

The Fe-CO Bond Energy in Myoglobin: A QM/MM Study of the Effect of Tertiary Structure

Nikki Strickland, Adrian J. Mulholland, and Jeremy N. Harvey

Centre for Computational Chemistry and School of Chemistry, University of Bristol, Bristol, BS8 1TS, United Kingdom

ABSTRACT The Fe-CO bond dissociation energy (BDE) in myoglobin (Mb) has been calculated with B3LYP quantum mechanics/molecular mechanics methods for 22 different Mb conformations, generated from molecular dynamics simulations. Our average BDE of 8.1 kcal/mol agrees well with experiment and shows that Mb weakens the Fe-CO bond by 5.8 kcal/mol; the calculations provide detailed atomistic insight into the origin of this effect. BDEs for Mb conformations with the R carbonmonoxy tertiary structure are on average 2.6 kcal/mol larger than those with the T deoxy tertiary structure, suggesting two functionally distinct allosteric states. This allostery is partly explained by the reduction in distal cavity steric crowding as Mb moves from its T to R tertiary structure.

Received for publication 18 November 2005 and in final form 13 December 2005.

Address reprint requests and inquiries to Jeremy N. Harvey, Tel.: 44-0-117-954 6991; Fax: 44-0-117-925-1295; E-mail: jeremy.harvey@bristol.ac.uk.

The relationship between protein structure and function is central to biochemistry and can be usefully probed with model proteins such as myoglobin (Mb). Despite intensive experimental study, it is still unclear exactly how Mb's tertiary structure affects the geminate recombination of carbon monoxide (1,2). We are interested in examining this problem at an atomistic level by using a quantum mechanical (QM) description of the heme group with a simpler molecular mechanics (MM) treatment of the remaining Mb (QM/MM method) (2). This allows us to treat the area of interest with accurate density functional theory while including effects from the rest of the protein. As a first step we report here our results concerning the thermochemistry of CO bonding in Mb, which highlight the presence of two functionally distinct allosteric states, corresponding to the tertiary structures of unligated Mb (deoxyMb, T) and CO-ligated Mb (MbCO, R). Such allosteric states are usually associated with multimeric proteins such as hemoglobin (Hb), where they allow cooperative binding of O₂. Monomeric Mb is often considered to be nonallosteric but this view is changing (3) based partly on how lactate reduces its O₂ affinity (4), differences in MbO₂ and deoxyMb O₂ equilibration curves (5), and Mb's control of pseudoenzymatic reactions (6).

Allosteric behavior in Mb is possible because ligand binding is accompanied by structural changes as Mb relaxes between ligated (R) and unligated (T) tertiary structures. This change in structure for Mb is similar, but not identical to that observed in the α - and β -subunits of Hb (7). As CO dissociates from the heme iron, the iron atom moves ~ 0.3 Å out of the porphyrin plane and the heme distorts from planarity. Concerted motion of the rest of Mb, notably the E and F α -helices (8,9), completes the transition between the T and R states. The impact of this relaxation on ligand binding is not yet fully understood.

We have explored the effect of Mb's tertiary structure on CO geminate reactivity by using QM/MM methods to calculate Fe-CO binding enthalpies. This has allowed us to probe Mb's allostery and to test our QM/MM methodology by comparing our bond energies to experiment (10). Calculating accurate metalloprotein bond energies is challenging, especially when structural relaxation accompanies ligand binding. Properties and energies of proteins vary slightly as they fluctuate between multiple conformational substates (11), within each tertiary state. We allowed for such variations and relaxation by calculating bond energies separately for Mb R and T states as averages of 11 values for different substates generated from molecular dynamics (MD) trajectory snapshots and minimized crystal structures. Previous QM/MM optimized geometries show that the Fe-CO bond is distorted $\leq 9^\circ$ by Mb's tertiary structure and slightly stabilized by a very weak hydrogen bond between the distal histidine (His-64) and oxygen of bound CO (12,13). The energetic impact of structural relaxation upon binding was not addressed in these studies (12,13). Crystal studies reveal that relaxation from the T state to the R state enables Val-68 to move from a position where it blocks CO binding to a sterically more favorable orientation (8). Such steric crowding in the distal cavity of T state Mb is verified here and helps explain Mb's allostery.

We started with 1.15-Å resolution x-ray structures of deoxyMb and MbCO (Protein Data Bank entries 1A6N and 1A6G (14)), then removed waters of crystallization and sulfate ions. The CHARMM all-atom force field (15) was modified by additional heme group parameters and used with CHARMM v27b2 (16) to prepare both proteins and carry out

MD simulations. Hydrogen atoms were added according to standard pK_A values for all amino acids except histidines. His-64 was chosen as the N_ϵ -protonated tautomer, in agreement with converging opinion (12,13), and the protonation states of other histidines were assigned according to their hydrogen bonding environment. Each protein was solvated with a 9-Å layer of preequilibrated TIP3 water to create two systems of $\sim 10,000$ atoms. Relaxation, heating to 300 K over 30 ps and equilibration until stable (~ 300 ps) prepared both systems for a 100-ps production run. No constraints or restraints were applied throughout. MM minimization of each solvated crystal structure and of 10 snapshots from each MD production run, generated 11 deoxyMb and 11 MbCO structures for QM/MM BDE calculations.

