* 1989; **99**: S38–S40.
- Wietholz H, Zysset Th, Kreiten K, Kohl D, Büchsel R, Matern S. Effect of phenytoin, carbamazepine, and valproic acid on caffeine metabolism. *Eur J clin Pharmac* 1989; **36**: 401–406.

(Received 24 February 1994,
accepted 12 July 1994)