The Fe-CO BDE is the energy difference between MbCO and the sum of gas phase CO and deoxyMb, if both Mbs have a similar conformation. Convergence to the same local substate for MbCO and deoxyMb is essential to obtain meaningful values, and was achieved by creating deoxyMb and MbCO “pairs” then carrying out a series of iterations, as described by Wirstam et al. (17) for O_2 binding to hemethyrin. Eleven T-state “pairs” were created by addition of CO to deoxyMbs and 11 R-state “pairs” were made by removal of bound CO from MbCO. The QM/MM calculations used the Jaguar QM program (18) and Tinker MM code (19), coupled by our QM/MM routines (2). The QM region, consisting of the heme porphine, the His-93 imidazole side chain and CO, were treated with the hybrid B3LYP functional and a flexible basis set. All other atoms made up the MM region, which was treated with the CHARMM all-atom force field (15). All results include a correction for zero-point energy (ZPE) calculated for a gas-phase model. Full computational details are in the Supplementary Material.

The overall BDE of Fe-CO Mb (shown as $\Delta E_{\text{overall}}$ in the thermodynamic cycle of Fig. 1) can be written in two different ways as a sum of two terms: $\Delta E_{\text{overall}} = \Delta E_1 + \Delta E_2$ or $\Delta E_3 + \Delta E_4$. One contribution is the average QM/MM bond energy for the R and T states of Mb (ΔE_1 and ΔE_3 , respectively). The other contribution is the relaxation energy for interconversion of R and T states in deoxyMb or MbCO (ΔE_2 or ΔE_4 , respectively). These cannot be readily calculated using our method. It is, however, apparent that ΔG_2 is negative, and ΔG_4

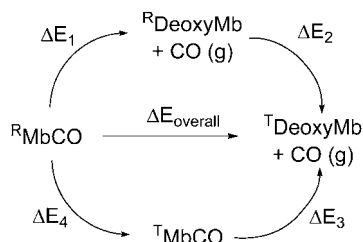


FIGURE 1 BDE thermodynamic cycle. “R”/“T” denote tertiary state and “MbCO”/“deoxyMb” indicates whether CO is bound or not to Mb.

is positive, since MbCO and deoxyMb are known to adopt R and T states, respectively. Assuming that entropic effects are small for the free energies ΔG_2 and ΔG_4 , we can therefore write that $\Delta E_1 > \Delta E_{\text{overall}} > \Delta E_3$. As a first approximation, we can take $\Delta E_{\text{overall}}$ as the average of ΔE_1 and ΔE_3 ; the difference between these values provides an estimate of the effect of the tertiary structure on the bond energy.

The 11 BDEs for R state Mb range between 8.1 and 10.8 kcal/mol (see Table S1 in Supplementary Material), with an average $\Delta E_1 = 9.4$ kcal/mol. The average for the T state is $\Delta E_3 = 6.9$ (range 3.0–9.0 kcal/mol). This yields an estimated BDE $\Delta E_{\text{overall}}$ of 8.1 kcal/mol, in good agreement with the experimental value of 10.8 kcal/mol (10). The BDE for the QM region alone is 13.9 kcal/mol (1, including ZPE correction), so Mb weakens the Fe-CO bond by 5.8 kcal/mol. This weakening is greater for Mb in the T state, ΔE_1 is 2.6 kcal/mol larger than ΔE_3 , showing that the two tertiary states are functionally different.

Structural analysis (Table S2 in Supplementary Material) reveals more steric hindrance in the distal cavity in the T state than in the R state. Distances between the CO oxygen and the hydrogen atoms Phe-43 $C_\gamma H$, His-64 $N_\epsilon H$, and Val-68 $C_\gamma H$ increase on average by 0.48, 0.53, and 0.05 Å during relaxation from the T to R. This does not, however, lead to significant changes in the tilt or bend of the Fe-CO bond (the Fe-C-O angle is $\sim 175^\circ$ in both states), in agreement with the known rigidity of this moiety (13). Increased amplitude of the motion of the iron into the heme plane upon CO binding lowers the BDE; this average movement is 0.33 Å for the T state and 0.25 Å for the R state. The polarizing effect of the protein environment on the QM region leads to an increase of 0.4 kcal/mol on average of the Fe-CO BDE (Table S3 in Supplementary Material). These electrostatic effects weakly destabilize the bond in the R state and weakly stabilize it in the T state, but the predominant contribution to the difference in bond energies between the two conformers is steric.

SUPPLEMENTARY MATERIAL

An online supplement to this article can be found by visiting BJ Online at <http://www.biophysj.org>.

ACKNOWLEDGMENTS

The authors are grateful for funding provided by the Engineering and Physical Sciences Research Council. A.J.M. also thanks the Biotechnology and Biological Sciences Research Council and the IBM High Performance Computing Life Sciences Outreach Program.

REFERENCES and FOOTNOTES

- Harvey, J. N. 2000. DFT computation of the intrinsic barrier to CO geminate recombination with heme compounds. *J. Am. Chem. Soc.* 122:12401–12402.

2. Harvey, J. N. 2004. Spin-forbidden CO ligand recombination in myoglobin. *Faraday Discuss.* 127:165–177.
3. Frauenfelder, H., B. H. McMahon, R. H. Austin, K. Chu, and J. T. Groves. 2001. The role of structure, energy landscape, dynamics, and allostery in the enzymatic function of myoglobin. *Proc. Natl. Acad. Sci. USA.* 98:2370–2374.
4. Giardina, B., P. Ascenzi, M. E. Clementi, G. DeSanctis, M. Rizzi, and M. Coletta. 1996. Functional modulation by lactate of myoglobin: a monomeric allosteric hemoprotein. *J. Biol. Chem.* 271:16999–17001.
5. Shibayama, N., and S. Saigo. 2003. Oxygen equilibrium properties of myoglobin locked in the liganded and unliganded conformations. *J. Am. Chem. Soc.* 125:3780–3783.
6. Fogel, U., M. W. Merx, A. Godecke, U. K. M. Decking, and J. Schrader. 2001. Myoglobin: a scavenger of bioactive NO. *Proc. Natl. Acad. Sci. USA.* 98:735–740.
7. Samuni, U., D. Dantsker, I. Khan, A. J. Friedman, E. Peterson, and J. M. Friedman. 2002. Spectroscopically and kinetically distinct conformational populations of sol-gel-encapsulated carbonmonoxy myoglobin: a comparison with hemoglobin. *J. Biol. Chem.* 277:25783–25790.
8. Kachalova, G. S., A. N. Popov, and H. D. Bartunik. 1999. A steric mechanism for inhibition of CO binding to heme proteins. *Science.* 284:473–476.
9. Srajer, V., Z. Ren, T. Y. Teng, M. Schmidt, T. Ursby, D. Bourgeois, C. Pradervand, W. Schildkamp, M. Wulff, and K. Moffat. 2001. Protein conformational relaxation and ligand migration in myoglobin: a nanosecond to millisecond molecular movie from time-resolved Laue X-ray diffraction. *Biochemistry.* 40:13802–13815.
10. Projahn, H. D., and R. Van Eldik. 1991. Volume profile analysis of the formation and dissociation of carboxymyoglobin: comparison with the corresponding oxymyoglobin system. *Inorg. Chem.* 30:3288–3293.
11. Frauenfelder, H., S. G. Sligar, and P. G. Wolynes. 1991. The energy landscapes and motions of proteins. *Science.* 254:1598–1603.
12. Sigfridsson, E., and U. Ryde. 2002. Theoretical study of the discrimination between O₂ and CO by myoglobin. *J. Inorg. Biochem.* 91:101–115.
13. Rovira, C., B. Schulze, M. Eichinger, J. D. Evanseck, and M. Parrinello. 2001. Influence of the heme pocket conformation on the structure and vibrations of the Fe-CO bond in myoglobin: a QM/MM density functional study. *Biophys. J.* 81:435–445.
14. Vojtechovsky, J., K. Chu, J. Berendzen, R. M. Sweet, and I. Schlichting. 1999. Crystal structures of myoglobin-ligand complexes at near-atomic resolution. *Biophys. J.* 77:2153–2174.
15. MacKerell, A. D., D. Bashford, M. Bellott, R. L. Dunbrack, J. D. Evanseck, M. J. Field, S. Fischer, J. Gao, H. Guo, S. Ha, D. Joseph-McCarty, L. Kuchnir, et al. 1998. All-atom empirical potential for molecular modeling and dynamics studies of proteins. *J. Phys. Chem. B.* 102:3586–3616.
16. Brooks, B. R., R. E. Bruccoleri, B. D. Olafson, D. J. States, S. Swaminathan, and M. Karplus. 1983. Charmm: a program for macromolecular energy, minimization, and dynamics calculations. *J. Comput. Chem.* 4:187–217.
17. Wirstam, M., S. J. Lippard, and R. A. Friesner. 2003. Reversible dioxygen binding to hemerythrin. *J. Am. Chem. Soc.* 125:3980–3987.
18. Schrodinger Inc. 1996–2001. Jaguar. Portland, OR.
19. Ponder, J. W. 2003. Tinker: Software Tools for Molecular Design. St. Louis, MO.